Egyptian Journal of Aquatic Biology & Fisheries Zoology Department, Faculty of Science, Ain Shams University, Cairo, Egypt. ISSN 1110 – 6131 Vol. 29(1): 2599 – 2609 (2025) www.ejabf.journals.ekb.eg



Profile of Secondary Metabolite Compounds of Seagrass *Enhalus acoroides* Extracted with Different Solvents from Suli Waters, Central Maluku Regency

Firman^{1*}, Fredrik Rieuwpassa², Meigy Nelce Mailoa²

¹Master of Marine Science Postgraduate Student, Pattimura University, Ambon, Indonesia ²Postgraduate Lecturer, Pattimura University, Ambon, Indonesia ***Corresponding Author: firmansyah0295@gmail.com**

ARTICLE INFO

Article History: Received: Jan. 13, 2025 Accepted: Feb. 20, 2025 Online: Feb. 25, 2025

Keywords: Enhalus acoroides, Extract, Solvent, Metabolite compound

ABSTRACT

Seagrass, especially Enhalus acoroides, is one of the important components in coastal ecosystems that provide various ecological and economic benefits. Secondary metabolite compounds produced by Enhalus acoroides can vary based on the environmental conditions where they grow. This study aimed to determine the profile of secondary metabolite compounds of Enhalus acoroides extract in the Suli Waters, Central Maluku Regency. This research was conducted experimentally in the laboratory and analyzed descriptively. Water quality was assessed based on temperature, dissolved oxygen levels, salinity, and pH, while identification of secondary metabolite compound groups was carried out qualitatively using phytochemical tests. Water quality showed that the seawater temperature in the waters of Suli Village reached 30°C; dissolved oxygen levels were 6.1mg/ L; salinity was 30% and pH was 6.8. It was observed that the methanol extract of Enhalus acoroides contains flavonoids, saponins, phenolics, tannins, steroids, terpenoids, and alkaloids. Meanwhile, flavonoids, phenolics, tannins, steroids, and terpenoids were detected in the ethyl acetate extract of Enhalus acoroides. Environmental conditions and the use of different types of solvents can affect the composition of the secondary metabolite compounds produced.

INTRODUCTION

Scopus

Indexed in

Coastal and marine resources in Indonesia are natural assets that have great potential for management and utilization in development (Oktawati *et al.*, 2018). In general, these resources consist of three main ecosystems: mangroves, seagrasses, and coral reefs (Kartika *et al.*, 2023). Each of these ecosystems has various benefits, namely coral reefs have the potential as ingredients for diarrhea medicine (Asih *et al.*, 2021); seagrass can be used in cosmetic products (Badriyah *et al.*, 2023a); and mangroves have the potential as herbal ingredients that are rich in antioxidants (Denny, 2024). Seagrass is a flowering plant (angiospermae) with single-lobed leaves (monocotyledons) that can grow well and permanently under the surface of the sea. Seagrass forms a carpet-like expanse in the sea, which can consist of one species or various species (Tangke, 2010). Ramadhan *et al.* (2021) explained that as an organism that lives permanently at the

ELSEVIER DO

IUCAT

bottom of the waters, seagrass has a greater potential to produce active compounds in its body.

Enhalus acoroides has been widely researched and reported to contain active compounds, including tannin and alkaloid compounds (Badriyah et al., 2023b). Enhalus acoroides from Sepanjang Beach-Yogyakarta contains tannins, saponins, triterpenoids, flavonoids, and steroids (Permana et al., 2020). Bioactive compounds in Enhalus acoroides have significant potential to be developed as antibacterials, anticancer, and antioxidants. Hitijahubessy et al. (2021) confirmed that Enhalus accordies extract is effective in inhibiting the growth of Vibrio sp. bacteria, thus showing its potential in controlling Vibrio parahaemolyticus bacteria. According to Nurfitriani et al. (2017), the content of secondary metabolites in marine organisms is influenced by several factors, where environmental factors are one of the main causes. Secondary metabolites are chemical compounds produced by organisms in response to various environmental conditions and stress, and they play an important role in ecological interactions, such as protection against predators, competition with other organisms, and adaptation to the environment (Petersen et al., 2020). According to Yang et al. (2018), environmental factors such as salinity, temperature, lighting, and others have significant impacts on the production and composition of secondary metabolites in marine organisms. Changes in salinity or temperature can affect the synthesis of bioactive compounds produced by marine organisms, such as algae, sponges, and coral reefs. This is also explained by Windyaswari et al. (2019) that water quality containing various nutrients and pollutants can also play a role in changing the secondary metabolite profile.

