



# Phytochemistry, Allelopathy And Anticancer Potentiality of *Melaleuca alternifolia* (Maiden and Betcher) Cheel and *Psidium guajava* L. (Myrtaceae)

Salama Mohamed El-Darier<sup>2</sup>, Mona Elsayed Mabrouk<sup>1</sup>, Nourhan Abas Zabady<sup>1</sup> and Kholod Ali Khattab<sup>1\*</sup>

<sup>1</sup>Botany Department, Faculty of Science, Damanhour University, Damanhour, Egypt

<sup>2</sup>Botany and Microbiology Department, Faculty of Science, Alexandria University, Alexandria, Egypt

**Citation:** El-Darier, S.M; Mabrouk, M.E; Zabady, N.A; Khattab, K.A (2025). Phytochemistry, Allelopathy And Anticancer Potentiality of *Melaleuca alternifolia* (Maiden and Betcher) Cheel and *Psidium guajava* L. (Myrtaceae). *Journal of Environmental Studies*, Vol. 38(1): 10-23.

## Article Information

Received 15 Decem. 2024,

Revised 20 Feb. 2025,

Accepted 23 Feb. 2025,

Published online. 1 June 2025

**Abstract:** *Melaleuca alternifolia* leaves ethanolic extract exert highest detection (++) of tannins, steroids, flavonoids and alkaloids. *Psidium guajava* ethanolic extract achieved strong detection (+++) of phenolics and flavonoids, while aqueous extract revealed high detection (++) of flavonoids. In petri dish experiment *P. guajava* leaves aqueous extract exert great inhibition effect on growth parameters of *Rumex dentatus*, *Solanum lycopersicum* (geaara 023) and *Solanum lycopersicum* (tomato extracted seed) under (control, 0.5%, 1%, 1.5%, 2%, 2.5%, 3%, 3.5%, 4%, 4.5% and 5%) concentration levels compared to *M. alternifolia* leaves aqueous extract. Pot experiment prepared to determine vegetative seedling storage protein profiling that showed a great decrease in genome template stability GTS (%) of *R. dentatus* under the effect of *P. guajava* leaves aqueous extract, however GTS (%) of *S. lycopersicum* (geaara 023) exert a great increase compared to *S. lycopersicum* (extracted seed) as crops under the effect of *M. alternifolia* leaves aqueous extract. Energy dispersive x-ray spectroscopy of *M. alternifolia* and *P. guajava* revealed that *M. alternifolia* leaves has high concentration levels of Cl, Ca, Al and Cu elements, whereas *P. guajava* leaves exert high concentration levels of Si, S, Fe and K elements. *M. alternifolia* ethanolic extract has a significant effect on lung (A549) and breast carcinoma (MCF7) cell lines with IC<sub>50</sub> of about 31 and 98.9 µg/ml respectively. On the other hand, *P. guajava* ethanolic extract showed a significant effect on lung (A549) and prostate carcinoma (PC3) cell lines of about IC<sub>50</sub> of 40 and 50.5 µg/ml respectively.

**Keywords:** Cell line, Carcinoma, Phytotherapy, Electrophoresis, Allelochemicals.

## Introduction

Myrtaceae family one of the most ecologically significant group of angiosperms that includes trees and shrubs this family comprises approximately of 140 genera and between 3800 and 5650 species valued for their edible fruits and use in traditional medicine (Mitra et al., 2012; Saber et al., 2023). *Melaleuca alternifolia* (tea tree) known for its medicinal values in treating wounds, fungal infections, sore throats and skin ailments due to their high levels of terpene

hydrocarbon contents (Shah et al., 2019; Kairey et al., 2023). *P. guajava* (guava) valued for its nutritional and medicinal benefits, is used to manage stomach aches, diabetes, and diarrhea due to high levels of saponins, quercetin, flavonoids, terpenes, and tannins (Kumar et al., 2021; Liu et al., 2024).

Allelopathy has inhibitory and stimulatory effects in all plant processes such as seed germination, growth parameters and weed management by releasing some phytochemicals (Rice, 1984; Bachheti et al., 2020).

\* Corresponding author E-mail: [kholodkhat26@yahoo.com](mailto:kholodkhat26@yahoo.com)

Allelopathy is also considered one of the indirect factors of regular cropping difficulties in the agriculture sector, so recently agricultural production management plans and ecological restoration depending on applications of allelopathy (Cheng & Cheng, 2015). *M. alternifolia* aqueous extracts have shown inhibitory effects on root growth in *Brachiaria brizantha* (Queiroz *et al.*, 2017) and have enhanced stress tolerance in crops (Yasin *et al.*, 2021). Likewise, *P. guajava* has demonstrated allelopathic effects, such as weed control (Kapoor *et al.*, 2019; Mabele & Ndong, 2019).

Cancer, defined as a complex disease arising from accumulated genetic mutations (Kumar *et al.*, 2021). *M. alternifolia* shows anticancer activity against prostate and breast cancer cell lines (Clark *et al.*, 2021). *P. guajava* extracts possess antiprostata cancer properties and contain pigments with antioxidant functions that may aid in cancer prevention during DNA repair, gene regulation and apoptosis (Chen *et al.*, 2010; Nalkiran & Nalkiran, 2024).

The present study is an attempt to maximize the ecological and medicinal benefits such as allelopathic interactions, antibacterial and anti-cancer potentiality of *Melaleuca alternifolia* and *Psidium guajava* with a view to assessing their contribution to human livelihood.

## Materials and Methods

### Collection of *Melaleuca alternifolia* and *Psidium guajava* Leaves

Mature green leaves of both *M. alternifolia* and *P. guajava* were collected from El- Beheira Governorate, Egypt during Summer season. *M. alternifolia* collected from Karam nursery (Damanhour City) at 31°2'47" N and 30°28'14" E, while *P. guajava* were collected from Mahalla Nasr Village (Shubrakhit Center) at 31°01'39" N and 30°42'46" E.

### Preparation of Aqueous and Ethanolic Extracts of *M. alternifolia* and *P. guajava* Leaves

Collected *M. alternifolia* and *P. guajava* mature green leaves were washed with tap water, then with distilled water for further cleaning and dried in an electric oven at 45° C. The dried leaves were ground to a fine powder. 0.5, 1, 1.5, 2, 2.5, 3, 3.5, 4, 4.5 and 5 g were transferred to labeled bottles, and then 100 mL of distilled water were added to each bottle. The mixture was shaken then the bottles were left for 48 hours at refrigerator and then filtered through very fine mesh and pressed carefully for full extraction to get extracts of 0.5%, 1%, 1.5%, 2%, 2.5%, 3%, 3.5%, 4%, 4.5% and 5%, the control (C) was (distilled water) (El-

Rokiek *et al.*, 2024), while ethanolic extract was prepared according to Sridhar *et al.* (2016).

### Germination Bioassay

Petri-dish experiment was applied to investigate the potential allelopathic effects of *P. guajava* and *M. alternifolia* on germination percentage (GP), radicle (RL) and plumule (PL) lengths of *R. dentatus*, *S. lycopersicum* (geaara 023) and *S. lycopersicum* (tomato extracted seed) as recipient species in pure culture. To accomplish this experiment 10 seeds of each recipient species were arranged in 9-cm diameter petri-dishes separately on disc of whatman No.1 filter paper under normal laboratory conditions. 10 ml of *P. guajava* and *M. alternifolia* leaves aqueous extract at Control, 0.5%, 1.0%, 1.5%, 2.0%, 2.5%, 3.0%, 3.5%, 4.0%, 4.5% and 5.0% were added daily to three replicates for thirteen days. The experiment was performed under normal laboratory conditions (20±2° C temperature, 75±2% relative humidity, and 14/10 h/dark photoperiod).

### Pot Experiment

Pot growth experiment was performed to test the allelopathic effect of 0.5%, 1.5% and 3.5% of *P. guajava* and *M. alternifolia* leaves aqueous extract on three replicates of *R. dentatus*, *S. lycopersicum* (geaara 023) and *S. lycopersicum* (tomato extracted seed) for three weeks with sandy clay soil (500 g in each pot) on seedling protein electrophoresis as molecular marker. The experiment was performed under normal laboratory conditions (20±2° C temperature, 75±2% relative humidity, and 14/10 h/dark photoperiod).

### Seedling Protein Electrophoresis

For assessing the allelopathic effect of *P. guajava* and *M. alternifolia* leaves aqueous extract at (0.5, 1.5 and 3.5) % concentration levels on protein content of *R. dentatus*, *S. lycopersicum* (geaara 023) and *S. lycopersicum* (tomato extracted seed) seedling compared to the effect of control (C) (distilled water), sodium dodecyl sulphate polyacrylamide gel electrophoresis (SDS-PAGE) was carried out using discontinuous buffer system according to Laemmli (1970).

