

#### VOL. 64, 2131:2140, JULY, 2018

I.S.S.N 0070-9484



#### Oral Medicine, X-Ray, Oral Biology and Oral Pathology

www.eda-egypt.org • Codex: 32/1807

## THE POTENTIAL ROLE OF CD31 AND BCL-2 IN THE ETIOPATHOGENESIS OF ORAL PYOGENIC GRANULOMA

Naglaa M. Kamal\*

#### **ABSTRACT**

Pyogenic granuloma (PG) is a vascular reactive lesion of the oral cavity which is caused by local trauma or irritation and has a distinct biological behavior. Up till now, the pathogenesis of PG is not adequately understood.

**Aim:** Assessment of the expression of CD31 and Bcl-2 in PG to clarify the etiopathogenesis of this lesion and to determine whether vascular proliferation in this lesion has a relation to its apoptotic status.

**Materials and Methods:**  $4\mu m$  sections were prepared from paraffin-embedded blocks of PG (20 samples) and normal gingiva as a control (10 samples). Staining of each specimen with hematoxylin and eosin (H&E), CD31 and Bcl-2 was done.

**Results:** all lesions of PG showed positive CD31 and Bcl-2 immunostaining in the endothelial cells of blood vessels and in the stromal ovoid cells. Control group showed positive CD31expression in the endothelial cells of blood vessels while Bcl-2 expression revealed positive expression mainly in the basal cells of the epithelium. A significant higher expression of CD31 and Bcl-2 was detected in PG than normal gingiva. A strong positive correlation was found between CD31 and Bcl-2 area percent of immune-expression.

**Conclusions:** The data of the present study suggest a role of Bcl-2 overexpression in the potentiation of angiogenic response and support the role of CD31 and Bcl-2 in the etiopathogenesis of PG.

Keywords: Pyogenic Granuloma, CD31, Bcl-2, Immunohistochemistry

#### INTRODUCTION

Pyogenic granuloma (PG) is a relatively common reactive lesion which mostly affects the oral mucosa and skin. The term pyogenic granuloma was first introduced by Hartzell in 1904 and is still being used

to represent this lesion.<sup>1</sup> But this term is confusing as neither it is caused by bacterial infection nor does it contain any pus. In addition, there is no granuloma formation histopathologically.<sup>2</sup> It is considered as an inflammatory hyperplasia.<sup>3</sup>

<sup>\*</sup> Lecturer, Oral Pathology Department, Faculty of Oral and Dental Medicine, Ahram Canadian University, 6th of October City, Giza Egypt.

(2132) E.D.J. Vol. 64, No. 3 Naglaa M. Kamal

PG arises as a painless nodular mass which may be rapidly growing in nature but rarely exceeding 2.5 cm.<sup>4</sup> It may have a smooth or lobulated surface and a sessile or pedunculated base. The lesion appears red due to the presence of numerous capillaries. Bleeding is a common finding.<sup>5,6,7</sup> Marked histopathological finding in pyogenic granulomas is a prominent capillary growth within a hyperplastic granulation reaction. A mixed inflammatory cell infiltrate of neutrophils, plasma cells, and lymphocytes is evident.<sup>4,8</sup>

A possible correlation has been found between PG and chronic physical irritations. These stimuli initiate the healing and repair response and may lead to inappropriate regulation of inflammatory reaction, hyperplasia, angiogenesis and hence, improper formation of granulation tissue which is the histopathological hallmark of PG. Though, PG is considered as a reactive lesion rather than a neoplastic one. Depending on these observations, it becomes obvious that pathogenesis of PG could be related to either inflammatory response or angiogenesis.<sup>4,9</sup> The most common etiologic factors are poor oral hygiene with accumulated plaque and calculus in the gingival crevice and overhanging restorations.<sup>10,11</sup> Hormonal disturbances could be also considered precipitating factors for pyogenic granuloma. The gingiva is the most frequently affected site. 12,13 Buccal mucosa, tongue and lips could also be affected. Incomplete surgical removal, failure of removing irritating factors and repeated trauma are considered reasons for recurrence.<sup>10</sup>

The pathogenesis of PG is not well established. Owing to the clinical behavior of this lesion as rapid growth rate and repeated recurrence some investigators consider it as a benign neoplasm but it is believed to be an inflammatory reactive lesion caused by different stimuli.<sup>3,14,15</sup>

