10.21608/avmj.2025.312727.1349

Assiut University web-site: www.aun.edu.eg

DETECTION OF HAEMOPROTOZOA IN TURKEYS AND CHICKEN

MANAL H. HASAN 1; ENAS S. HUSSEIN 2 AND WAAD KHALID KHALAF3

¹ Professor, Department of Microbiology; College of Veterinary Medicine; University of Mosul, Mosul, Iraq. Email: manalhimmadi@uomosul.edu.iq

orchid: 0000 -0003 -3466 -2320

² Assistant lecturer, Department of Microbiology; College of Veterinary Medicine; University of Mosul, Mosul, Iraq. Email: enassaddi@uomosul.edu.iq

orchid: 0000 -0002 -8227 -1538

³ Assistant lecturer, Department of Microbiology; College of Veterinary Medicine; University of Mosul, Mosul, Iraq. Email: waadkhalid@uomosul.edu.iq orchid: 0000 -0002 -0356 -6275

Received: 10 November 2024; Accepted: 18 Febuary 2025

ABSTRACT

Haemoprotozoa are protozoa parasites that inhabit the blood and tissue of their avian hosts, specifically within their intracellular and extracellular erythrocytes. This study was conducted from October 2021 to March 2022 to identify and diagnose various species of blood parasites infecting chickens and turkeys in Mosul and Erbil. A total of 120 blood samples were collected from local chickens and turkeys. Blood smears were used for diagnostic purposes. After the examination of blood smears, the results showed the overall infection rates were 60% (54 out of 90) in local chickens (Gallus gallus domesticus) and approximately 40% (12 out of 30) in turkeys. Notably, chickens had a higher prevalence of infection in males (55.6%), whereas turkeys showed a higher prevalence in females (50%) (P < 0.05). Four species of blood protozoa were identified among chickens and turkeys: Leucocytozoon spp., Plasmodium spp., Haemoproteus, spp. Additionally, Trypanosoma species were exclusively identified in the blood smears of chickens. These findings highlight a significant prevalence of haemoprotozoa infections in both local chickens and turkeys, indicating potential health risks for poultry populations. It is essential to implement monitoring and control measures to mitigate the impact of these parasites on avian health and productivity. Continued research is crucial for developing effective strategies to manage haemoprotozoa infections in poultry.

Key words: Haemoprotozoa, turkeys, Chickens, *Leucocytozoon* spp., *Plasmodium* spp., *Haemoproteus Trypanosoma* spp.

INTRODUCTION

In addition to being a principal source of proceeds for the country, the poultry business is crucial to the food financing of

Corresponding author: Enas S. Hussein *E-mail address:* enassaddi@uomosul.edu.iq *Present address:* Assistant lecturer, Department of Microbiology; College of Veterinary Medicine; University of Mosul, Mosul, Iraq.

animal protein (meat and eggs) to humans. One of the domesticated poultry types that is elevated the most vigorous is the chicken, which is also the most progressive and effective animal production industry. Until now a huge difference had been set up between the commercialized and village enterprise subsector of poultry farming in Africa and some regions of Asia, each with their characteristics (Nnadi and George, 2010). However, the majority of commercial

poultry is produced as chickens and turkeys. Haemosporidia are intracellular protozoan parasites located within the erythrocytes also tissues of their avian hosts. They are widely spread and subsist in an assortment of avian species, together with local chicken and turkey. Various types of bacterial, fungal, parasitic pathogens effortlessly infect local poultry (Andreina and Ananias, 2011;). The following genera the most common hemoparasites discovered in poultry in tropical regions: Leucytozoon Plasmodium spp., Haemoproteus Aegytinella spp., spp., Trypanosoma spp., and microfilariasis of worms' relevance to the suborder filariata (Permin and Hanson, 1998). There are reports that the majority of poultry hemoparasites have closely linked life cycles (Ogbaie et al., 2019). Relying on the species of the protozoa and the tissues impacted, hemoparasitic infection in birds primarily causes anemia, along with a host of other serious diseases. (Dey et al., 2008; Naqvi et al., 2017; Ogbaie et al., 2019). Numerous studies have demonstrated that a variety of vectors, such as biting midges, lice, fleas, black flies, and mosquitoes, aid in the spread of hemoparasites (Dey et al., 2008; Ogbaie et al., 2019). The existence of the protozoa may not lead to significant devision of the haematology of the chickens, but the haematological parameters can only be significantly affected by the severity and pathogenicity of the protozoan implicated (Ogbaie et al., 2019). Eggs and meat from birds are vulnerable to protozoal diseases like haemosporidia (Hasson, 2015). Shadan (2013) detected that Plasmodium spp. was the most widespread haemoparasites in local chickens in Sulaimani, Iraq 52.63% (70/133).

