

### **ORIGINAL PAPER**

# Efficacy of Pre-harvest Applications of Various Bio and Agrochemical Products on Anthracnose Disease of Mango Fruits and Keeping Quality During Storage Conditions.

Hassan, M. S. S.<sup>1</sup><sup>(1)</sup>, Shehata, A. S. F.<sup>1\*</sup><sup>(1)</sup> and Elmaghraby, I. M. K.<sup>2</sup><sup>(1)</sup>

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## ABSTRACT

Mango anthracnose disease is an essential problem in Egypt and one of the more significant economic restrictions of mango production. The economic losses caused by anthracnose disease are mostly attributed to reduce fruit quality and marketability. Anthracnose disease caused by Colletotrichum gloeosporioides is one of the most widespread fungal diseases that affects mango worldwide. This study was to verify the effects of pre-harvest applications of various bio and agrochemical products, Fungisei 1% (Bacillus subtilis strain IAB/BSO3), Biocontrol T34 12% (Trichoderma asperellum strain T34), Trifmine 15% (Triflumizole), Divide 60% and calcium chloride (CaCl<sub>2</sub>) against natural and artificial infection by C. gloeosporioides on two mango cultivars cv. Naomi and cv. Keitt after storage at 12±1 °C and 85-90% RH for 50 days and their effect on fruit quality parameters. C. gloeosporioides was able to induce anthracnose disease on mango fruits. Keitt cultivar was more susceptible than cv. Naomi under artificial inoculation in pathogenicity tests of the two years of the investigation. All tested products reduced C. gloeosporioides linear growth on PDA medium compared to control. Divide 60%, Trifmine 15% and CaCl<sub>2</sub> were the best effective treatments where each of them completely inhibited the growth of tested fungus. On the other hand, Biocontrol T34 12 % and Fungisei 1% recorded low efficacy in reducing the colony growth of the tested pathogen. All tested products were significantly better than the control in decrease the percentages of disease incidence and severity of anthracnose disease under natural and artificial infection with C. gloeosporioides. Calcium chloride, Trifmine 15% and Divide 60% their performance were the best compared with Fungisei 1% and Biocontrol T34 12 % for both natural and artificial infection during the two investigated seasons. Pre-harvest spraying any of CaCl<sub>2</sub>, Trifmine 15% EC and Divide 60% totally controlled and inhibited anthracnose disease on mango fruits which had been treated with the previous products in the field and were subjected to the natural infection and cold storage at 12±1 °C and 85-90% RH for 50 days during 2023 and 2024. All tested products kept fruit quality parameters and had significant effect due to total soluble solids, titratable acidity, fruit firmness and vitamin C (Ascorbic acid content).

Keywords: Pre-harvest spraying, bio and agrochemical products, anthracnose, mango fruits.

<sup>\*</sup>Correspondence: Shehata, A.S.F. E-mail: <u>arpp2022@arc.sci.eg</u>

Mabrouk S. S. Hassan https://orcid.org/0000-0002-3477-339X

#### Abou Ghanima, S. F. Shehata https://orcid.org/0000-0003-1698-7375

1. Plant Pathology Research Institute, Agricultural Research Center, 12619, Giza, Egypt.

#### Ibrahim M. K. Elmaghraby https://orcid.org/0000-0002-2728-9751

2. Central Lab. of Organic Agriculture, Agricultural Research Center, Giza 12619, Egypt.

#### **INTRODUCTION**

A tropical fruit known for its sweet and fresh flavor is the mango (Zakawa *et al.*, 2020). Mango trees are grown all throughout the world, but particularly in tropical nations. In 2019, 55.85 million metric tons of mangos were produced worldwide, a 37% growth over the previous ten years (FAO, 2021). Rich in proteins, vitamins A and C, carotene, malic and citric acids, and other elements, mango fruit has a high nutritional value. With an average vield of 10 tons per acre, Egypt's farmed acreage increased to 300,000 acres in 2022. With around 30,000 tons sent to markets in Europe, including France, Germany, the Netherlands, the UK, and Russia, as well as Gulf countries including the United Arab Emirates, Oman, Kuwait, and Saudi Arabia, in addition to Lebanon. Egypt is the tenthlargest exporter of mangos worldwide (Sharkawy et al., 2023). The mango fruit promotes the development of its color, flavor, and texture by increasing its respiration rate and ethylene production after harvest. Fruit quality is decreased

during storage due to the faster growth of these characteristics, and fungus cause postharvest losses. Anthracnose and other postharvest diseases are brought on by these Mango fruit is attacked by fungi. phytopathogens belonging to the genus Colletotrichum al., (Xu et 2017). Colletotrichum gloeosporioides has spread around the world and inflicted a significant economic damage. (Chung et al., 2010). Mango fruit anthracnose has been found to be caused by Colletotrichum species. According to reports, it is one of the most aggressive pathogens that attack the various mango fruit varieties in Egypt (Ismail and El-Ganainy, 2022), Mexico (Tovar-Pedraza et al., 2020), the Philippines (DelaCueva et al., 2021), Australia (Giblinet al., 2018), China (Mo et al., 2018) and Indonesia (Benatar et al., 2021). Colletotrichum symptoms may be in a latent phase prior to harvest; dark patches and tissue putrefaction are signs of the disease that emerges after harvest (Tovar-Pedraza et al., 2020). Although it can happen at any point in the fruits life cycle, anthracnose is most frequently found on mango fruits during storage and transportation. The infection on larger fruits may remain latent or inactive until the fruit ripens, at which point black, sunken necrotic lesions emerge on the fruit peel and rapidly enlarge (De Souza et al., 2013). The young fruits are either aborted or mummified. During the flowering and fruitsetting seasons, anthracnose disease is most harmful under damp conditions. Mango infections can occur during flowering and the early stages of fruit production (Sarkar, 2016). The disease shortens the fruits' shelf life by infecting them while they are in the field, during transit and in cold storage (Qin et al., 2019). The extent of disease resistance varies among cultivars. Two mango cultivars, "Keitt" and "Zill," were studied for their resistance to the anthracnose disease. When С. gloeosporioides infected commercially ripe or young fruit, the lesion sizes of "Keitt" fruit were less than those of "Zill" fruit (Gong et al., 2013)."Keitt" had a lower disease index than "Zill" when non-

inoculated fruits were harvested at commercial maturity, suggesting that "Keitt" was more disease resistant than "Zill" (Gong et al., 2013). Low temperatures and coatings are two methods that have been utilized to prevent deterioration, increase shelf life, and preserve mango fruit quality (Ravindra and Goswami, 2007). According to Tripathi and Dubey (2004), natural products are helpful and are being used as an alternative method of postharvest fruit deterioration and ripening delay. Most of the focus and effort in the fight against anthracnose has been on the use of fungicides. The application of fungicides reduces damage to fruit and inflorescence. Depending on the final destination of the exported fruit, different fungicides may be Early trials demonstrated used. the effectiveness of non-systemic fungicides such as zineb, maneb, or captan when treated weekly during flowering and subsequently monthly throughout fruit development (Sardrood and Goltapeh, 2018). In South Africa, it is advised to use copper oxychloride or copper oxychloride combined with zineb every 14 days in wet conditions and every 28 days in dry conditions of pre-harvest as part management to prevent anthracnose (Akem, 2006). Additionally, С. gloeosporioides has been reported to be resistant to systemic fungicides called benzimidazoles (Chung et al., 2010). Despite the excellent effectiveness of chemical treatment with several fungicides against C. gloeosporioides, reports of the establishment of fungicide-resistant isolates of this disease have been made (Kongtragoul et al. 2011). Agronomic significance could be greatly increased by investigating different bio-control agents to manage different plant diseases. Mango pre- and post-harvest anthracnose may be environmentally managed using an favorable future method that involves the bio-formulation application of *Trichoderma* harzianum, Bacillus subtilis, and Pichia (Sharma al., anomala et 2021). Chitinolytic activity was observed for Bacillus species and Trichoderma species

(Huang et al., 2005). Fungal mycelia are lysed by Bacillus species (Podile and Prakash 1996). Bacillus subtilis showed strong antagonistic action against C. gloeosporioides in the dual plate assay. Ashwini and Srividya According to microscopic investigations (2014),demonstrated a definite hyphallysis and fungal cell wall degradation. One of the key elements influencing the growth and quality of horticultural crops is mineral nutrition. Calcium (Ca), one of the mineral nutrients, is thought to be a major factor in determining the quality and shelf life of fruits and is known to have a significant impact on plant cell processes. According to Liu et al. (2017), postharvest calcium application has been shown to benefit fruit in a variety of ways, such as lowering disorders, physiological maintaining membrane permeability, preventing softening, slowing ripening processes, and reducing fruit decay as well as increasing fruit acceptability. It has previously been mentioned that CaCl<sub>2</sub> could be added to food to enhance the texture of the meat in goose meat preparations (Li et al.. study **2017**). This aimed to control anthracnose on mango fruits and keeping quality during storage conditions at 12±1 °C and 85-90% RH for 50 days during 2023 and 2024 on both Naomi and Keitt cultivars by performing pre-harvest spray of five bio and agrochemical products *i.e.* Fungisei 1% SC 150ml/100L, Biocontrol T34 12 %WP 200g/100L, Calcium chloride (CaCl<sub>2</sub>) 100g/100L, Trifmine 15% EC 25ml/100L and Divide 60% WG 100g/100L water.

### **MATERIALS AND METHODS**

This study was conducted using two mango varieties, Naomi and Keitt that were grown in El-Qanater El-Khayria, Qaliobiya governorate, Egypt, during 2022, 2023, and 2024 seasons.

#### Pathogen isolation:

In Qaliobiya governorate, mango fruits (cv. Naomi and cv. Keitt) exhibiting naturally occurring symptoms of anthracnose disease were collected from marketplaces, cold room storage and fields. After being chopped into small pieces, the diseased tissues were surface sterilized by immersing them in 1% sodium hypochlorite for 0.5 min. After being cleaned using sterile distilled water, the tissues were cultured on PDA medium. After that, the Petri plates were incubated for a week at 22°C. The hyphal tip method (HTM) was used to purify the developing fungal colonies. According to **Sutton (1980)** the purified isolates were identified depending on their morphological and cultural characters and maintained on PDA media.

