

(Review)

Recent advances in dual-targeting HDAC inhibitors for cancer treatment

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ABSTRACT

Histone deacetylases, often known as HDACs, are a class of enzymes that remove acetyl from the ϵ -N-acetyl lysine histones, thereby enabling histones to surround DNA with greater fidelity. HDACs are involved in a wide variety of biological activities, including gene regulation, transcription, cell proliferation, angiogenesis, migration, differentiation, and metastasis, among others. As a consequence, HDACs constitute an outstanding target for the discovery of anti-cancer drugs. The search for histone deacetylase inhibitors (HDACIs) has been expanded over the past decade with several B, and some have been offered. However, the HDACIs that are currently accessible are predominantly non-isoform selective and suffer from a number of problems, including limited efficacy, drug resistance, and toxicities. As a result, isoform-selective HDACIs and HDACIs with dual targeting capabilities have garnered a lot of attention from both academia and industry over the past five years. As a result, significant progress has been made in this field. In this paper, we summarize recent developments on HDACIs with dual targeting capabilities and their potential application in cancer treatment.

Keywords: HDAC; dual target; HDACIs.

Introduction

Cancer arises from alterations in the genetic material and variations in gene expression, including DNA methylation and histone modifications, which can impact the chromatin structure. [1],[2]. The nucleosome is the primary structural unit of chromatin, with a histone core that is coiled around it. A nucleosome consists of a pair of core histones, specifically H2A, H2B, H3, and H4 [3]. The main types of histone modifications include histone acetylation, phosphorylation, and methylation. Of these processes, histone acetylation is the predominant epigenetic process disrupted in cancer. [4]. The dysregulation of histone acetylation is intricately linked to the onset and progression of several malignancies. Typically, the process of acetylation of histones promotes the separation of DNA from histone octamers in nucleosomes, resulting in a more open and relaxed conformation. This allows for the binding of several transcription factors and cooperative transcription factors to DNA, resulting in the activation of specific genes that are engaged in different subsequent transcription processes [5]. Histone acetylation and histone deacetylation in the nucleus are regulated by histone acetyltransferases (HATs) and histone deacetylases (HDACs), respectively, to provide a stable and dynamic equilibrium. Histone acetyltransferases (HATs) catalyze the transfer of the acetyl group from acetyl-CoA to a specific lysine residue located at the amino terminus of histones. HDACs are enzymes that facilitate the removal of acetyl groups from histones, which have a strong affinity for negatively charged DNA. This leads to the condensation of chromatin and hinders the process of gene transcription. In addition, HDACs also specifically influence several non-histone substrates, including hormone receptors, chaperone proteins, and cytoskeleton proteins, to control essential physiological processes such as cell growth and cell apoptosis. Figure 2 illustrates the physiological roles of HDACs. HDACs are commonly acknowledged as transcriptional repressors because they can stabilize and condense chromosomes, hence reducing their accessibility to transcription factors [6]. In addition, HDACs act as suppressors of programmed cell death, whereas the administration of HDAC inhibitors boosts the synthesis of proteins that promote cell death, such as Bad, and decreases the amounts of proteins that prevent cell death, such as Bcl-2. HDAC inhibitors induce programmed cell death (apoptosis) in cancer cells through both internal (intrinsic) and external (extrinsic) routes [7]. In addition, HDACs have essential functions in DNA damage pathways since they act as central regulators of chromatin remodeling and the acetylation status of proteins involved in DNA damage. HDACs play a crucial role in regulating the cell cycle by influencing key components such as cyclin, CDK,

and nuclear material replication [8]. Recent research has confirmed that the process of autophagy is controlled by acetylation, specifically through the actions of HDAC6, which can trigger autophagy. Research has confirmed the connection between autophagy and the ubiquitinproteasome system [9],[10]. HDACs are often classified into four major categories: The protein HDAC 1 belongs to the class I. The categorization of histone deacetylases (HDACs) can be delineated into the subsequent classifications: The HDAC enzymes are categorized into four distinct classes: class I (HDAC 1, 2, and 8), class II (HDAC 4, 5, 6, 7, 9, and 10), class III (comprising of seven members: sirt1-sirt7, which rely on nicotinamide adenine dinucleotide (NADb)), and class IV (HDAC 11) [11], [12]. Class I and II HDACs are commonly referred to as "classic" HDACs. Class I HDACs are situated within the nucleus and possess the capacity to influence both histone and non-histone substrates. Class II histone deacetylases (HDACs) can be categorized into two subgroups: class IIa (comprising HDAC 4, 5, 7, and 9) and class IIb (consisting of HDAC 6 and 10). Class II HDACs have significant downstream effects. Divergent functions compared to class I HDACs [13]. Moreover, numerous studies have shown a clear link between the abnormal expression of HDACs and the development of particular cancerous tumors [14]. HDAC1 was discovered to be overexpressed in gastric [15], prostate [16], lung [17], and breast cancer [18]. Additionally, HDAC6 was increased specifically in breast cancer. In addition, there was an excessive presence of HDAC2 and HDAC3 in colorectal cancer [19],[20]. In cases of acute promyelocytic leukemia (PML), the target genes are inappropriately accessed by HDAC enzymes due to the presence of PML-RAR α (retinoic acid receptor- α) [21].

1 Compounds that inhibit histone deacetylase (HDAC) enzymes.

In recent years, a significant number of HDAC inhibitors have proven to be highly effective in treating cancer. Here are the names of five medications: The following are the names of five drugs: Vorinostat (SAHA), Romidepsin (FK228), Panobinostat (LBH-589), Chidamide (CS-055), and Belinostat (PXD101) (fig.1) I have been granted permission to provide medical treatment for hematological malignancies. HDAC inhibitors generally follow a traditional three-motif pharmacophore paradigm, consisting of a "Cap" group (an aromatic amide surface recognition group), a ZBG (zinc-binding group), and a linker that connects the ZBG and Cap. HDACIs can be categorized into four main groups based on their chemical structures: hydroxamic acids (such as Trichostatin A and Vorinostat), cyclic tetrapeptides (such as Romidepsin), benzamides (such as

Entinostat), and fatty acids (such as Valproic acid and Phenyl butyrate). TSA is a hydroxamic acid that is generated from the microorganisms Streptomyces hygroscopicus naturally. The US Food and Drug Administration (FDA) has officially approved SAHA, the first histone deacetylase (HDAC) inhibitor, exclusively for the treatment of cutaneous T cell lymphoma. Hydroxamates are commonly thought to have the capacity to affect all four types of HDAC enzymes. Fk228 is a tricyclic peptide derived from Bacillus violaceus and categorized as a natural hydroxamic acid. Entinostat (MS-275) is a benzamide HDAC inhibitor that has a prolonged duration of action and a high affinity for class I HDACs. Phenylbutyrate and valproic acid (VPA) are examples of HDAC inhibitors derived from fatty acids. Despite the efficacy of HDAC inhibitors in treating different cancer types, their utilization is restricted due to the incidence of adverse effects such as diarrhea, myelosuppression, and cardiovascular harm. The adverse consequences might be ascribed to the impact of HDACs on several cellular pathways, and the existing HDAC inhibitors primarily lack specificity for specific isoforms, sometimes known as pan-HDAC inhibitors. Hence, it is imperative to devise HDAC inhibitors that exhibit enhanced efficacy and selectively target certain isoforms. Isoform-specific HDAC inhibitors can mitigate the adverse effects linked to broadspectrum HDAC inhibitors. However, even HDAC inhibitors that specifically target a molecule can still encounter resistance in cancer treatments due to the activation of other pathways like AKT and CDK. Hence, it is crucial to develop HDAC inhibitors with the capability to selectively target several sites (known as dual-acting inhibitors) to successfully combat drug resistance caused by single-target HDAC inhibitors. During the last ten years, multiple evaluations have been conducted on medications that focus on two key regions related to HDAC. However, each study focuses on specific aspects of the dual inhibitors. Papavassiliou et al. [22] provided expert advice on the integration of HDAC inhibitors with other anti-tumor pharmacophores to create innovative tools for cancer treatment. Musso et al. provided a thorough summary of the progress made in the field of HDAC-based dual inhibitors up until 2015 [22]. In addition, they emphasized the significance of the pharmacological foundation for the logical progression of hybrid bifunctional drugs. Duan and his colleagues have performed a comprehensive analysis of dual-target inhibitors that are based on HDAC. Their review focused on analyzing the theoretical principles that underlie the creation of dual-target medications using HDAC, as well as the molecular interactions between HDAC and other target proteins. Furthermore, they investigated the methods by which compounds interact with their target proteins, yielding crucial insights into the relationships between the structure and function of these agents that target several locations. Given the ongoing advancements in the creation of dual-acting inhibitors that specifically target HDAC, it is imperative to provide a thorough summary of the latest accomplishments in this area from several angles [22].