Atun (2014) explains that the type of solvent used can also affect the secondary metabolite content of *Enhalus acoroides* because the solubility of these compounds varies depending on the solvent used. Polar solvents such as water or methanol are effective in extracting polar compounds, while non-polar solvents such as chloroform or ethyl acetate are better at extracting non-polar compounds. This is also emphasized by **Dewatasari** (2020) who elucidated that secondary metabolites that usually have different solubility properties will be easier to extract with solvents that match their polarity. The purpose of this study was to determine the profile of secondary metabolite compounds of *Enhalus acoroides* extracted with different solvents from the waters of Suli Village, Central Maluku.

MATERIALS AND METHODS

Research location description

Suli Village is one of the villages in Salahutu District, Central Maluku Regency, located on the coast. The coastal waters of Suli Village face the strait that separates Haruku Island and Seram Island. This causes the coastal waters of Suli Village to be directly influenced by the Seram Sea which enters through the strait, allowing it to be influenced by quite intensive tidal currents. However, due to the adaptation of seagrass morphology and the shape of its roots, the seagrass community can grow well in these waters. This is due to the type of basic water substrate which is dominated by muddy sand and carbonate substrates.

Tools and materials

The tools used during the experiment were a shaker (*Brushless DC motor*), autoclave (*SMIC model YX-28*), test tube (*Pyrex*®), analytical balance (A&D company *limited*), rotary evaporator (*BUCHI R-215*), aluminum foil, DO meter (*Hanna*), refractometer (*AMTAST DBS1*), pH meter (*Hanna*) and thermometer. While the materials used were *Enhalus acoroides*, methanol, ethyl acetate, distilled water, H₂SO₄ 2N, FeCl₃ 1%, HCl 2N and Mg.

Work procedures

Water quality measurement

Measurement of water quality parameters includes water temperature using a thermometer, dissolved oxygen (DO) using a DO meter (*Hanna*), pH using a pH meter (*Hanna*) and salinity using a refractometer (*AMTAST DBS1*).

Preparation of test materials and extraction of Enhalus acoroides

The test materials used in this study were *Enhalus acoroides* obtained from the water of Suli State, Central Maluku. The samples were taken to the laboratory to be washed and cut into small pieces, after which they were air-dried. After drying, the samples were ground with a blender to obtain powder.

50g of powder *Enhalus acoroides* were put into a 100mL Erlenmeyer flask. Then 50mL of methanol solvent was added. The mixture was shaken with a shaker for 3 hours at a speed of 120rpm (rotation per minute) and macerated for 24 hours. Then the maceration residue was filtered with a Buchner funnel and re-dissolved using the same solvent until it was clear. The filtrate obtained was combined in an extract storage container (**Ukratalo** *et al.*, **2023**). The above treatment was also repeated for the use of ethyl acetate solvent. The obtained methanol and ethyl acetate extracts were concentrated using a rotary evaporator at 30-40°C, and N₂ gas was passed through until the solvent evaporated completely. Then the yield was calculated using the equation:

$$Rendement = \frac{Weight of crude extract obtained}{Sample weight used} \times 100\%$$

Phytochemical test (Kaihena et al., 2022)

Identification of secondary metabolites using qualitative phytochemical tests with the principles of color reactions, precipitation, and foam formation includes testing of alkaloids, steroids/triterpenoids, polyphenols/tannins, flavonoids, and saponins. With the following work steps:

a. Alkaloid

2mL of extract solution was added with 3 drops of $2N H_2SO_4$ then heated, followed by testing with Mayer and Wagner reagents. Positive test results were obtained whenever a precipitate was formed on the addition of Mayer's reagent and a brownish precipitate was observed upon the addition of Wagner's reagent.

b. Triterpenoid/ Steroid

Lieberman-Burchard reagent was added to 2mL of the extract solution. Positive test results are obtained if a green precipitate is formed (containing steroids) or a red precipitate is formed (containing triterpenoids).

c. Polyphenols/ Tannins

1mL of extract solution was added with two drops of 1% FeCl₃ reagent. The formation of green or blue color indicates the presence of phenol compounds.

d. Flavonoid

Extract solution of 1mL was added with a little Mg powder and 2mL of 2N HCl. Flavonoid compounds will produce orange to red color.

e. Saponins

An extract solution of 1mL was added with distilled water and shaken vigorously. The presence of saponin compounds is indicated by the formation of 1-10cm foam that is stable and not less than 10 minutes.