## **Pathogenicity test:**

Mango fruits cv. Naomi and cv. Keitt were collected from the Agricultural Research Center (ARC), El-Qanater El-Khayriya Horticultural Research Station (EHRS) during 2023 and 2024. There were no wounds or rotting on the mango fruits. Three replicates of nine fruits from each cultivar were used. After being cleaned and surface sterilized, the mango fruits were slightly scraped and inoculated with spore suspension of Colletotrichum gloeosporioides  $(10^6 \text{ spores/ml})$  for one minute using 7 days old cultures (Hassan et al., 2021). Additionally, the same numbers of unwounded fruits were inoculated by spraying spore suspension  $(10^6 \text{ spores/ml})$ of Colletotrichum gloeosporioides (Hassan and Shehata, 2024). After 15 days of room temperature incubation at 24±1 °C, the inoculated and non-inoculated fruits were evaluated for disease symptoms using the disease incidence (%) and disease severity (%) scales according to Lakshmi et al. (2011) (Table 1). The causal fungus of anthracnose disease was re-isolated again for identity confirmation.

#### **Disease incidence (%)** = $n/N \ge 100$

Where, n= Number of infected mango fruits with anthracnose disease and N= the total number of all tested fruits.

# Disease severity (%) = $\frac{e n \times v}{5N} \times 100$

Where:

- 5 = Maximum disease severity grade,
- n = Number of infected fruits in each category (grade),
- N = Total number of the inspected fruits,
- v = Numerical value of each category as follows:

Hassan et al.,

Table	(1): Disease severity scale:
Rating	Where:
0	No of infected fruit.
1	5% of fruit affected area.
2	Between 5 and 10% of fruit affected
	area.
3	Between 10 and 20% of fruit affected
	area.
4	Between 20 and 50% of fruit affected
	area.
5	More than 50% of fruit affected area.

## In vitro experiment:

*In vitro*, the inhibitory efficacy of five bio and agrochemical products (Table 2); Fungisei 1% SC (*Bacillus subtilis* strain (IAB/BSO3  $1 \times 10^8$  CFU/ml)), Biocontrol T34 12% WP (*Trichoderma asperellum* strain T34), calcium chloride (CaCl<sub>2</sub>), Trifmine 15% EC (Triflumizole) and Divide 60% WG (Metiram 55% + Pyraclostrobin 5%) С. against gloeosporioides was assessed. Before solidifying, treatments were added each alone to conical flasks containing autoclaved PDA. They were then gently mixed and then poured into 90 mm plates. After solidification, equal disks 5 mm in diameter were cut from the periphery of the active mycelial growth of 7-day-old cultures of C. gloeosporioides to inoculate each plate separately at the center. At  $22\pm1$ °C, the plates were incubated. The control group consisted of untreated PDA plates that had been inoculated with fungus. For every treatment, six plates were used. The following formula was used to compute the average colony diameter (mm) and percentage inhibition once the control plates had completed full development (Hassan et al., 2021).

 $\% \text{ Inhibition} = \frac{\text{Mean of colony diameter in control plates} - \text{Mean of colony diameter of test plates}}{\text{Mean of colony diameter in control plates}} \times 100$ 

Table (2): Tested bio and agrochemical compounds.									
Product	Active ingredients	Status	Conc./100 L water						
Fungisei 1%	<i>Bacillus subtilis</i> strain (IAB/BSO3 1×10 <sup>8</sup> CFU/ml)	SC	150 ml						
Biocontrol T34 12%	Trichoderma asperellum strain T34	WP	200 g						
Calcium chloride	CaCl <sub>2</sub>	WP	100 g						
Trifmine 15%	Triflumizole	EC	25 ml						
Divide 60%	Metiram 55% + Pyraclostrobin 5%	WG	100 g						

## Field experiment:

During 2023 and 2024. field experiments were conducted on 12-year-old mango trees, cv. Naomi and cv. Keitt, at the Agricultural Research Center, El-Qanater El-Khayriya Horticultural Research Station (EHRS). Three mango trees were intended for each treatment, with three trees per replicate. Five sprays of each investigated and agrochemical products were bio applied; the first was sprayed at the start of fruit set, and the interval between sprays was 20 days. Water was sprayed on control mango trees. Mango fruits were harvested at the physiological maturity stage and split into two groups. Eighteen fruits were selected at random from each cultivar mentioned treatment. As in the pathogenicity test, the first group of mango fruits which were treated with Fungisei 1%, Biocontrol T34 12%, Calcium chloride (CaCl<sub>2</sub>), Trifmine 15%, and Divide 60% were surface sterilized and artificially inoculated with the prepared spore suspension Colletotrichum of gloeosporioides  $(10^6 \text{ spores/ml})$ . Without any artificial inoculation, the second group of mango fruits which had been treated with the identical bio and agrochemical products that had been tested in the field were subjected to a natural infection. For 50 days, mango fruits were kept in the EHRS Central Lab at 12±1 °C and 85-90% relative humidity (RH) in cardboard boxes. The disease incidence and severity of anthracnose, fruit firmness (Ib/inch<sup>2</sup>), total soluble solids (TSS), titratable acidity (TA) and vitamin C (Ascorbic acid as mg/100 g fresh weight) were assessed at the first and the end of the cold storage.

#### **Disease assessment:**

Disease incidence and disease severity (%):

Anthracnose disease incidence (%) and disease severity (%) were determined as mentioned before according to Lakshmi *et al.* (2011).

### Physical characteristics: Fruit firmness (Ib/inch<sup>2</sup>):

Using a hand pressure tester as  $Ib/inch^2$  fruit firmness was calculated according to **Hassan** *et al.* (2023).

## Chemical Properties: Total soluble solids (TSS):

Using Digital refractometer PR32 the percentage of TSS was calculated in mango fruit juice as recorded by Abdel-Rahman *et al.* (2021) and Hassan *et al.* (2023).

## Titratable acidity (TA) %:

TA was calculated by titrating the mango juice against 0.1 N NaOH using phenolphthalein indicators and expressed as percentage of citric acid according to **Abdel-Rahman** *et al.* (2021).

## Vitamin C (Ascorbic acid):

Vitamin C content (Ascorbic acid as mg/100g fresh weight) was evaluated according to **Shao** *et al.* (2013) using the method of 2, 6-dichlorophenol indophenol.

## RESULTS

#### Pathogen isolation:

Using its cultural and morphological characteristics, the isolated fungus from mango fruits (cv. Naomi and cv. Keitt) showing natural infection with anthracnose disease symptoms and were collected from fields, cold room storage and markets in Qaliobiya governorate during 2022 was purified and identified as *Colletotrichum gloeosporioides* Penz (Fig. 1).

## **Pathogenicity test:**

*Colletotrichum gloeosporioides* was able to cause anthracnose disease on mango fruits, according to the pathogenicity test (Fig. 2). Mango fruits cv. Naomi and cv. Keitt were subjected to a pathogenicity test, which was verified. According to data in the infection Table (3),bv С. gloeosporioides on cv. Keitt caused the highest disease severity (anthracnose) percentage on the cultivars examined during 2023 and 2024. According to the pathogenicity test conducted throughout the two-years of the investigation, cv. Keitt was more susceptible to artificial inoculation than cv. Naomi. Little amounts of the disease severity (%) on the unwounded fruits after inoculation by spore suspension using Colletotrichum gloeosporioides after the incubation period (Fig. 3).

 Colletotrichum gloeosporioides during the 2023 and 2024.

 Disease incidence (%)
 Disease severity (%)

 Cultivars
 Cultivars

Table (3): Pathogenicity test as anthracnose disease incidence (%) and disease severity of

<b>C</b> 14 <sup>2</sup>	Disease in	icidence (%)	Disease severity (%)			
Cultivars	2023	2024	2023	2024		
Naomi	100a	100a	37.77b	26.66b		
Keitt	100a	100a	71.11a	48.88a		

No significant differences between the treatments with the same letter/s in the same column at  $P \le 0.05$ 

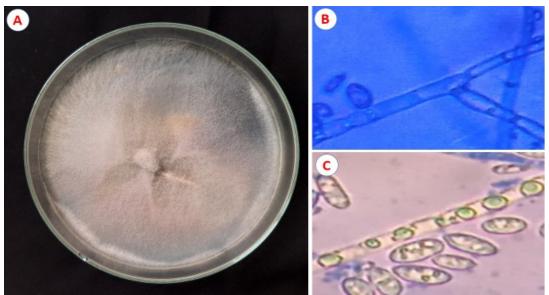
## In vitro experiment:

All tested bio and agrochemical products decreased *Colletotrichum gloeosporioides* linear growth on PDA medium compared to control, according to data in Table (4). The most successful treatments were Divide 60%, Trifmine 15% and Calcium chloride (CaCl<sub>2</sub>) where all tested materials inhibited the development of the tested fungus. However, the effectiveness of Biocontrol T34 (12%) and Fungisei 1% in lowering the studied pathogens colony growth was only 77.77 % and 72.77%, respectively.

**Table (4):** Effect of the tested bio and agrochemical products on the linear growth (mm) of Collectorichum gloeosporioides in vitro.

Tested chemical	Con./L	Colletotrichum gloeospor	<i>rioides</i> linear growth
rested chemical	Con./L	Mm	%Eff.
Fungisei 1%	1.5 ml	25.00 b	72.77
Biocontrol T34 12 %	2 g	20.00c	77.77
CaCl <sub>2</sub>	1 g	0.00d	100
Trifmine 15%	0.25ml	0.00d	100
Divide 60%	1 g	0.00d	100
Control		90.00a	00

No significant differences between the treatments with the same letter/s in the same column at  $P \le 0.05$  mm = Colony growth, millimeters. % Eff. = efficacy to control (untreated). Con. /L= concentration/ litre media



**Fig. (1):** *Colletotrichum gloeosporioides* isolated from mango fruits 7 days after incubation at 22±1 °C on PDA medium (A); (B) Conidiophore and (C) conidiospores.



