



## The Ameliorative Effect of Vitamin C Against Hematological, Biochemical, Oxidative and Immunosuppressive Effect of Cadmium Chloride in Rats



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**T**HIS current investigation's objective was to ascertain whether vitamin C has a safeguarding impact in opposition to the harmful effects of cadmium, a heavy metal with a high toxicity and a substantial environmental pollutant. The experimental group consisted of 20 rats divided into 4 groups. The current study was carried out for 30 days. A (Control), B (Cadmium group, 15 mg/kg CdCl<sub>2</sub>), Vitamin C group (150 mg/kg vitamin C), D (CdCl<sub>2</sub> 15 mg/kg +150 mg/kg b.wt) orally. After 4 weeks blood samples were taken and liver tissues. Findings revealed a substantial drop in RBC count, Hb concentration, and PCV ( $P > 0.05$ ) and a substantial rise in WBC count ( $P < 0.05$ ). On another hand, findings also revealed a substantial drop in total protein, albumin and an increase concentration of globulin, total and direct bilirubin. Also increase concentrations of ALP, AST and ALT. In addition to antioxidants GST, GSH, CAT, SOD, rise in MDA of cadmium group in comparison to A group but, when vitamin C combined by cadmium chloride, the results indicated a noticeable improvement in the parameters under study, indicating the therapeutic effect of vitamin C against cadmium chlorides toxicity in rats.

**Keywords:** Cadmium, Vitamin C, Antioxidant enzymes, Antioxidants parameters, Amelioration potential.

### Introduction

Earth frequently contains heavy metals, which are also referred to as metalloids and metals in the periodic Table [1]. Heavy metals (such as lead, cadmium, nickel, arsenic, chromium, and mercury) have a negative impact on human and animal health [2]. "A soft, silvery-white metal with an atomic weight of 113.41 and an atomic number of 48", cadmium (Cd) found in the earth's crust in combination with zinc also consists in the industries as an inevitable by-product of zinc, lead and copper. present in food, water, and cigarette smoke accumulates in the liver and causes a number of hepatic dysfunctions,

toxic, and carcinogenic. It belongs to chemical element group XII on the periodic Table [3]. The mechanisms and routes of Cd's toxicity at the tissue level in biological systems are still poorly understood. Reactive oxygen species (ROS) are produced more readily and oxidative stress is induced in several tissues as a result of Cd toxicity, according to a number of studies [4,5].

Furthermore, exposure to Cd promotes tissue damage brought on by lipid peroxidation [6]. Presently, numerous research have demonstrated that cadmium causes, hematological, biochemical effects, histopathological, immunosuppressive, and alterations in the gene expression pathway-

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induced damage to the antioxidant enzyme system [7,8]. It has been noted that Cd causes cell death, most likely via targeting mitochondria. Caspase-3 activity is a dynamic shape of cellular necrosis induced via the release of cytochrome C, and it was raised with mitochondrion-dependent apoptosis. Intracellular homeostasis and the redox system are affected by Cd. Apoptosis happens when the calcium equilibrium is disturbed [9], in addition a key biological sign in the severe hepatic tissue injury is tumor necrosis factor alpha (TNF- $\alpha$ ) [10].

Natural antioxidants are substances that prevent the chemical process of oxidation, which can result in the production of free radicals and trigger a cascade that may harm living organisms give a multitude of biological benefits of natural antioxidants, including the ability to fight cancer, ageing, inflammation, and atherosclerosis [11]. Ascorbate, C is a water-soluble vitamin. with potent antioxidant, chain-breaking, and hydrogen peroxide and superoxide scavenger properties, is found both within and outside of cells [12]. Given that humans lack a necessary enzyme in the biosynthetic route, which prevents them from synthesizing vitamin C, there has been a lot of interest in its application in the prevention and/or treatment of infections. The proper operation of the immune system depends on vitamin C [13] By repairing or neutralizing free radicals by donating one electron, followed by a proton, to produce dehydroascorbic acid, Vitamin C functions as a potential antioxidant utilized to lessen the liver and kidney toxicity brought on via certain medications [14].

Vitamin C demonstrated excellent therapeutic benefits against the rat toxicity-related problems and brought back these abnormalities to nearly normal levels. In the end, it was determined that cadmium has extreme destruction effects on liver cells, liver enzymes, and oxidative status of the liver. The current investigation's objective was to determine the vitamin C's protective effects against Cadmium chloride on hematological, biochemical, and immune-histopathological parameters as well as to track its potential to mitigate Cd-induced oxidative damage and hepatotoxicity in rats.

### **Material and Methods**

Cadmium chloride (CdCl<sub>2</sub>) (powder dissolved in distilled water immediately before use) and purchased vitamin C (powder) from the Egyptian

business Sigma. Catalase, GST, GSH, SOD, and MDA kits of the Bio-diagnostic, Egypt, human company kits for total protein, albumin, AST and ALT, ALP measured using kits from ELITech company (Paris, France). Diamond company (Cairo, Egypt) for Bilirubin total and direct. Recombinant of TNF-alpha and Caspase 3 from Scy Tek company.

#### *Experimental animal handling*

Twenty rats weighing between (150-200g) were bought from Tanta University's. They were housed in clean, well-ventilated plastic cages at a room temperature of 25 °C, with a relative humidity of 55 to 65%, and were exposed to light and darkness every 12 hours. Fourteen days before treatment, given fresh tap water and standard regular rodent diets to eat, ration used to feed rats composed of (yellow corn, soyabeans, vegetables oil, calcium carbonate salt and vitamins). According to the Mansoura University of Veterinary Medicine's recommendations, every experiment was approved and carried out.

#### *Experiment, tissue processing*

After the period of acclimation, the rats divided into 4 groups of five animals. Rats' LD50 value for cadmium determined by the oral route was 2696.54mg/kg for 90 days by Bliss method [15], via intraperitoneal injection was 5.98 and 3.9 mg/kg/ b.w for 24 h exposure and 48h exposure respectively [16].