Numerous researches have declaimed the geographical spreading of genetically obvious blood protozoal in various regions and locations (Chaser *et al.*, 2009). Since the presence of the vector is necessary for successful transmission, infections happen more frequently during the warmer months of the year. The sporozoite, which is found

in the salivary glands of the insect vector, is the infectious stage of the parasite (Friend and Franson, 1999; Tayler, 2015; Urquart *et al.*, 1996). In account of the important effects of avian haemoparasites on economic damage and mortalities, this study was conducted to diagnose *Leucocytozoon* spp., *Plasmodium* spp., Haemoproteus, *Trypanosoma* spp. in all affected local chickens and turkeys in Mosul and Erbil's city, Iraq.

MATERIAL & METHODS

1-Study area and sample collection

Local strains of domestic poultry (*Gallus gallus domesticus*) were acquired from various regions in Mosul and Erbil cities between October 2021 and March 2022. This included 90 chickens (48 males and 42 females) and 30 turkeys (18 males and 12 females). To collect blood samples, the wing vein of each bird was wiped with 70% ethyl alcohol, and approximately one milliliter of blood was drawn. For each individual sample, two blood smears were prepared to facilitate the morphological identification of hematozoa.

2- Samples Examination

The thin blood smears were prepared and fixed in absolute methanol for 5 minutes. Once the smears were fixed, they were stained using a 10% working solution of Giemsa's stain (Coles and Lea, 1986; Urquart et al, 1996). The prepared smears were examined under an oil immersion lens to identify blood protozoa. Different species, including Haemoproteus and Leucocvtozoon, were identified. *Plasmodium* infection confirmed bv recognizing intraerythrocytic schizonts or gametocyte stages of the parasites, while Trypanosoma spp. were identified as extracellular parasites (Coles and Lea, 1986; Fatima and Magboo, 2014; Tayler, 2015).

3-Statistical analysis

Chi-square analysis was performed on the data using the SPSS version to ascertain group differences at (P<0.05) (Niazi, 2001).

RESULTS

Microscopic examination revealed that the overall infection rates of haemoprotozoa in local strain domestic poultry and turkeys were 60% (54 out of 90) and 40% (12 out of 30), respectively, with a significance level of

P<0.05 for both single and double infections. The results identified four types of blood protozoa present among chickens and turkeys: *Leucocytozoon* spp., *Plasmodium* spp., *Haemoproteus*, and identified *Trypanosoma* spp.in chicken only (Table 1 & Fig 1).

Table 1: Infection rate of haemoprotozoa in chickens and turkeys.

			Infected	No. of	Positive	
Birds' species	Number examined	Number infected	rate %	negative samples	Number of single infections A	Number of double and mixed infections ^B
Chicken	90	54	60	36	42 a	12
Turkeys	30	12	40	18	12 ^b	0

Different letters mean there is a significant variance at a significant level P < 0.05