### Phytochemical Screening

Phytochemical screening of *P. guajava* and *M. alternifolia* leaves was analyzed in order to estimate the presence of carbohydrates, phenolic compounds, saponins, protein and amino acid (Rajendrabhai, 2017), vitamin C (Kumar *et al.*, 2011), tannins (Ramya *et al.*, 2019), alkaloids (Khalifa *et al.*, 2017), flavonoids (Baoduy *et al.*, 2015), phlobatannins and steroids (Ejikeme *et al.*, 2014). Total phenols, proteins and amino acids contents were determined by high

performance liquid chromatographic (HPLC). The quantity content of fatty acids and carbohydrates in the plant samples were performed by gas chromatography-mass spectrometry (GC-MS) using Agilent 6890N with mass detector 5973 inert and Agilent Technologies 6890 gas chromatograph (USA) with mass spectrometry detector 5973 and capillary column, respectively.

### Estimation Of Anti-Proliferative Activity

Three replicates of plant ethanolic extraction for cell line test (In Vitro) were determined by weighting 200 g of powdered sample of both *P. guajava* and *M. alternifolia* leaves were macerated in one liter of 99.6% denatured ethanol (1 Liter) for 24 hour at room temperature. The total volumes of solution were filtered then dried using rotary evaporator (buchiotavapor R114, Switzerland) under reduced pressure (Sergazy *et al.*, 2021).

### Measurement Of Potential Cytotoxicity By Sulforhodamine B (SRB) Assay

Three replicates of *P. guajava* and *M. alternifolia* leaves ethanolic extracts were tested using the method of Skehan *et al.* (1990) at the National Cancer Institute, Cairo, Egypt.

### Five Human Cancer Cell Lines Were Used In The Current Study

HCT (Colon carcinoma), A549 (Lung carcinoma), Hep-G2 (Liver hepatocellular carcinoma), MCF7 (Breast carcinoma) and PC3 (Prostate carcinoma). Surviving fractions of cells throughout drug exposure was characterized graphically by IC<sub>50</sub> values (drug concentration that yields 50% less cells than the drug-free control (Mothana *et al.*, 2009; Fithrotunnisa *et al.*, 2020), while Growth Inhibition Percentage (GIP) was calculated according Mosmann (1983).

### Energy Dispersive X-ray Spectroscopy

*M. alternifolia* and *P. guajava* leaves were mounted onto a stub with double-sided adhesive tape to determine the elements in *M. alternifolia* and *P. guajava* leaves with JEOL JSM-5300 SEM EDS at Faculty of Science, Alexandria University by using method of Scimeca *et al.*, (2018).

### Statistical Analysis

Some data of the present study were subjected to standard one way analysis of variance (ANOVA) using CoStat 6.303 (1998-2004) statistical analysis software manufactured by CoHort Software Company.

#### 1- Germination percentage (GP)

Germination percentage (GP) = (Number of germinated seeds/total number of seeds) X 100

#### 2- Inhibition percentage (IP)

The inhibition in seed germination as affected upon applying donor species extracts was calculated according to the formula of Cayuela *et al.* (2007).

Inhibition percentage (IP)=

[1- (allelopathic/control) 100]

#### 3- Mean Germination Time (MGT)

Mean germination time (MGT) was calculated according to the equation of Battle & Whittington (1969).

$MGT = \Sigma (G \times T) / F$

Where,

T = the day on which germination count was made, G = the number of seeds germinated on the day of the count, F = final number of seeds which germinated in each replicate.

#### 4- Seed Germination Index (SGI)

Seed germination index (SGI) was calculated according to the equation of Scott *et al.* (1984).

$SGI = \Sigma T_i N_i / S$

Where,

T<sub>i</sub> = is the number of days after sowing, N<sub>i</sub> = is the number of seeds germinated on day I and S = is the total number of seeds tested.

#### 5- Seedling Vigor Index (SVI)

Seedling vigour index (SVI) was calculated according to the equation of Islam *et al.* (2009) and Elouaer & Hannachi (2012).

$SVI = [\text{Seedling length (cm)} \times \text{germination percentage}] / 100$

## Results

### Phytochemical Screening

Phytochemical screening of *M. alternifolia* and *P. guajava* leaves represented in Table 1. and Table 2. respectively.

**Table 1.** Qualitative analysis of phytochemical constituents of *Melaleuca alternifolia* leaves.

Phytochemical classes	Tannins	Steroids	Flavonoids	Alkaloids	Carbohydrates	Glycosides
Aqueous extract	+	+	+	++	+	-
Ethanol extract	++	+	++	++	+	+

+: Detected; ++: highly detected ; -: Not detected.

**Table 2:** Qualitative analysis of phytochemical constituents of *Psidium guajava* leaves.

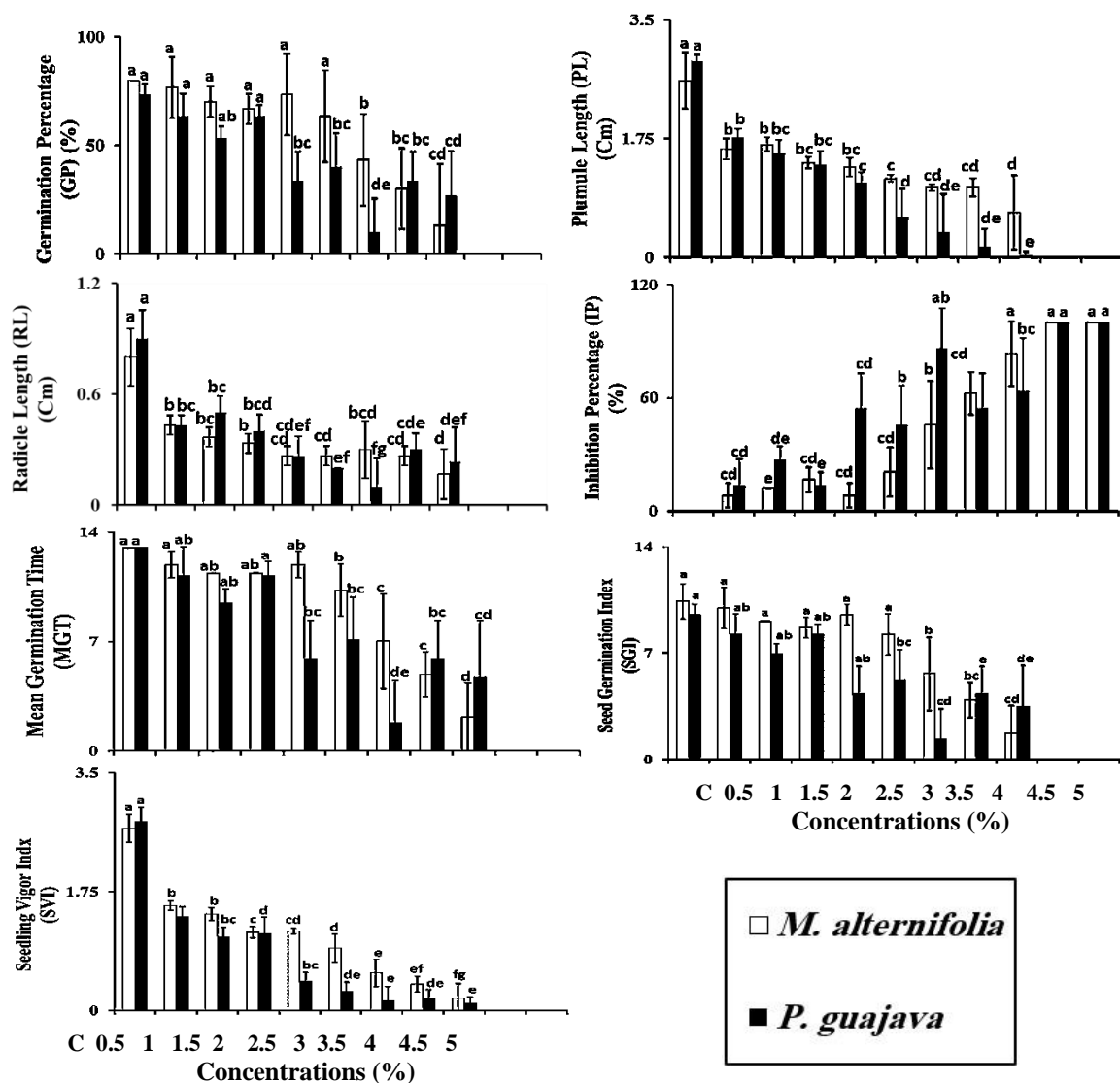
Chemical Constituents	Aqueous	Ethanol	Chemical Constituents	Aqueous	Ethanol
Carbohydrates	+	++	Alkaloids	+	++
Protein	-	+	Flavonoids	++	+++
Amino acid	+	+	Phlobatannins	+	+
Vitamin C	+	++	Steroids	+	-
Chloride	-	+	Phenolic compounds	+	+++
Tannins	+	++	Saponins	-	-

+++; Strongly positive; ++; positive; +; Trace; -: Not detected.