**Group A** (control): rats received normal diet and fresh water without cadmium chloride.

**Group B** (rats were only given cadmium chloride) in dose of 15mg/kg b.wt for 4 weeks orally.

**Group C** (rats received vitamin C only in dose 150 mg/kg b.wt over a period of 4 weeks orally.

**Group D** (vitamin C 150mg/kg/ b.wt and cadmium chloride 15mg/kg b.wt ).

#### *Sample preparation*

Blood samples collected individually from each rat in all group from the eye canthus with capillary tubes after anesthesia with i.p injection of ketamine & xylazine at a dose of 50-10mg/kg, respectively, in two different tubes for each sample tube with EDTA (50  $\mu$ l/ml blood), complete blood picture (CBC). The other blood sample was collected without anticoagulant, placed until complete clotting in room temperature then

centrifugated (3000 rpm/ 10 minutes) “separated serum in Eppendorf and kept at -20 °C till biochemical testing.

Tissue homogenate, “Rats dissected after cervical dislocation, liver was then removed, Phosphate-buffered saline (PBS) was used to clean the item before it between two filter sheets, where it dried. Half gram of liver grinded with 4.5 ml of phosphate buffer saline to create hepatic tissue homogenates centrifuged (4000 rpm/15 min) at 4 °C. The clear fluid aspirated and freezed at 20 °C. Another part of liver preserved in 10% formalin for conduction of histological and immunohistochemical analyses.

#### *Hematological estimation*

Numerated number of erythrocytes (RBCs) counts , hemoglobin (Hb) content , packed cell volume (PCV) % , leukocytes (TLC) counts . Calculations were made to determine the blood indices , mean corpuscular volume (MCV) , mean corpuscular haemoglobin (MCH), and mean corpuscular hemoglobin concentration (MCHC) [17].

#### *Biochemical estimation*

The collected frozen serum samples were analyzed for alanine transaminase (ALT), aspartate transaminase (AST) , alkaline phosphatase (ALP) , total bilirubin (TB) , direct bilirubin, total protein , and albumin using a sinostic automatic suction analyser , China . Globulin was calculated by subtracting albumin from total protein ; also calculate A/G ratio , [18] . The manufacturer’s instructions were followed while measuring the levels of MDA , SOD , Catalase , GSH and GST in the liver homogenates using commercially available kits (Bio-diagnostic, Egypt) [19].

#### *Histopathological estimation*

Hematoxylin and eosin (H&E) stain was applied to the formalin-fixed liver material, which was then embedded in paraffin, tissue section is cut by micrometer into 5-mm-thick sections, and viewed under a microscope. according to Bancroft and Gamble [20].

#### *Immunohistochemistry of TNF- $\alpha$ and Caspase-3 expression*

Formalin-fixed, paraffin-embedded liver tissue slices were used for immunohistochemistry. is significantly enhanced by treatment with citrate plus (Scy Tek catalog CPL500) were carried out as describe by[21]. Following deparaffinization, tissue sections were incubated with monoclonal rabbit anti-mouse primary antibody (1:100)

against caspase-3 (Biocare Medical, Pacheco, CA) and TNF- $\alpha$  overnight at 4 °C, then washed with TBS (tris buffer saline treated with goat anti-rabbit antibody for an hour at room temperature. (1:1000) and anti-mouse antibodies, developed for 1.5 minutes with 3, 3'-diaminobenzidine tetrahydrochloride (DAB), counterstained with hematoxylin, and covered. Using a light microscope (Nikon Eclipse TE2000-U, NIKON, Japan), Semi-quantitative research was done on the immuno-positive cells’ distribution pattern within the hepatic lobule. The number of immuno-positive cells per 1000 cells was also counted and analyzed using Image-J analysis software (Image J, 1.46a, NIH, USA).

#### *The analysis of cadmium residue in liver*

Using atomic absorption spectrophotometry (Perkin-Elmer A.A. Model 800) with the Zeeman-effect for background correction. In brief, 1:3 ultra-pure concentrated nitric acid and chlorine perchlorate employed to assimilate 0.5g at ratio (6:1). then complete dehydrated by a hot plate. Dilution with distilled water before being subjected to an atomic absorption spectrophotometry test for cadmium [22].

#### *Analytical Statistics*

Data presented as Mean  $\pm$  SEM for 5 rats in each group for the tables and figures. All data was analyzed using the SPSS application for (Windows, version 20 USA). ANOVA comparing the sets, Duncan employed to evaluate whether there was a significant difference ( $P < 0.05$ ) [23].

### **Results**

The experimental period in the current investigation was set at 30 days, and the nine rat batches were kept as such. No fatalities were noted during the experimental times.

#### *Cadmium concentration in liver of experimental rat*

According to (Table 1) , cadmium in the hepatic cadmium-toxicated tissues were substantially higher than the control group, whereas with co-treatment with vitamin C, the concentration of cadmium was significantly decline than cadmium intoxicated rats. The amount of cadmium residue in the liver of vitamin C-treated group and the control rats were the same.

**TABLE 1. the residues of cadmium in liver at 4<sup>th</sup> week post treatment with vitamin C in rats (Mean± SE).**

Group	Cadmium residues in liver ug/L (mean ±SD)
Cont.	0.00013 ±0.000042 <sup>b</sup>
Cd	0.024496±0.0036 <sup>a</sup>
Vit C	0.00033 ±0.00992 <sup>b</sup>
Cd+Vit C	0.027 ±0.007 <sup>a</sup>

**Cont.** (Control), **Cd** (Cadmium chloride), **Vit C** (Vitamin C), **Cd+VitC** (rats treated with Cadmium Chloride and vitamin C). Values with different superscript letters consider significance at ( $p < 0.05$ ).

#### *Hematological result*

According to the hematological results shown in (**Figure 1 A, B, C**), the cadmium-intoxicated group had a significantly lower in (RBCs×10<sup>6</sup>) counts, (Hb g/dl) concentration, and (PCV%) value than the control group. Meanwhile WBCs (×10<sup>3</sup>μL) counts showed significant increase in cadmium intoxicated group compared with control one (Mean ± SE). Our study showed prominent ameliorative effect of co-treatment of vitamin C with cadmium, via significantly up-regulate in RBCs counts, Hb conc, and PCV %, beside the significantly decreased in WBC counts (**Figure-1G**) (Mean ± SE). At the same study blood indices not changed significantly in different groups (**Figure 1 D, E, and F**) (Mean ± SE). from the previous data we found that cadmium chloride caused normocytic normochromic anemia with evidence of leukocytosis which correlated via vitamin C administration.