The marked growth of capillaries in PGs indicates a potent angiogenic action (formation of new blood vessels) which may play a role in their

etiopathogenesis.<sup>8,9,16,17</sup> Detection of blood vessels could be achieved by the utilization of antibodies that have an affinity for specific epitopes on the surface of endothelial cells as CD34, CD31 and factor VIII. <sup>18</sup>

CD31 or Platelet-endothelial cell adhesion molecule (PECAM-1) is a signaling receptor which is significantly expressed on the surface of the adult and embryonic endothelial cells. This molecule is a transmembrane glycoprotein that has a cytoplasmic domain. <sup>19,20</sup> CD31 is essential for angiogenesis due to its significant role in the adhesion between endothelial cells<sup>21</sup> and is considered one of the greatest valuable markers that detect endothelial cells.<sup>22</sup>

Apoptosis or programmed cell death is a physiologic normal function which maintains tissue homeostasis.<sup>23</sup> Consequently, disturbances in the function of the apoptotic system may lead to prolonged survival or excessive removal of cells and thus pathogenesis of different diseases. Various stimuli can modulate apoptosis such as growth factors, hormonal changes, cytokines, immunological responses and viral or bacterial infections.<sup>24</sup>

Bcl-2 protein is regarded as an apoptosis suppressor. It is a component of the outer mitochondrial membrane and a part of the endoplasmic reticulum.<sup>25</sup> This protein can interrupt apoptosis not only in the initial phase but also in the final one. That is because Bcl-2 stabilizes the mitochondrial membrane potential as well as it inhibits intracellular acidification and oxygen-reactive species formation.<sup>26</sup>

In the current work, the expression of CD31 and Bcl-2 in PG and normal gingiva (control) was investigated using immunohistochemical staining. The possible correlation between both markers was statistically analyzed. The aim is to gain a better understanding of the etiopathogenesis of PG that may have an impact on its biological behavior as

well as to determine whether vascular proliferation in this lesion has a relation to its apoptotic status.

#### MATERIALS AND METHODS

A total of twenty archival paraffin blocks of oral pyogenic granuloma were retrieved from:

- The pathological files of General Pathology Department, Nasser Institute for Research and Treatment, Ministry of health and population.
- The pathological files of General Pathology Department, Al Hussein Hospital, Al Azhar university.

Normal gingiva samples from 10 patients free of any systemic disorders were used as controls. Samples were taken after tooth extraction for orthodontic reasons. They were fixed in formalin and embedded in paraffin.

#### **Section preparation**

Three sections from each formalin-fixed, paraffin-embedded tissue block were cut into 4  $\mu m$  thickness and stained with the following:

- a) First section stained with hematoxylin and eosin to confirm the diagnosis of pyogenic granuloma.
- b) Second section stained with CD31 polyclonal antibody.
- c) Third section stained with Bcl-2 monoclonal antibody.

#### Immunohistochemical (IHC) staining procedure

The sections were deparaffinized with xylene and rehydrated in graded ethanol for IHC staining with CD31 and Bcl-2 antibodies. Heat mediated antigen retrieval was done using citrate buffer PH (6.0), then the sections were immersed in hydrogen peroxide (H2O2) to block the endogenous peroxidase activity, washed in phosphate-buffered saline (PBS), and then protein blocking reagent was added and incubated for 20 minutes at 37°C within

humid chamber to reduce the non-specific staining. The primary antibodies used in the present study were as follows:

- Concentrated polyclonal rabbit antibody for CD31 (Code No. PA5-16301 at dilution 1:50, Thermo Fisher Scientific USA).
- Concentrated monoclonal mouse antibody for Bcl-2 (code No. M0887 at dilution 1:50, Dako, Denmark).

Sections were incubated with the primary antibody overnight.

The sections were then washed twice in PBS and treated with the labelled streptavidin biotin complex (LSAB + System-HRP, Dako) at room temperature to bind the primary antibodies. Peroxidase activity was visualized by immersing the tissue sections in diaminobenzidine (Liquid DAB+ Substrate, Dako), which resulted in a brown reaction product. Finally, the sections were counterstained with Mayer's hematoxylin and cover-slipped.

