

Effects of Glucans and Nucleotides on the Immunity, Health, and Growth of Fish. A Review with Special Reference to the Egyptian Situation

Hanan A. Ghetas^{1*}, Ahmed Arafa¹, Alaa Eldin Eissa², and Mohamed A. Khallaf¹

- (1) Department of Aquatic Animals Medicine and Management, Faculty of Veterinary Medicine, University of Sadat City, Sadat City 32897, Egypt.
- (2) Department of Aquatic Animal Medicine and Management, Faculty of Veterinary Medicine, Cairo University, Giza 12211 Egypt.
- *Corresponding author: <u>hanan.ghetas@vet.usc.edu.eg</u> Received: 12/12/2024 Accepted: 29/12/2024

ABSTRACT

β-glucan and nucleotides have beneficial impacts on aquaculture species' health and growth. Studies on fish dietary nucleotides have revealed that may alter the structure of the intestines, increase resistance to stress, enhance the quality of the larvae by fortifying the brood supply, and promote growth. In the early phases of development and modify both innate and adaptive immunity. Nucleotides and β glucan supplementation have demonstrated very constant positive effects on a variety of fish species, despite sporadic variability in physiological responses. Despite the fact that research on fish nutrition based on nucleotides is just getting started, and there are a lot of fundamental questions have not yet been addressed, observations to date have supported the idea that fish require nucleotides as contingent or semi-essential nutrients, and that further study is required to improve fish culture by supplementing the diet with nucleotides. Scientists have demonstrated that β -glucan can alter some significant immunological (lysozyme, phagocytic, oxidative burst and phenoloxidase activities) and biochemical (serum hemoglobin, serum protein, and total hemocyte count) characteristics, giving fish and aquatic organisms a more effective immune profile. These beneficial effects are thought to be caused by the supply of physiologically necessary amounts of nucleotides by diet because some tissues have a restricted capacity for synthesis (e.g. lymphoid). Fish feed on diet supplemented with β -glucan exhibited decreased susceptibility to genes linked to acute inflammatory reactions. Research has demonstrated that fish's immune defense is enhanced by β-glucan of fish and other aquatic creatures, strengthening their resilience to disease by boosting beneficial responses and lowering harmful ones.

Key words: β-glucan, nucleotides, Immune response, Fish, Immunostimulant

INTRODUCTION

Natural or synthetic substances known as immunostimulants are used to alter

and strengthen an organism's immune response to diseases that usually come via disease-causing agents (Manoppo et

al.. 2015). These days immunestimulating agents are frequently used since they are inexpensive, safe, and cause minimal environmental contamination. Extracts from plants, microbial byproducts, and polysaccharides are examples of which immunostimulants are incorporated into the diets of aquaculture creatures. To safeguard fish from contagious diseases, it is essential to make choices about the most appropriate and potent immune-stimulating agent. One of the well-researched immunostimulants, β -glucans, is frequently utilized to enhance animal and human health (Fusté et al., 2019). Aquaculture is growing in most emerging countries to fulfill the yearly increase of need for aquatic food products. However, it is well known that one of the major obstacles that fish farmers encounter is the occurrence of disease outbreaks brought on by harmful organisms. Aquaculture is expected to rise by 62% between 2010 and 2030, meeting more than two-thirds of worldwide demand for fishes and crustaceans and Mollusca, as a result of changing eating habits and population expansion (Verdegem et al., 2023). A significant barrier is the prevalence of aquatic animal diseases, which are made worse by international trade, system intensification, and climate change (Singh and colleagues, 2023). Farmers are forced to use antibiotics and disinfectants due to the ease with which pathogen evolution and disease outbreaks are facilitated by high-density cultures and intensified systems (Li et al., 2024). However, excessive use of these substances weakens animal immune systems and encourages the growth of microorganisms resistant to

antibiotics, endangering human health worldwide (Devadas et al., 2024)

Furthermore, the usage of antibiotics has led to major issues, including the degradation of aquatic ecosystems and aquatic environments, the lingering presence of residues of antibiotics in fish meat, as well as the detrimental effects on human health. (Sumon et al., 2022). Alternative methods disease of prevention. like as vaccination and functional feed supplements that have been suggested in this area (Noman et al., 2024). Functional feed supplements, immunostimulants including and probiotics, have gained traction for their potential to enhance fish performance, immune systems, and feed utilization (Noman et al., 2024). Incorporating such supplements into fish diets could not only improve disease prevention but also contribute to better resource utilization, fostering sustainable aquaculture development (Mathan et al., 2024). The primary emphasis of this review is effect of dietary usage of β -glucan and nucleotides on the immunity, health and growth of fish and other aquatic species and current data on the applications in advanced aquaculture and the investigations that based on using of β glucan and nucleotides in Egypt in different species of fish.

The origins and properties of β-glucans

The main structural components of the cell wall the β -glucans that store carbohydrates and provide defense against invasive infections. They can be present in cereal grains like barley and oats. fungi such as *Pneumocystis carini* and *Agaricus subrufesuns*, and baker yeast, *Saccharomyces cerevisiae*(Shah et al., 2017). Additional sources of β -glucans include certain bacteria species that belong to the Rhizobiaceae family and edible mushrooms like shiitake,

lingzhi, and oyster mushrooms (Khan et al., 2017). The content of β -glucan varies based on the source. The amount of β -glucan in barley and oats and varies around 2% to 8%, but it makes up 50% to 60% of the entire polysaccharides found in the cell wall of the yeast (Nishantha et al., 2018). Additionally, the amount of β -glucan in various species of mushroom ranges from 8.608 g to 60.788 g/100 g Mass that is dry; wild-growing mushroom species such as bracket fungus have a larger amount of β -glucan (Sari et al., 2017).

 β -glucans' size, shape, and solubility can influence their activity all and, consequently, their immunomodulatory effects. As indicated by Suchecka et al. (2016), Higher molecular weight β glucans (like zymosan) are believed to have more significant cellular effects, such as activating leucocyte cells, the activity of phagocytes, antimicrobial properties, action of antioxidants, and activity of respiratory bursts. Conversely, β -glucans having a smaller molecular weight, such as glucan phosphate, have less influence on cells.

<u>Mechanism of action of β-glucans</u>

Both innate and acquired immune activity activation is significantly influenced by β -glucan. In addition to acting on invasive pathogens, b-glucanstimulated innate immune responses also support the activity and function of developed immunity (Sakai, 1999).