#### *Biochemical result*

##### *Liver function Tests*

Cadmium induced hepatocellular inflammation that manifested by clear rise in serum liver enzymes than control rats (**Figure 2 A, B and C**). Meanwhile the cadmium intoxicated treated with vitamin C showed a significant down-regulation in serum liver enzymes compared with cadmium intoxicated group, but not return to the normal value. Total bilirubin (TB) in Cd-intoxicated rats substantially superior to control rats. When compared to Cd-intoxicated rats, the level of bilirubin was dramatically reduced by vitamin C administration together with Cd. although they still vary from control levels. In rats exposed to Cd, there was a notable increase in the direct bilirubin. Co-treatment with vitamin C and Cd dramatically reduced the level of direct bilirubin, but it did not approach the normal

level. (**Figure 2 D, E, F**). Cadmium exposure led to a considerable decrease in total protein and albumin. Groups co-treated with vitamin C had significantly increase in total protein and albumin than groups exposed to cadmium only (**Figure 3 A, B**). Otherwise, globulin value in cadmium toxicated group rats were slightly decreased than control one, also with treatment with vitamin C, globulin significantly up-regulated but still less than normal level (**Figure 3, C**) In the group that had consumed cadmium, the A/G ratio had drastically decreased.

##### *Hepatic antioxidant and oxidative damage markers*

A considerable rise in liver MDA and considerable decrease in (Catalase, SOD, GSH, and GST) indicate that cadmium toxicity causes hepatic oxidative stress (**Table 2**). In comparison to cadmium-intoxicated rats, co-treatment with vitamin C brought about a considerable drop in liver MDA and prominent increase in (GSH, CAT, SOD and GST).

##### *Histopathological investigation of liver*

There were hepatic cords extending toward the peripheral normal portal area around each hepatic lobule's thin-walled core vein., divided by relatively little barriers as shown by H&E in control group (A) (liver portal vein, artery, and bile ductile) in (**Figure 4**). Hepatocytes hydropic degeneration, the hepatic sections of the cadmium group show localized zones of clot death infiltrated with inflammatory cells, occluded sinusoids, and congested veins in middle with inflammatory cell infiltration (B) (**Figure 5**). These histological abnormalities were lessened in rats given vitamin C along with these treatments, as evidenced by hepatocyte patterns that were noticeably normal, decreased portal inflammation, and a reduction in hepatocyte hydropic degeneration (**Figure6**)