Fig. (2): Symptoms of infection by *Colletotrichum gloeosporioides* on wounded artificially inoculated mango fruits. (A) cv. Naomi; (C) cv. Keitt and control (B & D).



**Fig. (3):** Little amounts of the disease severity (%) on the unwounded fruits after inoculation by spore suspension using *Colletotrichum gloeosporioides* after the incubation period. (A) cv. Naomi; (B) cv. Keitt and control (C & D).

#### **Field experiment:**

Disease incidence and disease severity (%):

Effect of pre-harvest spraying of the tested bio and agrochemical products on anthracnose disease incidence (%) and severity (%) under natural infection and artificially inoculated mango fruits by *C. gloeosporioides* after storage at  $12\pm1^{\circ}$ C and 85-90% RH for 50 days:

After 50 days storage period at 12±1 °C and 85-90% RH, the effects of bio and products. agrochemical Fungisei 1% (*Bacillus subtilis* strain (IAB/BSO3  $1 \times 10^8$ CFU/ml)), Biocontrol T34 12% (Trichoderma asperellum strain (T34)), Calcium chloride (CaCl<sub>2</sub>), Trifmine 15% EC (Triflumizole), and Divide 60% (Metriam 55% + Pyraclostrobin 5%) are shown on two mango cultivars in Table (5 and 6). In general, the highest percentage of disease incidence and severity was seen in the control treatment (Fig. 4 and 5). During the two seasons under investigation, calcium chloride (CaCl<sub>2</sub>), Trifmine 15% EC, and Divide 60% performed the best when compared to Fungisei 1% and Biocontrol T34 12% for both natural and artificial infections. On naturally infected mangos, CaCl<sub>2</sub>, Trifmine 15% EC, and Divide 60% completely inhibited and controlled anthracnose disease during 50 days of cold storage at 12±1 °C and 85-90% RH in 2023 and 2024 (Table 5). On the other hand in the artificial infections, CaCl<sub>2</sub> recorded the lowest disease incidence and severity % in both seasons on both tested cultivars.

Table (5): Effect of pre-harvest spraying of the tested bio and agrochemical products on two mango
cultivars on anthracnose disease incidence (%) and disease severity (%) caused by
Colletotrichum gloeosporioides under natural infection after storage at 12±1 °C and 85-
90% RH for 50 days during 2023 and 2024 seasons.

Treatments	Como		20	23		2024			
	Conc. /100L	Nac	omi	Ke	eitt	Nac	omi	Keitt	
		DI%	DS%	DI%	DS%	DI%	DS%	DI%	DS%
Fungisei 1%	150 ml	33.33b	13.33b	22.22b	8.88b	44.44b	8.88b	22.22b	8.88b
Biocontrol T34 12 %	200 g	33.33b	13.33b	22.22b	8.88b	44.44b	8.88b	22.22b	8.88b
CaCl <sub>2</sub>	100 g	00.00c							
Trifmine 15%	25 ml	00.00c							
Divide 60%	100 g	00.00c							
Control	Water	44.44a	17.77a	55.55a	15.55a	66.66a	26.66a	77.77a	33.33a

No significant differences between the treatments with the same letter/s in the same column at P $\leq$ 0.05 DI%=Disease incidence, DS%= disease severity.

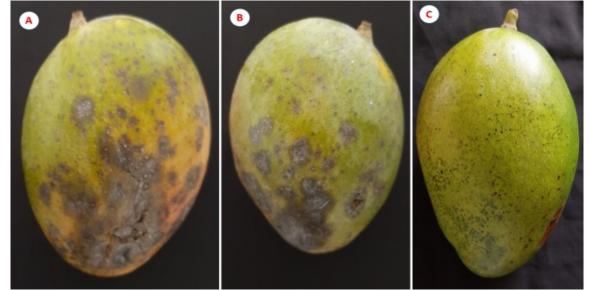
**Table (6):** Effect of pre-harvest spraying of the tested bio and agrochemical products on two mango cultivars on anthracnose disease incidence (%) and severity (%) caused by *Colletotrichum gloeosporioides* under artificial inoculation after storage at 12±1 °C and 85-90% RH for 50 days during 2023 and 2024 seasons.

	Como	-	20	23		2024				
Treatments	Conc. /100L	Naomi		Keitt		Naomi		Ke	itt	
	/100L	DI%	DS%	DI%	DS%	DI%	DS%	DI%	DS%	
Fungisei 1%	150 ml	66.66c	17.77c	77.77b	26.66b	77.77b	31.11b	77.77b	26.66b	
Biocontrol T34 12 %	200 g	88.88b	28.44b	77.77b	26.66b	77.77b	31.11b	77.77b	26.66b	
CaCl <sub>2</sub>	100 g	44.44e	11.11e	55.55d	15.55e	33.33d	20.00c	55.55d	15.55e	
Trifmine 15%	25 ml	55.55d	13.33d	66.66c	22.22d	66.66c	20.00c	66.66c	22.22d	
Divide 60%	100 g	55.55d	13.33d	77.77b	24.44c	66.66c	20.00c	77.77b	24.44c	
Control	Water	100.00a	40.00a	100.00a	60.00a	100.00a	33.33a	100.00a	31.33a	

No significant differences between the treatments with the same letter/s in the same column at P $\leq$ 0.05 DI%=Disease incidence, DS%= Disease severity.



Fig. (4): Symptoms of *Colletotrichum gloeosporioides* infection on artificially inoculated mango fruits cv. Naomi after storage at 12±1 °C and 85-90% RH for 50 days; (A) control; (B) treated fruit with bio products and (C) treated fruit with agrochemical products.



**Fig. (5):** Symptoms of *Colletotrichum gloeosporioides* infection on artificially inoculated mango fruits cv. Keitt after storage at 12±1 °C and 85-90% RH for 50 days; (A) control; (B) treated fruit with bio products and (C) treated fruit with agrochemical products.

## Fruit firmness (Ib/inch<sup>2</sup>):

Effect of pre-harvest spraying of the tested bio and agrochemical products on fruit firmness (Ib/inch<sup>2</sup>) under natural infection and artificially inoculated mango fruits by *C. gloeosporioides* after cold storage at  $12\pm1$  °C and 85-90% RH for 50 days:

After 50 days of cold storage at  $12\pm1^{\circ}$ C and 85-90% RH, the effect of the tested products on the fruit firmness of two mango cultivars against both natural and artificial infection with anthracnose (*C. gloeosporioides*) is shown in Tables (7 and 8). According to the data, fruit firmness was decreased as storage time increased. After 50 days of cold storage at  $12\pm1^{\circ}$ C and 85-90% relative humidity, the firmness of the mango fruit began to decline more noticeably than before. In comparison to the control treatment, all of the evaluated bio and agrochemical products increased the firmness values of mango fruit at harvest and after cold storage in 2023 and 2024. During the two seasons of the study, preharvest spraying with Divide 60%, Trifmine 15% EC, and calcium chloride (CaCl<sub>2</sub>) generally resulted in higher mango fruit firmness values than Fungisei 1% and Biocontrol T34 12% in both tested cultivars. Generally, after 50 days of cold storage at 12±1 °C and 85-90% RH, the mango fruits with artificial treated anthracnose infection had lower firmness values compared to the naturally infected mango fruits.

Table (7): Effect of pre-harvest spraying of the tested bio and agrochemical products on two
mango cultivars fruit firmness (Ib/inch <sup>2</sup> ) under natural infection of mango fruits
with anthracnose (Colletotrichum gloeosporioides) after cold storage at 12±1°C
and 85-90% RH for 50 days.

		Fruit firmness (Ib/inch <sup>2</sup> )									
Tuestanonta	Conc.		20	23		2024					
Treatments	/100L	Naomi		Keitt		Naomi		Keitt			
		0 day	50 day	0 day	50 day	0 day	50 day	0 day	50 day		
Fungisei 1%	150 ml	12.00a	4.99e	12.60b	3.50c	12.00b	4.00c	12.00b	4.50b		
Biocontrol T34 12 %	200 g	12.00a	5.50d	12.60b	4.00b	12.00b	4.00c	12.00b	4.50b		
CaCl <sub>2</sub>	100 g	12.33b	6.00a	12.66a	5.00a	12.50a	6.00a	12.50a	5.00a		
Trifmine 15%	25 ml	12.33b	5.90b	12.66a	5.00a	12.50a	5.50b	12.50a	5.00a		
Divide 60%	100 g	12.33b	5.70c	12.66a	5.00a	12.50a	5.50b	12.50a	5.00a		
Control	Water	11.30c	3.55f	11.70c	3.00d	11.11c	3.30d	11.50c	3.00c		

No significant differences between the treatments with the same letter/s in the same column at P $\leq$ 0.05

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Table (8): Effect of pre-harvest spraying of the tested products on two mango cultivars fruit firmness (Ib/inch<sup>2</sup>) under artificial infection of mango fruits with anthracnose (*Colletotrichum gloeosporioides*) after cold storage at 12±1°C and 85-90% RH for 50 days.