RESULTS AND DISCUSSION

1. Water quality of Suli State, Central Maluku

Measurement of the water quality of Suli Village, Central Maluku, referring to Government Regulation of the Republic of Indonesia Number 22 of 2021 concerning the Implementation of Environmental Protection and Management, can be seen in Table (1).

Parameter	Unit	Volume	Quality standards
Temperature	°C	30	28-30
Dissolved Oxygen/DO	mg/L	6.1	>5
Salinity	°/00	30	33-34
pH	-	6.8	7-8.5

Table 1. Measurement of water quality in Suli Village, Central Maluku

Table (1) shows that the sea water temperature in the waters of Suli State is 30°C, the dissolved oxygen content is 6.1mg/ L, the salinity is 300/00 and the pH is 6.8. The environment is one of the factors that influences the growth and development of *Enhalus acoroides*. Based on the measurement results, it can be said that the physical and chemical factors of the waters in Suli State do not inhibit the growth of *Enhalus acoroides*. This was emphasized in the study of **Sahertian and Wakano (2017)** who postulated that physical and chemical factors such as temperature, salinity, dissolved

oxygen, current speed, and pH do not interfere with or inhibit the growth of *Enhalus acoroides* because all of its physical components have not exceeded the established quality standards.

Sea water temperature is the temperature measured at the surface or at a certain depth of the ocean (**Akbari** *et al.*, **2017**). According to **Abraham** *et al.* (**2013**), this temperature varies depending on geographic location, depth, time of year, and other factors such as ocean currents and weather. Based on the quality standards (Government Regulation of the Republic of Indonesia Number 22 of 2021), it was deduced that the temperature of sea water in Suli Village which reaches 30°C is still optimal for seagrass growth.

According to **Frederick** *et al.* (2014), dissolved oxygen is a crucial element in waters (aquatic life) including seagrass which is a vital component of coastal ecosystems. Government Regulation of the Republic of Indonesia Number 22 of 2021 concerning Seawater Quality Standards for Biota shows that good DO levels for waters are >5 mg/L, with dissolved oxygen levels in the waters of Suli State at 6.1mg/ L, it can support optimal growth and health of *Enhalus acoroides*. Artika *et al.* (2020) explain that adequate dissolved oxygen allows *Enhalus acoroides* to get enough oxygen needed for metabolism and respiration processes.

Salinity, which measures the concentration of salt in seawater, plays a key role in determining the health and survival of seagrasses (**Cambridge** *et al.*, **2017**). The salinity in the waters of Suli State is $30^{\circ}/_{00}$ below the quality standard (Government Regulation of the Republic of Indonesia Number 22 of 2021) of $33-34^{\circ}/_{00}$. This is because the *Enhalus acoroides* sampling was carried out in the afternoon during the rainy season in May 2024, and there was a flow of fresh water from the Negeri Suli River which affected the salinity value. This is in line with the research results of **Sari** *et al.* (**2021**), who assessed that the distribution of salinity is influenced by several factors, namely water circulation, evaporation, rainfall, and river flow. *Enhalus acoroides* is a type of seagrass that can survive in a range of moderate to high salinity, so that a salinity of $30^{\circ}/_{00}$ can support relatively stable conditions for its growth. According to **Darui** *et al.* (**2024**), the salinity range that can be tolerated by seagrass is 10-40‰ and the optimum value is 35‰.

pH or acidity level has a big influence on aquatic plants and animals. The results of measuring the pH of seawater in the waters of Suli State were 6.8 below the quality standard set by the Government Regulation of the Republic of Indonesia Number 22 of 2021, with a range between 7-8.5. The pH value greatly affects the biochemical processes of water; at a pH range of <4.00, most aquatic plants will die because they cannot tolerate low pH (**Bongga** *et al.*, **2021**). **Salahuddin** *et al.* (**2022**) stated that low pH can occur due to the presence of dissolved organic matter and CO₂.