#### **Immunohistochemical Interpretation**

The immunoexpression of Bcl-2 was evaluated by presence of brown colored immunostaining reaction in the cytoplasm of the endothelial cells and stromal cells of the connective tissue, and in the basal cells of the epithelium.

Presence of brown colored immunostaining reaction in the cytoplasm and cell membrane of endothelial cells and stromal connective tissue cells was considered a positive reaction for CD31.

In each slide, 5 microscopic fields showing the highest immunopositivity were selected and photomicrographed using image analyzer.

Immunoreactivity, for CD 31 and Bcl-2 was assessed by estimating the area percentage of positive immunostained cells in relation to the area examined in each microscopic field using computerized image

(2134) E.D.J. Vol. 64, No. 3 Naglaa M. Kamal

analyzer (Leica Qwin - Germany) at research unit (Faculty of Dentistry Cairo University).

The image analyzer consisted of a colored video camera, colored monitor, and hard disk of *hp* personal computer connected to the microscope and controlled by Leica Qwin 500 software. The image analyzer was calibrated automatically to convert the measurement units (pixels) produced by the image analyzer program into actual micrometer units. The area and area percentage reaction were measured using a magnification x200. Mean values were then obtained for each specimen.

#### Statistical analysis

Statistical analysis was then performed using a commercially available software program (SPSS 19; SPSS, Chicago, IL, USA).

As data was parametric, significance of the difference between both groups was evaluated using unpaired t test.

Pearson correlation test was used to study correlation between different parameters. The Pearson correlation coefficient is used to measure the strength of a linear association between two variables, where the value r=1 means a perfect positive correlation and the value r=-1 means a perfect negative correlation. The level of significance was set at P < 0.05.

#### **RESULTS**

#### 1-Heamatoxylin and eosin stain findings

Histologically PG revealed a hyperplastic keratinized stratified squamous epithelium. The underlying connective tissue stroma revealed collagen fibers with numerous dilated capillaries engorged with red blood cells, inflammatory cell infiltration and plump ovoid endothelial cells.

For the control (normal gingival tissue), the

epithelium displayed a keratinized stratified squamous epithelium that covers a core of connective tissue. The epithelium revealed a normal arrangement of its layers (6–8 layers). The basement membrane was flat with no extended rete ridges. The underlying connective tissue revealed normal arrangement of its fibers, blood vessels, and few chronic inflammatory cells (Figure 1, A, B).

#### **Immunohistochemical findings**

#### CD31 immune-reactivity:

All lesions of PG showed CD31 positive immunereactivity in the endothelial cells of blood vessels and in the stromal ovoid cells. The expression was cytoplasmic and membranous.

For normal gingival tissues, all specimens showed staining of endothelial cells of blood vessels (figure 1, C, D).

#### **Bcl-2** immune-reactivity:

All lesions of PG showed cytoplasmic immunopositivity for Bcl-2 in the endothelial cells of blood vessels and in the stromal cells. Normal gingival tissues showed Bcl-2 expression mainly in the basal cells of the epithelium. (figure 1, E, F).

#### **Results of the immunohistochemical staining:**

# 1. Comparison between the expression of Bcl-2 and CD31 in pyogenic granuloma and normal gingiva

Regarding Bc1-2 area percent of immunoexpression, a higher mean value was recorded in pyogenic granuloma than the control group with a statistically significant difference (p<0.0001), (Table 1, Figure 2)

Regarding CD31 area percent of immunoexpression, a higher mean value was recorded in pyogenic granuloma than the control group with a statistically significant difference (p<0.0001), (Table1, Figure 2)