		Fruit firmness (Ib/inch <sup>2</sup> )									
Treatments	Conc. /100L		20	23		2024					
		Na	omi	Ke	eitt	Nao	omi	Keitt			
		0 day	50 day	0 day	50 day	0 day	50 day	0 day	50 day		
Fungisei 1%	150 ml	12.00b	4.00b	12.60b	3.50c	12.00b	4.00b	12.00b	4.00b		
Biocontrol T34 12 %	200 g	12.00b	4.00b	12.60b	4.50b	12.00b	4.00b	12.00b	3.50c		
CaCl <sub>2</sub>	100 g	12.33a	5.00a	12.66a	5.00a	12.50a	5.00a	12.50a	4.00b		
Trifmine 15%	25 ml	12.33a	5.00a	12.66a	5.00a	12.50a	5.50a	12.50a	4.00b		
Divide 60%	100 g	12.33a	5.00a	12.60b	5.00a	12.50a	5.00a	12.50a	5.00a		
Control	Water	11.30c	3.50c	11.70c	3.50c	11.11c	3.00c	11.50c	3.00d		

No significant differences between the treatments with the same letter/s in the same column at P $\leq$ 0.05

## Total soluble solids (TSS):

Effect of pre-harvest spraying of the tested bio and agrochemical products on total soluble solids (TSS) under natural infection and artificially inoculated mango fruits by *C. gloeosporioides* after cold storage at 12±1 °C and 85-90% RH for 50 days:

Data concerning total soluble solids (TSS) content in mango fruits cv. Naomi and cv. Keitt due to pre-harvest spraying of some bio and agrochemical products Fungisei 1%, Biocontrol T34 12 %, calcium chloride (CaCl<sub>2</sub>), Trifmine 15% EC and Divide 60% under natural and artificial infection by anthracnose (*C. gloeosporioides*) after cold storage at  $12\pm1$  °C and 85-90% RH for 50 days are reported in Tables (9 and 10). Data showed that, TSS

values were increased significantly with the progress of storage period and reached the highest level at the end of storage period compared with fruits at harvest. Partly, during 2023 and 2024, all tested products decreased TSS values at harvesting and after cold storage for 50 days compared with control treatment. In most cases, preharvest spraying of Fungisei 1% and Biocontrol T34 12 % decreased TSS values compared with Divide 60%, Trifmine 15% EC and CaCl<sub>2</sub> and control treatment in both tested cultivars during the two seasons of investigation. TSS values were increased under artificial infection than natural infection with C. gloeosporioides after cold storage at 12±1 °C and 85-90% RH for 50 days.

**Table (9):** Effect of pre-harvest spraying of the tested bio and agrochemical products on two mango cultivars total soluble solids (TSS) under natural infection with anthracnose (*Colletotrichum gloeosporioides*) after 50 days of cold storage at 12±1 °C and 85-90% RH.

		Total soluble solids (TSS)										
Tugatmonta	Conc.		2	023		2024						
Treatments	/100L	Naomi		Keitt		Naomi		Keitt				
		0 day	50 day	0 day	50 day	0 day	50 day	0 day	50 day			
Fungisei 1%	150 ml	11.30c	17.50e	11.50e	17.20c	11.60d	17.00d	11.30c	16.00c			
Biocontrol T34 12 %	200 g	11.30c	17.66d	11.55d	17.20c	11.60d	17.00d	11.30c	16.00c			
CaCl <sub>2</sub>	100 g	12.00b	18.33c	11.90c	18.00b	12.00c	18.00c	11.55b	17.50b			
Trifmine 15%	25 ml	12.00b	18.55b	11.95b	18.00b	12.00c	19.00b	11.55b	17.50b			
Divide 60%	100 g	12.00b	18.55b	11.95b	18.00b	12.30b	19.00b	11.55b	17.50b			
Control	Water	12.50a	19.00a	12.11a	18.66a	12.80a	19.22a	12.00a	18.00a			
No significant differences	hatwaan	the treat	monte u	ith the c	ama lattar/	s in the sor	na column	at D<0.05				

No significant differences between the treatments with the same letter/s in the same column at  $P \le 0.05$ 

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<b>Table (10)</b>	: Effect of	of pre-harv	est spi	raying of	the test	ed bio a	and agro	ochemical	products of	n two
	mango	cultivars	total	soluble	solids	(TSS)	under	artificial	infection	with
	anthrac	nose (Coll	etotric	hum glo	eospori	oides) a	fter col	d storage	at 12±1 °C	C and
	85-90%	6 RH for 50	) days.							

				To	tal solut	le solid	s (TSS)			
Treatments	Conc.		20	23		2024				
	/100L	Nac	omi	Ke	eitt	Na	omi	Keitt		
		0 day	50 day	0 day	50 day	0 day	50 day	0 day	50 day	
Fungisei 1%	150 ml	11.30c	17.90c	11.50e	18.50f	11.60d	18.70d	11.30c	19.00c	
Biocontrol T34 12 %	200 g	11.30c	17.90c	11.55d	18.80e	11.60d	18.70d	11.30c	19.00c	
CaCl <sub>2</sub>	100 g	12.00b	19.50b	11.90c	19.50c	12.00c	19.90b	11.55b	19.50b	
Trifmine 15%	25 ml	12.00b	19.50b	11.95b	19.00d	12.00c	19.50c	11.55b	19.50b	
Divide 60%	100 g	12.00b	19.50b	11.95b	19.80b	12.30b	19.50c	11.55b	19.50b	
Control	Water	12.50a	21.00a	12.11a	21.60a	12.80a	21.25a	12.00a	22.00a	

No significant differences between the treatments with the same letter/s in the same column at P≤0.05

#### Titratable acidity (TA) %:

Effect of pre-harvest spraying of the tested bio and agrochemical products on titratable acidity (TA) under natural infection and artificially inoculated mango fruits by *C. gloeosporioides* after cold storage at 12±1 °C and 85-90% RH for 50 days:

Concerning the response of titratable acidity (TA) content on mango fruits cv. Naomi and cv. Keitt to pre-harvest spraying of some bio and agrochemical products; Fungisei 1%, Biocontrol T34 12 %, calcium chloride (CaCl<sub>2</sub>), Trifmine 15% EC and Divide 60% under natural and artificial infection with anthracnose (*C. gloeosporioides*) after cold storage at  $12\pm1$  °C and 85-90% RH for 50 days, results are reported in Tables (11and 12). Data showed

that, during 2023 and 2024, all products caused a significant decrease in titratable acidity (TA) after cold storage for 50 days compared with 0 day. All tested products recorded increasing in TA compared with control in the two tested cultivars during the two years of the investigation. In most cases, pre-harvest spraying of Divide 60% gave the highest TA values followed by Trifmine 15% EC and CaCl<sub>2</sub> compared with control. Pre-harvest spraying of the products increased TA% values in cv. Keitt compared with cv. Naomi. TA values were decreased in mango fruits under artificial infection with anthracnose (*C*. gloeosporioides) compared to naturally infected one in both tested cultivars during the two seasons of investigation.

Table (11): Effect of pre-harvest spraying of the tested bio and agrochemical products on two								
mango cultivars titratable acidity (TA) under natural infection with anthracnose								
( <i>Colletotrichum gloeosporioides</i> ) after cold storage at $12 \pm 1$ °C and 85-90% RH								
for 50 days.								

					Titratal	ole acidity	(TA) %				
Treatments	Conc.	Conc. 2023					2024				
	/100L	Na	omi	i Keitt Naomi				Keitt			
		0 day	50 day	0 day	50 day	0 day	50 day	0 day	50 day		
Fungisei 1%	150 ml	0.95d	0.45d	0.98c	0.48d	0.90d	0.45e	0.99b	0.45c		
Biocontrol T34 12 %	200 g	0.97c	0.49c	1.90b	0.55c	0.99c	0.55d	1.99a	0.45c		
CaCl <sub>2</sub>	100 g	0.99b	0.69b	1.90b	0.75b	0.99c	0.60c	1.99a	0.80b		
Trifmine 15%	25 ml	0.99b	0.69b	1.90b	0.75b	1.03b	0.68b	1.99a	0.80b		
Divide 60%	100 g	1.03a	0.72a	1.96a	0.77a	1.11a	0.74a	1.99a	0.88a		
Control	Water	0.69e	0.29e	0.77d	0.33e	0.69e	0.45f	0.77c	0.36d		

No significant differences between the treatments with the same letter/s in the same column at  $P \le 0.05$ 

Table (12): Effect of pre-harvest spraying of the tested bio and agrochemical products on two mango cultivars titratable acidity (TA) under artificial infection with anthracnose (*Colletotrichum gloeosporioides*) after cold storage at 12±1 °C and 85-90% RH for 50 days.

Treatments					Titratab	le acidity	(TA) %		
	Conc.		2	023			20	24	
	/100L	Na	omi	ŀ	Keitt	Na	omi	Keitt	
		0 day	50 day	0 day	50 day	0 day	50 day	0 day	50 day
Fungisei 1%	150 ml	0.95b	0.25d	0.98c	0.45c	0.90d	0.35c	0.99b	0.25c
Biocontrol T34 12 %	200 g	0.94c	0.29c	1.96a	0.45c	0.99c	0.35c	1.99a	0.25c
CaCl <sub>2</sub>	100 g	0.94c	0.32b	1.96a	0.65b	0.99c	0.36b	1.99a	0.45b
Trifmine 15%	25 ml	0.94c	0.32b	1.96a	0.65b	1.03b	0.34d	1.99a	0.45b
Divide 60%	100 g	1.03a	0.38a	1.93b	0.77a	1.11a	0.38a	1.99a	0.47a
Control	Water	0.69d	0.20e	0.77d	0.33d	0.69e	0.35e	0.77c	0.30d

No significant differences between the treatments with the same letter/s in the same column at  $P \le 0.05$ 

# Vitamin C (Ascorbic acid):

Effect of pre-harvest spraying of the tested bio and agrochemical products on fruit vitamin C (Ascorbic acid) content under natural infection and artificially inoculated mango fruits by *C. gloeosporioides* after cold storage at 12±1 °C and 85-90% RH for 50 days:

Data in Tables (13 and 14) and Fig. (6) during 2023 and 2024, show a significant decrease in vitamin C (Ascorbic acid) mg/100g after cold storage for 50 days compared with 0 day. All tested bio and agrochemical products caused an increase in vitamin C (Ascorbic acid) compared with control in the two tested mango cultivars

during the two years of the investigation. In cases, pre-harvest spraying most of Fungisei 1% and Biocontrol T34 12 % gave the highest vitamin C values followed by calcium chloride (CaCl<sub>2</sub>) compared with control. Pre-harvest spraying of bio and agrochemical products increased vitamin C (Ascorbic acid) values in cv. Keitt compared with cv. Naomi. Under artificial infection with anthracnose (*C*. gloeosporioides), after cold storage at  $12\pm1$ °C and 85-90% RH for 50 days, vitamin C values were decreased compared with natural infected mango fruits in both tested cultivars during the two seasons of investigation.