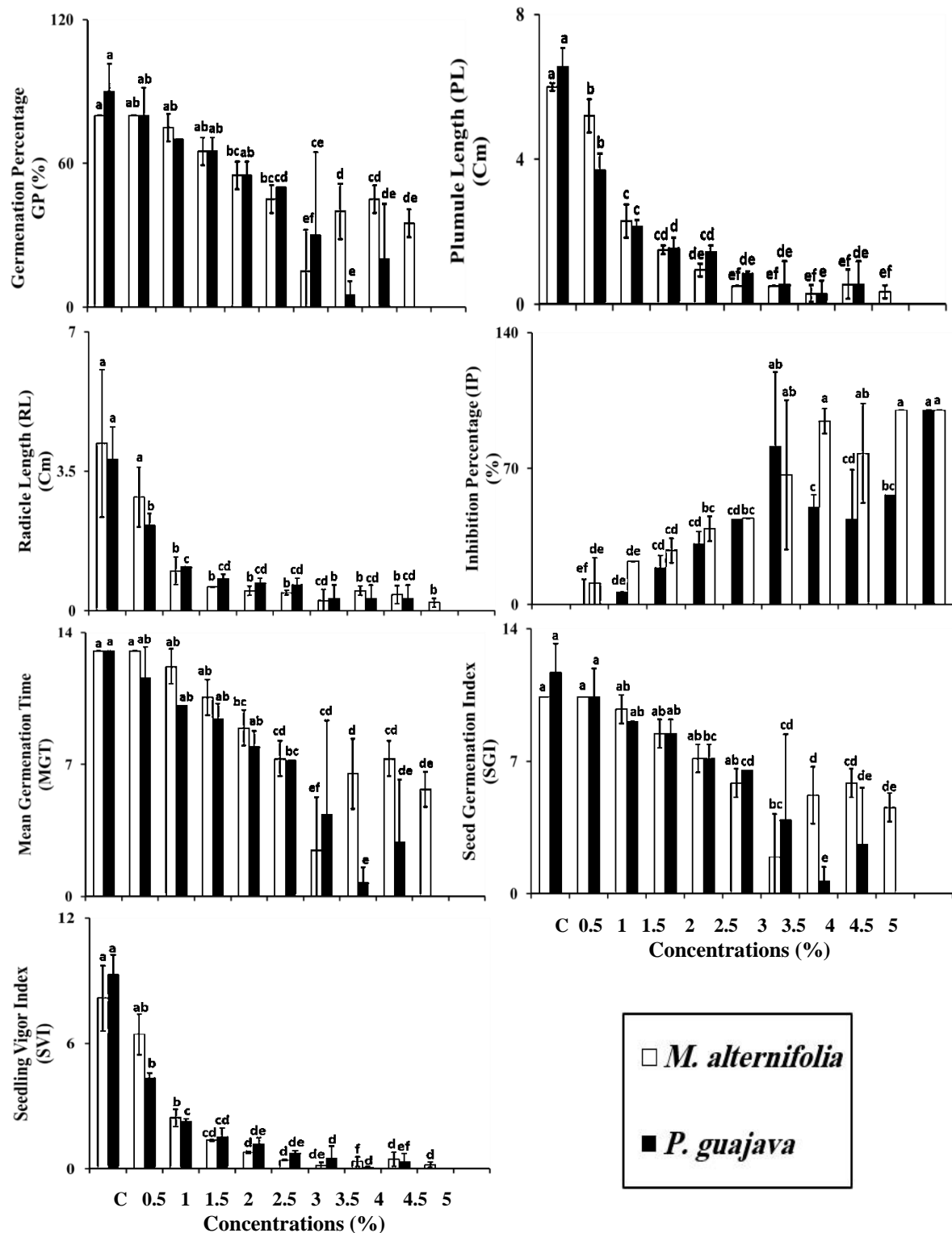
## Allelopathic Experiment

### A) Germination Bioassay Experiment

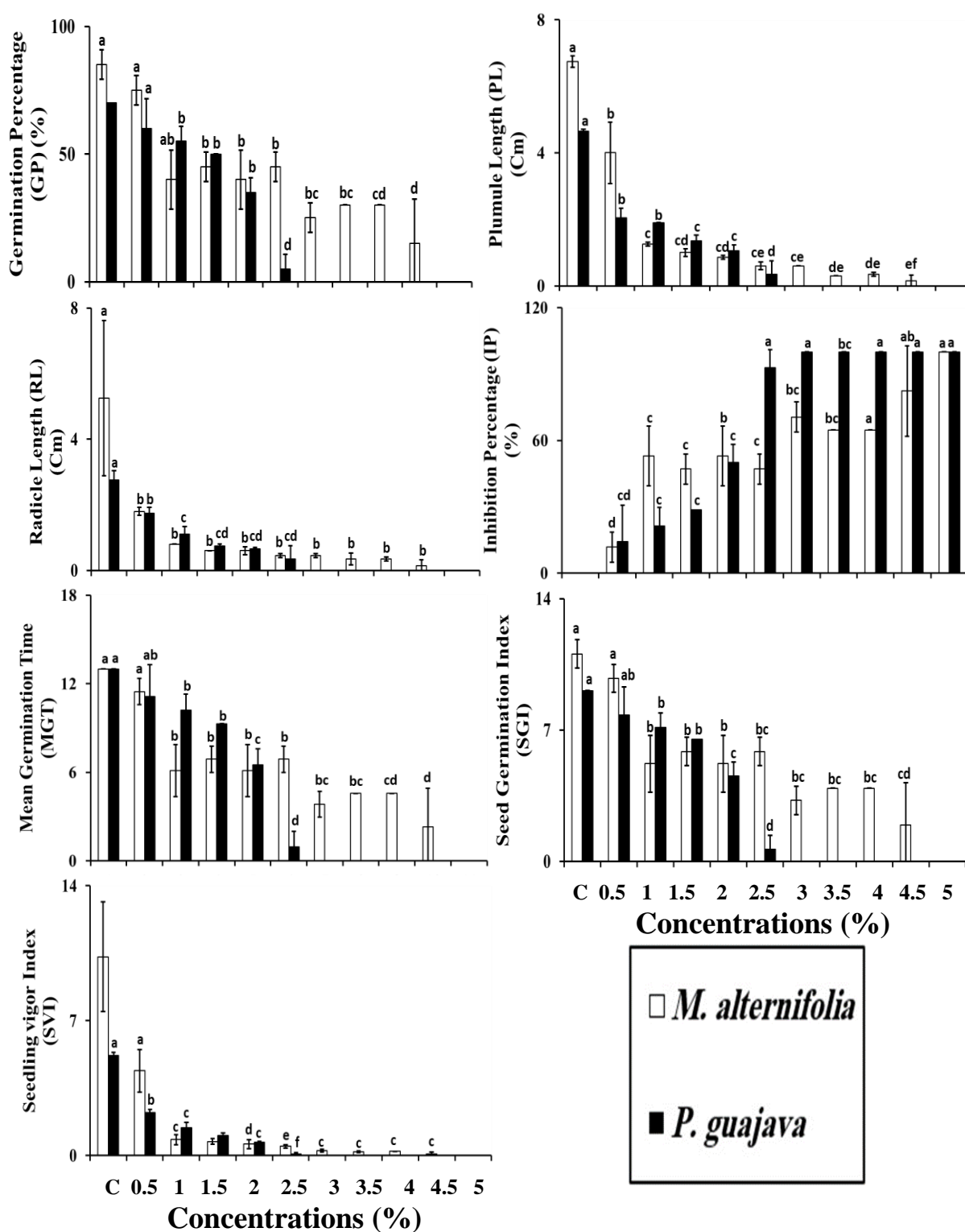
Data concerning germination percentage (GP), radicle length (RL), plumule length (PL), inhibition percentage (IP), mean germination time (MGT), seed germination index (SGI), seedling vigor index (SVI) of *R. dentatus*, *S. lycopersicum* (tomato extracted seeds) and *S. lycopersicum* (geaara 023) seeds as a recipient species that affected by *M. alternifolia* and *P. guajava* leaves aqueous extract are illustrated and statistically represented in Figure 1., Figure 2. and Figure 3. respectively.



**Figure 1:** Variation in germination percentage (GP), radicle length (RL), plumule length (PL), Inhibition percentage (IP), Mean Germination Time (MGT), Seed germination index (SGI), Seedling vigour index (SVI) of *R. dentatus* seeds affected by *M. alternifolia* and *P. guajava* leaves aqueous extract. Different letters within each column indicate a significant difference at  $p < 0.05$  according to one way ANOVA test. Error bars indicate standard error of means



**Figure 2:** Variation in germination percentage (GP), radicle length (RL), plumule length (PL), Inhibition percentage (IP), Mean Germination Time (MGT), Seed germination index (SGI), Seedling vigour index (SVI) of *S. lycopersicum* (tomato extracted seed) seeds affected by *M. alternifolia* and *P. guajava* leaves aqueous extract. Different letters within each column indicate a significant difference at  $p < 0.05$  according to one way ANOVA test. Error bars indicate standard error of means.