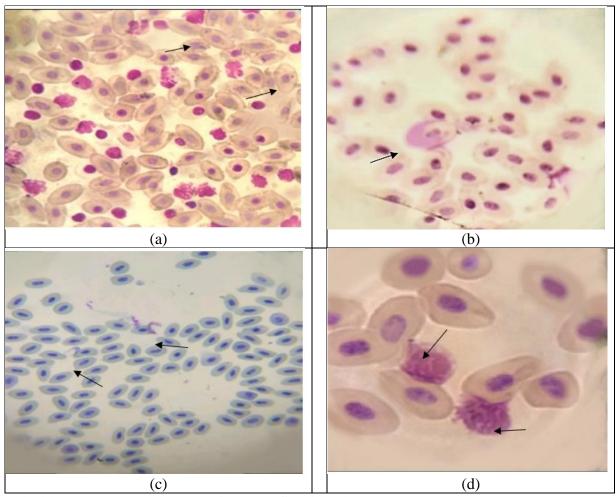


Fig (1) Haemoprotozoa stages in blood film in chicken

- (a) Plasmodium spp. 1000x (gametocyte stages ring shape)
- (b) Haemoproteus spp. 1000x gametocytes stage (discoid forms)
- (c) Trypomastigote form of Trypansoma spp. 1000x
- (d) Leucocytozoon spp. 1000x (gametocytes round shape)

In Mosul city, 33 out of 45 examined chickens tested positive for haemoprotozoa, resulting in an infection rate of 73.3%. Additionally, 6 out of 9 turkeys tested positive, yielding a rate of 66.6%. In Erbil City, the infection rates were lower, with 46.6% for chickens and 28.5% for turkeys,

showing a significant variation (P<0.05) (Table 2).

The present investigation recorded a high prevalence in males (62.5%) in chicken, while in turkeys recorded high prevalence in females (50%) P<0.05 (Table 3).

Table 2: Infection rate of haemoprotozoa in chickens and turkeys according to region.

	Mos	sul city	Erbil city		
Birds' species	number examined	number infected%	number examined	number infected%	
Chicken	45	33 (73.3%) ^a	45	21 (46.6%) ^a	
Turkeys	9	6 (66.6%) b	21	6 (28.5%) b	

Different letters mean there is a significant variance at a significant level P < 0.05

Table 3: infection rate of haemoprotozoa in chickens and turkeys according to sex

Birds' species	Total number examined (total number infected)	sex	number examined	number infected	Infection rate%
Chicken	90 (54)	Male	48	30 a	62.5%
		Female	42	24 ^b	57.1%
turkeys	30 (12)	Male	18	6 °	33.3%
		Famale	12	6 ^d	50%

Different letters mean there is a significant variance at a significant level P<0.05

The most commonly encountered haemoparasites in each chicken was Plasmodium (74.1%),Plasmodium Plasmodium Leucocytozoon (11.1%),Haemoproteus (9.3%),Plasmodium Haemoproteus, Leucocytozoon (1.8%)Trypansoma spp. (3.7%) (Table 4). While in turkeys was *Plasmodium* spp. (25%), Leucocytozoon spp. (33.3%). Haemoproteus spp. (41.6%) (Table 5).

DISCUSSION

The findings of our investigation have been detected the following genera of blood protozoa which affected the local hens and turkeys: *Trypanosoma*, *Plasmodium*, *Haemoproteus*, and Leucocytozoon. Our work's data is comparable to previous studies (Aiyedun *et al.*, 2022; Al-Kaabi, 2021; Al-Rubaie, and Ibrahim, 2020; Gimba

et al., 2014; Opara et al., 2014; Radfar et al., 2012).

Table 4: The percentage of infection with haemoprotozoa in chicken.

parasite spp.	infected number	infection %
plasmodium spp.	40	74.1 % ^a
Trypansoma spp.	2	3.7 % ^b
plasmodium spp. Haemoproteus spp.	5	9.3 % ^c
plasmodium spp. Leucocytozoon spp.	6	11.1 % ^d
plasmodium spp. Haemoproteus spp. Leucocytozoon spp.	1	1.8 % ^e
Total	54	

Different letters mean there is a significant variance at a significant level P < 0.05

Table 5: The percentage of infection with haemoprotozoa in turkey.

Parasites spp.	infected number	infection %
Plasmodium spp.	3	25 % ^a
Leucocytozoon spp.	4	33.3 % ^b
Haemoproteus spp.	5	41.6 % ^c
Total	12	

Different letters mean there is a significant variance at a significant level P < 0.05.