2. Extract yield of Enhalus acoroides

The liquid obtained was then evaporated to obtain a thick extract. The yield obtained in the study is depicted in Table (2).

Table 2. Extract/ yield of Enhalus acoroides					
Solvent	Extract (gr)	Yield (%)			
Methanol	7.52	15.04			
Ethyl Acetate	6.13	12.26			

Table (2) shows that the yield value in methanol solvent was higher (15.04%) than in ethyl acetate solvent (12.26%). **Ruhardi and Sahumena (2021)**, in their study, explained that the yield of ethyl acetate solvent is smaller than that of methanol solvent due to the presence of methoxy groups in the chemical structure of ethyl acetate which causes hydrogen bonds to form with compounds in the sample. The high and low yields can be influenced by several factors such as differences in methods, temperatures, types of solvents, and extraction times (**Novitasari & Jubaidah, 2018**). This was also stated by **Salamah and Widyasari (2015)** showing that other factors that may affect the yield value produced are the extraction method used, sample particle size, storage conditions and time, extraction time, comparison of the number of samples to the number of solvents used and the type of solvent used. The solvent polarity is an important factor as it equaled 5.2 for methanol, while that of ethyl acetate was 4.4 in polarity index.

3. Identification of secondary metabolite compounds of Enhalus acoroides extract

Identification of secondary metabolite compound groups was carried out qualitatively using phytochemical tests to determine the active compound groups contained in the *Enhalus acoroides* extract. The results of the qualitative secondary metabolite compound test are illustrated in Table (3).

declate extracts of Enhances decrotates								
No.	Compound groups	Methanol extract		Ethyl acetate extract				
		Color / sediment formed	Results	Color / sediment formed	Results			
1	Flavonoids	Yellow	+	Yellow	+			
2	Saponins	Foamy green	+	Green no foam	-			
3	Phenolic	Blue	+	Blue	+			
4	Tannin	Green	+	Green	+			
5	Steroid	Deep blue	+	Deep blue	+			
6	Terpenoid	Chocolate	+	Chocolate	+			
7	Alkaloid	Foamy green	+	Green no foam	-			

Table 3. Results of secondary metabolite compound content tests of methanol and ethyl

 acetate extracts of *Enhalus acoroides*

The results of the phytochemical test in Table (3) show that the methanol extract of *Enhalus acoroides* contains flavonoids, saponins, phenolics, tannins, steroids, terpenoids

and alkaloids, while the ethyl acetate extract of *Enhalus acoroides* contains flavonoids, phenolics, tannins, steroids and terpenoids. The results of this study differ from the findings of **Nurafni and Nur (2018)**, who also studied secondary metabolite compounds in methanol extracts of *Enhalus acoroides* in the waters of Morotai Island. In their study, the compounds found included alkaloids, flavonoids, saponins, and steroids. This difference indicates a variation in the results of the study related to the types of compounds identified in the *Enhalus acoroides* extract. Table (3) also shows that the ethyl acetate extract of *Enhalus acoroides* contains flavonoids, phenolics, tannins, steroids, and terpenoids. However, these results differ from research by **Antarsih (2017)**, which also tested ethyl acetate extract from *Enhalus acoroides* leaves and found compounds including saponins, tannins, alkaloids, steroids, and glycosides.

The difference in results between this study and the research of Antarsih (2017) and Nurafni and Nur (2018) may be due to several factors; one possibility is the difference in the extraction method used. According to Tjandrawinata and Nurkolis (2024), the extraction method can affect the type and concentration of compounds successfully extracted from Enhalus acoroides samples. This is in line with what was stated by Amin et al. (2023) that the extraction method used to obtain bioactive compounds from Enhalus acoroides involves techniques such as maceration with different solvents, viz. methanol, ethyl acetate, and n-hexane. Chemat et al. (2017) added that condition testing, such as temperature, pH, or extraction duration, may also contribute to differences in results. Another factor to consider is the difference in physical and chemical conditions of the waters where Enhalus acoroides grows. This was also confirmed by Fredley et al. (2019) elucidating that conditions such as salinity, temperature, and water quality can affect the chemical composition of seagrass. Research by Suriani and Lukman (2020) verified that differences in chemical compound composition can occur because the characteristics of seagrass habitats can impact the content of secondary metabolites found in seagrass. Differences in compound content between methanol and ethyl acetate extracts may be due to differences in extraction methods and how these compounds interact with environmental conditions. Research results by Florensia and Wijaya (2023) explain that the use of different solvents can affect the optimal extraction of secondary metabolite compounds. This finding is in line with that of research conducted by Hartati et al. (2019), which states that differences in phytochemical screening results can also be caused by differences in the sensitivity of the extraction method used to the amount of chemical content of the natural materials tested.