**Figure 3:** Variation in germination percentage (GP), radicle length (RL), plumule length (PL), Inhibition percentage (IP), Mean Germination Time (MGT), Seed germination index (SGI), Seedling vigour index (SVI) of *S. lycopersicum* (geaara 023) seeds affected by *M. alternifolia* and *P. guajava* leaves aqueous extract. Different letters within each column indicate a significant difference at  $p < 0.05$  according to one way ANOVA test. Error bars indicate standard error of means.



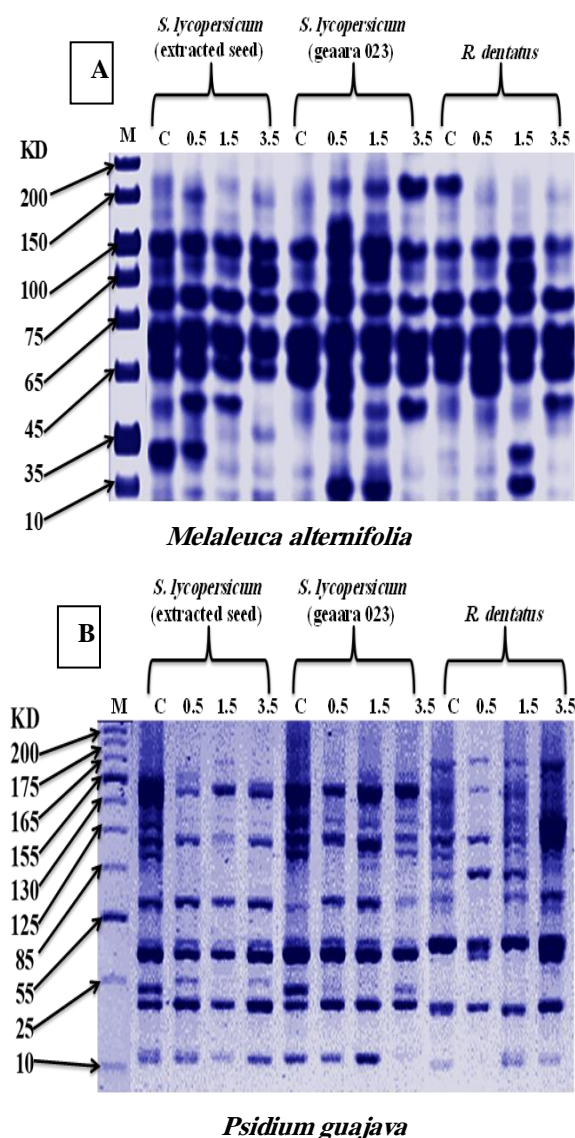
## B) Pot Experiment

### Seedling Protein Electrophoresis

The electrophenograms produced by SDS-PAGE (SDS-Polyacrylamide gel electrophoresis) of seedling proteins of *R. dentatus*, *S. lycopersicum* (extracted seed) and *S. lycopersicum* (geaara 023) affected by 0.5%, 1.5% and 3.5% of *M. alternifolia* leaves aqueous extract compared to control revealed a total of 22 bands come across all studied samples. The number of bands at 0.5% concentration level recorded values of about (12, 13 and 16), therefore the values varied to (12, 13 and 13) at 1.5% concentration level and lastly recorded values of about (14, 11 and 11) at 3.5%, compared to values of about (11, 13 and 13) at control level related to *R. dentatus*, *S. lycopersicum* (extracted seed) and *S. lycopersicum* (geaara 023) respectively. Protein profile exerts 3 common bands and absence of specific bands. The frequency of polymorphism at 0.5% concentration level recorded values of about (8% , 8% and 56%), while the values changed to (58%, 21% and 31%) at 1.5% concentration level and finally recorded values of about (43%, 36% and 27%) at 3.5% concentration level, compared to values of about (27%, 38% and 25%) at control level related to *R. dentatus*, *S. lycopersicum* (extracted seed) and *S. lycopersicum* (geaara 023) correspondingly Plate 1. A.

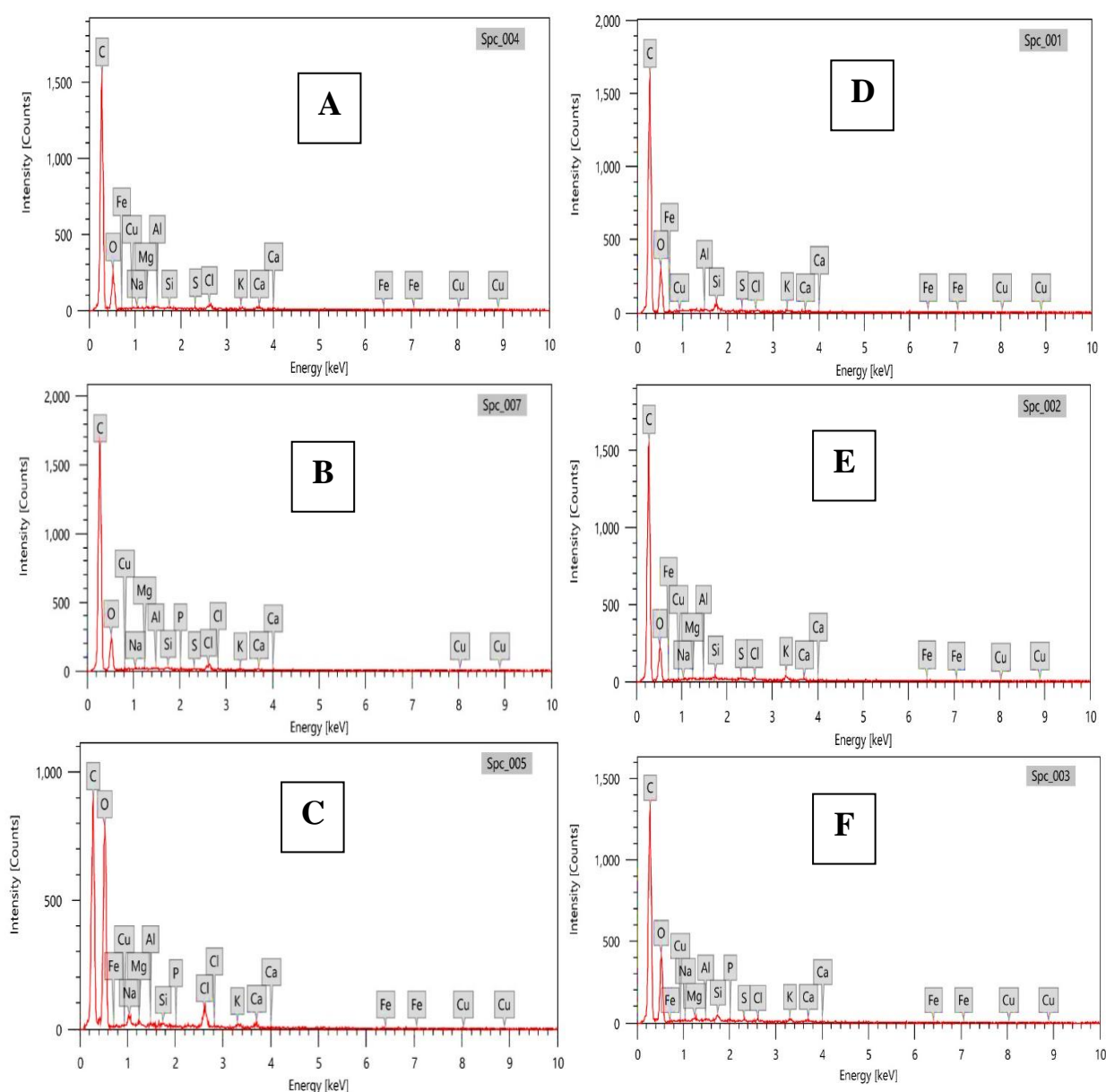
The electrophenograms produced by SDS-PAGE (SDS-Polyacrylamide gel electrophoresis) of seedling proteins of *R. dentatus*, *S. lycopersicum* (extracted seed) and *S. lycopersicum* (geaara 023) affected by 0.5%, 1.5% and 3.5% of *P. guajava* leaves aqueous extract compared to control showed a total of 18 bands come across all studied samples. The number of bands at 0.5% concentration level noted values of about (7, 10 and 9), therefore the values changed to (10, 7 and 13) at 1.5% concentration level and lastly recorded values of about (14, 11 and 10) at 3.5%, compared to values of about (11, 13 and 13) at control level related to *R. dentatus*, *S. lycopersicum* (extracted seed) and *S. lycopersicum* (geaara 023) respectively. Protein profile exerts 2 common bands and absence of specific bands. The frequency of polymorphism recorded null values at 0.5% concentration level, at 1.5% concentration level the frequency of polymorphism recorded values of about (20%, 14% and 23%) and finally recorded values of about (29%, 18% and 0%) at 3.5% concentration level, compared to values of about (0%, 25% and 14%) at control level related to *R. dentatus*, *S. lycopersicum* (extracted seed) and *S. lycopersicum* (geaara 023) respectively Plate 1. B.