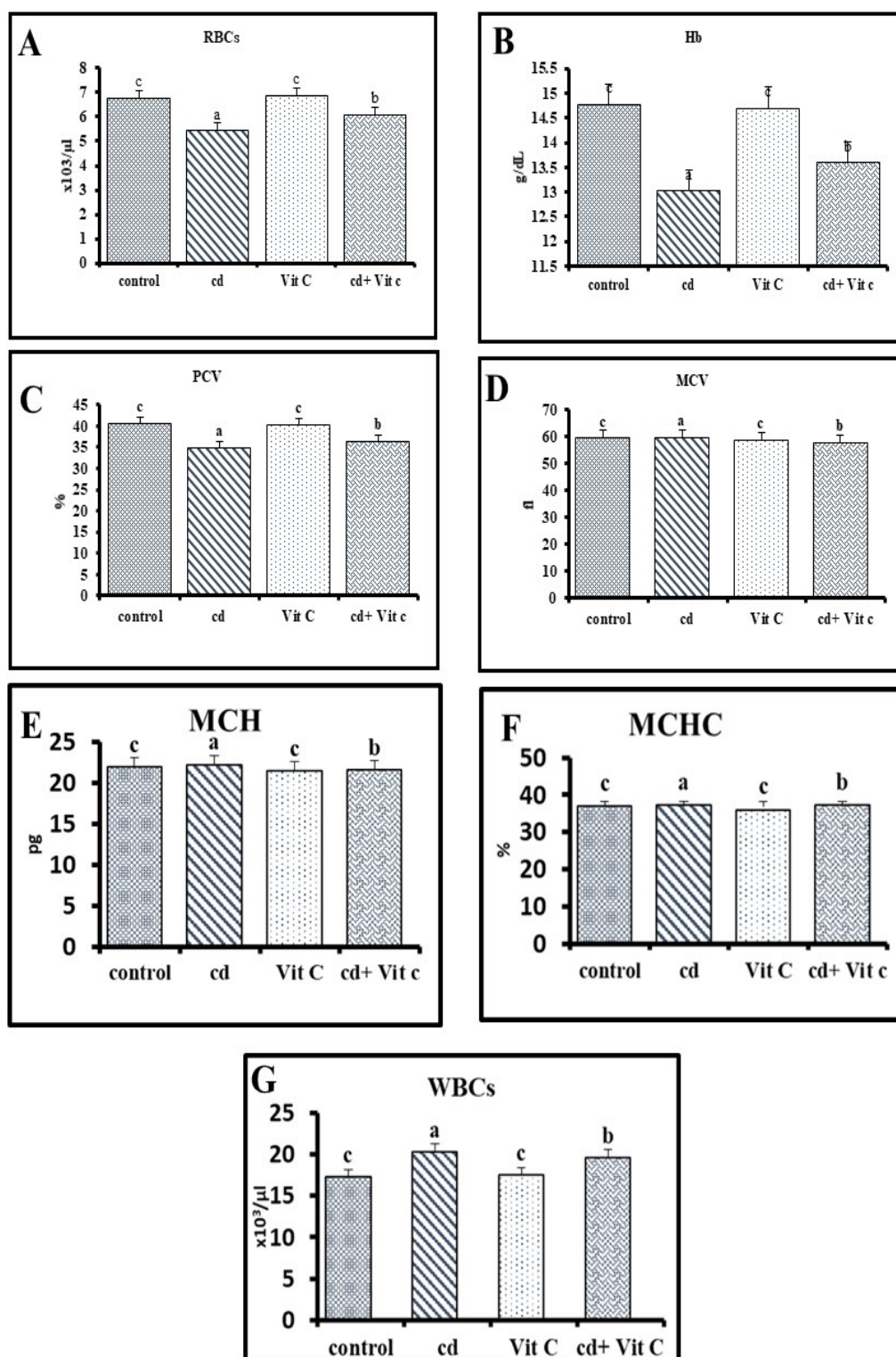


Fig. 1. Hematological result at the end of 4<sup>th</sup> weeks post treatment of vitamin C and in cadmium intoxicated rats (Mean  $\pm$  SE). (A) RBCs count, (B) Hb concentration, (C) PCV % (Packed cell volume), (D) MCV (Mean corpuscular volume), (E) MCH (Mean corpuscular hemoglobin, and (F) MCHC (Mean corpuscular hemoglobin concentration), (G) WBCs (White blood cells) count. Values with different superscript letters consider significance at (p<0.05).

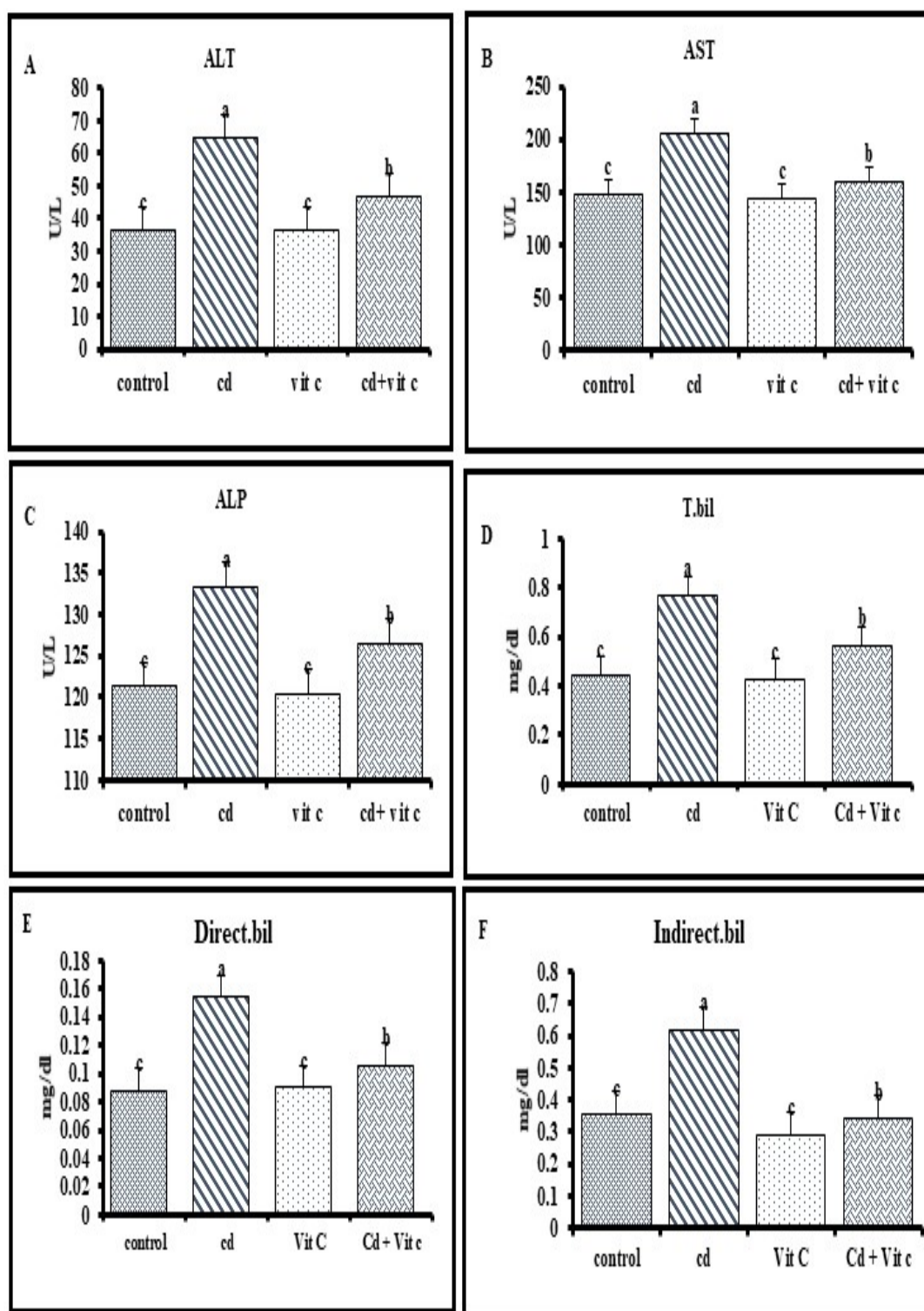


Fig. 2. Liver function tests at the end of 4<sup>th</sup> weeks post treatment of vitamin C in cadmium intoxicated rats (Mean  $\pm$  SE). (A) Alanine aminotransferase (ALT), (B) Aspartate aminotransferase (AST), (C) Alkaline phosphatase (ALP), (D) Total. bil (total bilirubin), (E) Direct. bil (direct bilirubin) and (F) and Indirect. bil (indirect bilirubin). Values with different superscript letters consider significance at (p<0.05).

