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ENTEROPATHOGENIC ESCHERICHIA COLI (EEC) IN SLAUGHTERED CATTLE AT A MODERN ABATTOIR

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Turns (eliterative) INTRODUCTION (foodifo) asses cals

The main objectives of establishing modern abattoirs with appropriate technology are the hygienic slaughtering and handling of meat, thus preventing the spread of zoonotic and meat borne diseases in both humans and animals, whilest keeping the environment free from pathogens of animal origin, and to achieve increased and improved meat production and handling.

characterized by goor indivinal bysicae (Minighay and

In Egypt, no studies till now are done are done on these newly built modern abattoirs to assess the effect of modernization on the presence of E.coli with special reference to enteropathogenic types. Therefore, one of the newly built abattoirs in the urban areas was selected and beef carcases are examined for surface contaminations with such microorganisms. Moreover, some factors in the abattoir as hands, clothes and transport in open cars which can contribute to E.coli contamination were also investigated.

The native habitat for E.coli is the enteric tract of man and animals, thus its presence in foods generally indicates direct or indirect pollution of faecal origin. E.coli is the classical indicator of the possible presence of enteric pathogenes in foods. The presence of E. coli in a meat does not denote directly

the presence of a pathogen, but only implies a certain risk that it may be present. In other words, the presence of E.coli in meat is not always closely correlated with the occurrence of salmonellae or other pathogenic microorganisms. E.coli in food have been associated with outbreaks of gastrointestinal diseases since 1903 (DELEPINE, 1903).

Definite E.coli serotypes are capable of causing various acute intestinal diseases in humans. Certain serotypes are responsible of colienteritis in children. They cause diseases in infants of the first months of life and in older infants; while others cause dysentery like diseases or cholera like diarrhoea. They produce also thermolabile and thermoresistant enterotoxine. Ecoli may also cause Colibacillosis in adults (peritonitis, meningitis, enteritis, toxinfections, cystitis, pyelitis, pyelonephritis, angiocholitis, salpingocophoritis, appendicitis, otitis and pureperal sepsis), PYATKIN and KRIVOSHEIN (1980).

E.coliois also judged to be a significant pathogen in casess of traveller's diarrhoea and in gastrointestinal illness in developing nations and other localized areas characterized by poor indiviual hygiene (MEHLMAN and ROMERO, 1982). Is such as the local areas of the local areas characterized by poor individual hygiene (MEHLMAN and ROMERO, 1982).

Several identifiable strains of E.coli have been known for many years to cause infantile diarrhoea (NETER et al., 1951; TAYLOR & CHARTER, 1952 and EWING et al., 1957 & 1963). In recent years it has become increasingly evident that E.coli strains also produce illness to a significant degree in adults (SAKAZAKI et al., 1974; WHO 1974 a; ANON, 1978 and SACK et al., 1986). There are two forms of the disease which differ clinically from each other to some degree (SOJKA, 1973). The first, caused by toxigenic strains, is characterized by excessive loss of fluid from profuse diarrhoea (the choleralike syndrome). The second frorm, caused by invasive strains, produces a syndrome closely resembling dysentry (the dysentry like syndrome). There are also forms of the disease in which these two clinical pictures are

intermixed. The incubation period is variable and short 6-36 hours (CDC, 1973 b), and tends to be shorter for invasive than for toxin producing strains (DUPONT et al., 1971). The duration of the disease is also generally short, 24 hours to few days, but it can be occasionly much longer. There are epidemics on record with a median duration of 9 days and a maximum of 77 days (CDC, 1973 a). Epidemics are particularly prove to occur in closed institutions for infants such as nurseries and maternity wards.

It has recently been convincingly demonstrated in volunteers that there sxist both invasive and enterotoxin producing strains, and strains combining these two capacities (DUPONT et al., 1971).

GLYES & BARNUM (1969) and SMITH & GYLES (1970) stated that there are two kinds of toxins; one is thermostable and the other is thermolabile. Preliminary immunological studies furnish evidence that the heatlabile toxins from different E.coli strains share antigenic determinants and also cross react with the cholera toxin (HOLMGREN et al., 1973).