 β -glucans are responsible for several functions that fortify and safeguard the immune system and provide the best defense against possible health risks due to their ability to directly bind and activate macrophage cells and other white blood cells such as the cells of neutrophils and natural killer (NK) cells (Herre et al., 2004).

Numerous vertebrate and invertebrate organisms have been the focus of a lot of study. On β-glucan as an immunostimulant; Soltanian et al. (2009) provided great details about this topic. When β -glucan was given to Atlantic salmon (Salmon salar) through diet, the activity of respiratory burst was stimulated. According to several studies, administration of β -glucan through diet temporarily boosts the ability to withstand infection in gilthead sea bream, (Ortuno et al., 2002), Clarias gariepinus (Yoshida et al., 1995), Catla catla, Rohu (Sahoo and Mukherjee, 2002), and Oncorhynchus tshawytscha (Nikl et al., 1993).

Resistance to the microsporidian developed by rainbow trout Salmonae Loma is significantly influenced by the timing of b-glucan administration (Guselle et al., 2007). Lysozyme activity, HSP concentrations of gill, and liver were increased at thirty days, However, sea bass fed glucans and alginic acid at 15 days showed a significant increase in serum complement activity. According to Bagni et al. (2005), who recorded the effect of dietary glucan that is depending on duration. Only fish fed the Ergosan diet showed increased complement activity. The parameters of innate and specific immunity, survival rate. performance of growth, the conversion index in fish under treatment and fish under control did not change substantially over time. A key factor in boosting immune responses is the purity of the immunostimulant agent (β -glucan) that is provided.

Usage of β glucan as an adjuvant (substances that enhance immune response) and/or immunostimulant (biologically active compounds) to strengthen the fish immune system has

grown (Filho et al., 2019). As stated by Bagni et al. (2005), β -glucan, which is from yeasts, derived mushrooms, seaweeds, and cereal plants, is the most beneficial bioactive component utilized aquaculture. Scientists in have demonstrated that β -glucans mainly strengthen fish's non-specific defenses, making them more resistant to infectious diseases. Fish have also shown signs of evolving specific defense, according to several research (Siwicki et al., 2004).

The immunological functions (lysozyme, complement, phagocytic, and serum bactericidal activity) of rohu (Labeo rohita) fed a diet containing 250 mg of β -glucan/kg peaked after 42 days of feeding, according to Misra et al. (2006), while the indices began to increase on day 28 of the trial. The experimental diet also increased phagocytic activity and lymphocyte proliferation. Chang et al. (2013) found that (*Epinephelus coioides*) exhibited significantly increased the lysozyme activity and complement activity against Vibrio alginolyticus when add in its diet mushroom β -glucan at 1 g and 2 g per kg diet. Beginning on day six and continuing until day thirty, the activities increased. According to Kaisa et al. (2018), the microbiota of the intestine plays essential function in preserving the shape of the intestine and barrier function; disturbances may result in inflammatory reactions (Przewł'ocka et al., 2020).

Prior studies have shown that inclusion of β -glucan in the diets of fish can affect gut microbiota's makeup. The composition of microbiota Solea senegalensis was altered by feeding βglucan, for instance, with a reduction in intestinal proportions the Vibrio (Carballo et al., 2019). Likewise, when sea bass were fed diets enriched with β the prevalence of glucan, the dysgonomonas species within the Bacteroidetes phylum was significantly lower. (Carda-Di'eguez et al., 2014). In a different study, β -glucan inclusion in the diet was associated with a reduction in the species richness and OTU count of the carp microbiota (Kühlwein et al., 2013). All these results are consistent with the findings of our ongoing study.

T1	Descent	T *	F ' I '		
The species of	Dose used	1 ime of	Findings	Challenge with pathogen	The References
fish		experiment			
Oreochromis	Dreochromis 0.5 g/kg One month		Lysozyme activity \uparrow , increasing	Deltamethrin	Dawood and others
niloticus			phagocytic activity and		(2020)
			phagocytic index and interleukin		
			8↑,		
			No change in Interleukin 1 beta		
Oreochromis	0.5 g/kg	2 months	Lysozyme activity ↑, increasing	Crowding stress	Dawood,
niloticus			phagocytic activity and		Metwally,
			phagocytic index and Tumor		et al.
			necrosis factor-alpha Interleukin		(2020)
			1 beta ↑,		
Oreochromis	1 BG and 0.5 BG g kg	60 days	Enhanced Immunoglobulin M,	Effect on Growth rate,	Dawood, Eweedah
niloticus	and 1 ASP, or 0.5 ASP		lysozyme, antioxidant enzymes,	Oxidative response and	et al.,
			bactericidal, and phagocytosis	Immune Response	(2020)
			which indicated improved		
			immunity		
Oreochromis	50 mg/kg)	12 weeks	hematocrit values increased,	Genetically Improved	Dawood, Magouz
niloticus			hemoglobin, and white blood	Farmed Tilapia	et al.,
			vessels, expression of genes and		(2020)
			growth rate		