The infection predominance for the aforesaid genres of protozoa in this investigation was 60% in chickens and 40% in turkeys. This upshot is not in agreement with by (Opara et al., 2014) were discovered in chickens that had low rates of blood infections, however turkeys had higher infection rates, with a frequency of 8.9% in chickens and 40% in turkeys in Nigeria. Okereke, (2010) also revealed 14% frequency among chicken raised locally in Imo State. Nonetheless. Our findings concur with study in Ghana (Poulsen et al., 2000) were recorded prevalence infection with haemoparasites 35% in chicken in Ghana West Africa and 71% for Malawi (Njuga, 2000). Moreover, a propagation of 61.9 % has been recorded in Uganda (Valkiunas et al., 2005). This significant disparity may be explained by the different environmental factors that affect birds These factors involved insect vectors existence, conflict to protozoa, specimen time, steward immune system resistance, habitation attribute such as temperature, and rain-fall and steward specify of protozoa (Opara et al., 2014, Garcia-Longoria et al., 2019; Gutierrez-Lopez et al., 2015; Rivero de Aguilar et al., 2018).

The 73.3%, 66.6 % The frequency of haemoprotozoa infections was found to be 46.6% and 28.5% in Erbil city and Mosul city, respectively, with a substantially greater prevalence in Erbil than in Mosul.

These findings are consistent with earlier research (Hasson, 2015; Valkiunas, 2005).

This indicates that haemoprotozoa infections, a class of illnesses produced by protozoan parasites that affect a variety of animals inclusively birds and their populations, may represent a wide range of geographical changes. In the present show significant differences association between sex and haemoprotozoa infection was detected in chicken and turkeys, However, the reason for different prevalence across bird's sexes might be due to the differences in their reproductive activities (Adamu, 2017; Opara and Nwokedi, 2011).

Chickens and turkeys inspected in this investigation were infected mainly with Haemoproteus spp. Leucocytozoon spp., Plasmodiumspp. and finally, Trypanosoma spp., which was infected with the chickens, this result is contract with a previous study (Permin and Hanson, 1998) that show the haemoparasites primarily set up attacking poultry in the humid district involved Leucocytozoon spp. Plasmodium spp, Aegyptinella sp., Haemoproteus sp. and, Trypanosoma spp. The result also agreement with (Opara et al, 2014) blood protozoa that recorded prevalence rate (8.9%) in located chickens while (40%) in turkeys, The chickens parasitemia were Leucocytozoon spp., by a propagation percentage of (8.9%) in Nigeria. Also, in Sulaimani, Iraq was 78.2% the prevalence rate of haemosporidium parasites chickens (Abdullah, 2013). Analogous genres of blood protozoa distinguished during examination were described for contaminate hens in Ethiopia (Adamu, 2017). In current study recorded infection with Trypanosoma sp. (3.7 %) in chicken this result agreement with a previous study (Saif et al., 2008) who showed A number of Trypanosoma species have been identified in poultry including T. avium, T. gallinarum, T.numidae and T. calmetti, however these parasites ensure minimal pathological effects on their

Additionally, the results avian host. recorded different rates of different haemoparasites during the study months in single, double and mixed infection with haemoprotozoa in different rates. It suggests that the parasite vectors in their environment and that insecticide use is minimal (Mirzaei et al., 2020). The reasons behind the discrepancies in results across studies remain unclear, but could include factors associated with parasites infection such as the interval of seasonal blood samples picked up; the species' environmental and behavioral traits, the weather, the dissemination of habitatrelied on vectors, the host type, the age of the host (Fecchio et al., 2011; Sebaio et al., 2012).

CONCLUSION

The documents acquired from this study indicated that haemoprotozoa infections are widespread between chicken and turkeys by Mosul and Erbil city, Iraq. Plasmodium spp., Haemoproteus spp., Leucocytozoon spp had been infected chicken and turkey as well as infection with Trypanosoma spp. Were recorded in chicken. Gender was significantly accompanying through the haemoprotozoa infections among chicken and turkeys in the investigation district. Single, double and mixed haemoprotozoa infections were recorded in chicken and single infection in Turkeys was confirmed by this study.

ACKNOWLEDGMENT

We are deeply grateful to the Veterinary Medicine College, University of Mosul, for their sincere support and encouragement of this manuscript.

CONFLICT OF INTEREST

The authors announced that there are no matches of concern about the publication of this manuscript.