CONCLUSION

It can be concluded that the sea water temperature in the waters of Suli State was recorded to be 30°C in the afternoon, with the dissolved oxygen content at 6.1 mg/ L, the salinity at $30^{\circ}/_{\circ\circ}$ and the pH at 6.8. The methanol extract of *Enhalus acoroides* contains flavonoids, saponins, phenolics, tannins, steroids, terpenoids and alkaloids, while the

ethyl acetate extract of *Enhalus acoroides* contains flavonoids, phenolics, tannins, steroids and terpenoids. Remarkably, environmental conditions and the use of different types of solvents can affect the composition of the secondary metabolite compounds produced.

ACKNOWLEDGEMENT

This research is part of projects titled "Korea-Indonesia Marine Technology Cooperation Research Center (20220512)" and "Ocean and Coastal Basic Survey and Capacity Enhancement in Cirebon, Indonesia (PG53340)" which are funded by the Ministry of Oceans and Fisheries, Korea.

REFERENCES

- Abraham, J.P.; Baringer, M.; Bindoff, N.L.; Boyer, T.; Cheng, L.J.; Church, J.A. and Willis, J.K. (2013). A review of global ocean temperature observations: Implications for ocean heat content estimates and climate change. Reviews of Geophysics., 51(3): 450-483.
- Akbari, E.; Alavipanah, S.K.; Jeihouni, M.; Hajeb, M.; Haase, D. and Alavipanah,
 S. (2017). A review of ocean/sea subsurface water temperature studies from remote sensing and non-remote sensing methods. Water., 9(12): 936.
- Amin, A.A.; Brawijaya, U.; Amrillah A.M. and Brawijaya, U. (2023). Antibacterial test of *Enhalus acoroides* seagrass against *Salmonella typhi* bacteria. Journal of Environmental Engineering and Sustainable Technology., 10(2): 95-103.
- Antarsih, N.R. (2017). Effect of ethyl acetate extract of seagrass leaves [*Enhalus acoroides* (LF) ROYLE] on mda and gsh levels of old male mice. Journal of Health Research Cell., 4(2): 56-65.
- Artika, S.R.; Ambo-Rappe, R.; Teichberg, M.; Moreira-Saporiti, A. and Viana, I.G. (2020). Morphological and physiological responses of *Enhalus acoroides* seedlings under varying temperature and nutrient treatment. Frontiers in Marine Science., 7: 325.
- Asih, E.N.N. and Kartika, A.G.D. (2021). Potential and characteristics of soft coral symbiotic bacteria *Sinularia* sp. as an anti- *Escherichia coli* bacterium from the waters of Gili Labak Island, Madura, Indonesia. Journal of Marine Research., 10(3): 355-362.
- **Atun, S.** (2014). Methods of isolation and structural identification of organic compounds from natural materials. Journal of Cultural Heritage Conservation., 8(2): 53-61.
- Badriyah, L.; Asih, E.N.N.; Ni'amah, S.N.; Ningrum, R.H.; Mardiyanti, Y. and Wulansari, D.R. (2023a). Detection of indications of erythema in hand body lotion preparations from seagrass extract (*Enhalus acoroides*) and sea urchin gonads (*Diadema setosum*). Unram Fisheries Journal., 13(1): 299-306.