Research on β-glucan's effects in Egypt

Oreochromis 0.5 g/kg 2 weeks		Lymphocytes↑, ↑ Atrazine,		Neamat-Allah	
niloticus			Immunoglobulin M Lysozymes [†] ,	Aeromonas	et al.
			Nitric oxide↑, Interleukin-8↔	sobria	(2020)
Oreochromis	Oreochromis 5 g/kg 2 weeks niloticus		Phagocytic activity [↑] , Phagocytic Aeromonas		Sherif and
niloticus			index [↑] , Respiratory burst	hydrophila	Mahfouz
			activity [†] , Tumor necrosis factor-		(2019)
			alpha↑		
Oreochromis	0.5 & 1 g/kg	Eighty-four days	<i>↑Immunoglobulin M</i> , serum	Aeromonas	Hosseny et al.
niloticus			Nitric oxide↔,	hydrophila	(2018)
			Lysozyme activity increased \leftrightarrow		
Oreochromis	2 g/kg	21 days	Interleukin-8 \leftrightarrow ↑, <i>Tumor</i>	Streptococcus iniae	Salah et al.
niloticus			necrosis factor-alpha ↑,		(2017)
			<i>Toll-like receptor</i> \uparrow , and		
			↑Immunoglobulin M		
Oreochromis	1 g/kg	Twenty-one days	Respiratory burst activity \uparrow ,	Aeromonas	El-Boshy et al.
niloticus			Phagocytic activity	hydrophila	(2010)
			\uparrow ,Bactericidal activity \uparrow ,		
			Lysozyme activity↑, Nitric		
			oxide↑		
Oreochromis	β-glucans + mannan oligosaccharides		killing \uparrow , Phagocytic activity \uparrow , \uparrow , Lysozyme activity \uparrow , Nitric		Selim & Reda,
niloticus			oxide ^{↑↑} , Lymphocytes ↑, Survival [↑] against Yersinia ruckeri		(2015)
Pagrus major	β-glucan + Vitamin C		<i>Respiratory burst activity</i> ↑, ↑Lysozyme Alternative complement		Dawood et al.
			activity \uparrow , Peroxidase activity \uparrow , Bactericidal activity \uparrow		(2017)
Pagrus major	β-glucan + <i>Lactobacillus plantarum</i> (heat-killed)		Bactericidal activity \uparrow , Alternative complement activity \uparrow ,		Dawood et al.
			Lysozyme activity [↑] , Peroxidase ac	(2015)	

Increasing (\uparrow), decreasing (\downarrow), and no significant changes were seen in the experimental fish when compared to the controls.

Nucleotides: Structure

Nucleotides are important metabolites which are significant for structural, metabolic, energetic, and regulatory processes (Rudolph, 1994). According to several studies, dietary nucleotide supplementation improved several physiological processes in numerous species, particularly in the early stages of life (Gil, 2002). It has been suggested that during periods of rapid development or high metabolism, particular organs, such as cells of immune system and cells of gastrointestinal system, may require more nucleotides than others.

Nucleotides (NT) are low molecular weight compounds composed of a nitrogenous base, one to three phosphate groups, and a pentose sugar. (Cosgrove, 1998). The nucleobases are categorized as either pyrimidines, which include cytosine (C), thymine (T), and uracil (U), or purines, which include adenine (A) and guanine (G). Ribose and deoxyribose are examples of pentose sugars. A nucleoside (NS) is made up of a pentose sugar connected to a purine or pyrimidine base, whereas an NT is a phosphate ester of NS (Li and Gatlin, 2006). NTs are the structural units used in the synthesis of DNA and RNA.

<u>The impact of dietary NTs on</u> performance of growth rate and the gastrointestinal tract

NT inclusion in diets is crucial for the establishment of healthy gastrointestinal (GI) tract functions in various species. For instance, the proximal, mid, and distal intestines of Commercially NT additions are added to the diet of Atlantic salmon. The mean fold heights of the fish fed the control diet were substantially lower than those of the fish fed ("Optimum" 0.03% of the overall feed). This could have resulted in a larger surface area of the gut (Burrells et al., 2001). Additionally, NT inclusion in diets may have an impact on the flora that lives in the gastrointestinal tract. After weaning pigs exposed to stresses,

Sauer et al. (2010) found that NT additions in diets could alter the composition of gastrointestinal microbiota (Moore et al., 2011). According to the authors, dietary NTs viable may be a substitute for antimicrobial growth enhancers in piglets. Dietary NTs have an impact on the performance of growth of some species some species particularly during the early stages of growth, like in Atlantic salmon. (Burrells et al., 2001), Oreochromis niloticus (Ramadan et al., 1994), rainbow trout (Mohebbi et al., 2013), grouper (Lin et al., 2009), Pacific white shrimp (Li et al., 2007), weanling pigs (Li et al., 2015) and mice (Xu et al., 2013). Even though nucleotide supplementation previously was demonstrated to enhance weaned rats' development on low-protein diets (Gyo"rgy, 1971), it has been accepted that under typical circumstances, de novo nucleotide synthesis is sufficient to sustain the growth rate. (Cosgrove, 1998). An outside source of nucleotides may stimulate fish growth and crustaceans to achieve their high rate of cell reproduction in their early phases, according to Borda et al. (2003), who application evaluated data on of nucleotides in the diets of to sea bream larvae. Additionally, Person-Le Ruyet et al. (1983) found that following 55-day feeding study, turbot larvae (around 100 mg/fish) fed supplemented diet with inosine- (1.3% of diet for 6 days, 0.13% for 45 days) showed considerably improve the growth and survival rate. According to their later research, giving turbot larvae diet contained 0.77% inosine for 10 or 20 days also markedly boosted their weight gain (approximate beginning weight of 230 mg/fish). According to Me'tailler et al. (1983), the enhancement of growth of inosine was

thought to be caused by increased feed consumption at the beginning of weaning, which encouraged quicker consumption of food and reduced waterborne nutrient leaching, or it might have had a role in metabolism. Along with inosine, combinations of nucleotides and metabolites (such Ascogen R. Chemoforma Co., Basal, Switzerland) have additionally been demonstrated to occasionally increase growth in fish, including young rainbow trout (Adamek et al., 1996) and tilapia larvae (Ramadan and Atef, 1991). On the other hand, most fish that are young or subadults seem to have a negligible growth-enhancing impact (Li et al., 2004).

The effect of dietary NTs on immune function

Dietary NT may be necessary to maintain adequate immunological function. (Carver et al., 1990). Li et al. (2005) investigated that Growth, stress tolerance (measured by plasma cortisol after confinement), and in situ challenge by co-habitation with A. ocellatum in juvenile red drums were not significantly impacted by the dietary addition of 0.2% NT (Optimum). The potential impact of pure NT mixes on juvenile red drums was also examined by Li et al. (2007). They discovered a brief growthenhancing effect that lasted for the first week before disappearing. However, the immune system function was improved by dietary supplementation of 0.5 or 1% NT (Ascogen), as indicated by the secretion of superoxide anion by head kidney macrophages, intestinal fold, and microvillus height in the same fish species. (Cheng et al., 2011). Burrells et al. (2001) found that adding dietary NT Optimum) increased (0.03%,the survival rates of rainbow trout after challenging the infectious salmon anemia virus and V. anguillarum.