Table (13): Effect of pre-harvest spraying of the tested bio and agrochemical products on two mango cultivars vitamin C content (Ascorbic acid) (mg/100g) under natural infection with anthracnose (*Colletotrichum gloeosporioides*) after cold storage at 12±1 °C and 85-90% RH for 50 days.

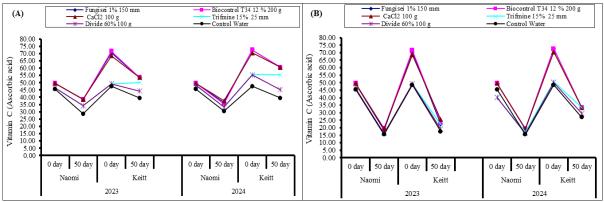
			V	itamin (	C (Ascor	rbic acid) mg/100g						
Treatments	Conc. /100L		20	23		2024						
		Na	omi	Ke	eitt	Na	omi	Keitt				
		0 day	50 day	0 day	50 day	0 day	50 day	0 day	50 day			
Fungisei 1%	150 ml	49.33c	38.50a	70.55b	53.50a	49.50a	36.50b	72.55a	60.50a			
Biocontrol T34 12 %	200 g	49.58a	38.50a	71.50a	53.50a	49.50a	35.50c	72.55a	60.50a			
CaCl <sub>2</sub>	100 g	49.50b	38.50a	68.50c	53.50a	49.50a	37.50a	70.50b	60.50a			
Trifmine 15%	25 ml	46.23e	34.20b	49.25e	50.20b	48.00c	33.20d	55.60c	55.20b			
Divide 60%	100 g	46.28d	34.20b	49.27d	44.20c	48.20b	33.20d	55.20d	45.20c			
Control	Water	45.53f	28.50c	47.56f	39.50d	45.57d	30.50e	47.57e	39.50d			

No significant differences between the treatments with the same letter/s in the same column at P≤0.05

Table (14): Effect of pre-harvest spraying of the tested bio and agrochemical products on two mango cultivars vitamin C content (Ascorbic acid) under artificial infection with anthracnose (*Colletotrichum gloeosporioides*) after cold storage at 12±1 °C and 85-90% RH for 50 days.

Treatments	-		r	Vitamin	C (Asc	orbic ac	id) mg/1	100g					
	Conc.	nc. 2023						2024					
	/100L	Nac	omi	Ke	eitt	Nac	eitt						
		0 day	50 day	0 day	50 day	0 day	50 day	e e e e e e e e e e e e e e e e e e e	50 day				
Fungisei 1%	150 ml	49.33c	18.50b	70.55b	22.50c	49.50a	19.50a	72.55a	33.50a				
Biocontrol T34 12 %	200 g	49.58a	19.50a	71.50a	23.50b	49.50a	19.00b	72.55a	33.50a				
CaCl <sub>2</sub>	100 g	49.50b	19.50a	68.50c	25.50a	49.50a	19.00b	70.50b	33.50a				
Trifmine 15%	25 ml	46.23e	16.20c	49.25e	22.20d	40.00d	17.20c	50.60c	33.50a				
Divide 60%	100 g	46.28d	16.20c	49.27d	1 9.20e	40.20c	16.20d	50.20d	29.50b				
Control	Water	45.53f	15.50d	48.56f	17.50f	45.57b	15.50e	48.57e	27.00c				

No significant differences between the treatments with the same letter/s in the same column at P≤0.05



**Fig. (6):** Effect of pre-harvest spraying of the tested bio and agrochemical products on two mango cultivars vitamin c (Ascorbic acid) mg/100g under natural infection (A) and artificial infection (B) with anthracnose (*Colletotrichum gloeosporioides*) after cold storage at 12±1°C and 85-90% RH for 50 days.

#### DISCUSSION

A tropical and seasonal fruit, mangos are typically accessible in Egypt during the summer months, from June through September serving as the primary harvest gathering time. With its superb exotic flavor, the mango (Mangifera indica L., Family Anacardiaceae) is regarded as the fruit's queen. About 60 million tons of mango fruits were produced worldwide (FAO, 2022). Mangos are the second most popular fruit crop in Egypt, after citrus. In the reclaimed land, the area planted with mango trees has grown very quickly. Keitt and Naomi, two previously introduced mango cultivars with exceptional fruit quality, were successfully cultivated in Egypt under various regional conditions (El-Agamy et al., 2018). Many diseases can affect mangos, some of which are quite

destructive and reduce the crop's yield and productivity. The most destructive of the mango diseases is anthracnose, which is caused by Colletotrichum gloeosporioides. As a post-harvest disease of harvested fruits, it mostly targets leaves, blossoms, young fruits, and twigs. Leaf spots, blossom blight, fruit spots, and eventually fruit rot are caused by black, irregularly shaped, and somewhat sunken spots that on leaves and fruits develop and progressively become larger. There is a significant prevalence of disease in moist or extremely humid environments, which are common in most mango-growing regions (Berardini et al. 2005). Mango availability, trade, and export are hampered by their high perish ability and vulnerability to postharvest infections. More than 25-30% of mangos are lost

during handling and storage, according to a nationwide survey (Prasad et al., 2019). According to Medlicott et al. (1990), mature green mango fruits should be kept between 10 and 12°C. In order to preserve fruit quality and prevent chilling injury, Pesis et al. (1997) kept mango fruits (cvs. Alphonso and Keitt) at various cold storage temperatures. They discovered that at 10°C. chilling injury symptoms manifested as pitting and red spots on the peel, while at lower temperatures, black spots emerged. When Pairi mango fruits were kept at 25°C, El-Khoreiby et al. (1998) found that the proportion of rotten fruits was 13% in the first week and 37% in the second. In the present study, C. gloeosporioides was able to induce anthracnose disease on mango fruits. Keitt cultivar was more susceptible than cv. Naomi under artificial inoculation in pathogenicity tests of the two years of the tested investigation. All bio and agrochemical products С. reduced gloeosporioides linear growth on PDA medium compared with control. Divide 60%, Trifmine 15% and calcium chloride (CaCl<sub>2</sub>) were the best effective treatments where they completely inhibited the growth of tested fungus. On the other hand, Biocontrol T34 12 % and Fungisei 1% recorded low efficacy in reducing the colony growth of the tested pathogen. All tested products were significantly better than the control in decrease the percentages of disease incidence and disease severity of anthracnose disease under natural and artificial infection with С. gloeosporioides. In general, the performance of CaCl<sub>2</sub>, Trifmine 15% and Divide 60% were the best compared with Fungisei 1% and Biocontrol T34 12 % for both natural and artificial infections during the two investigated season. Pre-harvest spraying any of CaCl<sub>2</sub>, Trifmine 15% EC and Divide 60% totally controlled and inhibited anthracnose disease on the second group of mango fruits which had been treated with the previous products in the field and were subjected to a natural infection after cold storage at 12±1 °C and 85-90% RH for 50 days during 2023 and

2024 growing seasons. These results are consistent with a prior study that found that C. gloeosporioides caused 20-30% of mango fruits in Hyderabad to decay. Mango anthracnose disease reduced Bangladesh's mango yield by 25-30%, according to Hossain and Ahmed (1994). In Thailand, Sardsud et al. (2003) found that mango anthracnose disease caused 62.8% of mangos loss during harvest and 63.2% in the markets. China's mango harvest can be reduced by 30-60% every year due to anthracnose (Li et al., 2019). Despite this, producers and sellers continue to offer low-quality fruits on the local market, with nations that export mangos, such as Ghana, suffering the most from the revenue gap between export and local markets (Kankam et al., 2022). Due to senescence and physiological changes, mango fruits are susceptible to post-harvest infections while being stored (Prusky and Keen, 1993). Khaliq et al. (2016) physiological the examined and characteristics of mango biochemical (Mangifera indica L. cv. Choke Anan) fruit in relation to the effects of 10% Arabic gum (AG) and 1% chitosan (CH) edible coatings. During the storage period, they found no discernible differences between fruits treated with CH 1% and fruits treated with AG 10% + CH 1%. At the conclusion of the storage period, the control samples had the highest decay percentage. Coating has been shown to be effective in inhibiting the growth of fungi in a variety of horticultural products (Tripathi and Dubey, 2004). A number of methods have been tested to prevent the development of rot and postpone the ripening of mango fruit by employing natural bio preservatives rather than fungicides. According to Elsabee and Abdou (2013), chitosan helps prevent postharvest fruit diseases and possesses antibacterial and antifungal potent qualities. The most promising fungicide in fields, according to Sharma and Verma (2007), was Saaf (carbendazim 12% + mancozeb 63%) @ 0.2%, which can be advised for the management of mango anthracnose. According to Gud and Raut

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(2008), the most effective fungicides for completely inhibiting the mycelial growth of mango anthracnose were propiconazole 0.1%, thiophanate methyl (0.2%), and Emisan 0.2%. The same authors (2008) mentioned that many Trichoderma species are effective at preventing the growth of the mycelia of C. gloeosporioides, which causes mango anthracnose. All of the isolated fungi linear growth was totally reduced by 0.5ml/L Difenoconazole 25% (Score 25% EC), 0.5g/L Boscalid 25.2% + Pyraclostrobin 12.8% (Bellis 38%WG), and 1ml/L Thiabendazole (Tekto 50% SC), (Hassan et al., 2024). In the two seasons of the study, using both tested cultivars, cv. Montakhab El-Qanater and cv. Ewais, Score 25% EC was the most effective at reducing mango fruit drop. Bellis 38% WG Tekto 50% SC came and next. Azoxystrobin 20% (25 ml/100 L water) + Tebuconazole 30% (Dovex 50% SC) and 40 ml/100 L water Difenoconazole 15% + Propiconazole 15% (Craft 30% EC) provided the lowest efficacy. Pre-harvest spraying of loquat trees with the best concentrations of Score 25% EC (Difenoconazole) 1 ml/litre, Scar Nat 70% (Thiophanate-methyl) WP 1 g/litre. Pyrmadol 40% SC (Pyrimethanil) 1 ml/litre. and Decent 32.5 EC (Azoxystrobin 20% + Difenoconazole 12.5%) 1 ml/litre helped control fruit rot, leaf spot, and inflorescence blight diseases (Hassan and Shehata 2024). The three main pathogens that cause avocado fruit rot in Egypt are Colletotrichum Lasiodiplodia gloeosporioides. theobromae, and Penicillium expansum.