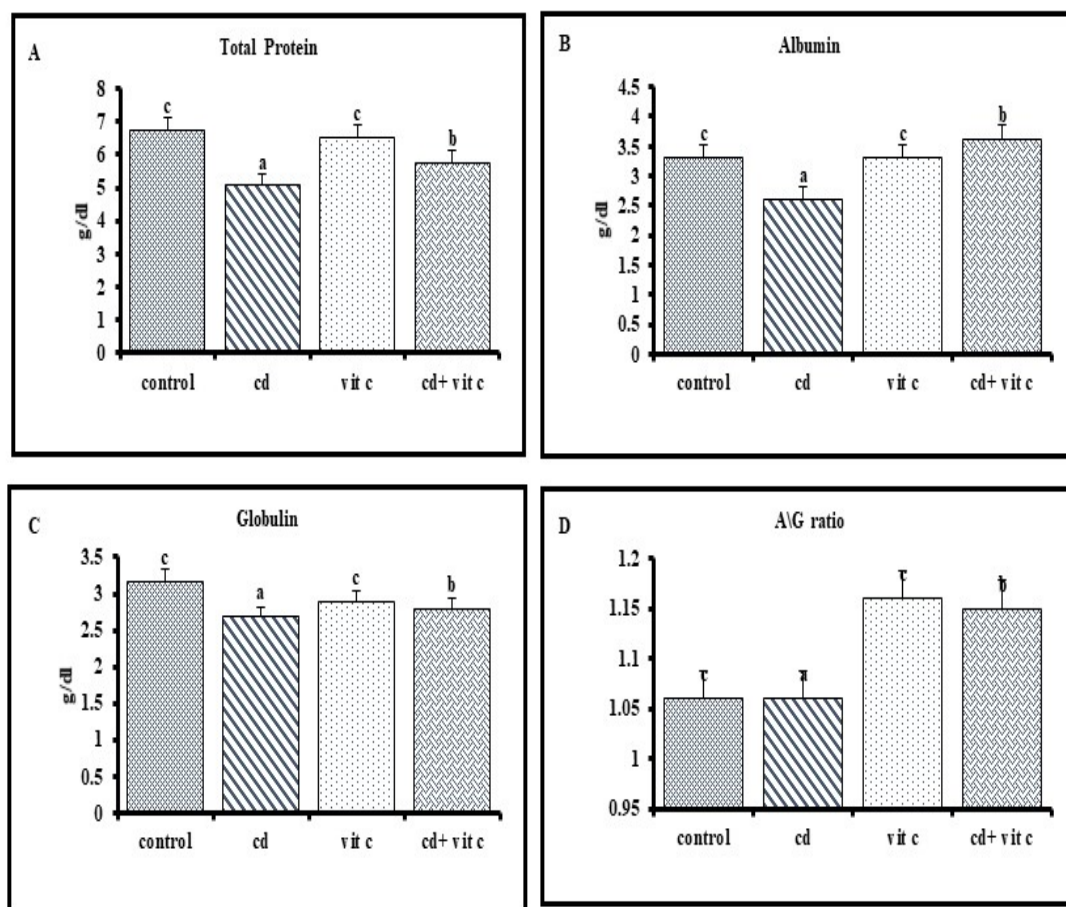


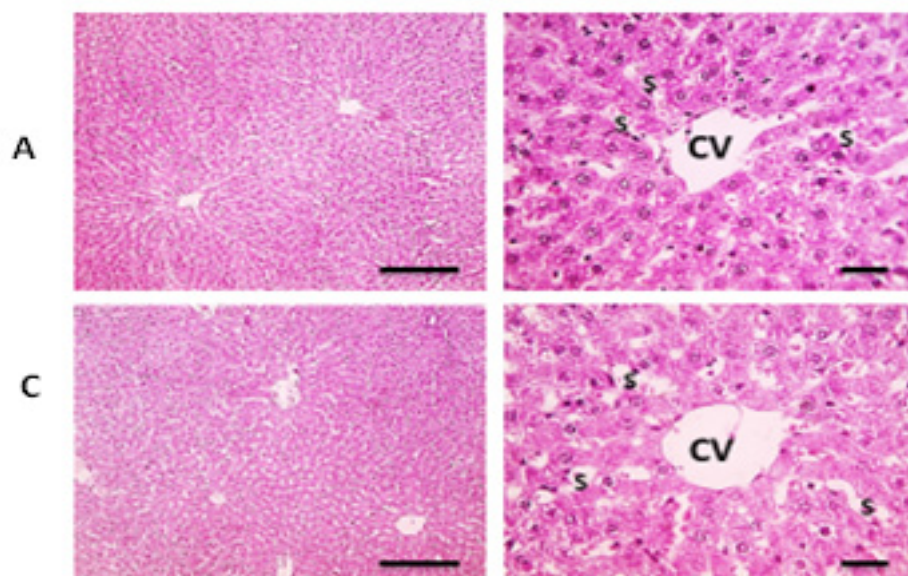
Figure 1. (A) Total protein, (B) Albumin, (C) Globulin and (D) A/G ratio. Values with different superscript letters consider significance at ( $p < 0.05$ ).

TABLE 2. Oxidative stress and antioxidant biomarkers in hepatic homogenate at 4th week post treatment of vitamin C in cadmium toxicated rats (Mean  $\pm$  SE).