Ethical of approval

This study follows the ethics guidelines of the College of Veterinary Medicine, University of Mosul 2022

REFERENCES

- Abdullah, S.H. (2013): Prevalence of blood parasites in local chickens in Qaradagh district, Sulaimani Iraq. The Iraqi J Vet Med, 37: 17–21. DOI: https://doi.org/10.30539/iraqijvm.v37i1.321.
- Adamu, A. (2017): Epidemiology of haemoparasite infection and their effects on hematological parameters in scavenging chickens of west Gojjam administrative zone, Amhara region, Ethiopia. MSc. Thesis. University of Gondar, Ethiopia. 71 p.
- Aiyedun, J.O.; Elagroudi, M.G.; OhFadunsin, S.D.; Nuhu, N.M.; Sanda, M.I.; Hussain, K.; Ganiyu, I.A. and Rabiu, M. (2022): Cross-species Prevalence and Risk Factors Associated with Avian Haemoparasitic Infections in Kwara Central, Nigeria. Zag Vet J, Volume 50, Number 2, p. 116-125, DOI: 10.21608/zvjz.2022.130463.1177.
- Al-Kaabi, H.K.J. (2021): Study of blood picture and some serum proteins in local chickens and domestic pigeon naturally infected by soft ticks. Al-Kufa University Journal for Biology. vol.13 (3): 26-31.DOI:10.36320 /ajb/v13.i3.8213.
- Al-Rubaie, H.M.A. and Ibrahim, R.M. (2020): Molecular Detection of Avian Malaria (Plasmodium gallinaceum) in Local Domesticated Breed Chickens (Gallus gallus domesticus) in Baghdad. Iraqi J. Vet. Med. Vol. 44(E0): 75-79. DOI: https://doi.org/10.30539/ijvm.v44i (E0). 1025.
- Andreína, M. and Ananias, A. (2011): Haemosporidian infection in captive

- masked bobwhite quail (Colinus virginianus ridgwayi), an endangered subspecies of the northern bobwhite quail. J. Vet. Parasitol.; (182): 113-120.DOI: 10.1016/j.vetpar.2011.06.006
- Chasar, A.; Loiseau, C.; Valkiūnas, G.; Iezhova, T. and Smith, T.B. (2009):
 Prevalence and diversity patterns of avian blood parasites in degraded African rainforest habitats. J. Mol. Ecol., (18): 4121-4133. DOI: 10.1111/j.1365-294X.2009.04346.x
- Coles, E.H. Lea Kocytes. (1986): In: Veterinary Clinical pathology.4 th edition. W.B.Saunders Company Philadelphia, London, Toronto, Mexico City, Rio de Janeiro, Sydney, Tokyo, llong Kong, pp.379-384.435.
- Dey, A.R.; Begum, N.; Anisuzzaman, M.; Khan, A.H.N.A.; Mondal, M.M.H. (2008): Haemoprotozoan infection in ducks: Prevalence and pathology. Bangladesh J. Vet. Med. 2008, 6, 53–58. DOI: 10.3329/bjvm. v6i1.1339
- Fatima A. and Maqboo A. (2014): Haemoproteus in wild and domestic birds. Sci. Int.; 26(1): 321–323. DOI: 0.37506/v20/i1/2020/mlu/194434
- Fecchio, A.; Lima, M.R.; Silveira, P.; Braga, É.M. and Marini, M.Â. (2011): High prevalence of blood parasites in social birds from a neotropical savanna in Brazil. Emu-Austral Ornithol. 111:132–138. DOI: 10.1071/MU10063.
- Friend, M. and Franson, J. (1999): Field Manual of Wildlife Diseases: General Field Procedures and Diseases of Birds. http://www.nwhc.usgs.gov/publications/ field manual.
- Garcia-Longoria, L.; Marzal, A.; De Lope, F. and Garamszegi, L. (2019): Host-parasite interaction explains variation in the prevalence of avian haemosporidians at the community level. PloS One 14, e0205624.
- Gimba, F.I.; Zakaria, A.; Mugok, L.B.; Siong, H.C.; Jaafar, N.; Moktar,