In Egypt, the characteristic symptoms of fruit rot were caused by artificial inoculation of these fungi. Under market conditions, avocado fruit rots may also be managed with pre-harvest treatments that contain the highest concentration of Imazalil and copper oxide nanoparticles. The research also demonstrated the viability of employing essential oils and alternative therapies as Serenade ASO (*Bacillus subtilis* QST 713) as management strategies for the prevention of avocado fruit rot disease both before and after harvest (Hassan et al., 2021). According to Begum et al. (2008), the Trichoderma isolates coiled around C. truncatum hyphae, further limiting its ability to develop and disseminate. Trichoderma species produce a variety of antibiotics that development suppress fungal and sporulation, including trichotoxin, trichodermin, harzianolide, and trichodermol (Yonas and Amare 2008).

Many Trichoderma species are known to produce siderophores and indole acetic acid, as well as a many of extracellular enzymes that aid in the breakdown of pathogenic fungi cell walls, such as chitinase, pectinase, amylase, and protease (Sallam et al., 2021). According to Yoshida et al. (2001), B. subtilis has the potential to produce antimicrobial peptide substances, such as subtilin, bacilysin and mycobacillisyn, which can alter hyphal morphology in C. acutatum and C. gloeosporioides hyphal cells, causing hyphal swelling, distortion, and cytoplasm aggregation. Anthracnose disease was successfully controlled on mango trees by spraving Streptomyces aureofaciens bacterial filtrate (Haggag et al., 2011). Korsten et al. (1991) recorded that *Bacillus subtilis*  $(10^7 \text{ cells/ml})$  has been used both before and after harvest to reduce avocado post-harvest infections. Additionally, B. subtilis decreased the severity of stem-end rot, the Dothiorella / Colletotrichum fruit rot complex, and anthracnose on Fuerte avocado fruit. Additionally, the fruit dipping treatment with B. subtilis was just as effective as or even more successful as a prochloraz. Mango anthracnose was managed by fruit dip and challenge inoculation techniques with promising bio-agents. Up to two days of incubation on the Alphonso variety, fruits with the treated bio-agents Trichoderma harzianum, T. asperellum, B. subtilis, P. fluorescens, and the yeasts Meyerozyma caribbica and Torulaspora delbrueckii showed no symptoms; nevertheless, following that, the fruits symptom expression was progressively increased. Bacterial bio-agents Ρ. fluorescens and B. subtilis had the lowest

disease incidence among the bio-agents examined, at 37.70% and 33.81%. respectively. In contrast, the disease index for fruits percentages treated with Trichoderma asperellum and T. harzianum were 43.75and 46.33%, respectively. On the tenth day of incubation, the disease index for the control (untreated fruits) recorded 85.50%, while the disease index for the yeast, M. caribbica, and T. delbrueckii, recorded 40.00 and 41.82 percent, respectively. Similar outcomes were obtained using the challenge inoculation strategy on the Alphonso variety (Sudha et al., 2021).

In the present study, all tested bio and agrochemical products kept fruit quality parameters and had significant effect due to total soluble solids TSS, TA and fruit firmness. All tested bio and agrochemical products increased mango fruit firmness values at harvesting and after cold storage at 12±1 °C and 85-90% RH for 50 days compared with control treatment. In most cases, pre-harvest spraying of Divide 60%, Trifmine 15% EC and CaCl<sub>2</sub> increased mango fruit firmness values compared with Fungisei 1% and Biocontrol T34 12 % in both tested cultivars during the two seasons of investigation. Mango fruit firmness values were decreased under artificial infection with anthracnose (C. gloeosporioides) than natural infected mangos after cold storage at  $12 \pm 1$  °C and 85-90% RH for 50 days. TSS values were increased significantly with the progress of storage and reached the highest level at the end of storage period compared with fruits at harvest. Partly, during 2023 and 2024, all tested bio and agrochemical products decreased TSS values at harvesting and after cold storage at 12±1 °C and 85-90% RH for 50 days compared with control treatment.

In most cases, pre-harvest spraying of Fungisei 1% and Biocontrol T34 12 % decreased TSS values compared with Divide 60%, Trifmine 15% EC and calcium chloride (CaCl<sub>2</sub>) and control treatment in both tested cultivars during the two seasons of investigation. TSS values were increased due to artificial

infection than under natural infection with C. gloeosporioides after cold storage at 12 ±1 °C and 85-90% RH for 50 days. All bio and agrochemical products caused a significant decrease in TA after cold storage for 50 days compared with 0 day. All tested bio and agrochemical products caused increase in TA compared with control in the two tested cultivars during the two years of the investigation. In most cases, pre-harvest spraying of Divide 60% gave the highest TA values followed by Trifmine 15% EC and calcium chloride (CaCl<sub>2</sub>) compared with control. Pre-harvest spraying of bio and agrochemical products increased TA% values in cv. Keitt compared with cv. Naomi. Under artificial infection with anthracnose (*C*. gloeosporioides) after cold storage at  $12\pm1$ °C and 85-90% RH for 50 days TA values were decreased compared to natural infected mango fruits in both tested cultivars during the two seasons of Significant decrease investigation. in vitamin C (Ascorbic acid) after cold storage for 50 days compared with 0 day. Pre-harvest spraying of Fungisei 1% and Biocontrol T34 12 % gave the highest vitamin C values followed by CaCl<sub>2</sub> compared to control. Pre-harvest spraying of bio and agrochemical products increased vitamin C (Ascorbic acid) values in cv. Keitt compared with cv. Naomi. Vitamin C values under artificially infected mango fruits with anthracnose (*C*. gloeosporioides), after cold storage at 12±1 °C and 85-90% RH for 50 days, were decreased compared to natural infection in both tested cultivars during the two seasons of investigation. These results are consistent with Ali et al. (2004), who found that ripening of mango fruits causes a decrease in firmness over the progress of the storage period. This is because ripening and cold storage increase the activity of that hydrolyze cell walls, enzymes including pectatelyases, polygalacturonase, pectinesterase, and pectin methylesterase. The hydrolysis of carbs to sugars due to moisture loss and a decrease in acidity may be the source of the larger increase in mango fruit content in TSS during the

progress of the storage period (Golding et al., 2005). Starch hydrolyzes into simple sugars throughout the ripening process; in ripe fruits, glucose, fructose, and sucrose predominate. The enzymes amylase, invertase, and sucrose synthase all boosted their activity and hydrolyzed the starch to produce sucrose (Kumar et al., 1994). According to a study by Mohamed et al. (2017) on the impact of chitosan coating on the storability of mango cv. "Zibda" fruit, the amount of mango fruit in titratable acidity (TA) dropped progressively and considerably over the course of the trial's two seasons. All preharvest coating applications considerably decreased the pace at which the titratable acidity (TA) of the fruit changed during cold storage. Fast senescence of mango fruits is linked to a rapid decrease in their TA content during storage (Abbasi et al., 2009). The conversion of citric acid into sugars and subsequent use of these sugars in different fruit metabolic processes may be the cause of the decrease in acidity over the storage period (Rathore et al., 2007). CaCl<sub>2</sub> has been shown to be a safe and efficient way to manage postharvest illness and prevent fruit decay, despite the fact that the use of some chemicals may have unintended lingering effects on human safety (Shen and Yang 2017). CaCl<sub>2</sub> and salicylic acid (SA) are signal response that activate molecules downstream reactions, such as the manufacture of antimicrobial compounds and cell wall reinforcement mechanisms, which may explain their impact on fruit disease prevention (Kumar 2014). The fungal pathogen C. gloeosporioides was stopped from growing by the applications of CaCl<sub>2</sub>, SA, and *M. pulcherrima* yeast. They also decreased the decay index, stopped the fruit from softening, stopped changes in the TSS and vitamin C contents, and increased the activity of crucial defense enzymes. Additionally, it was discovered the combination of SA, that М. pulcherrima yeast, and CaCl<sub>2</sub> was more effective than either treatment alone. Accordingly, we propose that SA, CaCl<sub>2</sub>, and *M. pulcherrima* yeast may be used as a

safe and efficient postharvest method to prevent anthracnose while also preserving and increasing mango postharvest life (Shao et al., 2019). Karemera et al. (2013) reported that mango fruits cv. Mallika exhibited a significant delay, with significantly higher fruit length, breadth, thickness, volume, weight, and pulp weight. Additionally, trees sprayed with 1.50% CaCl<sub>2</sub> 30 days before harvest showed also significantly higher TSS of fruits, a significantly higher percentage of total sugars and the lowest percentage of titratable acidity. Higher calcium levels in fruit can cause lower respiration and ethylene production rates, which may be the reason of the CaCl<sub>2</sub>-induced ripening delay (Hewajulige et al., 2003). CaCl<sub>2</sub> sprays may prolong the shelf life of fruits because, when fruits are picked at the right time, calcium plays a variety of roles, such as increasing fruit firmness compared to control Gore (2005). The fruit quality improvement that resulted from giving trees CaCl<sub>2</sub> may have been caused by its effects on improving the formation and transformation of carbohydrates and carbohydrate enzymes; other possible explanations include the reduction of abscission and the crucial role that calcium plays in maintaining the middle lamella cells (Kumar et al., 2006). The conversion of fruit organic compounds to TSS during enzymatic activities under the presence of CaCl<sub>2</sub> may be the cause of the increase in TSS during storage periods (Wahdan et al., 2011).