- Badriyah, L.; Asih, E.N.N.; Ni'amah, S.N.; Ningrum, R.H.; Mardiyanti, Y. and Wulansari, D.R. (2023b). Addition of seagrass extract (*Enhalus acoroides*) and sea urchin gonad (*Diadema setosum*) as a formulation of moisturizer body lotion. Indonesian Journal of Fisheries Product Processing., 26(1): 97-106.
- Bongga, M.; Sondak, C.F.; Kumampung, D.R.; Roeroe, K.A.; Tilaar, S.O. and Sangari, J. (2021). Study of the health conditions of seagrass meadows in Mokupa Waters, Tombariri District, Minahasa Regency. Journal of Tropical Coastal and Marine Affairs., 9(3): 44-54.
- Cambridge, M.L.; Zavala-Perez, A.; Cawthray, G.R.; Mondon, J. and Kendrick, G.A. (2017). Effects of high salinity from desalination brine on growth, photosynthesis, water relations and osmolyte concentrations of seagrass Posidonia australis. Marine Pollution Bulletin., 115(1-2): 252-260.
- Chemat, F.; Rombaut, N.; Sicaire, A.G.; Meullemiestre, A.; Fabiano-Tixier, A.S. and Abert-Vian, M. (2017). Ultrasound assisted extraction of food and natural products. Mechanisms, techniques, combinations, protocols and applications. A review. Ultrasonics sonochemistry., 34: 540-560.
- Darui, D.A.; Sondak, C.F.; Warouw, V.; Kemer, K.; Gerung, G.S. and Wagey, B.T. (2024). Condition of seagrass beds around the waters of Bowongkali Village, Central Tabukan District, Sangihe Islands Regency. Journal of Tropical Coastal and Marine Affairs., 12(1): 26-36.
- **Denny, B.** (2024). Molecular identification and potential of secondary antioxidant metabolites of endophytic fungi isolates from mangrove plants (*Rhizophora apiculata*). (Doctoral dissertation, Andalas University).
- **Dewatisari, W.F.** (2020). Comparison of chloroform and ethanol solvents on the yield of snake plant leaf extract (*Sansevieria trifasciata* Prain) using the maceration method. In Proceedings of the National Biology Seminar., 6(1): 127-132.
- Florensia, S. and Wijaya, A. (2023). The effect of different solvents on the results of phytochemical screening of Tapak Liman leaf extract (*Elephantopus scaber* L.). Scientific Journal of Pharmacy Simplisia., 3(2): 128-134.
- Fredley, J.; Durako, M.J. and Hall, M.O. (2019). Multivariate analyzes link macrophyte and water quality indicators to seagrass die-off in Florida Bay. Ecological Indicators., 101: 692-701.
- Friedrich, J.; Janssen, F., Aleynik, D.; Bange, H.W.; Boltacheva, N.; Çagatay, M.N. and Wenzhöfer, F. (2014). Investigating hypoxia in aquatic environments: diverse approaches to addressing a complex phenomenon. Biogeosciences., 11(4): 1215-1259.
- Hartati, Syamsuddin, B. and Karim, H. (2019). The effect of solvent type on the content of secondary metabolite compounds of Javanese Wood Click (*Lannea coromendelica*). Jurnal Sainsmat., 8(2): 19-27.