2 Categorization of HDAC inhibitors with multifaceted targeting capabilities

2.1 Dual Tubulin/HDAC Inhibitors

Microtubules, which are made up of tubulins, have a crucial function in preserving cellular structure and enabling intracellular transportation. They are regarded as a prominent target for anticancer drugs. Studies have shown that the simultaneous use of tubulin inhibitors, such as taxanes and vincristine, along with HDAC inhibitors can result in synergistic effects. This discovery provides evidence for the advancement of dual tubulin/HDAC inhibitors [23]. Wang et al. found the chalcone molecule as a dual inhibitor of tubulin and HDAC. Compound 6 demonstrated greater inhibitory efficacy against A459 cells (IC₅₀ value of 0.55 μ M) and HeLa cells (IC₅₀ value of 0.89 μ M) compared to SAHA. Furthermore, it induced apoptosis and G2/M cell cycle arrest in A549 cells in a way that depended on the dosage [24].



2.2 Combination inhibitors that target both protein tyrosine kinase (PTK) and histone deacetylase (HDAC) enzymes.

Protein tyrosine kinases (PTKs) catalyze the transfer of the γ -phosphate group from adenosine triphosphate (ATP) to the hydroxyl group on the tyrosine residues of substrate proteins. Protein tyrosine kinases (PTKs) exert a substantial influence on the processes of cell division, differentiation, and apoptosis. Protein tyrosine kinases (PTKs) have been a prominent area of interest in the field of cancer therapeutic research during the past two decades. HDAC inhibitors, such as the Cap and linker components, have a high degree of flexibility for structural modifications. Therefore, the scientific community has recently displayed substantial interest in the progress of dual PTK-HDAC inhibitors, as elaborated below [25].

2.2.1 Dual BCR/ABL and HDAC inhibitors

Individuals diagnosed with chronic myelogenous leukemia (CML) are typically treated with tyrosine-kinase inhibitors, specifically Bcr-Abl TKIs, as the first-line therapy. Imatinib is an orally administered tyrosine kinase inhibitor (TKI) that selectively inhibits the activity of BCR/ABL kinase. Imatinib acts by competitively binding to the ATP site, inducing the enzyme to assume a closed or self-suppressed conformation. Consequently, the enzyme's activity is hindered. Compound 7 was created by Thomas et al. as a dual inhibitor of BCR/ABL and HDAC. This was achieved by mixing Imatinib with MS-275. This compound demonstrates strong inhibitory activity against HDAC1, with an IC₅₀ value of 77nM. Furthermore, compound 7 exhibited similar inhibitory effects on cell proliferation as MS-275 and Imatinib when evaluated on HeLa cells [26].



2.2.2 EGFR-HDAC dual inhibitors

EGFR is a prominent target for PTK inhibitors, and there has been substantial progress in developing dual EGFR HDAC inhibitors by utilizing the pharmacophores of various EGFR inhibitors and the HDAC inhibitor SAHA [27]. Cai et al. combined the basic pharmacophores of erlotinib and SAHA to create a class of medicines that had strong anticancer properties. Among all the compounds, compound 8 (CUDC101) exhibited the highest level of activity, with IC₅₀ values of 4.4 and 2.4nM for HDAC and EGFR, respectively [28].



2.2.3 Combination of drugs that simultaneously target vascular endothelial growth factor receptor (VEGFR) and histone deacetylase (HDAC)

Anti-angiogenic therapy has been extensively utilized in the management of cancer. Angiogenesis is controlled by a range of angiogenic factors, with VEGFR being the primary target for anti-angiogenic therapy that has been extensively studied. Over 10 monoclonal antibodies and tiny compounds that specifically target VEGFR have been approved for the treatment of solid tumors. Nevertheless, the emergence of drug resistance has impeded the efficacy of VEGFR inhibitors in medical therapies, leading to the exploration of multi-targeted inhibitors derived from VEGFR. Peng et al. synthesized compounds 9 and 10 by merging the pharmacophore components of SAHA and Vandetanib. These compounds have dual inhibitory activity against VEGFR and HDAC. Compounds 9 and 10 demonstrated exceptional inhibitory efficacy, displaying an IC_{50} value of less than 3 nM for HDAC and less than 85 nM for VEGFR-2 [29, 30].



2.2.4 Combination of FGFR and HDAC inhibitors

The fibroblast growth factor (FGF) family, along with their four receptor tyrosine kinases (FGFR1, FGFR2, FGFR3, FGFR4), have a vital function in various physiological processes. FGFR overexpression has been detected in several malignancies, including breast cancer, gastric cancer, and ovarian cancer. Therefore, FGFR has become a pivotal target for cancer therapy. Liu et al. synthesized compound 11, which functions as a dual inhibitor of FGFR1 and HDAC. Compound 11 exhibited potent inhibitory activity against HDAC6, demonstrating an IC₅₀ value of 34 nM. Furthermore, compound 11 exhibited similar anti-proliferative effects as SAHA (IC₅₀ = 1.4μ M) on MCF-7 cells [31].



2.2.5 Compounds that inhibit both Janus kinase (JAK) and histone deacetylase (HDAC) simultaneously.

Janus kinases (JAKs) are frequently upregulated in many types of cancer and are considered a promising target for cancer therapy. Yang et al. synthesized Compound 12 by merging the pharmacophores of pacritinib and SAHA. This drug acts as a dual inhibitor of JAK and HDAC. The IC50 values for HDAC6 and JAK2 are 2.1 nM and 1.4 nM, respectively [32].



2.3 Compounds that simultaneously inhibit HDAC and other targets

Over the past few years, several chemicals that can block the activity of both HDAC and other targets have been developed. This section will specifically concentrate on the dual inhibitors that are being studied.

2.3.1 Dual inhibitors of MDM2 and HDAC

Murine Double Minute 2 (MDM2) regulates the anti-tumor activity of p53, a gene that suppresses tumor growth, by binding to the p53 protein and keeping its levels low. Compound 13 is a potent dual inhibitor that efficiently inhibits the activity of MDM2 and HDAC6 enzymes, with IC_{50} values of 0.11µM and 17.5 nM, respectively [33].



2.3.2 Dual inhibitors of LSD1/HDAC

LSD1 was the first histone demethylase to be discovered. It selectively eliminates methyl groups from histones H3 lysine 4 (H3K4) and H3 lysine 9 (H3K9) that are mono or di-methylated, hence either inhibiting or enhancing gene transcription [34, 35]. Duan et al. identified a cluster of tranylcypromine derivatives, particularly compound 15, which had exceptional inhibitory efficacy. The IC₅₀ values for HDAC1, HDAC2, and LSD1 were determined to be 15 nM, 23 nM, and 1.2 μ M, respectively [36]. Nitric oxide (NO) is an essential signaling molecule that has a vital role in the management of cancer and inflammation [37].



LSD1/HDAC dual inhibitor 14

2.3.3 Dual inhibitors IDO/HDAC

Indoleamine2,3-dioxygenase (IDO) is an enzyme that regulates the speed at which tryptophan is broken down. It exerts a beneficial influence on the proliferation of certain cancer cells via modulating the immune system [38]. Fang et al. synthesized compound 15 as a dual inhibitor that effectively targets both IDO1 and HDAC. The compound has IC_{50} values of 69.0 nM and 66.5 nM for IDO1 and HDAC, respectively [39].



2.3.4 Dual inhibitors DNMT/HDAC

DNA methyltransferase (DNMT) is the enzymatic catalyst responsible for the addition of methyl groups to DNA. Overexpression of DNMT can lead to the suppression of tumor suppressor genes. A study conducted by Yuan et al. discovered that compound 16, which consists of NSC-319745 and SAHA, exhibits dual inhibition of DNMT and HDAC. Compound 16 exhibited potent inhibition of these enzymes, with IC₅₀ values of 2.02, 0.93, and 4.16 μ M for DNMT1/3A and HDAC1, respectively [40].



2.3.5 Dual inhibitors ER/HDAC

The estrogen receptor (ER) is a transcription factor that is controlled by ligands. It has been discovered that it is over-expressed in around 80% of breast cancers. Compound 17 had a notable inhibitory effect against HDAC3/6 and ERs, with IC₅₀ values of 0.734, 0.3, and 0.82 μ M, respectively [41].



2.3.6 Dual Inhibitors of NO/HDAC

They synthesized a series of compounds that function as dual inhibitors of nitric oxide (NO) and histone deacetylase (HDAC) by connecting the NO donor component to the α -position of the pyridine ring in MS-275. The consequences of enzyme inhibition. Compound 18 showed higher levels of inhibition against HDAC1, 2, and 3 in comparison to MS-275, as indicated by the results. Compound 18 had IC₅₀ values of 1.02, 0.37, and 1.19 μ M for HDAC1, 2, and 3, respectively. In comparison, MS-275 had IC₅₀ values of 1.07, 0.74, and 1.23 μ M for the same HDAC enzymes [42].