**Plate 1:** Seedling protein electrophoresis attained from the seedling of *R. dentatus*, *S. lycopersicum* (tomato extracted seed) and *S. lycopersicum* (geaara 023) affected by 0.5%, 1.5% and 3.5% concentration of A: *Melaleuca alternifolia* and B: *Psidium guajava* leaves.



### Energy Dispersive X-ray Spectroscopy (EDS)

The analysis of *M. alternifolia* and *P. guajava* leaves was performed by using EDS method **Figure 4**. Ten elements were identified; C, O, Al, Si, S, Cl, K, Ca, Fe and Cu. The identified elements in *M. alternifolia* were detected in the following order C > O > Cl > Ca > K > Al > Cu > S > Fe > Si of about 70.1, 27.7, 0.63, 0.38, 0.32, 0.14, 0.13, 0.1, 0.07 and 0.06 respectively, while the identified elements in *P. guajava* revealed the following order C > O > Si > K > S > Fe > Al > Cu > Cl > Ca of about 67.9, 30.4, 0.5, 0.29, 0.2, 0.15, 0.11, 0.1, 0.09 and 0.08 correspondingly.



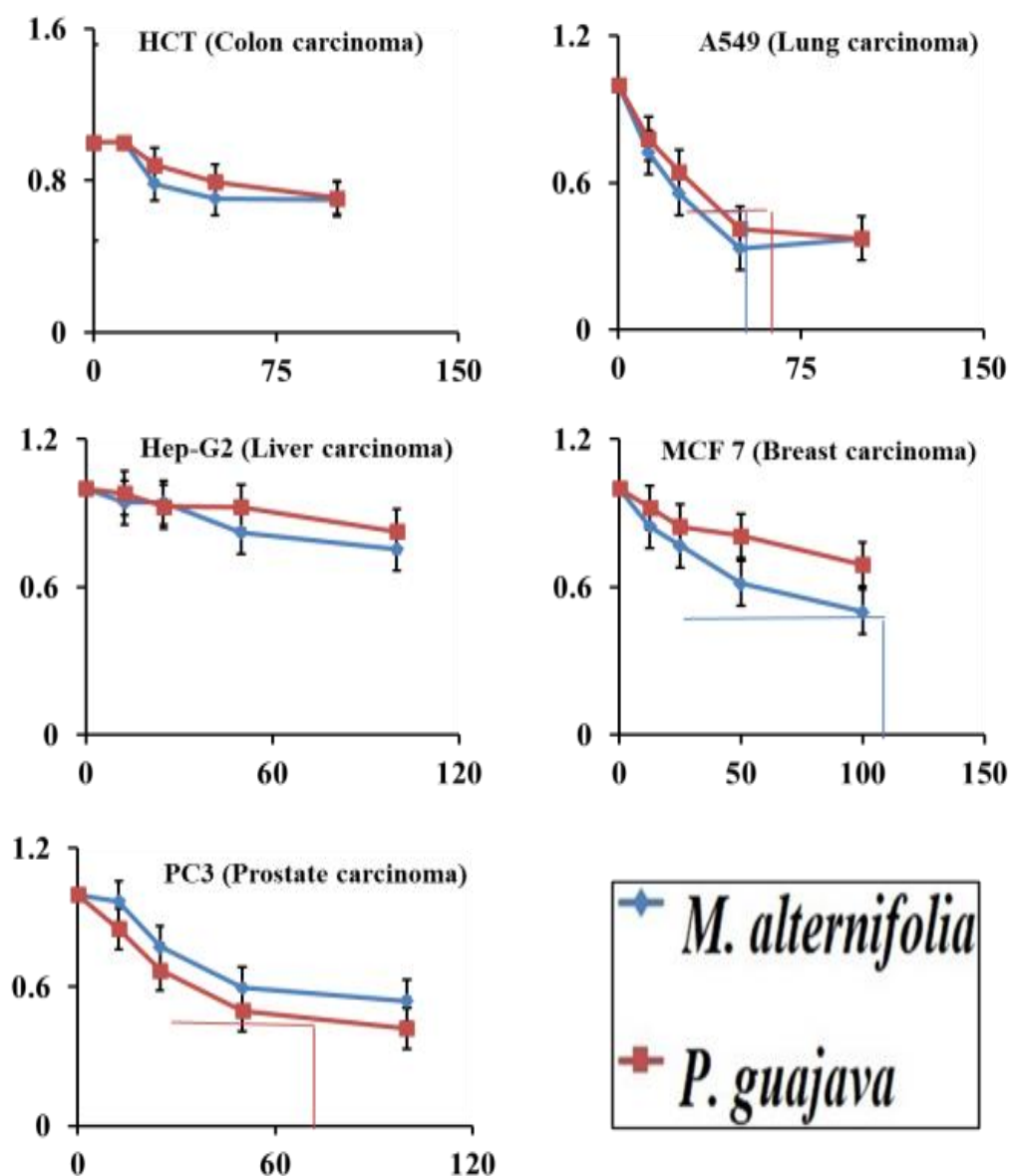
**Figure 4:** Analysis elements of *M. alternifolia* and *P. guajava* leaves by using Energy Dispersive X-ray Spectroscopy (EDS) method: (a-c) *M. alternifolia*; (d-f) *P. guajava*.

### Estimation of Anti-Proliferated Activity

The *in vitro* cytotoxic activity of *P. guajava* and *M. alternifolia* leaves ethanolic extract were determined on HCT (Colon carcinoma), A549 (Lung carcinoma), Hep-G2 (Liver hepatocellular carcinoma), MCF 7 (Breast carcinoma), and PC3 (Prostate carcinoma) carcinoma cell lines. The half-maximal inhibitory concentration IC<sub>50</sub> value (concentration of active compound needed to reduce the cell viability to 50%) was determined from dose-response curves of percent growth inhibition against test concentrations. To assess the toxicity of the extracts, each of the five cell lines was treated with four different concentrations of *P. guajava* and *M. alternifolia*. Visual observations

indicated that the viability of cancer cell lines was increasingly reduced while the concentrations of the extract were increased. *M. alternifolia* showed a significant effect on lung (A549) and MCF 7 (Breast carcinoma) cell lines with IC<sub>50</sub> of 31 and 98.9 µg/ml respectively. While HCT (Colon carcinoma), Hep-G2 (Liver hepatocellular carcinoma) and PC3 (Prostate carcinoma) attained no effect of IC<sub>50</sub>. *P. guajava* showed a significant effect on lung (A549) and PC3 (Prostate carcinoma) cell lines with IC<sub>50</sub> of 40 and 50.5 µg/ml respectively. While HCT (Colon carcinoma), Hep-G2 (Liver hepatocellular carcinoma) and MCF 7 (Breast carcinoma) attained no effect of IC<sub>50</sub> Figure 5.





**Figure 5:** Variation in The half-maximal inhibitory concentration value ( $IC_{50}$ ) through in vitro cytotoxic activity of *M. alternifolia* and *P. guajava* leaves ethanolic extract were determined on HCT (Colon carcinoma), A549 (Lung carcinoma), Hep-G2 (Liver hepatocellular carcinoma), MCF 7 (Breast carcinoma), and PC3 (Prostate carcinoma) carcinoma cell lines.

## Discussion

Myrtaceae is one of the dicotyledonous family belongs to order Myrtales and includes over 5650 species of about 130 to 150 genera and represents the eighth largest flowering plant family that comprises of several genera of extraordinary ecological and economic importance worldwide (Shah & Baghel, 2017). *M. alternifolia* and *P. guajava* are members of Myrtaceae family and have great ecological and economical roles in addition to their medicinal values (Carson *et al.*, 2006; Naseer *et al.*, 2018; Saber *et al.*, 2023).

*M. alternifolia* has therapeutic uses in traditional herbal medicine (Carson, *et al.* 2006; Refaey *et al.*, 2024). In the present study phytochemical screenings

of *M. alternifolia* and *P. guajava* leaves ethanolic extract exert high detection compared to aqueous extract. *M. alternifolia* leaves extract revealed presence of tannins, steroids, flavonoids and alkaloids that achieved high detection (++) compared to the others in both ethanolic and aqueous extracts. Concentration level of some secondary metabolites in *M. alternifolia* leaves disclosed presence of terpenes with high concentration level, such as terpinen-4-ol,  $\gamma$ -terpinene,  $\alpha$ -terpinene and methyl eugenol that acquires *M. alternifolia* various pharmacological activities such as anticancer, antioxidant, antimicrobial, and anti-inflammatory (Padovan *et al.*, 2017; Shah & Baghel, 2017; Shah *et al.*, 2019).