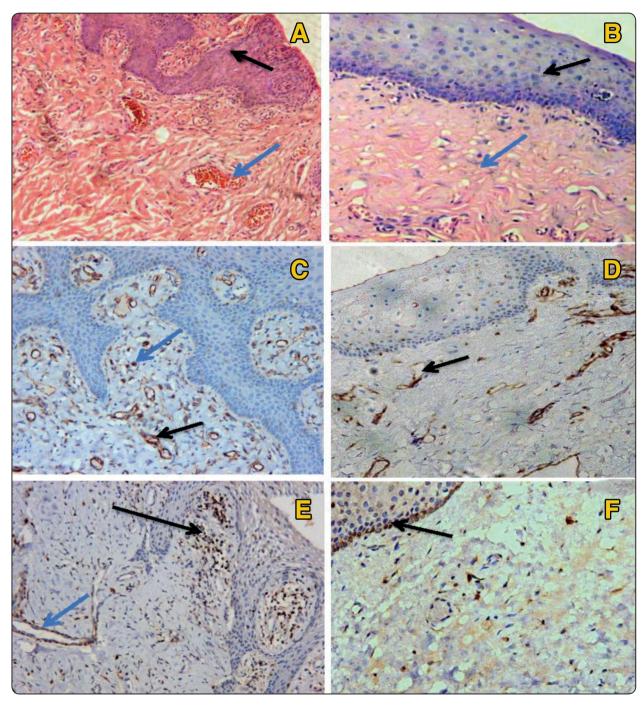


Fig. 1 (A): A photomicrograph of PG showing a hyperplastic keratinized stratified squamous epithelium (black arrow). The underlying connective tissue stroma revealed numerous dilated capillaries engorged with red blood cells (blue arrow) (H&E x100). (B): A photomicrograph of normal gingival tissue (control) showing a keratinized stratified squamous epithelium (black arrow) covering a core of connective tissue (blue arrow). The epithelium reveals normal arrangement of its layers (H&E x100). (C): A photomicrograph of PG showing immunopositivity for CD31 in the endothelial cells of the blood vessels (black arrow) and in the stromal ovoid cells (blue arrow) (CD31 x200). (D): A photomicrograph of normal oral epithelium showing CD31 expression in the endothelial cells of blood vessels (black arrow) (CD31 x100). (E): A photomicrograph of PG showing immunopositivity for Bcl-2 in the endothelial cells of blood vessels (blue arrow) and in the stromal cells (black arrow) (Bcl-2 x200). (F): A photomicrograph of normal gingival tissue showing Bcl-2 expression mainly in the basal cells of the epithelium (black arrow) (Bcl-2 x200).

(2136) E.D.J. Vol. 64, No. 3 Naglaa M. Kamal

TABLE (1) Comparison between the expression of Bcl-2 and CD31 in pyogenic granuloma and normal gingiva (unpaired t test)

	bcl-2 Area percent		CD31 Area percent	
	Pyogenic	Normal	Pyogenic	Normal
	Granuloma	gingiva	granuloma	gingiva
Mean	19.719	0.752	5.302	0.61
Std				
Dev	1.795	0.363	0.73	0.225
Max	22.714	1.222	6.4	1.299
Min	17.974	0.333	4.92	0.119
Т	32.75		3.22	
P value	<0.0001*		<0.0001*	

Significance level p<0.05, \*significant, ns=non-significant

### 2- Correlation between expression of Bcl2 and CD31 in pyogenic granuloma.

Pearson's correlation test revealed a strong positive correlation between CD31 and Bcl-2 area percent of immunoexpression (Table 2, Figure 3).

TABLE (2) Correlation between Bcl-2 and CD31 area percent of immunoexpression in PG

R	$\mathbb{R}^2$	Interpretation	P value
0.9683	0.9375	strong positive	<0.0001* (significant)

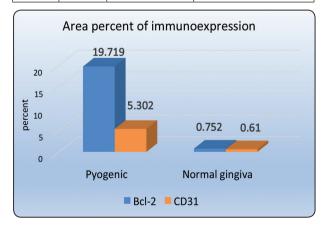


Fig. (2) Column chart showing mean Bcl-2 and CD31 area percent of immunoexpression in pyogenic granuloma and normal gingiva

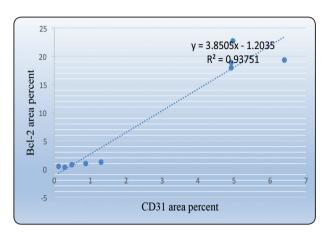


Fig. (3) Scatter plot showing correlation between CD31 and Bcl-2 area percent of immunoexpression in PG

#### DISCUSSION

Angiogenesis is controlled by the interaction between angiogenesis activators and inhibitors which are secreted by tumor cells and stromal cells. Inflammatory cells as macrophages and mast cells have an important role in angiogenesis and development of PG through the production of inflammatory mediators. Though the changes in the balance between angiogenesis stimulators and inhibitors can lead to the formation or regression of the blood vessels. <sup>27</sup>

The aim of the present study is to gain a better understanding of the etiopathogenesis of PG that may have an impact on its biological behavior as well as to determine whether vascular proliferation in this lesion is related to its apoptotic status.