2.3.7 Dual Inhibitors of mTOR/HDAC

mTOR, the mammalian target of rapamycin, is a vital protein kinase that plays a pivotal role in integrating signals from both the external environment and within the cell. It is responsible for overseeing cell growth and a range of other physiological tasks [43]. Compound 19 was created by merging the pharmacophores of an HDAC inhibitor with a mTOR inhibitor, resulting in a dual

inhibitor of both targets. It exhibited potent biological action by efficiently suppressing both HDAC1/10 and mTOR, with an IC₅₀ in the nanomolar range [44].



2.3.8 Dual inhibitors of Ras/Raf MAPK/HDAC

The Ras/Raf mitogen-activated protein kinase (MAPK) pathway is strongly associated with the development of tumors. Compound 20 was created by combining the properties of the Raf inhibitor sorafenib and the HDAC inhibitor Chidamide. This resulted in a dual inhibitor that can target both Raf and HDAC. Compound 20 showed significant inhibitory effects against HDAC1 (with an IC₅₀ value of 1.17 μ M) and B-RafV600E (with an IC₅₀ value of 0.073 μ M), as demonstrated by the enzyme inhibition assay [<u>33</u>].



2.3.9 Dual Inhibitors of HMT/HDAC

G9a is a methyltransferase enzyme that catalyzes the addition of a methyl group to the lysine residue on the histone protein H3. Methylation specifically targets lysine 9 on histone H3 and is involved in regulating gene expression by suppressing transcription. Zanget et al. have developed a novel pharmaceutical compound, 21, that acts as a dual inhibitor of G9a/HDAC. This compound combines the quinazoline core of a G9a inhibitor with the hydroxamic acid moiety of an HDAC inhibitor. It effectively inhibits the function of both HDAC and G9a enzymes, with IC₅₀ values of 5.7 and 7.136 μ M, respectively [45].



2.3.10 Dual Inhibitors of COX/HDAC

Cyclooxygenase (COX) is divided into two different subtypes: COX-1 and COX-2. COX-2 is essential for the growth of cancer cells as it consistently supplies prostaglandin E2 (PGE2) to these cells [46]. The synthesized compound 22 by employing SAHA and indomethacin. This medication exhibited strong suppression of HDAC6 activity, with an IC₅₀ value of 5 nM [47].



2.3.11 Dual inhibitors of proteasome/HDAC

The proteasome, operating as a multi-enzyme complex, possesses the capacity to eradicate misfolded or unfolded proteins that pose a threat to cells [48]. RTX-V5 (24) is a potent dual inhibitor that efficiently blocks the activity of HDAC6 and HDAC8 enzymes. It achieves this by exhibiting an IC₅₀ value of 0.27 μ M for HDAC6 and 0.53 μ M for HDAC8 [49].



2.3.12 Dual Inhibitors of VDR/HDAC

The Vitamin D receptor (VDR) is a nuclear receptor regulated by ligands. VDR, acting as a gene transcription factor, plays a crucial role in controlling the cell cycle and differentiation. Compound 24 is a highly effective inhibitor of both the vitamin D receptor (VDR) and histone deacetylase (HDAC). The compound demonstrates notable inhibitory effects on cell proliferation, with an IC₅₀ value of 10 μ M against 4T1 cells [50].



2.3.13 Dual Inhibitors of Bcl-2/HDAC

The B-cell lymphoma-2 (Bcl-2) gene acts as an oncogene and can inhibit apoptosis. Compound 25 showed the highest level of enzymatic inhibition against HDAC6 (IC₅₀ = 28 nM) and Bcl-2 (IC₅₀ = 0.23μ M) [51].



2.3.14 Dual Inhibitors of HMGR/HDAC

Compound 26 was specifically engineered to target both HDAC and HMGR (3-hydroxy-3methylglutaryl coenzyme A reductase) by connecting the hydroxamate group with the essential structural components of statin, allowing it to bind to both proteins. Compound 26 demonstrated strong inhibitory activity against HDACs and HMGR, with IC_{50} values in the nanomolar range. Furthermore, compound 26 efficiently decreased the activity of HMGR and facilitated the acetylation of histone and tubulin in cancer cells. Remarkably, these effects were observed without causing any harm to normal cells [52].



2.3.15 Dual inhibitors of prototype PL/HDAC

In 2016, the discovery of that a hybrid compound 27, which contains SAHA and PL, had a strong ability to cause programmed cell death in AML cells by working together beneficially. The IC₅₀ values of 27 in different AML cell lines, including aggressive and chemotherapy-resistant cell types, were much lower than the IC₅₀ values of PL, SAHA alone, or their combination. Both PL and SAHA were demonstrated to synergistically promote DNA damage in U937 cells. This hybrid molecule serves as a beginning stage in the investigation of more potent and precise chemicals that selectively attack tumor cells. PL-HDAC is utilized for the treatment of hematological malignancies [53].



2.4 Dual cyclin-dependent kinases (CDKs) and HDAC inhibitors

Cyclin-dependent kinases (CDKs) can regulate the progression of the cell cycle and induce apoptosis, proliferation, and cellular expansion [54, 55]. Due to the excessive activation of CDKs in numerous cancers, researchers have created small molecule inhibitors to specifically target

CDKs in cancer treatment. Zhao et al. significantly inhibited HDAC1 (with an IC₅₀ of 17 nM) and CDK4 (with an IC₅₀ of 1.2 nM) by employing SAHA and abemaciclib, resulting in the synthesis of compound 28 [56].



Dual inhibitors of phosphatidylinositol 3-kinase (PI3K) and histone deacetylase (HDAC)

Studies have shown that cancer cells exhibit atypical amounts of phosphatidylinositol 3-kinase (PI3K) expression [57], Multiple PI3K inhibitors are currently being developed to treat solid tumors and hematological malignancies. Chen et al. have created compound 29, which acts as a dual inhibitor of HDAC and PI3K. It has demonstrated strong inhibitory activity against HDAC1 and PI3Ka, with IC₅₀ values of 2.7 and 50 nM, respectively [58]. Grewal and his team have created a novel drug named Inhibitor 30. This chemical combines the HDAC hydroxamate pharmacophore with Idelalisib, a PI3K inhibitor, utilizing a meticulously constructed linker (-benzyl). Compound 30 demonstrated strong inhibitory effects on many cancer cell lines and displayed remarkable selectivity and effectiveness (IC50 < 10 nM) against the PI3Kg, d, and HDAC6 enzymes. Crucially, compound 30 did not demonstrate any deleterious impacts on healthy cells, including PBMCs, NIH3T3, and HEK293. Nevertheless, it caused necrosis in numerous cell lines, including mutant and FLT3-resistant AML cells, as well as raw blasts obtained from AML patients [59].



2.5 Dual topoisomerase/HDAC inhibitors

Topoisomerase, often known as topo, is a frequently occurring ribozyme that can manipulate the structure of DNA's superhelix by creating reversible covalent complexes with the DNA [60, 61]. Several topoisomerase inhibitors, such as camptothecin and doxorubicin, have been specifically targeted targeting topoisomerase due to its vital biological functions in the proliferation of cancer cells. SAHA and the topo I inhibitor SN-38 served as the basis for the development of the dual-acting HDAC-topoisomerase I inhibitor 31, as reported by Oyelere and colleagues. Based on biological testing, 31 demonstrated strong suppression of cancer cell proliferation and maintained its effectiveness against HDAC (with IC_{50} values of 37 nM and 81 nM for HDAC1/6) and Topo I [62].



2.6 Dual nicotinamide phosphoribosyl transferase (NAMPT)/HDAC Inhibitors

Nicotinamide adenine dinucleotide (NAD⁺), an essential signaling molecule, is involved in the redox processes of several cells [63]. Nicotinamide phosphoribosyl transferase (NAMPT) controls the synthesis of NAD⁺. Recent research has discovered that cancer cells have a substantial dependency on NADP. Thus, by blocking NAMPT, the production of NAD⁺ in cancer cells can be limited, thereby affecting the growth and division of cancer cells [64]. Chen et al. synthesized Compound 32 by adding the hydroxamate group to the NAMPT inhibitor, resulting in a dual inhibitor of NAMPT and HDAC. Compound 32 demonstrated potent inhibitory activity against both HDAC1 and NAMPT, with IC₅₀ values of 2 and 15 nM, respectively [64].



Dual inhibitors of bromodomain and extra terminal (BET) proteins and histone deacetylases (HDACs)

The BET family, consisting of bromodomain and extra terminal proteins, is a prominent set of epigenetic regulators that control gene transcription and impact cell proliferation [65]. HDACs exert influence on gene transcription by participating in several epigenetic mechanisms. Considering the similar biological effects of HDACs and BETs, it is possible to construct a combination of both, known as dual BET/HDAC. Shao et al. developed Compound 33 as a dual inhibitor of BET and HDAC, using RVX-208 and SAHA as the basis. Compound 33 exhibited potent inhibitory effects on HDAC1 and BRD4/BD2, with IC₅₀ values of 0.2 and 0.4 μ M, respectively [66].