According to Leonardi et al. (2003), dietary NT (Optimum) enhanced the number of B lymphocytes and decreased the release of plasma cortisol. It also improved the cells' resistance to the infectious pancreatic necrosis virus. Tahmasebi-Kohyani et al. (2012) recently discovered that feeding fingerling rainbow trout 0.15-0.2% NT supplementation improved their performance in handling and crowding stress (plasma cortisol, glucose, and ion concentration), immune function, lysozyme activity, and IgM level), as well as their resistance to *S. iniae*.

		Period of			
Nucleotide used	Dose incorporated in	administratio	Species	Results	Authors
	diet	n			
	(kg-1 diet)				
Ascogen	2 and 5 g	4 months	Hybrid tilapia	↑growth and survival	Ramadan and Atef (1991)
Ascogen	5 g	4 months	Hybrid tilapia	↑ antibody titer after vaccination and mitogenic response of lymphocyte	Ramadan et al., (1994)
nucleotides and/or beta-glucan	(200 mg/kg diet)		Broiler chicken	Improving parameters growth of, morphology of intestine, and a number of biochemical parameters of broiler chicken	Aya et al., (2024)
Nucleotides	0.05%, 0.15%, and 0.25% NTs	30 days	Nile tilapia	improved cytokine gene expression, leukocyte cells, antioxidant activity, non-specific immunity, and resistance to disease in addition to blood proteins.	Reda et al., (2018)
nucleotide	0.5, 1.5 and 2.5 g	60 days	Nile tilapia	highest growth performance, as well as the expression of the genes for Ghrelin, insulin-like growth factor, and growth hormone	Selim et al. 2020
a commercial nucleotide Nucleoforce Fish™	250mg/kg or 500mg/kg	150 days	Gilthead seabr eam	Enhancement of the mitochondrial enzyme in the liver activity , state of antioxidants, expression of immune genes, and gut microbial ecology	El-Nokrashy Et al., (2021)

Studies on the immunostimulatory effects of Nucleotides in Egypt

The consequences of combiningnucleotidesandβ-glucanonaquaculture

It has been demonstrated that using two or more supplements together has a synergistic effect that produces better results than using them separately (Hany et al., 2024). According to earlier studies, pigs' growth performance might be enhanced by applying nucleotides and β -glucan together (Huang et al., 2023).

Nucleotides and β -glucan have prebiotic anti-inflammatory benefits and by strengthening fish's resilience to stress and boosting their immune systems. 2021).Numerous (Pogue et al., physiological and metabolic processes depend on nucleotides. (Krüger and van der Werf, 2018). In periods of rapid growth or when certain disorders are present. endogenously produced nucleotides may not be enough to meet the needs of tissues and cells with high metabolic activity, even if they can typically meet the needs of a normal metabolism. (Norton et al., 2001). As a result, more nucleotides are necessary to promote the best possible cellular growth and function. It has been demonstrated that dietary nucleotides improve the growth of fish and utilization of feed. (Liu, 2016), immune response and stress resistance (Reda et al., 2018), health of the intestine (Cheng et al., 2011), particularly the gut microbiota's homeostasis (El-Nokrashy et al., 2021), and quality of flesh (Tie, 2018). Numerous natural sources contain the chemically diverse and highly conserved carbohydrate known as β-Several research glucan. have documented more beneficial effects for fish due to of β -glucan addition, such as improving health of intestine (Carballo et al., 2019), enhancing immunological response (Ching et al., 2021), increasing stress tolerance (Salah et al., 2017), and promoting growth (Song et al., 2020). Furthermore, it has been discovered that dietary glucan enhances the quality of pig meat by raising intramuscular fat content and muscle pH, decreasing muscle drip losses, and altering the ratio of saturated to unsaturated fatty acids. (Luo et al., 2019), However, it is unclear how β -glucan supplementation affects the quality of fish meat. Fish flesh

quality is determined by several physicochemical factors as pH, waterholding capacity, and the texture of the flesh (Jiang et al., 2016). Previous research has demonstrated that supplements containing β -glucan and nucleotides can both enhance the quality of flesh. For example, it has been discovered that feeding broilers β -glucan improves the quality of their flesh. (Cho et al., 2013) and pigs (Luo et al., 2019) by lowering the amount of cooking loss. In a similar way it has been observed that dietary nucleotides enhance the quality of grass carp flesh by preventing the decrease in muscle pH and minimizing muscle cooking loss. (Tie, 2018). As a strong adjuvant, β -glucan successfully elicit can host immunological responses, both innate adaptive. (Qi et and al.. 2011). Additionally, it can lessen the inflammatory response by inhibiting TLRs and lowering the synthesis and release of proinflammatory cytokines such as IL-1 β and tumor necrosis factor- α (TNF- α). (Kankkunen et al., 2010). Nucleotides' immunomodulatory qualities have also been demonstrated in fish, as demonstrated by their capacity to inflammation reduce intestinal in zebrafish when taken as supplements. (Guo et al., 2017), as well as suppress the grass carp's TNF-α mRNA expression level. (Tie et al., 2021).

CONCLUSIONS AND PERSPECTIVES

It is widely acknowledged that the use of immunostimulants in fish trials results in positive outcomes like protection against diseases because they boost humoral and cellular responses. However, care must be given regarding problems like tolerance, undesirable side effects like immunosuppression from using excessive amounts of immunostimulants,

or undesirable effects brought on by long-term usage of such substances. The use of β -glucan and nucleotides in aquaculture may improve financial outcomes and lower production costs (volume of investment). The cost-benefit analysis of these feed additives needs to be conducted for other commercially significant marine species as well as in a broader perspective across the production cycle. Combined application of nucleotides and β-glucan might advantages provide more than supplementing with β-glucan or nucleotides alone. It is hoped that several difficulties, focusing especially the immunological response on following binding polarization of immunostimulants to receptors, would become public knowledge following the development of proteomic and genomic methods for several fish species.

REFERENCES

- Adamek, Z., Hamackova, J., Kouril, J., Vachta, R.andStibranyiova, I. (1996). Effect of Ascogen probiotics supplementation on farming success in rainbow trout (*Oncorhynchus mykiss*) and wels (*Silurusglanis*) under conditions of intensive culture. Krmiva, 38, 11–20.
- Aya, E., Shabaan, H., Fatma, E., Samar, B. and Olla, K. (2024). Analysis of the liver transcriptome in broiler chicken fed with dietary nucleotides and/or beta-glucan revealed enhancement in growth parameters, intestinal morphology, and some biochemical parameters. J. Adv. Vet. Res. 14, 4.
- Bagni, M., Romano, N., Finola, M.G.,Abelli, L., Scapigliati, G., Tiscar,P.G., Sarti, M.and Marino, G.(2005). Short- and long-term effects

of a dietary yeast β -glucan (Macrogard) and alginic acid (Ergosan) preparation on immune response in sea bass (*Dicentrarchus labrax*). Fish & Shellfish Immunology, 18, 311–325.