# CONCLUSION

study, Colletotrichum In this gloeosporioides was able to induce anthracnose disease on mango fruits. Keitt cultivar was more susceptible than cv. Naomi under artificial inoculation in pathogenicity tests. All tested bio and agrochemical products reduced С. gloeosporioides linear growth on PDA medium compared with control. Divide 60%, Trifmine 15% and Calcium chloride (CaCl<sub>2</sub>) were the best effective treatments where each completely inhibited the growth of the tested fungus. On the other hand, Biocontrol T34 12 % and Fungisei 1% recorded low efficacy in reducing the colony growth of the tested pathogen. All tested bio and agrochemical products were significantly better than the control in decrease the percentages of disease incidence and disease severity of anthracnose disease under natural and artificial infection with C. gloeosporioides. The performance of Calcium chloride, Trifmine 15% and Divide 60% were the best compared with Fungisei 1% and Biocontrol T34 12 % for both natural and infections during artificial the two investigated seasons. Pre-harvest spraying using CaCl<sub>2</sub>, Trifmine 15% and Divide 60% totally controlled and inhibited anthracnose disease on naturally infected mango during cold storage at 12±1 °C and 85-90% RH for 50 days during 2023 and 2024. All tested products kept fruit quality parameters and had significant effect due to total soluble solids, titratable acidity, fruit firmness and vitamin C values.

# Author's contribution

Majority contribution for the whole article belongs to the author(s). The authors read and approved the final manuscript.

# **Competing interests**

The author declares that he has no competing interests.

# **REFERENCES:**

- Abdel-Rahman, F. A.; Monir, G. A.; Hassan, M. S. S.; Ahmed, Y.; Refaat, M. H.; Ismail A.I. and El-Garhy, H. A.
  S. (2021). Exogenously applied chitosan and chitosan nanoparticles improved apple fruit resistance to blue mold, upregulated defense-related genes expression, and maintained fruit quality. Horticulturae, 7(8) 224.
- Abbasi, N. A.; Iqbal, Z. M.; Maqbool, M. and Hafiz, I. A. (2009). Postharvest quality of mango (*Mangifera indica* L.) fruit as affected by chitosan coating. Pakistan Journal of Botany 41, 343-357.
- Akem, C. N. (2006). Mango anthracnose disease: present status and future research priorities. Plant Pathol. J. 5,

266-273. 10.3923/ppj.2006.266.273.

- Ali, Z. M.; Chin, L.H. and Lazan, H. (2004). A comparative study on wall degrading enzymes, pectin modifications and softening during ripening of selected tropical fruits. Plant Science, 167, 317-327.
- Ashwini, N. and Srividya, S. (2014). Potentiality of *Bacillus subtilis* as bio control agent for management of anthracnose disease of chilli caused by *Colletotrichum gloeosporioides* OGC1. 3 Biotech 4:127-136.
- Begum, M. M.; Sariah, M.; Abidin, Z. M.
  A.; Puteh, B. A. and Rahman, A. M.
  (2008). Antagonistic potential of selected fungal and bacterial biocontrol agents against *Colletotrichum truncatum* of soybean seeds. Pertanica J. Trop AgricSci 31:45-53.
- Benatar, G. V.; Wibowo, A. and Suryanti, (2021). First report of *Colletotrichum asianum* associated with mango fruit anthracnose in Indonesia. *Crop Prot.* 141:105432. doi: 10.1016/j.cropro.2020.105432.
- Berardini, N.; Fezer, R.; Conrad, J.; Beifuss, U.; Carle, R. and Schieber, A. (2005).
  Screening of mango (*Mangifera indica* L.) cultivars for their contents of flavonol O- and xanthone C-glycosides, anthocyanins, and pectin. J. Agric Food Chem. 53:1563-1570.
- Chung, W. H.; Chung, W. C.; Peng, M. T.; Yang, H. R. and Huang, J. W. (2010). Specific detection of benzimidazole resistance in *Colletotrichum gloeosporioides* from fruit crops by PCR-RFLP. New Biotechnol. 27, 17-24. doi: 10.1016/j.nbt.2009.10.004.
- DelaCueva, F. M.; Laurel, N. R.; Dalisay, T. U. and Sison, M. L. J. (2021).
  Identification and characterization of *Colletotrichum fructicola*, *C. tropicale* and *C. theobromicola* causing mango anthracnose in the Philippines. Arch. Phytopathol. 54, 1989-2006. doi: 10.1080/03235408.2021.1968234.
- De Souza, A.; DelphinoCarboni, R. C.; Wickert, E.; de MacedoLemos, E. G. and de Goes, A. (2013). Lack of host

doi:

specificity of *Colletotrichum* spp. isolates associated with anthracnose symptoms on mango in Brazil. Plant Pathol. 62, 1038-1047. doi: 10.1111/ppa.12021.

- El-Agamy, M. K.; Aly, H. H. and Hosny, S. (2018). Flowering and fruiting behavior of some introduced mango cultivars grown in Giza governorate conditions. Middle East Journal of Agric. Research, 7: 559-56.
- Elsabee, M. Z. and Abdou, E. S. (2013). Chitosan based edible films and coatings: A review. Materials Science and Engineering C, 33 (4), 1819-1841.
- El- Khoreiby, A. M. K.; Abbas, M. T.; Habib, S. S. and Serry, N. K. H. (1998). Effect of some postharvest treatments on mango fruit storability. Menofiya J. Agric. Res., 23(4): 055-1079.
- FAO, (2021). Food and Agriculture Organization. World Food and Agriculture - Statistical Yearbook 2020. Rome. pp. 368.
- FAO, (2022). Food and Agriculture Organization. World Food and Agriculture - Statistical Yearbook 2022. Rome. pp. 380.
- Giblin, F. R.; Tan, Y. P.; Mitchell, R.; Coates, L.M.; Irwin, J. A. G. and Shivas. R. G. (2018). *Colletotrichum* species associated with pre-and postharvest diseases of avocado and mango in Eastern Australia. Australasian Plant Pathology, 47(3), 269-276. <u>https://doi.org/10.1007/s13313-018-0553-0</u>.
- Golding, J. B.; Ekman, J. H. and Glasson, M.C. (2005). Regulation of fruit ripening.Stewart Postharvest Review.Postharvest Biology and Technology, 3, 1-5.
- Gong, D. Q.; Zhu, S. J.; Gu, H.; Zhang, L. B.; Hong, K. Q. and Xie, J. H. (2013). Disease resistance of 'Zill'and 'Keitt' mango fruit to anthracnose in relation to defense enzyme activities and the content of anti-fungal substances. J. Hortic. Sci. Biotechnol. 88, 243–250. doi: 10.1080/14620316.2013.11512962.
- Gore, S. P. (2005). Studies on extending the shelf-life of fig (*Ficus corica* L.) fruits

Cvs. "Poona" fig and "Dinkar". M. Sc. (Agri.) Thesis, Mahatma PhuleKrishi Vidyapeeth, Rahuri.

- Gud, M. A. and Raut, S. P. (2008). Control of mango anthracnose and stem end rot fungi by fungicides and bio agent. J. Maharashtra Agri. Uni., 33(1): 120-122.
- Haggag, W. M.; Mohamed, E. M. and Azzazy, A. M. E. (2011). Optimization and production of antifungal hydrolysis enzymes by *Streptomyces aureofaciens* against *Colletotrichum gloeosporioides* of mango. Agricultural Sci 2(2):146-157.
- Hassan, M. S. S.; Monir, G. A. and Radwan, M. A. (2021). Efficacy of certain essential oils, copper oxide, copper oxide nanoparticle, Imazalil and *Bacillus subtilis* to control fruit rot of avocado. Egypt. J. Phytopathol., 49(1): 166-181.
- Hassan, M. S. S.; Shehata, A. S. F. and Abdel-Wahed, G. A. (2023). Efficacy of some chemicals on controlling pear fruit rot and fruit quality under storage condition. Egyptian Journal of Phytopathology, Vol. 51, No. 1, pp 47-64.
- Hassan, M. S. S.; Shehata, A. S. F. and Banora, M. Y. (2024). Impact of some agrochemical products on early fruit drop of certain Egyptian mango cultivars induced by fungal infection. Egyptian Journal of Phytopathology, Vol. 52, No.1, pp 67-82.
- Hassan, M. S. S. and Shehata, A. S. F. (2024). Influence of some agrochemical products on Loquat inflorescence blight, leaf spot and fruit rot diseases. Egyptian Journal of Phytopathology, Vol. 52, No.2, pp 56-68.
- Hewajulige, I. G. N.; Wilson, W. R. S., R.
  L.; Wijesundera, C. and Abeysekere,
  M. (2003). Fruit calcium concentration and chilling injury during low temperature storage of pineapple. J. Sci. Food Agri., 83: 1451-1454.
- Hossain, A. A. and Ahmed, A. (1994). A monograph on mango varieties of Bangladesh. Dhaka: Bangladesh

Agricultural Research Institute. Horticulture Research Centre, BARI, Joydebpur, Gazipur, pp 155.

- Huang, C. J.; Wang, T. K.; Chung, S. C. and Chen, C. Y. (2005). Identification of an antifungal chitinase from a potential biocontrol agent, *Bacillus cereus*. J. Biochem Mol Biol Sci 38:82-88.
- Ismail, A. M. and El-Ganainy, S. M. (2022). Characterization of *Colletotrichum* species associating with anthracnose disease of mango in Egypt. J. Plant Dis. Prot. 129, 449–454. doi: 10.1007/s41348-021-00538-8.
- Kankam, F.; Larbi-Koranteng, S.; Adomako, J.; Kwodaga, J. K.; Akpatsu, I. B., and Danso, Y., *et al.* (2022). "Anthracnose disease of mango: epidemiology, impact and management options" in *Current and emerging challenges in the diseases of trees* (London: IntechOpen).
- Karemera, U. N. J.; Mukunda, G. K.; Ansar, H. and Amreen, T. (2013).
  Effect of pre-harvest sprays of calcium chloride on post-harvest behavior in mango fruits (*Mangifera indica* L.) Cv. Mallika. Plant Archives Vol. 13 No. 2, pp. 925-928.
- Khaliq, G.; Mohamed, M.T.M.; Ding, P.; Ghazali, H. M. and Ali, A. (2016).
  Storage behaviour and quality responses of mango (*Mangifera indica* L.) fruit treated with chitosan and gum Arabic coatings during cold storage conditions. International Food Research Journal 23(Suppl): S141- S148.
- Kongtragoul, P.; Nalumpang, S.; Miyamoto, Y.; Izumi, Y. and Akimitsu, K. (2011). Mutation at codon 198 of Tub2 gene for carbendazim resistance in *Colletotrichum gloeosporioides* causing mango anthracnose in Thailand. Journal of Plant Protection Research 51 (4): 377-384.
- Korsten, L.; de Villiers, E. E.; Jager, E. E.S.; Cook, de N. and Kotze, J. M. (1991). Biological control of avocado postharvest diseases. South African Avocado Growers' Association Yearbook, 14:57-59.