2.7 Dual poly (ADP-ribose) polymerase (PARP)/HDAC inhibitors

The DNA repair enzyme poly (ADP-ribose) polymerase (PARP) is essential for DNA replication, chromatin remodeling, and apoptosis [67, 68]. PARP is overexpressed in numerous solid and hematological malignancies [69]. Studies have shown that HDAC inhibitors and PARP inhibitors can enhance the responsiveness of other anticancer drugs [70, 71]. The development of dual inhibitors targeting both PARP and HDAC holds promise for cancer therapy. Yuan and

coworkers have created compound 34, which exhibits dual inhibitory effects for PARP and HDAC. Compound 34 has an IC₅₀ value of 8.21 nM for HDAC6 and an IC₅₀ value of 5.02 nM for PARP2.



2.8 Dual inhibitors of HSP90/HDAC

Heat shock protein 90 (Hsp90) is an ATP-dependent molecular chaperone that regulates the folding of oncoproteins. Studies have shown that the presence of HSP90 is elevated in cancer cells [72] Furthermore, HDAC6 specifically acts on HSP90 in a subsequent step. Dual inhibitors of HDAC and Hsp90 have a strong basis in the chaperone activity of Hsp90, which can be influenced by the reduction of HDAC6 [73]. Yao et al. revealed that Compound 35, which combines the pharmacophores of roxolitinib, BEP800, and SAHA, has a powerful inhibitory effect on JAK2, HDAC6, and HSP90. The IC₅₀ values for these inhibitory activities are 3.76 μ M, 6.30 μ M, and 20.2 μ M, respectively [74].



2.9 HDAC/FAK drugs that target both HDAC and FAK simultaneously

Focal adhesion kinase (FAK) is an intracellular enzyme that belongs to the class of non-receptor tyrosine kinases [75]. It plays a pivotal role in the processes of cancer cell proliferation, survival, and migration. FAK inhibitors that specifically target cancer cells have emerged as a promising treatment option for various types of malignancies. Research indicates that the combination of FAK and HDAC inhibitors has a synergistic effect in preventing cell growth, and HDAC inhibitors promote the deactivation of FAK caused by FAK inhibitors [76, 77]. Mustafa et al. discovered new compounds that can inhibit both HDAC2 and FAK. These compounds are based on 5-pyridyl-1,2,4-triazoles. Compound 36, which has the o-aminobenzamide group as a ZBG, has been found as a possible inhibitor of HDAC2 with an IC₅₀ value of 1.28 μM and FAK with an IC₅₀ value of 15.6 nM [78].



2.10 Dual targeting HDAC/proteasome inhibitors

The proteasome is a crucial enzyme in the ubiquitin-proteasome system (UPS) that is responsible for regulating and breaking down cellular proteins [79]. Inhibiting the proteasome has a significant impact on several signaling pathways within the cell. Currently, five known categories of drugs have been recognized as proteasome inhibitors. However, only a small number of compounds, like bortezomib, have progressed to the clinical development stage. This is because of issues related to metabolic instability, problems with effectiveness, or a lack of selectivity. In 2020, Zhou et al. developed and created new HDAC/Proteasome inhibitors using the fundamental pharmacodynamic structures of MS-275 and bortezomib [80]. Compound 37, which has the o-aminobenzamide group as its ZBG, demonstrated a remarkable inhibitory impact on the proteasome with an IC₅₀ of 1.1 nM. Additionally, it exhibited effective targeting of HDAC1 with an IC₅₀ of 0.255 μ M. In comparison to bortezomib, the molecule has shown greater effectiveness in inhibiting the growth of bortezomib-resistant multiple myeloma (MM) cell line KM3/BTZ. The

 IC_{50} values for the compound were 8.98 nM, whereas for bortezomib it was 226 nM. This difference was statistically significant (P < 0.01).



2.11 Dual targeting HDAC/Snail inhibitors

Snail, a zinc finger transcription factor [81], is the primary catalyst of epithelial-mesenchymal transition (EMT) and effectively suppresses the production of E cadherin. Research has demonstrated that Snail can interact with chromatin remodeling elements through its distinct domain (SNAG), hence performing its function of inhibiting transcription [82]. Founded Snail interacts with the histone complex (Sin3A-HDAC1/2) through this domain to deacetylate histone H3 and H4. This process inhibits the production of E-cadherin. To enhance the effectiveness of cancer treatment, Cui et al. developed a range of HDAC/Snail dual inhibitors, taking advantage of the significant influence that the combined application of Snail and HDACs has on cancer treatment [83]. One of the compounds, compound 38, with o-aminobenzamide as its ZBG, demonstrated the most potent inhibitory effects against HDAC1 (IC₅₀ = 0.405 μ M) and Snail (K_d = 0.180 μ M).



2.12 Dual targeting HDAC/MIF inhibitors

Macrophage migration inhibitory factor (MIF), often referred to as glycosylation inhibitory factor (GIF), is a cytokine with pro-inflammatory properties. It has a significant function in regulating the first stages of both innate and adaptive immune responses. Studies indicate that the MIF signal can enhance cell survival via the Akt pathway and hinder cell apoptosis by suppressing the tumor suppressor protein P53 [84]. Furthermore, MIF is abundantly present in several types of cancers such as gastric cancer, pancreatic cancer, and lung cancer, among others [85, 86]. There is significant potential to create small compounds that can simultaneously inhibit both HDACs and MIFs. In 2021, Cao et al. published a study introducing novel small molecule inhibitors that can simultaneously target HDAC/MIF [87]. Compound 39 showed outstanding inhibitory activity against MIF, with an IC₅₀ value of 0.18 μ M. It also demonstrated effective inhibition of HDAC, with IC₅₀ values of 0.2 μ M, 1.1 μ M, and 0.6 μ M for HDAC1, HDAC2, and HDAC3, respectively.



2.13 Dual targeting HDAC/estrogen inhibitors

Estrogen is a lipid-soluble steroid hormone that exists in three forms in the human body: estrone (E1), 17β -estradiol (E2), and estriol (E3). Multiple investigations have demonstrated a strong correlation between endogenous estrogen and the development of breast cancer [88]. Estrogen imbalances have the potential to lead to diseases including breast cancer and ovarian cancer. Gastric cancer and other types of cancer can result in the inhibition of important estrogen signaling.

This blockage, caused by chemotherapy, might lead to death in cancer patients due to toxicityrelated complications [89]. The combination of the HDAC inhibitor Vorinostat and fulvestrant is more efficient than fulvestrant alone in controlling the expression of cyclin and promoting estrogen oogenesis. The down-regulation of the receptor α (Er α) and the inhibition of its transcription is achieved. Mendoza Sanchez et al. [90] created and produced estrogen/HDAC inhibitors that target two areas simultaneously by merging the structural features of ICI-164384 and MS-275. One of the compounds, compound 40, had potent anti-estrogen activity (IC₅₀ = 0.21 µM) and outstanding HDAC3 inhibitory activity with an IC₅₀ value of 3.18 µM.



2.14 Dual targeting HDAC/androgen inhibitors

Prostate cancer, often known as PCa, is the predominant form of cancer found in males. The prevalence and development of this phenomenon are intricately linked to androgen, primarily testosterone, and are propelled by the androgen signaling system [90]. The androgen receptor (AR), which is a transcription factor activated by ligands, is currently a primary focus for therapeutic interventions in prostate cancer [91]. Due to the rapid increase in the number of prostate cancer cells, Barrett et al. created, produced, and assessed a range of anti-androgen/HDAC hybrids [92]. Out of all the compounds examined, those based on o-aminobenzamide exhibited superior performance. These compounds have greater inhibitory efficacy compared to hydroxamic acid-based drugs. Compound 41 demonstrated strong inhibitory effects against androgen and HDAC, with IC₅₀ values of 0.98μ M and 1.33μ M, respectively.



2.15 Dual targeting TRAIL/HDAC inhibitors

Tumor necrosis factor-related apoptosis-inducing ligand (TRAIL) is a type II transmembrane protein that belongs to the tumor necrosis factor superfamily [93]. Research has demonstrated that the concurrent use of histone deacetylases (HDACs) and TRAIL agonists can enhance the sensitivity of TRAIL to colon cancer, liver cancer, breast cancer, and other types of solid tumors [94-96]. Cui et al. developed and created a range of dual-target inhibitors for TRAIL/HDAC by including potent fragments and merging them with the HDAC inhibitor Chidamide [97]. One of the compounds is compound 42. The compound demonstrated potent inhibitory activity against HDAC1, with an IC₅₀ value of 151 nM. Additionally, it displayed increased sensitivity towards the production of the TRAIL protein.