- Borda, E., Martinez-Puig, D. and Cordoba, X. (2003). A balanced nucleotide supply makes sense. Feed Mix, 11, 24–26.
- Burrells, C., Williams, P. and Forno, P. (2001). Dietary nucleotides: A novel supplement in fish feeds. Effects on resistance to disease in salmonids. Aquaculture, 199(1-2), 159–169.
- Carballo, C., Pinto, P.I., Mateus, A.P., Berbel, C., Guerreiro, C.C., Martinez-Blanch, J.F., Codo ner, F.M., Mantecon, L., Power, D.M. and Manchado, M. (2019). Yeast βglucans and microalgal extracts modulate the immune response and gut microbiome in Senegalese sole (*Solea senegalensis*). Fish. Shellfish Immunol. 92, 31–39.
- Carda-Diéguez, M., Mira, A.and Fouz,
 B. (2014). Pyrosequencing survey of intestinal microbiota diversity in cultured sea bass (*Dicentrarchus labrax*) fed functional diets. FEMS Microbiology Ecology, 87, 451–459.
- Carver, J.D., Cox, W.I. and Barness, L.A. (1990). Dietary nucleotide effects upon murine natural killer cell activity and macrophage activation. Journal of Parenteral and Enteral Nutrition, 14, 18–22.
- Chang, C.S., Huang, S.L., Chen, S. and Chen, S.N. (2013). Innate immune responses and efficacy of using mushroom beta-glucan mixture (MBG) on orange-spotted grouper, *Epinephelus coioides*, aquaculture. Fish & Shellfish Immunology, 35(1), 115–125.

- Cheng, Z., Buentello, A. and Gatlin, D. M.III. (2011). Dietary nucleotides influence immune responses and intestinal morphology of red drum (*Sciaenops ocellatus*). Fish & Shellfish Immunology, 30(1), 143– 147.
- Ching, J.J., Shuib, A.S., Abdul Majid, N. and Taufek, N. (2021). Immunomo dulatory activity β glucans in fish: Relationship betwee n β glucan administration parameters an d immune response induced. Aquac ulture Research, 52, 1824–1845.
- Cho, J., Zhang, Z. and Kim, I. (2013). Effects of single or combined dietary supplementation of β-glucan and kefir on growth performance, blood characteristics and meat quality in broilers. Br. Poult. Sci. 54, 216–221.
- Cosgrove, M. (1998). Nucleotides. Nutrition, 14(10), 748–751.
- Devadas, S., Zakaria, Z., Shariff, M., Bhassu, S., Karim, M.andNatrah, I. (2024). Methodologies and standards for monitoring antimicrobial use and antimicrobial resistance in shrimp aquaculture. Aquaculture, 579, 740216.
- Dawood, M.A. O., Abdo, S.E., Gewaily, M.S., Moustafa, E.M., SaadAllah, M.S., AbdElkader, M.F., Hamouda, A.H., Omar, A.A. and Alwakeel, R.A. (2020). The influence of dietary β-glucan on immune, transcriptomic, inflammatory and histopathology disorders caused by deltamethrin toxicity in Nile tilapia (Oreochromis niloticus). Fish& Shellfish Immunology, 98, 301–311.
- Dawood, M.A., Eweedah, N.M.,
 Moustafa, E.M. and Shahin, M.G.
 (2020). Synbiotic effects of *Aspergillus oryzae* and β-glucan on

growth and oxidative and immune responses of Nile Tilapia, *Oreochromis niloticus.Probiotics and Antimicrobial Proteins*, 12(1), 172–183. https:// doi.org/10.1007/s1260 2-018-9513-9.

- Dawood, M.A., Koshio, S., El-Sabagh, M., Billah, M.M., Zaineldin, A.I., Zayed, M.M. and Omar, A.A.E.D. (2017). Changes in the growth, humoral and mucosal immune responses following β -glucan and vitamin C administration in red sea bream, *Pagrusmajor*. *Aquaculture*, 470, 214–222.
- Dawood, M.A., Koshio, S., Ishikawa, M. and Yokoyama, S. (2015).Interaction effects of dietary supplementation of heat-killed Lactobacillus plantarum and β glucan on growth performance, digestibility and immune response of juvenile red sea bream, Pagrus major. Fish Shellfish k Immunology, 45(1), 33–42.
- Dawood, M.A., Magouz, F.I., Salem, M.
 F., Elbialy, Z. I. and Abdel-Daim,
 H.A. (2020). Synergetic effects of *Lactobacillus plantarum* and β-glucan on digestive enzyme activity, intestinal morphology, growth, fatty acid, and glucose-related gene expression of genetically improved farmed tilapia. *Probiotics and Antimicrobial Proteins*, 12, 389–399.
- Dawood, M.A., Metwally, A.E.S., El-Sharawy, M.E., Atta, A.M., Elbialy, Z.I., Abdel-Latif, H.M.R. and Parav, B. A. (2020). The role of β -glucan in the growth, intestinal morphometry, and immune-related gene and heat shock protein expressions of Nile tilapia (Oreochromis under *niloticus*)

different stocking densities. *Aquaculture*, 523, 735205.