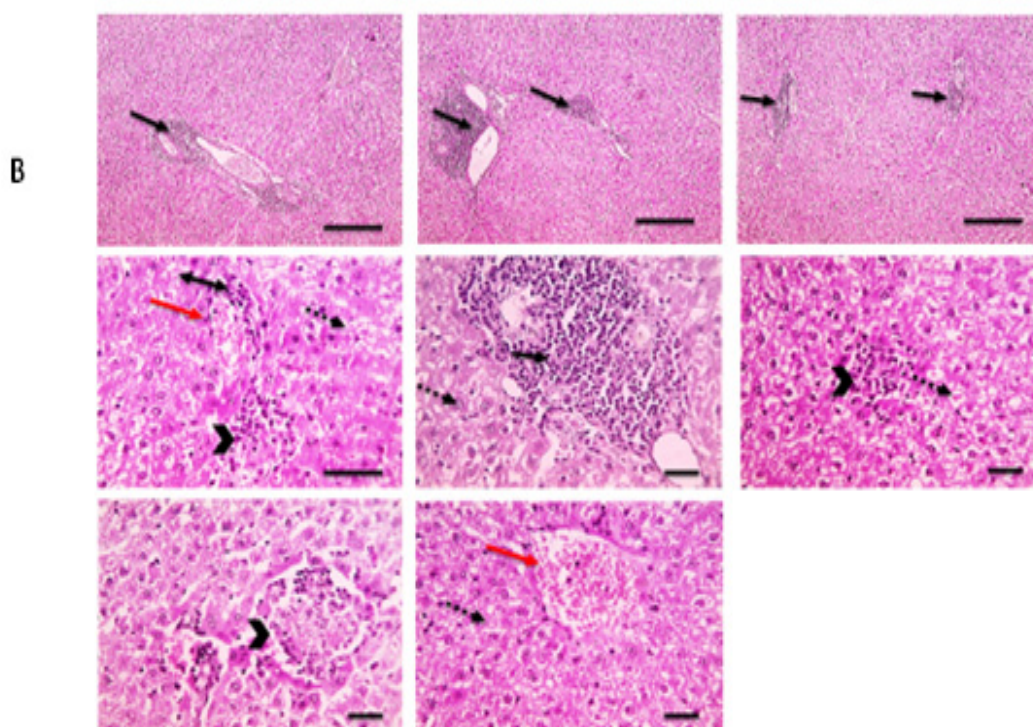
Group	MDA (n mol/g)	SOD (U/tissue)	CAT (U/tissue)	GSH (mg/g.tissue)	GST (mg/g tissue)
Cont.	24.54 $\pm$ 2.87 <sup>c</sup>	611.60 $\pm$ 0.92 <sup>a</sup>	1.93 $\pm$ 0.07 <sup>a</sup>	19.54 $\pm$ 1.42 <sup>a</sup>	24.49 $\pm$ 0.59 <sup>b</sup>
Cd	72.93 $\pm$ 6.19 <sup>a</sup>	320.24 $\pm$ 44.09 <sup>c</sup>	1.07 $\pm$ 0.16 <sup>c</sup>	9.68 $\pm$ 0.55 <sup>c</sup>	18.67 $\pm$ 0.56 <sup>d</sup>
Vit C	24.00 $\pm$ 0.8 <sup>c</sup>	610.20.00 $\pm$ 1.49 <sup>a</sup>	1.97 $\pm$ 0.007 <sup>a</sup>	19.68 $\pm$ 2.32 <sup>a</sup>	26.24 $\pm$ 0.54 <sup>a</sup>
Cd+Vit C	40.23 $\pm$ 2.82 <sup>b</sup>	505.4 $\pm$ 14.96 <sup>b</sup>	1.74 $\pm$ 0.02 <sup>b</sup>	14.94 $\pm$ 0.86 <sup>b</sup>	20.41 $\pm$ 0.29 <sup>c</sup>

Cont. (control), Cd (cadmium chloride), Vit C (vitamin C), Cd +Vit C (Cadmium & vitamin C). MDA (Malondialdehyde), SOD (Superoxide dismutase), CAT (Catalase), GSH (Reduced glutathione), GST (Glutathione-S-transferase). Values with different superscript letters consider significance at ( $p < 0.05$ ).





**Fig. 4. .** Microscopic pictures of H&E-stained hepatic sections showing normal radially arranged hepatic cords around central veins (CV) with normal portal areas and sinusoids (S) in control group (A) and groups (C) (group of vitamin C only Low magnification X: 100 bar 100 and high magnification X: 400 bar 50



**Fig. 5.** Microscopic pictures of H & E stained hepatic sections showing marked portal inflammation (black arrows) , hepatocytes hydropic degeneration (dashed black arrows) and focal areas of coagulative necrosis ( black arrowheads ) infiltrated with leukocytic cells, occluded sinusoids, congested central veins (CV) ( red arrows ) with leukocytic cells infiltration around CV ( double headed black arrows ) in group (B) ( group intoxicated with cadmium ) Low magnification X: 100 bar 100 and high magnification X: 400 bar.



### Immunohistochemistry result

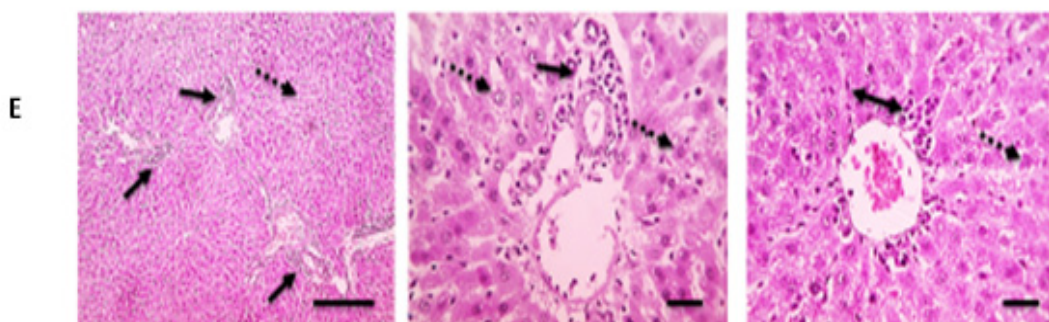
According to immunohistochemistry of caspase-3 & TNF- $\alpha$  proteins of hepatic slices of all rat sets increased with toxicity (**Figures 7 and 8**) compared to the control group. The expression of them were considerably decreased when vitamin C and cadmium were coupled (**Table 3**).

### Discussion

Scientists and physicians have long been interested in how natural anti-oxidants can reverse the oxidative damage brought on by xenobiotic intoxication. The primary factor in acute cadmium toxicity was a shift in cellular redox balance, which led to oxidative stress, hepatocellular damage, and cadmium buildup in the liver [24]. The route, dose, length of exposure, and species sensitivity all have an impact on how quickly metals are absorbed, distributed, and accumulated in tissues and organs [25]. Liver is first organ affected by toxicity of cadmium, and it sustains substantial damage and accumulation [26]. In present study found sets intoxicated with

cadmium showed clear accumulation of cadmium of hepatic cells. Numerous researches looked at health advantages of vitamin C to lessen the toxicity caused by cadmium. The current study discovered that supplementing with vitamin C lessen accumulation in hepatic tissue due to its antioxidant effect [27]. The current study demonstrated that cadmium-intoxicated rats displayed prominent anemia with a decrease in PCV, RBCs and hemoglobin (Hb). This anemia was explained by hemolysis, increased permeability, and decreased erythropoietin hormone synthesis [28-30]. Although blood indices (MCV, MCH, and MCHC) show no change (normocytic normochromic anemia) and this agreed with some authors [8]. As opposed to that, systemic inflammation was evident in the Cd intoxicated rats, where leukocytes significantly increased and that agreed with many investigators [31,32].