Current challenges and future perspectives

Treatments that target a single agent or single target are often deemed ineffective in the setting of cancer due to the presence of redundant disease pathways and the development of drug resistance. Therefore, to improve the combined effectiveness and reduce the development of drug resistance, many drugs are used in several anti-cancer treatment processes [98]. An alternative strategy could be developing a singular medicine that has customized numerous pharmacodynamics. Several studies have been conducted on the utilization and incorporation of multi-target pharmacology in anti-tumor medications, including both small molecules and protein biologics [99, 100]. The notion of synergistic pharmacology involves targeting two enzymes simultaneously using a single complex or small molecule inhibitor. These enzymes are known to work together as epigenetic modulators, and by targeting them, gene expression can be effectively suppressed [101]. The discovery of dual inhibitors that selectively target different epigenetic modulators could offer a novel strategy for the advancement of epigenetic medications. Dual-targeting medications offer several potential advantages over conventional therapies, such as single-target or combination therapy. These advantages include more predictable pharmacokinetics, increased patient compliance, lower dosage requirements for delivery, and

enhanced potency with reduced toxicity. Nevertheless, there are numerous difficulties and obstacles involved in effectively implementing the concept of simultaneously targeting HDAC and other epigenetic regulators in laboratory experiments. The primary obstacle is the problem of optimizing the combined therapeutic effectiveness of the medicine within a living organism while also maximizing its safety profile. The issue arose in achieving a balance between the potencies of the medication and the desired levels of epigenetic modulators. The drug's in vitro activity should preferably be within a range of one order of magnitude. Attaining this objective is challenging, as it may ultimately compromise the effectiveness of either target. The development of dual epigenetic inhibitors that have a balanced effect on living organisms is made more complex by differences in how the inhibitors bind to their targets, how they are distributed throughout the body, and the levels of enzymes in different tissues. Additionally, the timing of when the inhibitors start to block their targets is also a factor [102]. The second obstacle is to achieve a satisfactory pharmacokinetic profile without compromising the actions of inhibitors. Most dual epigenetic inhibitors that are now known are created using either a pharmacophore-linked or -fused design technique. However, this approach often leads to the production of compounds with high molecular weight and unfavorable pharmacokinetic properties. The observed failure of certain dual epigenetic inhibitors is likely due to their suboptimal pharmacokinetics. This additional loophole must be promptly resolved by thoroughly investigating the SAR (Structure-Activity Relationship) of the parent target, while also considering all the advantages and disadvantages of the pharmacophore-merged and fragment-based screening techniques. The objective is to develop a small molecule dual epigenetic inhibitor with favorable pharmacokinetics. Dual action inhibitors intelligently aim to achieve equilibrium in inhibiting both enzymes simultaneously at the cellular level, while also overcoming the substantial regulatory obstacles posed by the two parent molecules. Efficiently arranged for a clinical trial. The robust correlation between histones and other epigenetic modulators, along with the demonstrated effectiveness of dual epigenetic inhibitors in treating cancer, will drive the exploration of dual epigenetic target treatments as a prominent area of research in the next years. It is expected that the continued examination of cancer's epigenetic networks will soon result in the discovery of inhibitors that target two epigenetic targets simultaneously[103].

Conclusion

As more intricate mechanisms are unveiled, it is fascinating to observe that epigenetic processes play a vital role not only in regular cellular activities but also in the formation of malignancies. The alteration in The manifestation of the epigenetic state becomes increasingly evident at the onset of cancer, so rendering therapy that focuses on the epigenome a possible avenue for cancer treatment. However, the main treatment strategy for malignancies revolves around methods that cause DNA damage. Nevertheless, the development of resistance and the lack of response impede the efficacy of the treatment. After the approval of vorinostat, belinostat, panobinostat, romidepsin, and chidamide, HDAC inhibitors (HDACI) have become generally suggested as a standard therapeutic method for treating a growing number of cancers. The concurrent targeting of HDAC inhibitors (HDACI) and medicines that specifically target other epigenetic pathways, such as BET, DNA/DNMT, G9a, LSD1, and EZH2, holds great potential. While many medications that specifically target these epigenetic pathways are currently undergoing clinical trials, their interactions with HDACi have not been studied yet. The simultaneous utilization of epigenetic regulators and HDACi as dual inhibitors has exhibited substantial synergy in both preclinical and clinical investigations. This synergistic combination has significant potential to advance the utilization of HDAC inhibitors in the fight against cancer.

• Conflict of interest

The Authors declare no conflict of interest

References

[1] A. Suraweera, K.J. O'Byrne, D.J. Richard, Combination therapy with histone deacetylase inhibitors (HDACi) for the treatment of cancer: achieving the full therapeutic potential of HDACi, Frontiers in oncology 8 (2018) 92.

[2] Z. Liu, T. Chen, Q. Han, M. Chen, J. You, F. Fang, L. Peng, B. Wu, HDAC inhibitor LMK-235 promotes the odontoblast differentiation of dental pulp cells, Molecular Medicine Reports 17(1) (2018) 1445-1452.

[3] X. Liao, Y. Liao, Y. Zou, G. Li, C. Liao, Epigenetic modifications of histone H3 during the transdifferentiation of Thy-1 (+) Lin (-) bone marrow cells into hepatocytes, Molecular Medicine Reports 12(5) (2015) 7561-7567.

[4] F. Meng, G. Sun, M. Zhong, Y. Yu, M.A. Brewer, Inhibition of DNA methyltransferases, histone deacetylases and lysine-specific demethylase-1 suppresses the tumorigenicity of the ovarian cancer ascites cell line SKOV3, International journal of oncology 43(2) (2013) 495-502.

[5] Y. Zhang, H. Lin, X. Guo, X. Zou, A case series pilot study on the combination of 5aminolevulinic acid and photodynamic therapy (ALA-PDT) for treatment of vitiligo*, Anais Brasileiros de Dermatologia 93(4) (2018) 539-545.

[6] G.D. Kim, Y.H. Choi, A. Dimtchev, S.J. Jeong, A. Dritschilo, M. Jung, Sensing of Ionizing Radiation-induced DNA Damage by ATM through Interaction with Histone Deacetylase*, Journal of Biological Chemistry 274(44) (1999) 31127-31130.

[7] B.E. Bernstein, J.K. Tong, S.L. Schreiber, Genomewide studies of histone deacetylase function in yeast, Proceedings of the National Academy of Sciences 97(25) (2000) 13708-13713.

[8] R. Mathew, V. Karantza-Wadsworth, E. White, Role of autophagy in cancer, Nature Reviews Cancer 7(12) (2007) 961-967.

[9] E. White, R.S. DiPaola, The double-edged sword of autophagy modulation in cancer, Clinical cancer research 15(17) (2009) 5308-5316.

[10] S. Li, X. Liu, X. Chen, L. Zhang, X. Wang, Histone deacetylase 6 promotes growth of glioblastoma through inhibition of SMAD2 signaling, Tumor Biology 36 (2015) 9661-9665.

[11] S. Yoon, G.H. Eom, HDAC and HDAC inhibitor: from cancer to cardiovascular diseases, Chonnam medical journal 52(1) (2016) 1.

[12] J. Ma, T. Luo, Z. Zeng, H. Fu, Y. Asano, Y. Liao, T. Minamino, M. Kitakaze, Histone deacetylase inhibitor phenylbutyrate exaggerates heart failure in pressure overloaded mice independently of HDAC inhibition, Scientific Reports 6(1) (2016) 34036.

[13] M. Yoshida, M. Kijima, M. Akita, T. Beppu, Potent and specific inhibition of mammalian histone deacetylase both in vivo and in vitro by trichostatin A, Journal of Biological Chemistry 265(28) (1990) 17174-17179.

[14] X.-Y. Shi, W. Ding, T.-Q. Li, Y.-X. Zhang, S.-C. Zhao, Histone deacetylase (HDAC) inhibitor, suberoylanilide hydroxamic acid (SAHA), induces apoptosis in prostate cancer cell lines via the Akt/FOXO3a signaling pathway, Medical science monitor: international medical journal of experimental and clinical research 23 (2017) 5793.

[15] J.H. Choi, H.J. Kwon, B.I. Yoon, J.H. Kim, S.U. Han, H.J. Joo, D.Y. Kim, Expression profile of histone deacetylase 1 in gastric cancer tissues, Japanese Journal of Cancer Research 92(12) (2001) 1300-1304.

[16] K. Halkidou, L. Gaughan, S. Cook, H.Y. Leung, D.E. Neal, C.N. Robson, Upregulation and nuclear recruitment of HDAC1 in hormone refractory prostate cancer, The Prostate 59(2) (2004) 177-189.

[17] S. Shuai, X. Yan, J. Zhang, S. Kang, F. Chen, R. Luo, A. Li, TIP30 nuclear translocation negatively regulates EGF-dependent cyclin D1 transcription in human lung adenocarcinoma, Cancer letters 354(1) (2014) 200-209.

[18] Z. Zhang, H. Yamashita, T. Toyama, H. Sugiura, Y. Ando, K. Mita, M. Hamaguchi, Y. Hara, S. Kobayashi, H. Iwase, Quantitation of HDAC1 mRNA expression in invasive carcinoma of the breast, Breast cancer research and treatment 94 (2005) 11-16.