- Dawood, M.A.O., Moustafa, E.M., Elbialy, Z.I., Farrag, F., Lolo, E.E.E., Abdel-Daim, H. A., Abdel-Daim, M. M. and Van Doan, H. (2020). Lactobacillus plantarum L-137 and/or β -glucan impacted the histopathological, antioxidant. immune-related genes and resistance of Nile tilapia (Oreochromis *niloticus*) against Aeromonashydrophila. Research in Veterinary Science, 130, 212–221.
- M.E., Ahmed, El-Boshy, M.. AbdelHamid, F.M. and Gadalla, H.A. (2010). Immunomodulatory effect of dietary Saccharomyces *cerevisiae*, β -glucan and laminaran in mercuric chloride treated Nile tilapia (Oreochromis niloticus) and experimentally infected with Aeromonashydrophila. Fish & Shellfish Immunology, 28(5-6),802-808
- El-Nokrashy, A., El-Banna, R., Edrise, B., Abdel-Rahim, M., Jover-Cerdá, M., Tomás-Vidal, A., Prince, A., Davies, S., El-Haroun, E. and Goda, A.S (2021). Impact of nucleotideenriched diets on the production of gilthead seabream, Sparus aurata fingerlings by modulation of liver mitochondrial enzyme activity. antioxidant status., immune gene and expression. gut microbial ecology. Aquaculture, 535, 736398.
- Filho, F.D.O.R., Koch, J.F.A., Wallace, C. and Leal, M.C. (2019). Dietary β-1,3/1,6-Glucans Improve the Effect of a Multivalent Vaccine in Atlantic Salmon Infected with *Moritella viscosa* or Infectious Salmon Anemia Virus. *Aquaculture Int.* 27, 1825–1834.

- Fusté, N.P., Guasch, M., Guillen, P., Anerillas, C., Cemeli, T., Pedraza, N. and Garí, E. (2019). Barley βglucan accelerates wound healing by favoring migration versus proliferation of human dermal fibroblasts. Carbohydrate Polymers, 210, 389–398.
- Gil, A. (2002). Modulation of the immune response mediated by dietary nucleotides. European Journal of Clinical Nutrition, 56 (Suppl. 3), S1–S4.
- Guo, X., Ran, C., Zhang, Z., He, S., Jin, M. and Zhou, Z. (2017). The growth promoting effect of dietary nucleotides in fish is associated with an intestinal microbiota mediated reduction in energy expenditure. Journal of Nutrition, 147, 781–788.
- Guselle, N. J., Markham, R. J. F. and Speare, D.J. (2007). Timing of intraperitoneal administration of β-1,3/1,6 glucan to rainbow trout, *Oncorhynchus mykiss* (Walbaum), affects protection against the microsporidian *Loma salmonae*. Journal of Fish Diseases, 30(2), 111–116.
- György, P. (1971). Biochemical aspect of human milk. American Journal of Clinical Nutrition, 24, 970.
- Hany, S.A., Eman, Y.M., Hayam, D.T., Abdelkrim, I. and Mohamed, S.H. (2024). The potential synergistic action of quercetin and/or *Pediococcus acidilactici* on Nile tilapia (*Oreochromis niloticus*) performance. Aquaculture, 581, 740353.
- Herre, J., Gordon, S. and Brown, G. D. (2004). Dectin-1 and its role in the recognition of beta-glucans by macrophages. Molecular Immunology, 40, 869–876.

- Hosseny, H. M., Motaal, S. M. A., Kamel, M. A. and El-Murr, A. H. I. (2018). Ameliorative effect of beta glucan diet in Oreochromis Niloticus against Aeromonas hydrophila. Research Journal of Pharmaceutical Biological and Chemical Sciences, 9(6), 391–404.
- Huang, W., Xiao, X., Hu, W., Tang, T., Bai, J., Zhao, S., Ao, Z., Wei, Z., Gao, W. and Zhang, W. (2023). Effects of dietary nucleotide and wall veast cell on growth performance, feed utilization, antioxidative and immune response of (Ctenopharyngodon carp grass idella). Fish & Shellfish Immunology, 134, 108574.
- Kaisa, H., Hanne, J., Aki, R., Anna, H., Veera, K., Jonna, J. and Reetta, S. (2018). The potential of gut commensals in reinforcing intestinal barrier function and alleviating inflammation. Nutrients, 10, 988.
- Kankkunen, P., Teiril[•]a, L., Rintahaka, J., Alenius, H., Wolff, H. and Matikainen, S. (2010). (1, 3)-βglucans activate both dectin-1 and NLRP3 inflammasome in human macrophages. J. Immunol. 184, 6335–6342.
- Khan, A. A., Gani, A., Masoodi, F. A., Mushtaq, U. and Naik, A. S. (2017). Structural, rheological, antioxidant, and functional properties of β glucan extracted from edible Agaricus mushrooms bisporus, Pleurotus ostreatus and Coprinus atramentarius. Bioactive Carbohydrates and Dietary Fibre, 11, 67–74.
- Krüger, D. and van der Werf, M. (2018). Benefits of nucleotide supplementation in aquaculture: Fish. Ohly Application Note, 1–4.

- Kühlwein, H., Emery, M., Rawling, M., G., Merrifield, D.and Harper. Davies, S. (2013). Effects of a dietary β -(1,3) (1,6)-D-glucan supplementation intestinal on microbial communities and intestinal ultrastructure of mirror carp (Cyprinus carpio L.). Journal of Applied Microbiology, 115. 1091-1106.
- Leonardi, M., Sandino, A.M. and Klempau, A. (2003). Effect of a nucleotide-enriched diet on the immune system, plasma cortisol levels and resistance to infectious pancreatic necrosis (IPN) in juvenile rainbow trout (*Oncorhynchus mykiss*). Bulletin of the European Association of Fish Pathologists, 23, 52–59.
- Li, H., Zhao, P., Lei, Y., Li, T. and Kim, I. (2015). Response to an Escherichia coli K88 oral challenge and productivity of weanling pigs receiving a dietary nucleotides supplement. Journal of Animal Science and Biotechnology, 6(1).
- Li, P. and Gatlin, D.M. (2006). Nucleotide nutrition in fish: Current knowledge and future applications. Aquaculture, 251, 141–152.
- Li, P., Burr, G.S., Goff, J., Whiteman, K. W., Davis, K.B., Vega, R.R., Neill, W. H. and Gatlin, D.M. (2005). A preliminary study on the effects of dietary supplementation of brewer's yeast and nucleotides, singularly or in combination, on juvenile red drum (*Sciaenops ocellatus*). Aquaculture Research, 36(11), 1120–1127.
- Li, P., Gatlin, D. M. and Neill, W.H. (2007). Dietary supplementation of a purified nucleotide mixture transiently enhanced growth and feed utilization of juvenile red

drum, *Sciaenops ocellatus*. Journal of the World Aquaculture Society, 38(2), 281–286.