*P. guajava* has many medicinal values (Otuoma *et al.*, 2020; Kareem & Kadhim, 2024). *P. guajava* leaves ethanolic extract achieved high detection (+++) related to phenolic and flavonoids, while aqueous extract attained high detection (++) related to flavonoids that acquires *P. guajava* leaves various pharmacological activities such as antimicrobial, antimalarial, antimutagenic, anticancer, antitumor, anti-hyperglycemic and antinociceptive (Molla & Azene, 2017; Hashemi *et al.*, 2018; Camarena-Tello *et al.*, 2018; Lok *et al.*, 2023).

Allelopathy occurred by releasing many phytochemicals into the environment which is responsible for suppressing germination and growth of neighboring plants by modification their metabolism or degradation their soil communities (Shan *et al.*, 2023). In petri dish experiment *P. guajava* leaves aqueous extract exert great inhibition effect on some growth parameters such as GP, RL, PL, IP, MGT, SGI and SVI of *R. dentatus*, *S. lycopersicum* (geaara 023) and *S. lycopersicum* (tomato extracted seed) under all concentration levels (Control, 0.5%, 1%, 1.5%, 2%, 2.5%, 3%, 3.5%, 4%, 4.5% and 5% compared to *M. alternifolia* leaves aqueous extract that agree with Queiroz *et al.* (2017); Mabele & Ndong (2019) and Kapoor *et al.* (2019). The great allelopathic effect of *P. guajava* leaves aqueous extract due to presence of phenolic and flavonoids with excessive concentration according to Kawawa *et al.* (2016) and Motalebnejad *et al.* (2023).

Phenolics are recognized as one of the main promises allelochemicals in the ecosystem that is responsible for inhibition of seed germination and seedling growth (Madany & Saleh, 2015; Khan *et al.*, 2017; Patanè *et al.*, 2023), by inhabiting cell division and alter cells structures (Gomaa *et al.*, 2014; El-Metwally *et al.*, 2022) and suppress protein biosynthesis (Shahidi & Dissanayaka, 2023) and inactivate several enzymes (Singh *et al.*, 2021). Flavonoids exert a great inhibitory effect on seed germination and seedling growth (Gomaa *et al.*, 2014; Quy *et al.*, 2024).

Phenolics and flavonoids with excessive concentrations are described as eco-friendly so used as bioherbicides for weeds management (Alghamdi *et al.*, 2022; El-Metwally *et al.*, 2022). High concentration levels of phenolics and flavonoids interfere with cell division, hormone biosynthesis and photosynthesis (Alghamdi *et al.*, 2022), protein synthesis (Kuljarusnont *et al.*, 2024), which causes significant inhibition of weeds. *P. guajava* leaves aqueous extract causes a great inhiation in seed germination and seedling growth of *R. dentatus*, whereas *S. lycopersicum* (geaara 023) and *S. lycopersicum* (tomato extracted seed) growing safely

under low concentration levels of *M. alternifolia* leaves aqueous extract.

The analysis of *M. alternifolia* and *P. guajava* leaves was performed by using energy dispersive x-ray spectroscopy (EDS). *M. alternifolia* leaves revealed high concentration levels of Cl, Ca, Al and Cu elements, whereas *P. guajava* leaves exert high concentration levels of Si, S, Fe and K elements. High concentration levels of Si, S, Fe and K elements promote production of phenolics and flavonoids in *P. guajava* leaves that apparent as an ordinary defensive component against heavy elements toxicity (Shi *et al.*, 2018; Vega *et al.*, 2019).

Vegetative seedling storage protein profiling (VSPS) can be used for many objectives, such as supporting cropping polyculture ecosystems (Marzouk *et al.*, 2017) weed management (Khatab & El-Darier, 2020). The study evaluates the VSPS in *R. dentatus*, *S. lycopersicum* (tomato extracted seed) and *S. lycopersicum* (geaara 023) seedlings affected by 0.5, 1.5 and 3.5% concentration of *M. alternifolia* and *P. guajava* leaves aqueous extract. *P. guajava* leaves aqueous extract decreased GTS (%) of *R. dentatus* as a weed in concordance with Khatab and El-Darier (2020), however GTS (%) of *S. lycopersicum* (geaara 023) increase more than *S. lycopersicum* (tomato extracted seed) as crops under the effect of *M. alternifolia* leaves aqueous extract according to Marzouk *et al.* (2017). *P. guajava* has phenolics and flavonoids with excessive concentration that decrease the incorporation of phosphorus into DNA and RNA and reduce the incorporation of certain amino acid into proteins that decrease protein synthesis rate according to Khatab & El-Darier (2020), so *P. guajava* leaves aqueous extract decrease GTS (%) more stronger than *M. alternifolia* leaves aqueous extract.

The current study intended to explore and manipulate the cytotoxic and anti-proliferative effects of *M. alternifolia* and *P. guajava* leaves ethanolic extract on five human tested cell lines HCT (Colon carcinoma), A549 (Lung carcinoma), Hep-G2 (Liver hepatocellular carcinoma), MCF7 (Breast carcinoma) and PC3 (Prostate carcinoma).

*M. alternifolia* leaves ethanolic extract exhibited a strong significant anticancer activity against lung carcinoma A549 (IC<sub>50</sub>= 31 µg/ml), moderate significant anticancer activity against breast carcinoma MCF 7 (IC<sub>50</sub>= 98.9 µg/ml) and weak significant anticancer activity against HCT, Hep-G2 and PC3 carcinoma. Nalkiran & Nalkiran (2024) showed that the extracts of *M. alternifolia* induced the cell growth arresting apoptosis by down regulating NF-kb signaling in lung cancer cells. The anticancer activities of *M. alternifolia* are mainly due to its plentiful

steroidal saponins (Neychev *et al.*, 2007). Clark *et al.* (2021) stated that the extract of *M. alternifolia* has many biological activities such as cytotoxic, anti-proliferative and proapoptotic activities against prostate cancer and breast cancer. *M. alternifolia* has high antitumor potential due to presence of high steroidal saponins act as potential candidates can be used for this purpose (Sobolewska *et al.*, 2020; Elekofehinti *et al.*, 2021; Cui *et al.*, 2024).

According to Anggrelia *et al.* (2024) anticancer activities of *P. guajava* is due to excessive concentration of phenolics and flavonoids so considered as a free radical scavengers that help in gene expression regulation, DNA damage repair, cell proliferation and apoptosis (Luo *et al.*, 2014; Kadhim & Kareem, 2024). In the present study *P. guajava* leaves ethanolic extract exhibited a strong significant anticancer activity against lung carcinoma A549 (IC<sub>50</sub>=40 µg/ml), moderate anticancer activity against prostate carcinoma PC3 (IC<sub>50</sub>=50.5 µg/ml) and a weak anticancer activity against HCT, Hep-G2 and Mc7 carcinoma that agree with Lok *et al.* (2023), who demonstrated that *P. guajava* ethanolic extract has a significant cytotoxic activity against lung, breast, colon, prostate, leukemia, kidney and ovarian cancer cells.

## Conclusion

In the present study we can conclude that, *M. alternifolia* and *P. guajava* leaves ethanolic extract have high concentration level of terpenes and flavonoids respectively, which acquire them high pharmacological activities. *M. alternifolia* leaves revealed high concentration levels of Cl, Ca, Al and Cu elements, whereas *P. guajava* leaves exert high concentration levels of Si, S, Fe and K elements that promote production of phenolics and flavonoids which appear as an ordinary defensive component against heavy elements toxicity.

*P. guajava* leaves aqueous extract exert strongly decrease in genome template stability GTS (%) of *R. dentatus* so may be used as a powerful bioherbicide tool in controlling this pernicious weed, while *M. alternifolia* leaves aqueous extract revealed increase in GTS (%) of *S. lycopersicum* (geaara 023) more than more than *S. lycopersicum* (extracted seed) as crops so may be used as a powerful tool in mixing polyculture ecosystems.

*M. alternifolia* leaves ethanolic extract revealed a strong significant anticancer activity compared to *P. guajava* leaves ethanolic extract against lung carcinoma A549 (IC<sub>50</sub>=31 µg/ml) and (IC<sub>50</sub>=40 µg/ml) respectively. *M. alternifolia* exerts moderate anticancer activity against breast carcinoma MCF7 (IC<sub>50</sub>=98.9 µg/ml), while *P. guajava* showed a

moderate anticancer activity against prostate carcinoma PC3 (IC<sub>50</sub>=50.5 µg/ml).