MATSIEVSKII et al., (1971) found that an acute outbreak of food poisoning caused by E.coli O 124 was observed among 198 children and 90 personnel members of a children's sanatorium. In 1982, BRYAN stated that the serogroups of E.coli that have caused the invasive type illness are 0₂₅, 0₂₈, 0₁₁₂, 0₁₂₄, 0₁₃₆, 0₁₄₃, 0₁₄₇, 0₁₄₄ and 0₅₁₂. while those that have been shown to elaborate enterotoxins are: 0₆, 0₁₅, 0₁₈, 0₂₀, 0₂₇, 0₄₄, 0₅₅, 0₇₈, 0₁₁₁, 0₁₁₄, 0₁₁₉, 0₁₂₅, 0₁₂₆, 0₁₂₇, 0₁₂₈, 0₁₄₂, 0₁₄₈, 0₁₅₄, 0₁₅₅ and 0₁₅₆. He stated that the incubation period of the disease (E.coli Diarrhoeas) was 8-24 hours, mean 11 hours for invasive type.

The symptoms are fever, chills, headache, myalgia, abdominal cramps, profuse watery diarrhoea, it is similar to Shigellosis. In the enterotoxigenic type the incubation period was 8-44 hours with a mean 26 hours. The symptoms are diarrhoea (rice water stools), vomiting, dehydration, shock. It is similar to cholera. Recently, FOSTER (1987) stated that another type, E.coli 0

H₇ cause a very serious hemorrhagic colitis, or bloody diarrhoea, that sometimes spreads into the urinary tract and causes hemolytic uremic syndrome. This organism resiudues in the animal population and sometimes appears in raw meat or milk.

The aim of the present study is to investigate the sanitary status of a modern abattoir by detection of EEC and its serotypes existing on the surface of beef carcases and meat contact surfaces.

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MATERIAL AND METHODS

One hundred and fifty swabs were taken from the surface of beef carcases and meat contact surfaces at a modern abattoir for monitoring the enteropathogenic E.coli.

The swabs were taken from the surface of twenty carcases at two sites (shoulder and thigh) directly after skinning and after preparation as well.

Moreover, other ten carcases were also swabbed twice at the same sites; one directly after preparation and stamping as well as after delivery in the open cars used for distribution of carcases to butchers in the city.

To investigate contaminants on the meat contact surfaces, thirty swabs were taken; ten from each of right hand of meat handlers, clothes of the workers and inner surface of the open cars used for meat distribution.

The Enteropathogenic E.coli isolation was carried out according to ICMSF (1978). The isolated E.coli colonies were identified biochemically using the Enterotube II "Roche". The isolates were identified serologically by using Diagnostic sera, "Wellcome E.coli agglutination sera for diagnosis the Enteropathogenic types".

RESULTS AND DISCUSSION

From the results achieved in Table (1) it was found that E.coli was isolated from 20% of shoulder samples after skinning and from 55% of samples after preparation. IT was also found in 20% of thigh samples after skinning and 30% after preparation. E.coli could be also detected in 70% of shoulder samples and 40% of thigh samples after preparation and stamping while after delivery in cars from 50% of shoulder and 40% of thigh samples. Isolated E.coli from cars were more than those from hands and clothes of workers.

Table (1): Isolated E.coli from carcases and meat contact surfaces

Site	Sample	Isolates		
		No.	bns Ac % Steeps	ng his parte
Shoulder	A	and 4 tlls	20	55 n
	B	11 pr	oduce55 trea	SE I
	C	7	70	18.0
lety i	D	5	a (City) same	F12 13
Thigh	(A) (A	4	20	getq Les 1
unn (Seithe e	В	6	30	
11 14 11 11	C	4	40	
is white	D	4	40	
Hands Clothes Cars		1	10 10 40	

A = After skinning

B = After preparation

C = After preparation and stamping

D = After delivery in cars

Certain serotypes of E.coli have a primary pathogenicity in the intestine and cause gastroenteritis, mainly in infants. Most outbreaks take place in infants under 18 months, though many cases are in children up to 5 years old some infections cause diarrhoea in adults.