PG may be regarded as an enormous expression of granulation tissue formation during an extended healing procedure after trauma or inflammation.<sup>28,29</sup> Local inflammation or trauma activates the release of cytokines (as angiogenic factors and basic fibroblast growth factor) from the macrophages and mast cells at the site of the injury. These cytokines stimulate angiogenesis process and neovascularization.<sup>30</sup>

CD31 immune-reactivity in the present study was detected in the endothelial cells of the blood vessels and in the ovoid stromal cells of PG. Similar

expression pattern was obtained in the previous studies of Saghafi et al., 2011, Freitas et al., 2005 and Rezvani et al., 2010.<sup>9, 31,32</sup> Normal gingival tissues revealed CD31 expression mainly in the endothelial cells of blood vessels which meet the results of the previous study of Kasprzak et al., 2012. <sup>33</sup>

The results of the current work showed a higher mean area percentage of CD31 immunoexpression in pyogenic granuloma in comparison with normal gingiva with a statistically significant difference that are in accordance with the results of the study of Seyedmajidi et al., 2015<sup>34</sup> and could be explained by the results of the study of Rezvani et al., 2010. They found CD31immunoreactivity in the lining endothelial cells and the proliferating stromal cells of PG and suggested that these stromal ovoid cells are mostly immature endothelial cells.<sup>32</sup> Hematopoietic stem cells or monocytic endothelial cell progenitors could be the origin of this immature cells in PG. The endothelial cells that derive from monocytes express endothelial cell markers including CD31.<sup>35</sup>

Moreover, the study of Blackwell et al., 2016 confirmed the presence of immature cells in PG. It was the first study that demonstrates embryonic stem cell (ESC) markers expression in PG and revealed the presence of hematopoietic stem cells in this lesion. They displayed the expression of the ESC markers SOX2, pSTAT3 and NANOG on the lining endothelial cells and the stromal endothelial cells.<sup>36</sup>

One of the regulators of endothelial cell progenitors is vascular endothelial growth factor (VEGF). It can stimulate the transformation of the progenitor cells to endothelial cells (both hematopoietic stem cell and monocytic endothelial cell progenitors). VEGF mediated this action by increasing both cell differentiation and proliferation.<sup>37</sup> It also acts as a chemotactic factor for endothelial cell progenitors and stimulates vascular permeability, assisting the entrance and migration of progenitor cells at the injured site.<sup>38,39</sup>

The most common cells that synthesize and secrete VEGF are macrophages, fibroblasts, mast cells and epithelial cells. Their role is manifested in case of chronic inflammation.<sup>40,41</sup>

Taken together, the results obtained in the previous studies and the data reported in the current work suggest that angiogenesis has an essential role in the etiopathogenesis of pyogenic granuloma. The increased expression of CD31 in PG may correspond to the reactive or inflammatory character of this lesion.<sup>32</sup> Though, estimation of the expression of angiogenesis markers can assist the diagnosis and treatment of vascular lesions and may play a role in the production of new therapies that depend on antiangiogenic drugs.<sup>17,34</sup>

Bcl-2 is an apoptosis suppressor protein.<sup>25</sup> It is a part of the outer mitochondrial membrane and the endoplasmic reticulum.<sup>42</sup>

Studies that had been done on the expression of Bcl-2 in oral reactive lesions are scarce. So, in the current work, Bcl-2 expression in oral pyogenic granuloma and normal gingiva was assessed. The study of Nakamura, 2000 is the only study that had been done to investigate the expression of Bcl-2 marker in PG.<sup>14</sup>

In the present study, PG showed increased expression of Bcl-2 than normal gingiva. Cytoplasmic immunopositivity for Bcl-2 was detected in the endothelial cells of blood vessels and in the stromal cells. Similar results were obtained in the study of Nakamura, 2000. He reported that PG showed less apoptosis and higher expression of Bcl-2 in comparison to conventional granulation tissue (GT) and suggested that suppression of apoptosis in PG played a role in its rapid growth.<sup>14</sup> The difference in the behavior between PG and GT could be related to apoptosis regulation. Granulation tissue in wound healing process shows regression to scar tissue due to apoptosis of endothelial cells and fibroblasts.43,44