- M.A.; Rahman, A.R.R.; Amzah, A.; Abu, J.; Sani, R.A.; Amin-Babjee, S.M. and Sharma, R.S.K. (2014): Haemoparasites of domestic poultry and wild birds in Selangor, Malaysia. Malaysian J Vet Res, 5: 43–51.
- Gutierrez-Lopez, R.; Gangoso, L.; Martinez-de la Puente, J.; Fric, J.; Lopez-Lopez, P.; Mailleux, M.; Munoz, J.; Touati, L.; Samraoui, B. and Figuerola, J. (2015): Low prevalence of blood parasites in a long-distance migratory raptor: the importance of host habitat. Parasites Vectors 8, 189.
- Hasson, RH. (2015): Haemosporidians parasites of Gallus domesticus, poultry in Iraq. Int. J. Adv Res.; 3(8): 1046-1054.
- Mirzaei, F.; Siyadatpanah, A.; Norouzi, R.; Pournasir, S.; Nissapatorn, V. and Pereira, M.L. (2020): Blood Parasites in Domestic Birds in Central Iran.Vet Sci.; 7(3): 126. doi: 10.3390/vetsci7030126.
- Nagvi, M.A.U.H.; Khan, M.K.; Igbal, Z.; Rizwan, H.M.; Khan, M.N.; Naqvi, S.Z.; Zafar, A.; Abbas, R.Z. and Abbas, A. (2017): Prevalence and associated risk factors haemoparasites, and their effects on hematological profile domesticated chickens in District Layyah, Punjab, Pakistan. Prev. Vet. Med. 143, 49-53. DOI: 10.1016/j.prevetmed.2017.05.001
- Niazi, A.D. (2001): Statistical analysis in medical research Uni. Nahrein Republic of Iraq. 148.
- Njuga, G.R. (2000): Ecto- and Haemoparasites of Chickens in Malawi with Emphasis on the Effects of the Chicken Louse, Menacanthus cornutus. M.Sc. Thesis, University of Malawi, Lilongwe.
- Nnadi, P.A. and George, S.O. (2010): A Cross-Sectional Survey on Parasites of Chickens in Selected Villages in the Subhumid Zones of South-

- Eastern Nigeria. Journal of Parasitological Research, ID: 141824. DOI: 10.1155/2010/141824
- Ogbaje, C.I.; Okpe, J.A. and Oke, P. (2019): Haemoparasites and Hematological parameters of Nigerian indigenous (local) and exotic(broiler) chickens slaughtered in Makurdi major markets, Benue State, Nigeria. Alex. J. Vet. Sci, 63, 90–96. DOI: 10.5455/ajvs.53637.
- Okereke, R.E. (2010): Occurrence of Blood Parasites in Local and Exotic Chickens Slaughtered in Owerri, Imo State. B-Agri-Technology Project Report Submitted to the Department of Animal Science and Technology, Federal University of Technology, Owerri.
- Opara, M.N. and Nwokedi, C.C. (2011):
 Occurrence of Haemoparasites
 among Small Ruminants Reared
 under Traditional Husbandry System
 in Owerri, Southeast Nigeria.
 Bulletin of Animal Health and
 Production in Africa, 59, 393-398.
- Opara, M.N.; Osowa, D.K. and Maxwell, J.A. (2014): Blood and Gastrointestinal Parasites of Chickens and Turkeys Reared in the Tropical Rainforest Zone of Southeastern Nigeria. Open Journal of Veterinary Medicine, 4, 308-313 http://dx.doi.org/10.4236/ojvm.2014.41 2037
- Permin, A. and Hanson, J.W. (1998): Epidemiology, Diagnosis and Control of Poultry Parasites. FAO Animal Health Manuals 4 Rome, Food and Agriculture Organization of the United Nation.
- Poulsen, J.; Permin, A.; Hindsho, O.; Yelifari, L.; Nansen, P. and Bloch, P. (2000): Prevalence and Distribution of Gastrointestinal Helminthes and Haemoparasites in Young Scavenging Chickens in Upper Eastern Region of Ghana West Africa. Preventive Veterinary Medicine,45,237-245.http://dx.doi.org/10.1016/S0167-5877(00)00125-2.