## References

- Alghamdi, S.A., Al-Nehmi, A.A., Ibrahim, O.H. (2022). Potential allelopathic effect of wheat straw aqueous extract on Bermuda grass noxious weed. *Sustainability*, 14(23): 15989.
- Anggrelia, T.P., Dewi, I.C., Herawati, R.L., Azkiyah, S.M., Zahro, A.F., Maulana R.P., Seran, A.A., Klau, I.C., Ningsih, A.Y. (2024). Anticancer activity of guava leaf extract (*Psidium guajava*). *Al Makki Health Informatics Journal*, 2(1): 93-98.
- Bachheti, A., Sharma, A., Bachheti, R.K., Husen, A., Pandey, D.P. (2020). Plant allelochemicals and their various applications. *Co-Evolution of Secondary Metabolites* (Eds.). *Springer International Publishing* pp. 441-465.
- Baoduy, N., Trang, D.T., Trang N.P. (2015). Preliminary phytochemical analysis of leaf extracts *Ofthuja orientalis* (L.) Endl. *International Journal of Research Science and Management*, 2(1):21- 25.
- Battle, J.P., Whittington, W.J. (1969). The influence of genetic and environmental factors on the germination of sugar beet seed. *The Journal of Agricultural Science*, 73(3): 329-335.
- Camarena-Tello, J.C., Martínez-Flores, H.E., Garnica-Romo, M.G., Padilla-Ramírez, J.S., Saavedra-Molina, A., Alvarez-Cortes, O., Rodiles-López, J.O. (2018). Quantification of phenolic compounds and in vitro radical scavenging abilities with leaf extracts from two varieties of *Psidium guajava* L. *Antioxidants*, 7(3): 34.
- Carson, C.F., Hammer, K.A., Riley, T.V. (2006). *Melaleuca alternifolia* (tea tree) oil: A review of antimicrobial and other medicinal properties. *Clinical microbiology reviews*, 19(1): 50-62.
- Cayuela, M., Millner, P., Slovin, J., Roig, A. (2007). Duckweed (*Lemna gibba*) growth inhibition bioassay for evaluating the toxicity of olive mill wastes before and during composting. *Chemosphere*, 68(10): 1985-1991.
- Chen, K.C., Peng, C.C., Chiu, W.T., Cheng, Y.T., Huang, G.T., Hsieh, C.L. (2010). Action mechanism and signal pathways of *Psidium guajava* L. Aqueous extract in killing prostate cancer Incap cells. *Nutrition Cancer Journal*, 62(2): 260-270.
- Cheng, F., Cheng, Z. (2015). Research progress on the use of plant allelopathy in agriculture and the physiological and ecological mechanisms of allelopathy. *Frontiers in Plant Science*, 6:1020.

- Clark, A.M., Magawa, C., Pliego-Zamora, A., Low, P., Reynolds, M., Ralph, J. (2021). Tea tree oil extract causes mitochondrial superoxide production and apoptosis as an anticancer agent, promoting tumor infiltrating neutrophils cytotoxic for breast cancer to induce tumor regression. *Biomedicine and Pharmacotherapy*, 140: 111790.
- Cui, A., Liu, H., Liu, X., Zhang, M., Xiao, B., Wang, B., Yang, J. (2024) Steroidal saponins: Natural compounds with the potential to reverse tumor drug resistance (Review). *Oncology Letters*, 28: 585.
- Ejikeme, C., Ezeonu, C.S., Eboatu, A.N. (2014). Determination of physical and phytochemical constituents of some tropical timbers indigenous to Niger delta area of Nigeria. *European Scientific Journal*, 10(18): 247-270.
- Elekofehinti, O., Iwaloye, O., Olawale, F., Ariyo, E. (2021). Saponins in cancer treatment: current progress and future prospects. *Pathophysiology*, 28(2):250-272.
- El-Metwally, I.M., Saady, H.S., Elewa, T.A. (2022). Natural plant by-products and mulching materials to suppress weeds and improve sugar beet (*Beta vulgaris* L.) yield and quality. *Journal of Soil Science and Plant Nutrition*, 22(4): 5217-5230.
- Elouaer, M., Hannachi, C. (2012). Seed priming to improve germination and seedling growth of safflower (*Carthamus tinctorius*) under salt stress. *Eurasian Journal of Bioscience*, 6: 76-84.
- El-Rokiek, K.G., Shehata, A.N., Saad El-Din, S.A., Tarraf, S.A. (2024). An allelopathic evaluation of aqueous *Aloe vera* leaf and root extracts on the weed *Sonchus oleraceus* associated *Vicia faba* L. *Journal of Plant Protection Research*, 64(1): 11-18.
- Fithrotunnisa, Q., Arsianti, A., Kurniawan, G., Qorina, F., Tejaputri, N.A., Azizah, N.N. (2020). In vitro cytotoxicity of *Hibiscus sabdariffa* Linn extracts on A549 lung cancer cell line. *Pharmacognosy Journal*, 12(1):14-9.
- Gomaa, N.H., Hassan, M.O., Fahmy, G.M., González, L., Hammouda, O., Atteya, A.M. (2014). Allelopathic effects of *Sonchus oleraceus* L. on the germination and seedling growth of crop and weed species. *Acta Botanica Brasilica*, 28(3): 408-416.
- Hashemi. J.M., Haridy, L.A., Qashqari, R.J. (2018). Total phenolic, flavonoid and antioxidant compounds of guava whey juice fortified by *Moringa olifera* aqueous extract to extend shelf-life. *International Journal of Pharmaceutical Research and Allied Sciences*, 7(2): 86-100.
- Kadhim, E.J., Kareem, A.T. (2024). Isolation and identification of phenolic compounds in guava leaves and assessment of their cytotoxic effects against AMJ-13 and MCF-7 breast cancer cell lines. *Journal of the Faculty of Medicine Baghdad*, 66(3): 334-343.
- Kairey, L., Agnew, T., Bowles, E.J., Barkla, B.J., Wardle, J., Lauche, R. (2023). Efficacy and safety of *Melaleuca alternifolia* (tea tree) oil for human health-A systematic review of randomized controlled trials. *Frontiers in Pharmacology*, 14: 1116077.
- Kapoor, D., Rinzim, V., Tiwari, A., Sehgal, A., Landi, M., Brestic, M., Sharma, A. (2019). Exploiting the allelopathic potential of aqueous leaf extracts of *Artemisia absinthium* and *Psidium guajava* against *Parthenium hysterophorus*, a widespread weed in India. *Plants*, 8(12): 552.
- Kareem, A.T., Kadhim E.J. (2024). *Psidium guajava*: A review on its pharmacological and phytochemical constituents. *Biomedical and Pharmacology Journal*, 17(2): 1079-1090.
- Kawawa, R.C., Muyekho, F.N., Obiri, J.F., Agevi, H., Obiet, L. (2016). The allelopathic impact of *Psidium guajava* L. leaf extracts on the germination and growth of *Cassia occidentalis* L. seeds. *IOSR Journal of Agriculture and Veterinary Science*, 9(7): 101-105.
- Khalifa, A.A., Alnade, H.S., Kollab, W.A., Alafid, F., Edrah, S.M. (2017). Qualitative and quantitative phytochemical analysis and antimicrobial activity of *Retama* extract grown in Zliten Libya. *International Journal of Medical Science and Clinical Inventions*, 4(4): 2861-2866.
- Khan, A.U., Ullah, F., Mehmood, S., Irshad, M., Khan, F.U. (2017). Allelopathic effects of *Jatropha curcas* L. leaf aqueous extract on early seedling growth of *Parthenium hysterophorus* L. Pakistan *Journal of Agricultural Research*, 30(1): 1-8.
- Khattab, K., El-Darier, S. (2020). Allelopathic management of *Avena fatua* L. (Wild Oat) pernicious weed growing in *Triticum aestivum* (Wheat) L. crop fields. *Journal of Biodiversity and Environmental Sciences*, 16(1): 1-14.
- Kuljarusnont, S., Iwakami, S., Iwashina, T., Tungmunthum, D. (2024). Flavonoids and other phenolic compounds for physiological roles, plant species delimitation, and medical benefits: A promising view. *Molecules*, 29(22): 5351.
- Kumar, K.R., Kumar, P.P., Mallikarjuna, R.N. (2011). Development and validation of RP\_HPLC method for the estimation of ascorbic acid in health drink.