(2138) E.D.J. Vol. 64, No. 3 Naglaa M. Kamal

All the samples of normal gingiva of the present work revealed positive staining with Bcl-2 that was detected in the basal cells of the epithelium and are consistent with previous studies.<sup>23,45</sup> This pattern of expression in normal epithelium revealed the role of the progenitor basal cells, which need protection against apoptosis to maintain the survival of the whole epithelium.<sup>46</sup>

Bcl-2 area percent of immunoexpression, showed a higher mean value in pyogenic granuloma than normal gingiva, with a statistically significant difference.

Previous studies revealed that decreased apoptosis of endothelial cells may have a role in the rapid growth of PG. <sup>8,14</sup> The increased expression of Bcl-2 in the endothelial cells can facilitate the generation of a sustained potent angiogenesis through the expression of angiogenic stimulators as VEGF.<sup>47</sup> In addition, Bcl-2 may act to enhance the maturation of the newly developed vessels.<sup>48</sup>

Altogether these data explain the strong positive correlation between CD31 and Bcl-2 area percent of immunoexpression in the present study and suggest that Bcl-2 overexpression may have a role in angiogenesis process.

#### **CONCLUSION**

The results of the current work are suggestive of a role of Bcl-2 overexpression in the potentiation of angiogenic response. They also support the reactive nature of pyogenic granuloma and the role of both angiogenesis and apoptosis in the etiopathogenesis of this lesion.

#### REFERENCES

- 1. Hartzell ME. "Granuloma pyogenicum." J Cutan Dis Syph,1904; 22:520–525.
- Kamal R, Dahiya P and Puri A. Oral pyogenic granuloma: Various concepts of etiopathogenesis. J Oral Maxillofac Pathol. 2012; 16:79-82.

- Newadkar UR, Khairnar S and Dodamani A. Pyogenic granuloma: A clinicopathological analysis of fifty cases. J Oral Res Rev. 2018; 10:7-10.
- 4. Jafarzaddeh H, Sanatkhani M and Mohtasham N. Oral pyogenic granuloma: a review. J Oral Sci. 2006; 48:167-175.
- Gomes SR, Shakir QJ, Thaker PV and Tavadia JK. Pyogenic granuloma of the gingiva: A misnomer? - A case report and review of literature. J Indian Soc Periodontol. 2013: 17:514-9.
- Vassilopoulos SI, Tosios KI, Panis VG, et al. Endothelial cells of oral pyogenic granulomas express eNOS and CD105/endoglin: an immunohistochemical study. J Oral Pathol Med. 2011; 40:345-51.
- 7. Pagliai KA and Cohen BA. Pyogenic granuloma in children. Pediatr Dermatol. 2004, 21:10-13.
- Yuan K, Jin, YC and Lin M.T. Pathogenetics factors in pyogenic granulomas in pregnancy are modulated by female sex hormones. J. Peridontolol. 2002; 73:701 – 708.
- Saghafi S, Amoueian S, Montazer M, et al. Assessment of VEGF, CD-31 and Ki-67 immunohistochemical markers in oral pyogenic granuloma: a comparison with hemangioma and inflammatory gingivitis. Iranian J Basic Med Sci. 2011; 14:185-9.
- Regezi JA, Sciubba JJ and Jordan RC. Oral pathology: clinical pathologic correlations. Elsevier Health Sci. 2012;118-20.
- Martins-Filho PR, Piva MR, Da Silva LC, Reinheimer DM and Santos TS. Aggressive Pregnancy Tumor (Pyogenic Granuloma) with Extensive Alveolar Bone Loss Mimicking a Malignant Tumor: Case Report and Review of Literature. Int. J. Morphol. 2011; 29:164-167.
- 12. Al-Khateeb T and Ababneh K. Oral pyogenic granuloma in Jordanians: a retrospective analysis of 108 cases. J Oral Maxillofac Surg. 2003; 61:1285-1288.
- Gordón-Núñez MA1, de Vasconcelos Carvalho M, Benevenuto TG, Lopes MF, Silva LM and Galvão HC. Oral pyogenic granuloma: a retrospective analysis of 293 cases in a Brazilian population. J Oral Maxillofac Surg. 2010; 68:2185-8.
- Nakamura T. Apoptosis and expression of Bax/Bcl-2 proteins in pyogenic granuloma: A comparative study with granulation tissue and capillary hemangioma. J Cutan Pathol 2000; 27:400-5.