[19] A.J. Wilson, D.-S. Byun, N. Popova, L.B. Murray, K. L'Italien, Y. Sowa, D. Arango, A. Velcich, L.H. Augenlicht, J.M. Mariadason, Histone Deacetylase 3 (HDAC3) and Other Class I HDACs Regulate Colon Cell Maturation and p21 Expression and Are Deregulated in Human Colon Cancer*, Journal of Biological Chemistry 281(19) (2006) 13548-13558.

[20] X. Liu, J.H. Wang, S. Li, L.L. Li, M. Huang, Y.H. Zhang, Y. Liu, Y.T. Yang, R. Ding, Y.Q. Ke, Histone deacetylase 3 expression correlates with vasculogenic mimicry through the phosphoinositide3-kinase/ERK–MMP–laminin5 γ 2 signaling pathway, Cancer Science 106(7) (2015) 857-866.

[21] R.J. Lin, T. Sternsdorf, M. Tini, R.M. Evans, Transcriptional regulation in acute promyelocytic leukemia, Oncogene 20(49) (2001) 7204-7215.

[22] X. Peng, Z. Sun, P. Kuang, J. Chen, Recent progress on HDAC inhibitors with dual targeting capabilities for cancer treatment, European Journal of Medicinal Chemistry 208 (2020) 112831.

[23] M.-W. Chao, M.-J. Lai, J.-P. Liou, Y.-L. Chang, J.-C. Wang, S.-L. Pan, C.-M. Teng, The synergic effect of vincristine and vorinostat in leukemia in vitro and in vivo, Journal of hematology & oncology 8 (2015) 1-15.

[24] B. Wang, X. Chen, J. Gao, L. Su, L. Zhang, H. Xu, Y. Luan, Anti-tumor activity evaluation of novel tubulin and HDAC dual-targeting inhibitors, Bioorganic & medicinal chemistry letters 29(18) (2019) 2638-2645.

[25] N. Zhou, W. Xu, Y. Zhang, Histone deacetylase inhibitors merged with protein tyrosine kinase inhibitors, Drug discoveries & therapeutics 9(3) (2015) 147-155.

[26] S. Mahboobi, S. Dove, A. Sellmer, M. Winkler, E. Eichhorn, H. Pongratz, T. Ciossek, T. Baer, T. Maier, T. Beckers, Design of chimeric histone deacetylase-and tyrosine kinase-inhibitors: a series of Imatinib hybrides as potent Inhibitors of wild-type and mutant BCR-ABL, PDGF-R β , and histone deacetylases, Journal of medicinal chemistry 52(8) (2009) 2265-2279.

[27] C.-J. Lai, R. Bao, X. Tao, J. Wang, R. Atoyan, H. Qu, D.-G. Wang, L. Yin, M. Samson, J. Forrester, CUDC-101, a multitargeted inhibitor of histone deacetylase, epidermal growth factor receptor, and human epidermal growth factor receptor 2, exerts potent anticancer activity, Cancer research 70(9) (2010) 3647-3656.

[28] X. Cai, H.-X. Zhai, J. Wang, J. Forrester, H. Qu, L. Yin, C.-J. Lai, R. Bao, C. Qian, Discovery of 7-(4-(3-ethynylphenylamino)-7-methoxyquinazolin-6-yloxy)-N-hydroxyheptanamide (CUDc-101) as a potent multi-acting HDAC, EGFR, and HER2 inhibitor for the treatment of cancer, Journal of medicinal chemistry 53(5) (2010) 2000-2009.

[29] F.-W. Peng, T.-T. Wu, Z.-W. Ren, J.-Y. Xue, L. Shi, Hybrids from 4-anilinoquinazoline and hydroxamic acid as dual inhibitors of vascular endothelial growth factor receptor-2 and histone deacetylase, Bioorganic & medicinal chemistry letters 25(22) (2015) 5137-5141.

[30] F.-W. Peng, J. Xuan, T.-T. Wu, J.-Y. Xue, Z.-W. Ren, D.-K. Liu, X.-Q. Wang, X.-H. Chen, J.-W. Zhang, Y.-G. Xu, Design, synthesis and biological evaluation of N-phenylquinazolin-4amine hybrids as dual inhibitors of VEGFR-2 and HDAC, European journal of medicinal chemistry 109 (2016) 1-12.

[31] J. Liu, C. Qian, Y. Zhu, J. Cai, Y. He, J. Li, T. Wang, H. Zhu, Z. Li, W. Li, Design, synthesis and evaluate of novel dual FGFR1 and HDAC inhibitors bearing an indazole scaffold, Bioorganic & medicinal chemistry 26(3) (2018) 747-757.

[32] E.G. Yang, N. Mustafa, E.C. Tan, A. Poulsen, P.M. Ramanujulu, W.J. Chng, J.J. Yen, B.W. Dymock, Design and synthesis of janus kinase 2 (JAK2) and histone deacetlyase (HDAC) bispecific inhibitors based on pacritinib and evidence of dual pathway inhibition in hematological cell lines, Journal of medicinal chemistry 59(18) (2016) 8233-8262.

[33] A. Geng, H. Cui, L. Zhang, X. Chen, H. Li, T. Lu, Y. Zhu, Discovery of novel phenoxybenzamide analogues as Raf/HDAC dual inhibitors, Bioorganic & medicinal chemistry letters 29(13) (2019) 1605-1608.

[34] Y. Duan, W. Qin, F. Suo, X. Zhai, Y. Guan, X. Wang, Y. Zheng, H. Liu, Design, synthesis and in vitro evaluation of stilbene derivatives as novel LSD1 inhibitors for AML therapy, Bioorganic & medicinal chemistry 26(23-24) (2018) 6000-6014.

[35] S. Hayami, J.D. Kelly, H.S. Cho, M. Yoshimatsu, M. Unoki, T. Tsunoda, H.I. Field, D.E. Neal, H. Yamaue, B.A. Ponder, Overexpression of LSD1 contributes to human carcinogenesis

through chromatin regulation in various cancers, International journal of cancer 128(3) (2011) 574-586.

[36] Y.-C. Duan, Y.-C. Ma, W.-P. Qin, L.-N. Ding, Y.-C. Zheng, Y.-L. Zhu, X.-Y. Zhai, J. Yang, C.-Y. Ma, Y.-Y. Guan, Design and synthesis of tranylcypromine derivatives as novel LSD1/HDACs dual inhibitors for cancer treatment, European journal of medicinal chemistry 140 (2017) 392-402.

[37] D. Hirst, T. Robson, Targeting nitric oxide for cancer therapy, Journal of Pharmacy and Pharmacology 59(1) (2007) 3-13.

[38] J. Godin-Ethier, L.-A. Hanafi, C.A. Piccirillo, R. Lapointe, Indoleamine 2, 3-dioxygenase expression in human cancers: clinical and immunologic perspectives, Clinical cancer research 17(22) (2011) 6985-6991.

[39] K. Fang, G. Dong, Y. Li, S. He, Y. Wu, S. Wu, W. Wang, C. Sheng, Discovery of novel indoleamine 2, 3-dioxygenase 1 (IDO1) and histone deacetylase (HDAC) dual inhibitors, ACS medicinal chemistry letters 9(4) (2018) 312-317.

[40] Z. Yuan, S. Chen, C. Gao, Q. Dai, C. Zhang, Q. Sun, J.-S. Lin, C. Guo, Y. Chen, Y. Jiang, Development of a versatile DNMT and HDAC inhibitor C02S modulating multiple cancer hallmarks for breast cancer therapy, Bioorganic chemistry 87 (2019) 200-208.

[41] A.F. Palermo, M. Diennet, M. El Ezzy, B.M. Williams, D. Cotnoir-White, S. Mader, J.L. Gleason, Incorporation of histone deacetylase inhibitory activity into the core of tamoxifen–A new hybrid design paradigm, Bioorganic & medicinal chemistry 26(15) (2018) 4428-4440.

[42] S. Atlante, K. Chegaev, C. Cencioni, S. Guglielmo, E. Marini, E. Borretto, C. Gaetano, R. Fruttero, F. Spallotta, L. Lazzarato, Structural and biological characterization of new hybrid drugs joining an HDAC inhibitor to different NO-donors, European Journal of Medicinal Chemistry 144 (2018) 612-625.

[43] C.S. Takeuchi, B.G. Kim, C.M. Blazey, S. Ma, H.W. Johnson, N.K. Anand, A. Arcalas, T.G. Baik, C.A. Buhr, J. Cannoy, Discovery of a novel class of highly potent, selective, ATP-competitive, and orally bioavailable inhibitors of the mammalian target of rapamycin (mTOR), Journal of Medicinal Chemistry 56(6) (2013) 2218-2234.

[44] Y. Chen, X. Yuan, W. Zhang, M. Tang, L. Zheng, F. Wang, W. Yan, S. Yang, Y. Wei, J. He, Discovery of novel dual histone deacetylase and mammalian target of rapamycin target inhibitors as a promising strategy for cancer therapy, Journal of Medicinal Chemistry 62(3) (2019) 1577-1592.