- Hitijahubessy, H.; Susiyanto, A.Y.; Samid, A. and Cesar, O. (2021). The effect of seagrass extract *Enhalus acoroides* in vitro as an antibacterial against *Vibrio* sp. which causes ice-ice disease in *Eucheuma cottoni* seaweed. Molluca Journal of Chemistry Education (MJoCE)., 11(2): 93-98.
- Kaihena, M.; Mas'uth Pratomo, M.S. and Ukratalo, A.M. (2022). Isolation and identification of secondary metabolite compounds of Laor worm extract as antibacterial. International Journal of Social Services and Research., 2(5): 441-452.
- Kartika, A.G.D.; Asih, E.N.N.; Nuzula, N.I. and Dewi, K. (2023). Counseling on the introduction of marine biota and the environment at SDN 61 Gresik-East Java. Journal of Community Service Sakai Sambayan., 7(3): 169-174.
- Novitasari, N. and Jubaidah, S. (2018). Comparison of extraction methods on the yield of sea rambai leaf extract (*Sonneratia caseolaris* L. Engl). Manuntung Scientific Journal., 4(1): 79-83.
- Nurafni, N. and Nur, R.M. (2018). Antifouling activity of bioactive compounds from seagrass in Morotai Island Waters. Journal of Archipelago Marine Science., 1(2): 107-112.
- Nurfitriani, E.; Mulyani, Y. and Agung, M.U.K. (2017). Relationship between water quality and secondary metabolite profile of *Holohuria atra* meat extract in Lampung Bay and Garut Waters. Indonesian Aquatics., 2(2): 146-154.
- Oktawati, N.O.; Sulistianto, E.; Fahrizal, W. and Maryanto, F. (2018). Economic value of seagrass ecosystem in Bontang City. EnviroScienteae., 14(3): 228-236.
- Permana, R.; Aulia, A.; Nora, A. and Pringgo, K.D.N.Y.P. (2020). Identification of bioactive compounds and potential antioxidant activity of seagrass *Enhalus* acoroides (Linn. F). Jurnal Aquatek., 1(1): 66-72.
- Petersen, L.E.; Kellermann, M.Y. and Schupp, P.J. (2020). Secondary metabolites of marine microbes: From natural products chemistry to chemical ecology. In Youmares 9-The Oceans: Our Research, Our Future: Proceedings of the 2018 conference for Young Marine Research in Oldenburg, Germany (pp. 159-180). Springer International Publishing.
- Ramadhan, R.; Dirgayusa, I.G.N.P. and Faiqoh, E. (2021). Phytochemical screening and toxicity test of seagrass leaf extract *Halophila ovalis* in Karma Beach and Serangan Beach, Bali. Journal of Marine and Aquatic Sciences., 7(2): 140-147.
- Ruhardi, A. and Sahumena, M.H. (2021). Identification of flavonoid compounds in sembung leaves (*Blumea balsamifera* L.). Journal Syifa Sciences and Clinical Research (JSSCR)., 3(1): 29-36.
- Sahertian, D.E. and Wakano, D. (2017). The growth rate of *Enhalus acoroides* leaves on different substrates in the coastal waters of Poka Village, Ambon Island.

BIOSEL (Biology Science and Education): Journal of Science and Education Research., 6(1): 61-68.

- Salahuddin, S.; Apriadi, T. and Muzammil, W. (2022). Growth of seagrass *Enhalus acoroides* in the waters of Pangkil Village, Teluk Bintan District, Bintan Regency. Indonesian Journal of Marine Science and Technology., 15(1): 31-38.
- Salamah, N. and Widyasari, E. (2015). Antioxidant activity of methanol extract of longan leaves (*Euphoria longan* (L) Steud.) with 2, 2'-diphenyl-1picrylhydrazyl radical scavenging method. Pharmaciana., 5(1): 25-34.
- Sari, R.M.; Kurniawan, D. and Sabriyati, D. (2021). Density and distribution patterns of seagrass based on community activities in Pengujan Waters, Bintan Regency. Journal of Marine Research., 10(4): 527-534.
- Suriani, S. and Lukman, M. (2020). Relationship between physical and chemical environmental factors and antioxidant activity of seagrass seeds *Enhalus* acoroides in Jikumerasa Village, Buru Regency, Maluku Province. KASA Journal., 1(1): 28-35.
- **Tangke, U.** (2010). Seagrass ecosystem (benefits, functions and rehabilitation). Agrikan: Journal of Fisheries Agribusiness., 3(1): 9-29.
- Tjandrawinata, R.R. and Nurkolis, F. (2024). A comparative analysis on the impact of extraction methods on carotenoids composition, antioxidants, antidiabetic, and antiobesity properties in Seagrass *Enhalus acoroides*: In Silico and In Vitro Study. Marine Drugs, 22(8): 365.
- Ukratalo, A.M.; Kakisina, P. and Mailoa, M.N. (2023). The effect of Eucheuma cottonii extract on body weight and blood sugar levels of Mouse (Mus musculus) Diabetes Mellitus Type 1. Journal of Tropical Biology., 23(3): 554-563.
- Windyaswari, A.S.; Elfahmi, E.; Faramayuda, F.; Riyanti, S.; Luthfi, O.M.; Ayu, I.P. and Magfirah, R. (2019). Phytochemical profile of sea lettuce (*Ulva lactuca*) and filamentous micro algae (*Spirogyra* sp.) as potential marine natural materials from Indonesian waters. Kartika: Pharmaceutical Scientific Journal., 7(2): 88-101.
- Yang, L.; Wen, K.S.; Ruan, X.; Zhao, Y.X.; Wei, F. and Wang, Q. (2018). Response of plant secondary metabolites to environmental factors. Molecules., 23(4): 762.