- Journal of Chemical and Pharmaceutical Research*, 3(3): 363-374.
- Kumar, M., Tomar, M., Amarowicz, R., Saurabh, V., Nair, M. S., Maheshwari, C., Satankar, V. (2021). Guava (*Psidium guajava* L.) leaves: *Nutritional composition, phytochemical profile, and health-promoting bioactivities*. *Foods*, 10(4): 752.
- Laemmli, U.K. (1970). Cleavage of structural proteins during the assembly of bacteriophage T4. *Nature*, 227(5259): 680-684.
- Liu, C., Jullian, V., Chassagne, F. (2024). Ethnobotany, phytochemistry and biological activities of *Psidium guajava* in the treatment of diarrhea: a review. *Frontiers in Pharmacology*, 15:1459066.
- Lok, B., Babu, D., Tabana, Y., Dahham, S., Adam, M., Barakat, K., Sandai, D. (2023). The anticancer potential of *Psidium guajava* (guava) extracts. *Life*, 13(2): 346.
- Luo, H., Cai, Y., Peng, Z., Liu, T., Yang, S. (2014). Chemical composition and in vitro evaluation of the cytotoxic and antioxidant activities of supercritical carbon dioxide extracts of pitaya (dragon fruit) peel. *Chemistry Central Journal*, 8(1):1-7.
- Mabele, A.S., Ndonga, M.F. (2019). Efficacy of guava (*Psidium guajava*) mulch allelopathy in controlling tomato (*Solanum lycopersicum*) weeds. *East African Journal of Agriculture and Biotechnology*, 1(1):7-11.
- Madany, M.M., Saleh A.M. (2015). Phytotoxicity of *Euphorbia helioscopia* L. on *Triticum aestivum* L. and *Pisum sativum* L. *Annals of Agricultural science*, 60(1):141-151.
- Marzouk, R.I., El-Darier, S.M., Mabrouk, M., Khattab, K.A. (2017). Growth and molecular expression of Okra seeds interacted with fourteen Mango cultivars in mixed cropping system. *Journal of Agricultural Science*, 9(8): 193.
- Mitra, S.K., Irenaeus, T.K., Gurung, M.R., Pathak, P.K. (2012). Taxonomy and importance of Myrtaceae. *Acta Horticulturae*, 959: 23-34.
- Molla, T., Azene, H. (2017). A Systemic review on antioxidant and hepato protective effect of *Psidium guajava* leaf and fruit extract. *Journal of Diseases and Medicinal Plants*. 3(2): 42-57.
- Mosmann, T. (1983). Rapid colorimetric assay for cellular growth and survival: application to proliferation and cytotoxicity assays. *Journal of Immunological Methods*, 65(1-2): 55-63.
- Motalebnejad, M., Karimmojeni, H., Majidi, M.M., Mastinu, A. (2023). Allelopathic effect of aqueous extracts of grass genotypes on *Eruca Sativa* L. Plants, 12(19): 3358.
- Mothana, R.A., Lindequist, U., Gruenert, R., Bednarski, P.J. (2009). Studies of the *in vitro* anticancer, antimicrobial and antioxidant potentials of selected Yemeni medicinal plants from the island Soqatra. *BMC Complementary and Alternative Medicine*, 9(7):22-29.
- Nalkiran, I., Nalkiran, H.S. (2024). Effects of *Melaleuca alternifolia* (tea tree) oil and vitamin E combination on the viability of non-small cell lung cancer cells. *Rize Medical Journal*, 2(1):34-48.
- Naseer, S., Hussain, S., Naeem, N., Pervaiz, M., Rahman, M. (2018). The phytochemistry and medicinal value of *Psidium guajava* (guava). *Clinical phytoscience*, 4(1): 1-8.
- Neychev, V.K., Nikolova, E., Zhelev, N., Mitev, V.I. (2007). Saponins from *Tribulus terrestris* L. are less toxic for normal human fibroblasts than for many cancer lines: influence on apoptosis and proliferation. *Experimental Biology and Medicine (Maywood)*, 232(1): 126 -33.
- Otuoma, J., Nyongesah, J.M., Owino, J., Onyango, A.A., Okello, V.S. (2020). Ecological manipulation of *Psidium guajava* to facilitate secondary forest succession in tropical forests. *Journal of Ecological Engineering*, 21(7): 210-221.
- Padovan, A., Keszei, A., Hassan, Y., Krause, S.T., Köllner, T.G., Degenhardt, J., Gershenzon, J., Külheim, C., Foley, W.J. (2017). Four terpene synthases contribute to the generation of chemotypes in tea tree (*Melaleuca alternifolia*). *BMC Plant Biology*, 17(160): 1-14.
- Patanè, C., Pellegrino, A., Cosentino, S.L., Testa, G. (2023). Allelopathic effects of *Cannabis sativa* L. aqueous leaf extracts on seed germination and seedling growth in durum wheat and barley. *Agronomy*, 13(2): 454.
- Queiroz, R.L., Pires, V., Bartelega, A., Carvalho, J.W., Serafim, A. (2017). Evaluation of *Melaleuca alternifolia* (Cheel) extract in the germination of *Brachiaria brizantha*. *Fitos Magazine*, 10(4): 397-403.
- Quy, N.T., Ngan, N.L., Tram, N.N., Men T.T. (2024). Investigating plant growth and germination inhibition of extract from *Leucaena leucocephala* (Lamk.) de Wit. *International Journal of Agricultural Technology*, 20(4):1527-1544.
- Rajendrabhai, V.D. (2017). Detection of phytochemical and pharmacological properties of crude extracts of *Tribulus terrestris* collected from tribal regions of Baglan (M.S.), India.



- Ramya, P., Vasanth, P.M., Prasad, P.V., Babu, V.S. (2019). Qualitative phytochemical screening tests of *Alpinia galanga* L. *World Journal of Pharmaceutical Research*, 8(5):1064-1077.
- Refaey, M.S., Abosalem, E.F., El-Basyouni, R.Y., Elsheriri, E.S., Elbehary, S.H., Fayed, M.A. (2024). Exploring the therapeutic potential of medicinal plants and their active principles in dental care: A comprehensive review. *Heliyon*, 10(18): e37641.
- Rice, E.L. (1984). "Allelopathy." 2nd Ed. *Academic Press, New York*. pp. 421.
- Saber, F.R., Munekata, P.E., Rizwan, K., El-Nashar, H.A., Fahmy, N.M., Aly, S.H., Lorenzo, J.M. (2023). Family Myrtaceae: The treasure hidden in the complex/diverse composition. *Critical Reviews in Food Science and Nutrition*, 7: 1-19.
- Scimeca, M., Bischetti, S., Lamsira, H.K., Bonfiglio, R., Bonanno, E. (2018). Energy dispersive X-ray (EDX) microanalysis: A powerful tool in biomedical research and diagnosis. *European journal of histochemistry*, 62(1): 2841.
- Scott, S.J., Jones, R.A., Williams, W.A. (1984). Review of data analysis methods for seed germination. *Crop science*, 24(6): 1192-1199.
- Sergazy, S., Vetrova, A., Orhan, I.E., Deniz, F.S. (2021). Antiproliferative and cytotoxic activity of Geraniaceae plant extracts against five tumor cell lines. *Future Science OA*, 3;8(2): FSO775.
- Shah, G., Baghel, U.S. (2017). Pharmacognostic standardization of the leaf of *Melaleuca alternifolia* (Maiden & Betcher) Cheel. *African Journal of Traditional, Complementary and Alternative Medicines*, 14(3): 1-11.
- Shah, G., Dhawan, R.K., Singh, S., Baghel, U.S. (2019). Antioxidant activity of methanol extract leaves of *Melaleuca alternifolia* (Maiden & Betcher) Cheel. *Journal of Materials and Environmental Sciences*, 10(12): 1286-1295.
- Shahidi, F., Dissanayaka, C. (2023). Phenolic-protein interactions: Insight from in-silico analyses - a review. *Food Production, Processing and Nutrition*, 5(1):2.
- Shan, Z., Zhou, S., Shah, A., Arafat, Y., Rizvi, S.A., Shao, H. (2023). Plant allelopathy in response to biotic and abiotic factors. *Agronomy*, 13(9):2358.
- Shi, P., Song, C., Chen, H., Duan, B., Zhang, Z., Meng, J. (2018). Foliar applications of iron promote flavonoids accumulation in grape berry of *Vitis vinifera* cv. Merlot grown in the iron deficiency soil. *Food chemistry*, 253: 164-170.
- Singh, A.A., Rajeswari, G., Nirmal, L.A., Jacob, S. (2021). Synthesis and extraction routes of allelochemicals from plants and microbes: A review. *Reviews in Analytical Chemistry*, 40(1): 293-311.
- Skehan, P., Storeng, R., Scudiero, D., Monks, A., McMahon, J., Vistica, D., Warren, J.T., Bokesch, H., Kenney, S., Boyd, M.R. (1990). New colorimetric cytotoxicity assay for anticancer-drug screening. *Journal of the National Cancer Institute*, 82(13): 1107-1112.
- Sobolewska, D., Galanty, A., Grabowska, K., Makowska-Wąs, J., Wróbel-Biedrawa, D., Podolak, I. (2020). Saponins as cytotoxic agents: An update (2010-2018). Part I-steroidal saponins. *Phytochem Rev*, 19:139-189.
- Sridhar, K., Rajesh, B., Sangeetha, K. (2016). Phytochemical screening and GC-MS analysis of ethanolic extract of *Tribulus Terrestris*. *International Journal of Clinical Pharmacology Research*, 6(1): 44-50.
- Vega, I., Nikolic, M., Pontigo, S., Godoy, K., Mora, M., Cartes, P. (2019). Silicon improves the production of high antioxidant or structural phenolic compounds in barley cultivars under aluminum stress. *Agronomy*, 9(7): 388.
- Yasin, M., Younis, A., Javed, T., Akram, A. (2021). River tea tree oil: Composition, antimicrobial and antioxidant activities and potential applications in agriculture. *Plants*, 10(10): 2105.