[45] L. Zang, S.M. Kondengaden, Q. Zhang, X. Li, D.K. Sigalapalli, S.M. Kondengadan, K. Huang, K.K. Li, S. Li, Z. Xiao, Structure based design, synthesis and activity studies of small hybrid molecules as HDAC and G9a dual inhibitors, Oncotarget 8(38) (2017) 63187.

[46] X.-C. Xu, COX-2 inhibitors in cancer treatment and prevention, a recent development, Anticancer drugs 13(2) (2002) 127-137.

[47] I. Raji, F. Yadudu, E. Janeira, S. Fathi, L. Szymczak, J.R. Kornacki, K. Komatsu, J.-D. Li, M. Mrksich, A.K. Oyelere, Bifunctional conjugates with potent inhibitory activity towards cyclooxygenase and histone deacetylase, Bioorganic & medicinal chemistry 25(3) (2017) 1202-1218.

[48] C. Wang, B. Ding, L. Jiang, C. Yin, Q. Zhong, G. Yu, X. Li, F. Meng, Increased expression of amyloid precursor protein promotes proliferation and migration of AML1/ETO-positive leukemia cells and be inhibited by panobinostat, Neoplasma 62(6) (2015) 864-871.

[49] S. Bhatia, V. Krieger, M. Groll, J.D. Osko, N. Reßing, H. Ahlert, A. Borkhardt, T. Kurz, D.W. Christianson, J. Hauer, Discovery of the first-in-class dual histone deacetylase–proteasome inhibitor, Journal of medicinal chemistry 61(22) (2018) 10299-10309.

[50] K. Bijian, D. Kaldre, T.-T. Wang, J. Su, M. Bouttier, A. Boucher, M. Alaoui-Jamali, J.H. White, J.L. Gleason, Efficacy of hybrid vitamin D receptor agonist/histone deacetylase inhibitors in vitamin D-resistant triple-negative 4T1 breast cancer, The Journal of Steroid Biochemistry and Molecular Biology 177 (2018) 135-139.

[51] R. Zhou, S. Fang, M. Zhang, Q. Zhang, J. Hu, M. Wang, C. Wang, J. Zhu, A. Shen, X. Chen, Design, synthesis, and bioactivity evaluation of novel Bcl-2/HDAC dual-target inhibitors for the treatment of multiple myeloma, Bioorganic & medicinal chemistry letters 29(3) (2019) 349-352.

[52] J.-B. Chen, T.-R. Chern, T.-T. Wei, C.-C. Chen, J.-H. Lin, J.-M. Fang, Design and synthesis of dual-action inhibitors targeting histone deacetylases and 3-hydroxy-3-methylglutaryl coenzyme A reductase for cancer treatment, Journal of medicinal chemistry 56(9) (2013) 3645-3655.

[53] Y. Liao, X. Niu, B. Chen, H. Edwards, L. Xu, C. Xie, H. Lin, L. Polin, J.W. Taub, Y. Ge, Synthesis and antileukemic activities of piperlongumine and HDAC inhibitor hybrids against acute myeloid leukemia cells, Journal of medicinal chemistry 59(17) (2016) 7974-7990.

[54] P. Nurse, Genetic control of cell size at cell division in yeast, Nature 256(5518) (1975) 547-551.

[55] A. Errico, K. Deshmukh, Y. Tanaka, A. Pozniakovsky, T. Hunt, Identification of substrates for cyclin dependent kinases, Advances in enzyme regulation 50(1) (2010) 375-399.

[56] B. Zhao, Z. Huang, Z. Qin, Y. Li, T. Wang, L. Wang, W. Zhou, C. Yu, X. Wang, S. Yang, Enhancement of histone deacetylase inhibitor sensitivity in combination with cyclin-dependent kinase inhibition for the treatment of oral squamous cell carcinoma, Cell Physiol Biochem 53(1) (2019) 141-156.

[57] S.Z. Millis, S. Ikeda, S. Reddy, Z. Gatalica, R. Kurzrock, Landscape of phosphatidylinositol-3-kinase pathway alterations across 19 784 diverse solid tumors, JAMA oncology 2(12) (2016) 1565-1573.

[58] D. Chen, C.K. Soh, W.H. Goh, H. Wang, Design, synthesis, and preclinical evaluation of fused pyrimidine-based hydroxamates for the treatment of hepatocellular carcinoma, Journal of medicinal chemistry 61(4) (2018) 1552-1575.

[59] A. Thakur, G.J. Tawa, M.J. Henderson, C. Danchik, S. Liu, P. Shah, A.Q. Wang, G. Dunn, M. Kabir, E.C. Padilha, Design, synthesis, and biological evaluation of quinazolin-4-one-based hydroxamic acids as dual PI3K/HDAC inhibitors, Journal of medicinal chemistry 63(8) (2020) 4256-4292.

[60] Y. Pommier, Drugging topoisomerases: lessons and challenges, ACS chemical biology 8(1) (2013) 82-95.

[61] Y. Pommier, Y. Sun, S.-y.N. Huang, J.L. Nitiss, Roles of eukaryotic topoisomerases in transcription, replication and genomic stability, Nature reviews Molecular cell biology 17(11) (2016) 703-721.

[62] W. Guerrant, V. Patil, J.C. Canzoneri, L.-P. Yao, R. Hood, A.K. Oyelere, Dual-acting histone deacetylase-topoisomerase I inhibitors, Bioorganic & medicinal chemistry letters 23(11) (2013) 3283-3287.

[63] L. Galluzzi, O. Kepp, M.G.V. Heiden, G. Kroemer, Metabolic targets for cancer therapy, Nature reviews Drug discovery 12(11) (2013) 829-846.

[64] W. Chen, G. Dong, Y. Wu, W. Zhang, C. Miao, C. Sheng, Dual NAMPT/HDAC inhibitors as a new strategy for multitargeting antitumor drug discovery, ACS Medicinal Chemistry Letters 9(1) (2018) 34-38.

[65] J. Li, P. Wang, B. Zhou, J. Shi, J. Liu, X. Li, L. Fan, Y. Zheng, L. Ouyang, Development of 4, 5-dihydro-benzodiazepinone derivatives as a new chemical series of BRD4 inhibitors, European Journal of Medicinal Chemistry 121 (2016) 294-299.

[66] M. Shao, L. He, L. Zheng, L. Huang, Y. Zhou, T. Wang, Y. Chen, M. Shen, F. Wang, Z. Yang, Structure-based design, synthesis and in vitro antiproliferative effects studies of novel dual BRD4/HDAC inhibitors, Bioorganic & Medicinal Chemistry Letters 27(17) (2017) 4051-4055.

[67] A. Sonnenblick, E. De Azambuja, H.A. Azim Jr, M. Piccart, An update on PARP inhibitors moving to the adjuvant setting, Nature reviews Clinical oncology 12(1) (2015) 27-41.

[68] P. Bai, Biology of poly (ADP-ribose) polymerases: the factorums of cell maintenance, Molecular cell 58(6) (2015) 947-958.

[69] V.B. Gandhi, Y. Luo, X. Liu, Y. Shi, V. Klinghofer, E.F. Johnson, C. Park, V.L. Giranda, T.D. Penning, G.-D. Zhu, Discovery and SAR of substituted 3-oxoisoindoline-4-carboxamides as potent inhibitors of poly (ADP-ribose) polymerase (PARP) for the treatment of cancer, Bioorganic & medicinal chemistry letters 20(3) (2010) 1023-1026.

[70] N.J. Curtin, C. Szabo, Therapeutic applications of PARP inhibitors: anticancer therapy and beyond, Molecular aspects of medicine 34(6) (2013) 1217-1256.

[71] L. Zhang, Y. Han, Q. Jiang, C. Wang, X. Chen, X. Li, F. Xu, Y. Jiang, Q. Wang, W. Xu, Trend of histone deacetylase inhibitors in cancer therapy: isoform selectivity or multitargeted strategy, Medicinal research reviews 35(1) (2015) 63-84.

[72] L. Whitesell, S.L. Lindquist, HSP90 and the chaperoning of cancer, Nature Reviews Cancer 5(10) (2005) 761-772.

[73] P. Bali, M. Pranpat, J. Bradner, M. Balasis, W. Fiskus, F. Guo, K. Rocha, S. Kumaraswamy, S. Boyapalle, P. Atadja, Inhibition of histone deacetylase 6 acetylates and disrupts the chaperone function of heat shock protein 90: a novel basis for antileukemia activity of histone deacetylase inhibitors, Journal of Biological Chemistry 280(29) (2005) 26729-26734.

[74] L. Yao, S. Ohlson, B.W. Dymock, Design and synthesis of triple inhibitors of janus kinase (JAK), histone deacetylase (HDAC) and Heat Shock Protein 90 (HSP90), Bioorganic & medicinal chemistry letters 28(8) (2018) 1357-1362.