- Li, H., Zhao, P., Lei, Y., Li, T. and Kim, I. (2015). Response to an *Escherichia coli* K88 oral challenge and productivity of weanling pigs receiving a dietary nucleotides supplement. Journal of Animal Science and Biotechnology J Animal Sci Biotechnol, 6(1).
- Li, P., Lawrence, A.L., Gastille, F.L. and Gatlin, D.M. (2007). Preliminary evaluation of a purified nucleotide mixture as a dietary supplement for Pacific white shrimp *Litopenaeus vannamei* (Boone). Aquaculture Research, 38, 887–890.
- Li, P., Lewis, D.H. and Gatlin, D.M. (2004). Dietary oligonucleotide from yeast RNA influences immune responses and resistance of hybrid striped bass (*Morone chrysops* × *M. saxatilis*) to *Streptococcus iniae* infection. Fish & Shellfish Immunology, 16, 561–569.
- Li, W., Zeng, J., Li, Y., Ge, C., Su, J. and Yao, H. (2024). A fascinating finding: The application of traditional Chinese medicine in aquaculture prevents the spread and diffusion of antibiotic resistance genes among gut microbes. Aquaculture, 583,740573.
- Lin, Y., Wang, H. and Shiau, S. (2009). Dietary nucleotide supplementation enhances growth and immune responses of grouper, *Epinephelus malabaricus*. Aquaculture Nutrition, 15(2), 117–122.
- Liu, B. (2016). The effect of dietary nucleotide supplementation on growth and feed efficiency of rainbow trout (*Oncorhynchus mykiss*) fed fish meal-free and

animal protein-free diets. Doctoral Dissertation, University of Guelph.

- Luo, J., Zeng, D., Cheng, L., Mao, X., Y u, J., Yu, B. and Chen, D. (2019). D ietary beta glucan supplementation improves growth performance, carcass traits and meat quality of finishing pigs. Animal Nutrition, 5, 380–385.
- Mathan Muthu, C.M., Vickram, A.S., Bhavani Sowndharya, B.. Saravanan, A., Kamalesh, R. and Dinakarkumar, Y.A (2024).Comprehensive review on the utilization of probiotics in towards aquaculture sustainable shrimp farming. Fish Shellfish Immunol.147:109459.
- Manoppo, H., Manurung, U.N. andTumbol, R. A. (2015). Efficacy of baker's yeast as immunostimulant in Nile tilapia (*Oreochromis niloticus*). International Journal of Chem. Tech. Research, 8(2), 559–565.
- Métailler, R., Cadena-Roa, M. and Person-Le Ruyet, J. (1983). Attractive chemical substances for the weaning of Dover sole (*Solea vulgaris*): qualitative and quantitative approach. Journal of the World Mariculture Society, 14, 679–684.
- Misra, C.K., Das, B.K., Mukherjee, S.C. and Pattnaik, P. (2006). Effect of multiple injections of beta-glucan on non-specific immune response and disease resistance in *Labeo rohita* fingerlings. Fish & Shellfish Immunology, 20(3), 305–319.
- Mohebbi, A., Nematollahi, A., Gholamhoseini, A., Tahmasebi-Kohyani, A. and Keyvanshokooh, S. (2013). Effects of dietary nucleotides on the antioxidant status and serum lipids of rainbow trout

(Oncorhynchus mykiss). Aquaculture Nutrition, 19(4), 506– 514.

- Moore, K., Mullan, B., Pluske, J., Kim, J. and D'souza, D. (2011). The use of nucleotides, vitamins and functional amino acids to enhance the structure of the small intestine and circulating measures of immune function in the post-weaned piglet. Animal Feed Science and Technology, 165(3-4), 184–190.
- Neamat-Allah, A.N., Abd El Hakim, Y. Mahmoud. and E.A. (2020).Alleviating effects of β -glucan in Oreochromis niloticus on growth performance, immune reactions. antioxidant. transcriptomics resistance disorders and to Aeromonas sobria caused by atrazine. Aquaculture Research, 51(5), 1801–1812.
- Nikl, L., Evelyn, T.P.T. and Albright, L.J. (1993). Trials with an orally and immersion-administered beta-1,3 glucan as an immunoprophylactic against *Aeromonas salmonicida* in juvenile chinook salmon *Oncorhynchus tshawytscha*. Diseases of Aquatic Organisms, 17(3), 191–196.
- Nishantha, M.D.L.C., Zhao, X., Jeewani, D.C., Bian, J., Nie, X. and Weining, S. (2018). Direct comparison of β-glucan content in wild and cultivated barley. International Journal of Food Properties, 21(1), 2218–2228.
- Noman, M., Kazmi, S.S.U.H., Saqib, H.S.A. (2024). Harnessing probiotics and prebiotics as ecofriendly solution for cleaner shrimp aquaculture production: state-of-theart scientific consensus. Sci Total Environ.;915:169921.

- Norton, R., Leite, J., Vieira, E., Bambirra, E., Moura, C., Penna, G. and Penna, F. (2001). Use of nucleotides in weanling rats with diarrhea induced by a lactose overload: effect on the evolution of diarrhea and weight and on the histopathology of intestine, liver and spleen. Brazilian Journal of Medical and Biological Research, 34, 195– 202.
- Ortuno, J., Cuesta, A., Rodriguez, A., Esteban, M.A. and Meseguer, J. (2002). Oral administration of yeast, Saccharomyces cerevisiae, enhances the cellular innate immune response of gilthead seabream (*Sparus aurata L.*). Veterinary Immunology and Immunopathology, 85, 41–50.
- Person-Le Ruyet, J., Menu, B., Cadena-Roa, M. and Métailler, R. (1983). Use of expanded pellets supplemented with attractive chemical substances for the weaning of turbot (*Scophthalmus maximus*). Journal of the World Mariculture Society, 14, 676–678.
- Pogue, R., Murphy, E.J., Fehrenbach, G. W., Rezoagli, E. and Rowan, N.J. (2021). Exploiting immunomodulatory properties of beta-glucans derived from natural products for improving health and sustainability in aquaculture-farmed concise organisms: review of existing knowledge, innovation and future opportunities. Current Opinion in Environmental Science & Health, 21, 100248.
- Przewłócka. K., Folwarski. M., Kaźmierczak-Siedlecka. K., Skonieczna-Żydecka, Κ. and Kaczor, J.J. (2020). Gut-muscle axis exists and may affect skeletal muscle adaptation to training. Nutrients, 12, 1451.