- Kamal R, Dahiya P, Palaskar S and Shetty VP. Comparative analysis of mast cell count in normal oral mucosa and oral pyogenic granuloma. Journal of Clinical and Experimental Dentistry. 2011; 3:1-4.
- Epivatianos A, Antoniades D, Zaraboukas T, et al. Pyogenic granuloma of the oral cavity: comparative study of its clinicopathological and immunohistochemical features. Pathol Int. 2005; 55:391-7.
- 17. Altamirano E and Drut R. Pyogenic granuloma of the transverse colon. Report of a pediatric case. Patologia. 2008; 46:263-5.
- Vermeulen PB, Gasparini G, Foks SB, et al. Second international consensus on the methodology and criteria of evaluation of angiogenesis quantification in solid human tumours. Eur. J. Cancer. 2002; 38:1564–1579.
- Newman PJ and Newman DK. Signal transduction pathways mediated by PECAM-1: new roles for an old molecule in platelet and vascular cell biology. Arterioscler. Thromb. Vasc. Biol. 2003; 23:953-964.
- Privratsky JR, Paddock CM, Florey O, Newman DK, Muller WA and Newman PJ. Relative contribution of PECAM-1 adhesion and signaling to the maintenance of vascular integrity. J Cell Sci. 2011; 124:1477-85.
- Pusztaszeri MP, Seelentag W and Bosman FT. Immunohistochemical expression of endothelial markers CD31 CD34 Von Willebrand factor and Fli-1 in normal human tissues. J Histochem Cytochem. 2006; 54:385-95.
- Rosai J and Ackerman's Surgical Pathology. 10th ed. St Louis: Saunders Elsevier, 2012; 37-41.
- Bulut Ş, Uslu H, Özdemir BH and Bulut ÖE. Expression of caspase-3, p53 and Bcl-2 in generalized aggressive periodontitis. Head Face Med. 2006; 2:17.
- 24. Singh N. "Apoptosis in health and disease and modulation of apoptosis for therapy: an overview," Indian Journal of Clinical Biochemistry 2007;22: 6–16.
- Kummoona R, Mohammad Sámi S, Al-Kapptan I and Al-Muala H. Study of antiapoptotic gene of oral carcinoma by using Bcl-2 oncogene. J Oral Pathol Med. 2008; 37:345-51.
- Juneja S, Chaitanya NB and Agarwal M. Immunohistochemical expression of Bcl-2 in oral epithelial dysplasia and oral squamous cell carcinoma. Indian J Cancer. 2015; 52:505-10.

- Kapoor P and Deshmukh RS. VEGF: A critical driver for angiogenesis and subsequent tumor growth: An IHC study.
  J Oral and maxillofacial Pathol. 2012; 16:330–37.
- Vara JT, Gurudu VS, Ananthaneni A, Bagalad BS, Kuberappa PH and Ponnapalli HP. Correlation of Vascular and Inflammatory Index in Oral Pyogenic Granuloma and Periapical Granuloma – An Insight into Pathogenesis. J Clin Diagn Res. 2017;11: ZC25-ZC28.
- Verma PK, Srivastava R, Baranwal HC, Chaturvedi TP, Gautam A and Singh A. Pyogenic granuloma hyperplastic lesion of the gingiva: Case reports. Open Dent J 2012; 6:153 6.4.
- Kheur S, Patekar D, Bagul N, Kulkarni M, Routray S and Dhas V. Role of mast cell in oral pathology. Oral & Maxillofacial Pathology Journal. 2013; 4(1).
- Freitas TM, Miguel MC, Silveira EJ, Freitas RA and Galvão HC. Assessment of angiogenic markers in oral hemangiomas and pyogenic granulomas. Exp Mol Pathol. 2005; 79:79-85.
- Rezvani G, Azarpira N, Bita G and Zeynab R. Proliferative activity in oral pyogenic granuloma: A comparative immunohistochemical study. Indian J Pathol Microbiol. 2010; 53:403-407.
- 33. Kasprzak A, Surdacka A, Tomczak M, Przybyszewska W, et al. Expression of angiogenesis-stimulating factors (VEGF, CD31, CD105) and angiogenetic index in gingivae of patients with chronic periodontitis. Folia Histochemica Et Cytobiologica. 2012; 50: 554–564
- 34. Seyedmajidi M, Shafaee S, Hashemipour G, Bijani A and Ehsani H. Immunohistochemical evaluation of angiogenesis related markers in pyogenic granuloma of gingiva. Asian Pacific J Cancer Prev. 2015; 16:7513-7516.
- Schmeisser A, Garlichs CD, Zhang H, Eskafi S, Graffy C, Ludwig J, Strasser RH and Daniel WG. Monocytes coexpress endothelial and macrophagocytic lineage markers and form cord-like structures in Matrigel under angiogenic conditions. Cardiovasc Res. 2001; 49:671–680.
- Blackwell MG, Itinteang T, Chibnall AM, Davis PF and Tan ST. Expression of embryonic stem cell markers in pyogenic granuloma. J Cutan Pathol. 2016; 43: 1096–1101.
- Fernandez Pujol B, Lucibello FC, Gehling UM, Lindemann K, Weidner N, Zuzarte ML, Adamkiewicz J, Elsasser HP, Muller R and Havemann K. Endothelial-like cells derived from human CD14 positive monocytes. Differentiation. 2000; 65:287–300.