- Radfar, M.H.; Khedri, J.; Adinehbeigi, K.; Nabavi, R. and Rahmani, K. (2012):
 Prevalence of parasites and associated risk factors in domestic pigeons (Columba livia domestica) and free-range backyard chickens of Sistan region, east of Iran. J Parasit Dis, 36: 220–225., doi: 10.1007/s12639-012-0112-5.
- Rivero de Aguilar, J.; Castillo, F.; Moreno, A.; Penafiel, N.; Browne, L.; Walter, S.T.; Karubian, J. and Bonaccorso, E. (2018): Patterns of avian haemosporidian infections vary with time, but not habitat, in a fragmented Neotropical landscape. PloS One 13, e0206493.
- Saif, Y.M.; Fadly, A.M.; Glisson, J.R.; McDougald, L.R.; Nolan, L.K. and Swayne, D.E. (2008): Diseases of Poultry (12th ed). Wiley-Blackwell. 1352pp.
- Sebaio, F.; Braga, É.M.; Branquinho, F.; Fecchio, A. and Marini, M.Â. (2012): Blood parasites in passerine birds from the Brazilian Atlantic Forest. Rev. Bras. Parasitol. Veterinária; 21: 7–15. doi: 10.1590/S1984-2961201200010 0003.
- Taylor, MA.; Coop RL. and Wall RL. (2015): Veterinary parasitology. 4th ed. Wiley-Blackwell, Singapore: Wiley-Blackwell.
- Urquhart, G.M.; Armour, J.; Duncan, J.L.; Dunn, A.M. and Jennings, F.W. (1996): Veterinary Parasitology. 2nd Edition, Blackwell Science Ltd., Oxford, 224-234.
- Valkiunas G. (2005): Avian malaria parasites and other haemosporida. London. Boca Raton, Florida: CRC Press;. p 936.
- Valkiunas, G.; Sehgal, R.N.; Iezhova, T.A. and Smith, T.B. (2005): Further Observations on the Blood Parasites of Birds in Uganda. Journal of Wildlife Diseases, 41, 580-587. http://dx.doi.org/10.7589/0090-3558-41.3.580.

الكشف عن الأوالي الدموية في الديوك الرومية والدجاج منال حسن ، ايناس سعدى حسين ، وعد البغياني

Email: enassaddi@uomosul.edu.iq Assiut University web-site: www.aun.edu.eg

الطفيليات الدموية هي طفيليات دموية أولية توجد في كريات الدم الحمراء داخل الخلية وخارج الخلية بالإضافة الى للاسحة المحلوم وتشخيص طفيليات بالسحة الحالية الى الكشف وتشخيص طفيليات للعروم تهدف الدراسة الحالية الى الكشف وتشخيص طفيليات المحلي والديوك الرومية التي تم فحصها في مدينة الموصل ومدينة أربيل. ولتحقيق هذا الهدف من الدراسة التي اجريت للفترة من أكتوبر 2021 حتى مارس 2022 من خلال فحص مسحات الدم للدجاج والديك الرومي والتي اظهرت ارتفاع معدلات الإصابة بطفيليات الدم في شهر مارس يصل الى 100 %. من 120 عينة دم تم جمعها من الدجاج المحلي والديوك الرومية في مناطق مختلفة من مدينتي الموصل وأربيل, بلغت معدلات الاصابة بتطفل الدم بالأوالي الدموية في كلا المنطقتين في الدجاج المنزلي (Gallus الموصل وأربيل, بلغت معدلات الاصابة بتطفل الدم بالأوالي الدموية في كلا المنطقتين في الديوك الرومية حوالي 40% ((20/12)) . أظهرت النتائج ارتفاع نسبة الانتشار عالية في ذكور الدجاج (6.25%) ، بينما سجلت في الديوك الرومية نسبة انتشار عالية في ذكور الدجاج (6.25%) ، بينما سجلت في الديوك الرومية مسحات الدم التي تم فحصها في الدجاج فقط.

Leucocytozoon spp., Plasmodium spp., Haemoproteus spp. Trypanosoma الكلمات العقاحية: spp.