Table (2): Serotypes of isolated Enteropathogenic E.coli from surface of beef carcases and meat contact surfaces.

le 70% and to lear	Seroty	No. of	
Samples	poly valent	nds and clothes	
Shoulderons seasoass	il from	lsolated E.pc	Table (1)
-After skinning	2 2	0 ₅₅ : K ₅₉ (B 5)	12
-After preparation	2	0 ₁₁₁ : K ₅₈ (B 4)	2
-After preparation and stamping -After delivery in car	2	0 ₁₁₉ : K ₆₉ (B14)	l l Nil Shoulder
Thigh	ir		stoph Lwise
-After skinning -After preparation -After preparation and		- p	Nil Nil
stamping	-	- 1	Nil
-After delivery in car	2	0 ₁₁₉ : K ₆₉ (B14)	ot act 53irT
Hands Clothes	2	0 ₂₆ : K ₆₀ (B 6)	of right of and linear Nil
Cars	2	0 ₅₅ : K ₅₉ (B 5)	9,10

The two serotypes, alpha (BRAY, 1945) and beta (SMITH, 1949), first shown to be the cause of outbreaks of infantile diarrhoea were later typed as $^{
m O}_{111}$: $^{
m K}_{58}$ (B 4)

and O_{55} : K_{59} (B 5). The biotype Dyspepsikoli A IV and Dyspepsikoli A I which were identified as cause of infantile gastroenteritis by ADAM (1927) were also later typed as O_{111} : K_{58} and O_{55} : K_{59} respectively. Such serotypes were isolated from the shoulder of beef carcases after skinning and after preparation and from cars used for transport of meat from the abattoir to butcher's shops.

Further serotypes have since been found in cases and outbreaks of infection, e.g. in 0-group 26, 44, 86, 112, 119, 124, 125, 126, 127, 128, 142 and 158 (ROWE et al., 1974) and 0₁₅₇ (FOSTER, 1987). Not all strains in these enteropathogenic serotypes are able to cause gastroenitis. The particular strains that cause gastroenteritis are distinguished from others in the same serotype by their ability to form an enterotoxin demonstrable by the production of fluid secertion and distension in ligated loops of intestine in rabbits; most strains of the same sertypes found in symptomless excretors do not form enterotoxin (TAYLOR et al., 1961; SMITY & HALLS, 1967 and SACK et al., 1971). The power to produce enterotoxin can be acquired by a non toxigenic strain by the receipt of a plasmid transmitted on conjugation from a toxigenic strain (SMITH and HALLS, 1968).

The enteropathogenic E.coli strains produce disease not only in man but also in a number of domesticated animals, such as calves, pigs, poultry and lambs (COOKE, 1974). A variety of foods have been implicated in outbreaks: stewed meat and gravy (KORETSKAIA and KOVALEVSKAIA, 1958); roast mutton (COSTIN et al., 1964); pork and chicken (SMITH et al., 1965) and ham and pie (WHO, 1974 b). The detection of even low numbers of Enteropathogenic E.coli in food, particulary in baby foods, reveals a public health hazard as significant as the demonstration

of Salmonellae in such foods (ICMSE, 1978).

The sanitary importance of Enteropathogenic E.coli has been emphasized by many other authors as it has been implicated in human cases of gastroenteritis, epidemic diarrhoea in infants, sporadic summer diarrhoea in children as well as in cases of food poisoning (FERGUSON & JUNE, 1952; JUNE et al., 1953; ROGERS & CARACKELL, 1956; EWING& DAIRS, 1961; MACKIE & MACCARTNEY, 1962; EWING. 1963; BODILY et al., 1970; ETKIN & GORBACH, 1971; MATSIEVSKII et al., 1971; LOVE et al., 1972; CRUICKSHANK et al., 1975; MARIER et al., 1973; SINGH & RANGNATHAN. 1974 & 1977; MOSSEL, 1975; KORNACKI & MARTH, 1982; KRIEG & HOLT, 1984 and FOSTER, 1987). Moreover, organism was the most frequent cause cystitis, pyelitis, pyelonephritis, appendicitis and peritonitis (GRONROSS et al., 1955; LEPPANEN, 1958 & 1959; SMITH & CONANT, 1960; RYATKIN, 1967 and MOSSEL, 1975).

SUMMARY

One hundred and fifty swabs were taken from the surface of beef carcases and meat contact surfaces at a modern abattoir for monitoring the EEC. Moreover, other swabs were taken from hands of meat handlers, clothes and inner surfaces of open cars used for meat distribution.

nost strains of

E.coli was isolated more oftenly from the shoulder than from the thigh and from cars more than from hands and clothes of workers.

Serotypes of EEC from surface of beef carcases were 055: K 59 (B 5) from shoulder after skinning and 0111: K 58 (B 4) after preparation and 0 119: K 69 (B 14) from shoulder after preparation and stamping and from thigh after delivery in cars. EEC serotype 026: K 60 (B 6) was isolated from hands and 0 55: K 59 (B 5) from cars.

Public health significance of E.coli and isolated serotypes was discussed.

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