(2140) E.D.J. Vol. 64, No. 3 Naglaa M. Kamal

 Jordan RC, Daniels TE, Greenspan JS, et al. Advanced diagnostic methods in oral and maxillofacial pathology. Part II: immunohistochemical and immunofluorescent methods. Oral Surg., Oral Med., Oral Pathol. 2002; 93:207–212.

- Sedivy R, Beck-Managetta J, Haverkam PF., et al. Expression of vascular endothelial growth factor-C correlates with the lymphatic microvessel density and the nodal status in oral squamous cell cancer. J. Oral Pathol. Med. 2003; 32:455–460.
- 40. Niczyporuk M, Hermanowicz A, Matuszczak E, Dziadziuszko R, Knaś M, Zalewska A, et al. A lack of correlation between mast cells, angiogenesis, and outcome in nonsmall cell lung cancer. Exp Lung Res. 2012; 38:281-5.
- 41. El Hayderi L, Paurobally D, Fassotte MF, André J, Arrese JE, Sadzot-Delvaux C, et al. Herpes simplex virus Type-I and pyogenic granuloma: a vascular endothelial growth factor-mediated association? Case Rep Dermatol. 2013; 5:236–43.
- 42. Lindsay J, Esposti MD and Gilmore AP. Bcl-2 proteins and mitochondria—Specificity in membrane targeting for death. Biochim Biophys Acta. 2011; 1813:532-9.
- 43. Kantarci A, Augustin P, Firatli E, Sheff MC, Hasturk H, et

- al. Apoptosis in gingival overgrowth tissues. J Dent Res. 2007; 86: 888-892.
- 44. Mitic K, Popovska M, Belazelkoska A and Jovanovic R. Gingival Tissue and Apoptosis. J Interdiscipl Med Dent Sci. 2014; 2: 141.
- 45. Garewal J, Garewal R and Sircar K. Expression of Bcl-2 and MIB-1 markers in oral squamous cell carcinoma (OSCC)-A comparative study. Journal of Clinical and Diagnostic Research. 2014; 8: QC01.
- 46. Abou Elkhier MT, EL- Zehary RR, Mourad MI et al. Immunohistochemical assessment of Bcl-2 and Ki-67 in gingival tissues of normal and immunosuppressed patients as predictors of neoplasia. Annals of Oral & Maxillofacial Surgery. 2014;10; 2:14.
- Bufalo DD, Trisciuoglio D, and Milella M. Crosstalk Between VEGF and Bcl-2 in Tumor Progression and Angiogenesis. In: Harmey JH, editor. VEGF and Cancer book. Texas:Kluwer Academic / Plenum Publishers. 2004; Chap.4:26-39.
- 48. Schechner JS, Nath AK, Zheng L. et al. In vivo formation of complex microvessels lined by human endothelial cells in an immunodeficient mouse. Proc Natl Acad Sci USA. 2000; 97:9191–9196.