Co-treatment (vitamin C+ cadmium) ameliorated the level of PCV %, RBCs counts and Hb conc. due to reduction of oxidative damage to vascular walls, and renal cells which produced



**Fig. 6.** Microscopic pictures of H&E-stained hepatic sections showing markedly decreased portal inflammation (black arrows) and hepatocytes hydropic degeneration (dashed black arrows) few leukocytic cells infiltration around CV (double headed black arrows) in group (E) groups co treated with vitamin C). Low magnification X: 100 bar 100 and high magnification X: 400 bar 50

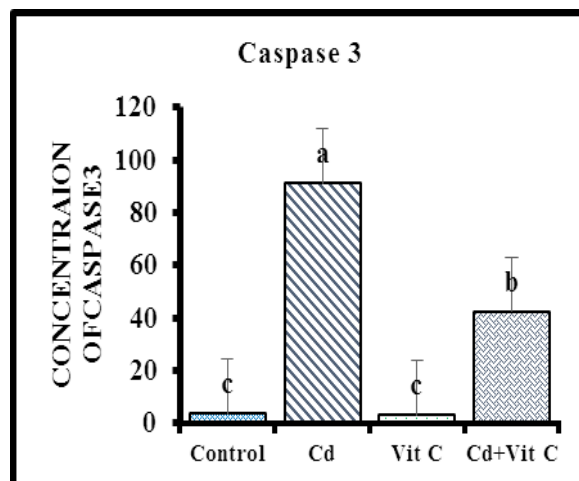
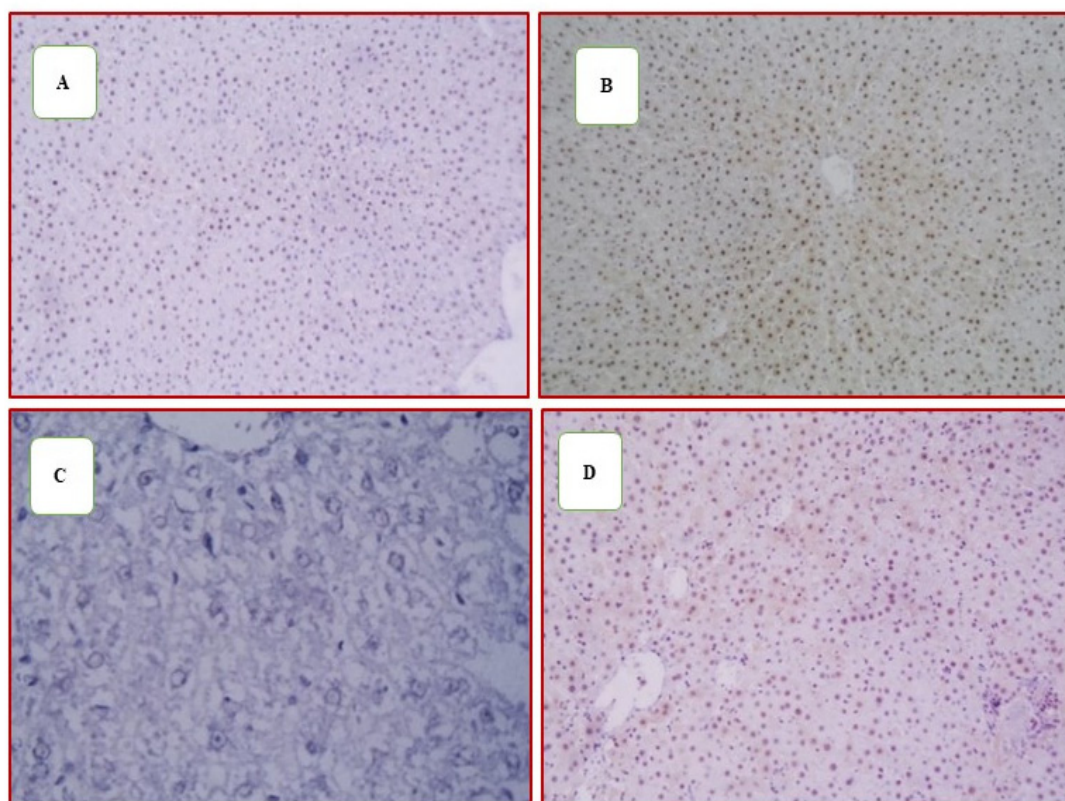
**TABLE 3.** Immunohistochemistry result:

Group	Caspase 3	TNF- $\alpha$
Cont.	0.25 $\pm$ 0.049 <sup>c</sup>	0.25 $\pm$ 0.05 <sup>a</sup>
Cd	91.13 $\pm$ 2.07 <sup>a</sup>	79.41 $\pm$ 4.49 <sup>c</sup>
Vit C	0.21 $\pm$ 0.04 <sup>c</sup>	0.06 <sup>a</sup> $\pm$ 0.25
Cd + Vit C	42.17 $\pm$ 3.96 <sup>b</sup>	19.13 $\pm$ 1.48 <sup>b</sup>