- Kumar, D. (2014). Salicylic acid signaling in disease resistance. Plant Science, 228, 127–134.
- Kumar, S.; Das, D. K.; Singh, A. K.; and Prasad, U. S. (1994). Sucrose metabolism during maturation and ripening of mango cultivars. Plant Physiology and Biochemistry 21(1): 27-32.
- Kumar, M. R.; Reddy, Y. N. and Srihari, D. (2006). Effect of calcium and plant growth regulators on flowering and yield of mango (*Mangifera indica* L.) Cv. Baneshan. J. Res. Angrau., 34: pp.11-15.
- Lakshmi, B. K. M; Reddy P. N and Prasad R. D (2011). Cross infection potential of *Colletotrichum gloeosporioides* Penz, isolates causing anthracnose in subtropical crops. *Tropical Agricultural Research* 2:183-193
- Li, X.; Sun, Y. Y.; Pan, D. D.; Wang, Y. and Cao, J. X. (2017). The effect of CaCl<sub>2</sub>marination on the tenderizing pathway of goose meat during conditioning. Food Research International, 102, 487-492.
- Li, Q.; Bu, J.; Shu, J.; Yu, Z.; Tang, L. and Huang, S. (2019). *Colletotrichum* species associated with mango in southern China. Sci. Rep. 9:18891. doi: 10.1038/s41598-019-54809-4.
- Liu, H.; Chen, F. S.; Lai, S. J.; Tao, J. R.; Yang. H. S and Jiao, Z. G. (2017). Effects of calcium treatment and low temperature storage on cell wall polysaccharide nanostructures and quality of postharvest apricot (*Prunus armeniaca*). Food Chemistry, 225, 97-116.
- Medlicott, T. G.; Sigarist, J. M. M. and Sy, O. (1990). Ripening of mangoes following low-temperature storage. J. Amer. Soc. Hort. Scie., 115(3): 430-434.
- Mo, J.; Zhao, G.; Li, Q.; Solangi, G. S.; Tang, L. and Guo, T. (2018). Identification and characterization of *Colletotrichum* species associated with mango anthracnose in Guangxi, China. Plant Dis. 102, 1283–1289. doi: 10.1094/PDIS-09-17-1516-RE.

- Mohamed, M. A. A.; Abd El-Khalek, A. F. and Elmehrat, H. G. (2017). Pre-harvest applications of potassium silicate, chitosan and calcium chloride to improve mango cv. 'Zibda' fruit quality and storability. Egypt. J. Hort., 44(1):17 - 32.
- Pesis, E.; Faure M.; Marinansky B. A.; Degani, C.; Gazit, S.; Lahav, E.; Pesis, E.; Prusky, D.; Tomer, E. and Wysoki, M. (1997). Induction of chilling tolerance in mango by temperature conditioning, heat, low 02 and ethanol vapours. Acta Hort., 455: 629-634.
- Podile, A. R. and Prakash, A. P. (1996). Lysis and biological control of *A. niger* by *Bacillus subtilis* AF1. Can J Microbiol 42:533-538
- Prasad, K.; Sharma, R. R.; Sethi, S. and Srivastav, M. (2019). Influence of harvesting method on postharvest loss, shelf-life and quality of mango (*Mangifera indica*) fruits, Indian J. Agric. Sci. 89, 445-449, https://doi.org/10.56093/ijas.v89i3.8758 7.
- Prusky, D. and Keen, N. T. (1993). Involvement of preformed antifungal compounds and the resistance of subtropical fruits to fungal decay. Plant Disease, 77, 114-119.
- Qin, L. P.; Zhang, Y.; Su, Q.; Chen, Y. L.; Nong, Q. and Xie, L. (2019). First report of anthracnose of *Mangifera indica* caused by *Colletotrichum scovillei* in China. Plant Dis. 103:1043. doi: 10.1094/PDIS-11-18-1980-PDN.
- Rathore, H. A.; Masud, T.; Sammi, S. and Soomro, A. H. (2007). Effect of storage on physico-chemical composition and sensory properties of Mango (*Mangifera indica* L.) variety Dosehri. Pakistan Journal of Nutrition 6, 143– 148.
- Ravindra, M. R. and Goswami, T. K. (2007). Post-harvest handling and storage of mangos and overview. Journal of Food Science and Technology. 44 (5): 449-458.
- Sallam, N.; Ali, E. F.; Seleim, M. A. A. and Bagy, H. (2021). Endophytic fungi associated with soybean plants and their

antagonistic activity against *Rhizoctonia solani*. Egypt J. Biol. Pest Control 31:54

- Sardrood, B. P. and Goltapeh, E. M. (2018). Effect of agricultural chemicals and organic amendments on biological control. Fungi in Sustainable Agriculture Reviews. ed. E. Lichtfouse, Vol. 12 (Berlin: Springer), 217-359.
- Sardsud, U.; Sardsud, V.; and Singkaew, S. (2003). Postharvest loss assessment of mango cv. Nam Dok Mai. Warasan WitthayasatKaset. Available at: <u>https://agris.fao.org/agris-</u> <u>search/search.do?recordID=TH2005000</u> <u>687</u> (Accessed February 1, 2023).
- Sarkar, A. K. (2016). Anthracnose diseases of some common medicinally important fruit plants. J. Med. Plant Res. 4, 233-236.
- Shao, Y. Z.; Xie, J. H.; Chen, P.and Li, W. (2013). Changes in some chemical components and in the physiology of rambutan fruit (*Nephelium lappaceum* L.) as affected by storage temperature and packing material. Fruits, 68, 15-24.
- Shao, Y. Z.; Zeng, J.; Tang, H.; Zhou, Y. and Li, W. (2019). The chemical treatments combined with antagonistic yeast control anthracnose and maintain the quality of postharvest mango fruit. Journal of Integrative Agriculture 18(5): 1159–1169.
- Sharkawy, S. A.; Maklad, T. N.; Mostafa, F. A. and Alkolaly, A. M. (2023).
  Integrated control program of powdery mildew on mango trees in Egypt by foliar sprays of fungicides combined with fertilizers. Alex. J. Agric. Sci. Vol. 68, No.2, pp. 101-107. DOI: 10.21608/alexja.2023.210045.1037.
- Sharma, A. and Verma, K. S. (2007). *In vitro* cross pathogenicity and management of *Colletotrichum gloeosporioides* causing anthracnose of mango. *Ann. Plant Prot. Sci*15, 186–188.
- Sharma, A.; Sharma, I. M.; Sharma, M.; Sharma, K. and Sharma, A. (2021). Effectiveness of fungal, bacterial and yeast antagonists for management of mango anthracnose (*Colletotrichum*)

*gloeosporioides*). Egyptian Journal of Biological Pest Control.pp.1-11.

- Shen, Y. F. and Yang, H. Q. (2017). Effect of pre-harvest chitosan-salicylic acid treatment on postharvest table grape quality, shelf life, and resistance to *Botrytis cinerea*-induced spoilage. Scientia Horticulture, 224, 367-373.
- Sudha, S.; Narendrappa, T. and Sivakumar, G. (2021). Management of post-harvest anthracnose disease in mango using promising bio control agents. The Pharma Innovation Journal; SP-10(3): 210-214.
- Sutton, B.C. (1980). The Coelomycetes. Commonwealth Mycological Institute, Kew. pp. 696.
- Tovar-Pedraza, J. M.; Mora-Aguilera, J. A.; Nava-Diaz, C.; Lima, N. B.; Michereff, S. J. and Sandoval-Islas, J. S. (2020). Distribution and pathogenicity of *Colletotrichum* species associated with mango anthracnose in Mexico. Plant Dis. 104, 137–146. doi: 10.1094/PDIS-01-19-0178-RE
- Tripathi, P. and Dubey, N. K. (2004). Exploitation of natural products as an alternative strategy to control postharvest fungal rotting of fruit and vegetables. Postharvest Biology and Technology. 32(3): 235–245.
- Wahdan, M. T.; Habib, S. E.; Bassal, M. A. and Qaoud, E. M. (2011). Effect of some chemicals on growth, fruiting,

yield and fruit quality of Cv. "Succary Abiad" mango. J. Hort. Sci., 7(2): pp. 651-658.

- Xu, X., Lei, H., Ma, X., Lai, T., Song, H., Shi, X. and Li, J. (2017). Antifungal activity of 1-methylcyclopropene (1-MCP) against anthracnose (*Colletotrichum gloeosporioides*) in postharvest mango fruit and its possible mechanisms of action. International Journal of Food Microbiology, 241, 1-6. <u>https://doi.org/10.1016/j.ijfoodmicro.20</u> <u>16.10.002</u>
- Yonas, K. and Amare, A. (2008). Postharvest biological control of anthracnose (*Colletotrichum* gloeosporioides) on mango (*Mangifera* indica). Postharvest Biol Technol 50:8– 11
- Yoshida, S.; Hiradate, S.; Tsukamato, T.; Hatakeda, K. and Shirata, A. (2001). Antimicrobial activity of culture filtrate *Bacillus amyloliquefaciens* RC-2 isolated from mulberry leaves. Phytopathology 91:181–187
- Zakawa, N. N.; Oyebanji, E. O.; Timon, D. and Batta, K. (2020). Anti-fungal activities of aqueous leaf extracts of *Moringa oleifera* Lam. on *Mangifera indica* L. post-harvest fruit-rot pathogens from some markets in Yola North, Adamawa state. World Journal of Pharmaceutical Research, 9(6), 1675-1687.



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