- Qi, C., Cai, Y., Gunn, L., Ding, C., Li, B., Kloecker, G., Qian, K., Vasilakos, J., Saijo, S., Iwakura, Y., Yannelli, J.R. and Yan, J. (2011). Differential pathways regulating innate and adaptive anti-tumor immune responses by particulate and soluble yeast-derived betaglucans. Blood, 117, 6825–6836.
- Ramadan, A., Atef, M. and Afifi, N. (1991). Effect of the biogenic performance enhancer (Ascogen "S") on growth rate of tilapia fish. Acta Veterinaria Scandinavica, 304–306.
- Reda, R.M., Selim, K.M., Mahmoud, R. and El-Araby, I.E. (2018). Effect of dietary yeast nucleotide on antioxidant activity, non-specific immunity, intestinal cytokines, and disease resistance in Nile tilapia. Fish & Shellfish Immunology, 80, 281–290.
- Rudolph, F.B. (1994). The biochemistry and physiology of nucleotides. The Journal of Nutrition, 124, 124S– 127S.
- Sahoo, P.K. and Mukherjee, S.C. (2002). The effect of dietary immunomodulation upon *Edwardsiella tarda* vaccination in healthy and immunocompromised Indian major carp (*Labeo rohita*). Fish & Shellfish Immunology, 12, 1–16.
- Sakai, M. (1999). Current research status of fish immunostimulants. Aquaculture, 172, 63–92.
- Salah, A.S., El Nahas, A.F. and Mahmoud, S. (2017). Modulatory effect of different doses of β -1,3/1,6-glucan on the expression of antioxidant, inflammatory, stress and immune-related genes of *Oreochromis niloticus* challenged

with *Streptococcus iniae*. Fish & Shellfish Immunology, 204–213.

- Sari, M., Prange, A., Lelley, J.I. and Hambitzer, R. (2017). Screening of beta-glucan contents in commercially cultivated and wild growing mushrooms. Food Chemistry, 216, 45–51.
- Sauer, N., Bauer, E., Vahjen, W., Zentek, J. and Mosenthin, R. (2010). Nucleotides modify growth of selected intestinal bacteria in vitro. Livestock Science, 133(1-3), 161–163.
- Shah, A., Gani, A., Masoodi, F.A., Wani, S.M., and Ashwar, B.A. (2017). Structural, rheological and nutraceutical potential of β-glucan from barley and oat. Bioactive Carbohydrates and Dietary Fibre, 10, 10–16.
- Selim, K.M. and Reda, R.M. (2015). Beta-glucans and mannan oligosaccharides enhance growth and immunity in Nile tilapia. North American Journal of Aquaculture, 77(1), 22–30.
- Singh, B.K., Delgado-Baquerizo, M., Egidi, E. (2023). Climate change impacts on plant pathogens, food security and paths forward. Nature Reviews Microbiology, 21(10), 640–656.
- Siwicki, A.K., Kazun, K., Gtabski, E., Terech-Majewska, E., Baranowski, P. and Trapkowska, S. (2004). The effect of beta-1.3/1.6—glucan in diets on the effectiveness of antiyersinia ruckeri vaccine an experimental study in rainbow trout (Oncorhynchus mykiss). Polish Journal of Food and Nutrition Sciences, 13-54(SI 2), 69–61.
- Soltanian, S., Stuyven, E., Cox, E., Sorgeloos, P. and Bossier, P. (2009). β-Glucan as

immunostimulant in vertebrates and invertebrates. Critical Reviews in Microbiology, 35(2), 109–138.

- Song, L., Zhou, Y., Ni, S., Wang, X.,
 Yuan, J., Zhang, Y. and Zhang, S. (2020). Dietary intake of β-glucans can prolong lifespan and exert an antioxidant action on aged fish *Nothobranchius guentheri*. Rejuvenation Research, 23, 293–301.
- Suchecka, D., Harasym, J.P., Wilczak, J., Gajewska, M., Oczkowski, M., Gudej, S., Błaszczyk, K., Kamola, D., Filip, R. and Gromadzka-Ostrowska, J. (2015). Antioxidative and anti-inflammatory effects of beta-glucan high concentration purified aqueous extract from oat in experimental model of LPS-induced chronic enteritis. Journal of Functional Foods, 14, 244–254.
- Sumon, M.A.A., Molla, M.H.R., Hakeem, I.J. (2022). Epigenetics and probiotics application toward the modulation of fish reproductive performance. Fishes, 7(4), 189.
- Tahmasebi-Kohyani, A., Keyvanshokooh, S., Nematollahi,
 A., Mahmoudi, N, and Pasha-Zanoosi, H. (2012). Dietary administration of nucleotides to enhance growth, humoral immune responses, and disease resistance of the rainbow trout (Oncorhynchus mykiss) fingerlings. Fish & Shellfish Immunology, 30(1), 189–193.
- Tie. H.M. (2018).Effects and mechanisms of dietary nucleotides on growth performance, functional organs health and flesh quality of voung grass carp (Ctenopharyngodon idellus). Master's Thesis. Sichuan Agricultural University.

- Tie, H.M., Jiang, W.D., Feng, L., Wu, P., , Liu, Y., Kuang, S.Y., Tang, L. and Zhou, X.Q. (2021).
 Dietary nucleotides in the diets of o ngrowing grass carp (*Ctenopharyng odon idella*) suppress *Aeromonas hy drophila* induced intestinal inflamm ation and enhance ntestinal diseaseresistance via NFκB and TOR signaling. Aquaculture , 533, 736075.
- Verdegem, M., Buschmann, A.H., Latt, U.W., Dalsgaard, A.J.T. and Lovatelli, A. (2023). The contribution of aquaculture systems to global aquaculture production. Journal of the World Aquaculture Society, 54(2), 206–250.
- Xu, M., Liang, R., Guo, Q., Wang, S., Zhao, M. and Zhang, Z. (2013).
 Dietary nucleotides extend the life span in sprague-dawley rats. The Journal of Nutrition, Health & Aging, 223–229.
- Yoshida, T., Kruger, R. and Inglis, V. (1995). Augmentation of nonspecific protection in African catfish, *Clarias gariepinus* (Burchell), by long term oral administration of immunostimulants. Journal of Fish Diseases, 18, 195–198.