[75] V.M. Golubovskaya, Targeting FAK in human cancer: from finding to first clinical trials, Frontiers in bioscience (Landmark edition) 19 (2014) 687.

[76] J.C. Dawson, B. Serrels, A. Byron, M.T. Muir, A. Makda, A. García-Muñoz, A. von Kriegsheim, D. Lietha, N.O. Carragher, M.C. Frame, A synergistic anticancer FAK and HDAC inhibitor combination discovered by a novel chemical–genetic high-content phenotypic screen, Molecular cancer therapeutics 19(2) (2020) 637-649.

[77] J. Song, X. Liu, Y.-F. Zhang, X.-Y. Tian, M.-Y. Deng, C.-Z. Huang, S.-Y. Zhang, The dual FAK-HDAC inhibitor MY-1259 displays potent activities in gastric cancers in vitro and in vivo, Bioorganic chemistry 131 (2023) 106328.

[78] M. Mustafa, A.A. Abd El-Hafeez, D. Abdelhamid, G.D. Katkar, Y.A. Mostafa, P. Ghosh, A.M. Hayallah, G.E.-D.A. Abuo-Rahma, A first-in-class anticancer dual HDAC2/FAK inhibitors bearing hydroxamates/benzamides capped by pyridinyl-1, 2, 4-triazoles, European journal of medicinal chemistry 222 (2021) 113569.

[79] L.R. Dick, P.E. Fleming, Building on bortezomib: second-generation proteasome inhibitors as anti-cancer therapy, Drug discovery today 15(5-6) (2010) 243-249.

[80] Y. Zhou, X. Liu, J. Xue, L. Liu, T. Liang, W. Li, X. Yang, X. Hou, H. Fang, Discovery of peptide boronate derivatives as histone deacetylase and proteasome dual inhibitors for overcoming bortezomib resistance of multiple myeloma, Journal of medicinal chemistry 63(9) (2020) 4701-4715.

[81] Y. Wu, Y. Zhang, D. Wang, Y. Zhang, J. Zhang, Y. Zhang, L. Xu, T. Wang, S. Wang, Q. Zhang, USP29 enhances chemotherapy-induced stemness in non-small cell lung cancer via stabilizing Snail1 in response to oxidative stress, Cell Death & Disease 11(9) (2020) 796.

[82] W.-X. Zhang, J. Huang, X.-Y. Tian, Y.-H. Liu, M.-Q. Jia, W. Wang, C.-Y. Jin, J. Song, S.-Y. Zhang, A review of progress in o-aminobenzamide-based HDAC inhibitors with dual targeting capabilities for cancer therapy, European journal of medicinal chemistry (2023) 115673.

[83] H. Cui, J. Huang, Y. Lei, Q. Chen, Z. Hu, J. Niu, R. Wei, K. Yang, H. Li, T. Lu, Design and synthesis of dual inhibitors targeting snail and histone deacetylase for the treatment of solid tumour cancer, European Journal of Medicinal Chemistry 229 (2022) 114082.

[84] H. Lue, M. Thiele, J. Franz, E. Dahl, S. Speckgens, L. Leng, G. Fingerle-Rowson, R. Bucala, B. Lüscher, J. Bernhagen, Macrophage migration inhibitory factor (MIF) promotes cell survival by activation of the Akt pathway and role for CSN5/JAB1 in the control of autocrine MIF activity, Oncogene 26(35) (2007) 5046-5059.

[85] K. Mangano, E. Mazzon, M.S. Basile, R. Di Marco, P. Bramanti, S. Mammana, M.C. Petralia, P. Fagone, F. Nicoletti, Pathogenic role for macrophage migration inhibitory factor in glioblastoma and its targeting with specific inhibitors as novel tailored therapeutic approach, Oncotarget 9(25) (2018) 17951.

[86] B. Otvos, D.J. Silver, E.E. Mulkearns-Hubert, A.G. Alvarado, S.M. Turaga, M.D. Sorensen, P. Rayman, W.A. Flavahan, J.S. Hale, K. Stoltz, Cancer stem cell-secreted macrophage migration inhibitory factor stimulates myeloid derived suppressor cell function and facilitates glioblastoma immune evasion, Stem cells 34(8) (2016) 2026-2039.

[87] F. Cao, Z. Xiao, S. Chen, C. Zhao, D. Chen, H.J. Haisma, F.J. Dekker, HDAC/MIF dual inhibitor inhibits NSCLC cell survival and proliferation by blocking the AKT pathway, Bioorganic Chemistry 117 (2021) 105396.

[88] H. Samavat, M.S. Kurzer, Estrogen metabolism and breast cancer, Cancer letters 356(2) (2015) 231-243.

[89] J. Blakemore, F. Naftolin, Aromatase: contributions to physiology and disease in women and men, Physiology (2016).

[90] R. Mendoza-Sanchez, D. Cotnoir-White, J. Kulpa, I. Jutras, J. Pottel, N. Moitessier, S. Mader, J.L. Gleason, Design, synthesis and evaluation of antiestrogen and histone deacetylase inhibitor molecular hybrids, Bioorganic & medicinal chemistry 23(24) (2015) 7597-7606.

[91] C.E. Massie, A. Lynch, A. Ramos-Montoya, J. Boren, R. Stark, L. Fazli, A. Warren, H. Scott, B. Madhu, N. Sharma, The androgen receptor fuels prostate cancer by regulating central metabolism and biosynthesis, The EMBO journal 30(13) (2011) 2719-2733.

[92] R.R. Barrett, C. Nash, M. Diennet, D. Cotnoir-White, C. Doyle, S. Mader, A.A. Thomson, J.L. Gleason, Dual-function antiandrogen/HDACi hybrids based on enzalutamide and entinostat, Bioorganic & Medicinal Chemistry Letters 55 (2022) 128441.

[93] B. Sun, Y. Liu, D. He, J. Li, J. Wang, W. Wen, M. Hong, 中药及其有效成分对 TRAIL 引起肿瘤细胞凋亡具有增敏作用, Journal of Zhejiang University-SCIENCE B 22 (2021) 190-203. [94] B. Zhang, B. Liu, D. Chen, R. Setroikromo, H.J. Haisma, W.J. Quax, Histone deacetylase inhibitors sensitize TRAIL-induced apoptosis in colon cancer cells, Cancers 11(5) (2019) 645. [95] Z.-J. Li, Y.-J. Hou, G.-P. Hao, X.-X. Pan, H.-R. Fei, F.-Z. Wang, CUDC-907 enhances TRAIL-induced apoptosis through upregulation of DR5 in breast cancer cells, Journal of Cell Communication and Signaling 14(4) (2020) 377-387.

[96] M.W. Shin, S.L. Kim, H.C. Yang, S.K. Yim, S.Y. Seo, S.T. Lee, H.-K. Kim, S.-W. Kim, The HDAC1 inhibitor CBUD-1001 enhances TRAIL-induced apoptosis in colorectal cancer cells, Anticancer Research 41(9) (2021) 4353-4364.

[97] H. Cui, Z. Hu, K. Yang, J. Huang, Y. Wu, Q. Chen, R. Wei, P. Wang, H. Wang, H. Li, Design and synthesis of highly TRAIL expression HDAC inhibitors based on ONC201 to promote apoptosis of Colorectal cancer, European Journal of Medicinal Chemistry 238 (2022) 114484.

[98] B. Al-Lazikani, U. Banerji, P. Workman, Combinatorial drug therapy for cancer in the postgenomic era, Nature biotechnology 30(7) (2012) 679-692.

[99] B. Apsel, J.A. Blair, B. Gonzalez, T.M. Nazif, M.E. Feldman, B. Aizenstein, R. Hoffman, R.L. Williams, K.M. Shokat, Z.A. Knight, Targeted polypharmacology: discovery of dual inhibitors of tyrosine and phosphoinositide kinases, Nature chemical biology 4(11) (2008) 691.

[100] M.P. Martin, S.H. Olesen, G.I. Georg, E. Schönbrunn, Cyclin-dependent kinase inhibitor dinaciclib interacts with the acetyl-lysine recognition site of bromodomains, ACS chemical biology 8(11) (2013) 2360-2365.

[101] J.H. Kalin, M. Wu, A.V. Gomez, Y. Song, J. Das, D. Hayward, N. Adejola, M. Wu, I. Panova, H.J. Chung, Targeting the CoREST complex with dual histone deacetylase and demethylase inhibitors, Nature communications 9(1) (2018) 53.

[102] J. Zhou, X. Jiang, S. He, H. Jiang, F. Feng, W. Liu, W. Qu, H. Sun, Rational design of multitarget-directed ligands: strategies and emerging paradigms, Journal of medicinal chemistry 62(20) (2019) 8881-8914.

[103] Y.-C. Duan, S.-J. Zhang, X.-J. Shi, L.-F. Jin, T. Yu, Y. Song, Y.-Y. Guan, Research progress of dual inhibitors targeting crosstalk between histone epigenetic modulators for cancer therapy, European Journal of Medicinal Chemistry 222 (2021) 113588.