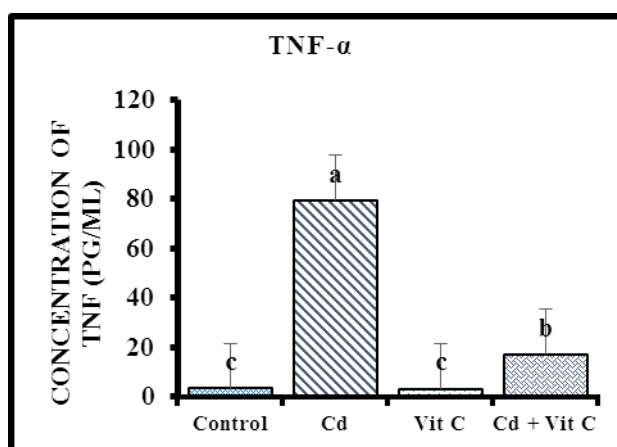
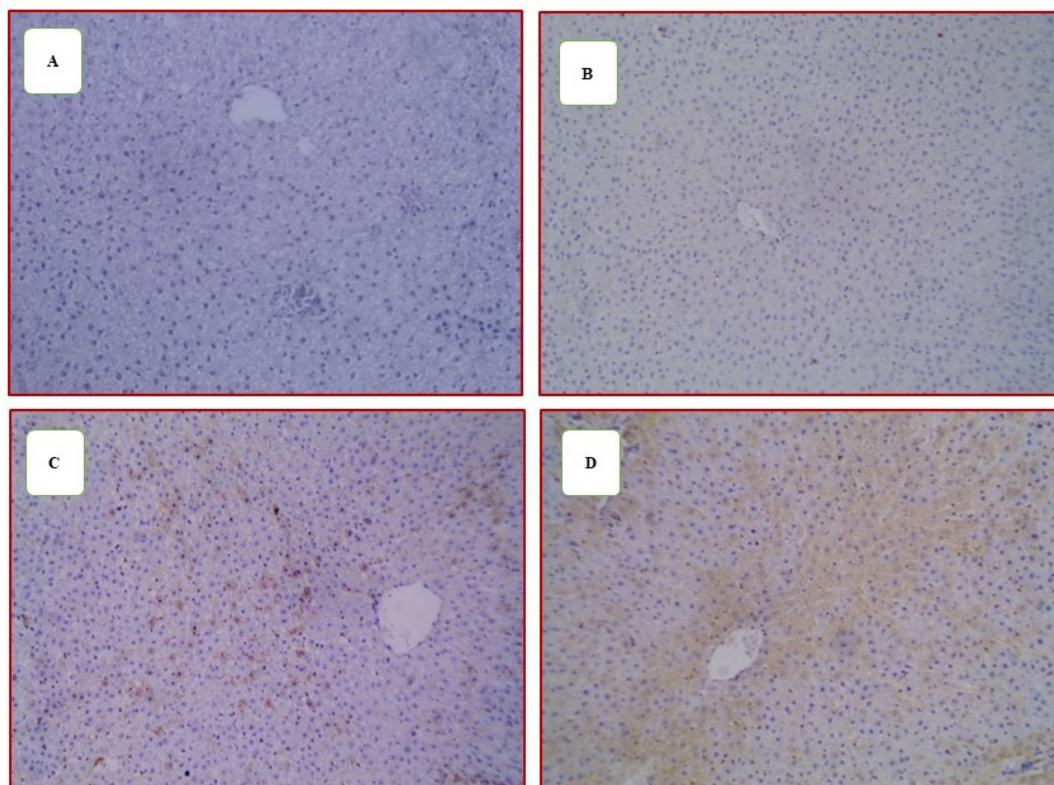
Cont. (control), Cd (cadmium chloride) Cd.

Values with different superscript letters consider significance at ( $p < 0.05$ ).

## I-Caspase

**Fig.7.**

- A - Control: Liver displayed negative immunostaining against Caspase 3. (IHC, DAB immunostaining, hematoxylin as a counter stain) .
- B - Cadmium group: Liver displayed strong immunostaining against Caspase 3. (IHC, DAB immunostain, hematoxylin as a counter stain, 100x) .
- C – vitamin C group: Liver displayed negative immunostaining against Caspase 3. (IHC, DAB immunostaining, hematoxylin as a counter stain) .
- D - Cadmium group with vitamin C: Liver displayed moderate immunostaining against Caspase 3. (IHC, DAB immunostaining, hematoxylin as a counter stain, 100x)
- E – Immunohistopathological result of rats liver at the end of 4th week post treatment of vitamin C in cadmium intoxicated rats Caspase 3 (Mean ± SE) Values with different superscript letters consider significance at ( $p < 0.05$ ) .

**2-TNF- $\alpha$ :****Fig. 8.**

- A - Control: Liver displayed negative immunostaining against TNF $\alpha$ . (IHC, DAB immunostaining, hematoxylin as a counter stain, 100x).
- B - Cadmium group: Liver displayed strong immunostaining against TNF $\alpha$ . (IHC, DAB immunostaining, hematoxylin as a counter stain, 100x).
- C - Vitamin C group: Liver displayed negative immunostaining against TNF $\alpha$ . (IHC, DAB immunostaining, hematoxylin as a counter stain).
- D - Cadmium group with vitamin C: Liver displayed moderate immunostaining against TNF $\alpha$ . (IHC, DAB immunostaining, hematoxylin as a counter stain, 100x).
- E - Immunohistopathological result of rat's liver at the end of 4th week post treatment of vitamin C in cadmium intoxicated rats TNF- $\alpha$  (Mean  $\pm$  SE). Values with different superscript letters consider significance at ( $p < 0.05$ ).



erythropoietin hormone, maintains physiological homeostasis, and encourages the small intestine to absorb more iron [33]. The anti-inflammatory properties of vitamin C, on the other hand, prevented leukocytes from increasing [34]. Liver is important location to biotransformation & cadmium metabolism [35]. This indicated by Changes in levels of (ALT, AST, and ALP), which are markers to hepatitis and other conditions [36]. The rate of metabolic protein degradation reflects the impact of cadmium on ALT and AST activity [37] due to enhanced plasma membrane permeability or cellular necrosis lead to enter the bloodstream from the cytosol, as well as the use of amino acids for oxidation or gluconeogenesis as a result of toxicity [38], and all results confirmed with histopathological findings in liver tissue. As founded degeneration and necrosis in hepatic cells. Our results showed that vitamin C reduces the hepatotoxicity brought on by Cd, likely by lowering oxidative damage and lipid peroxidation while maintaining the antioxidant defense system and that agreed with some researchers [39]. Liver which is body's only location for protein synthesis, was impacted by oxidative stress, which may be the main cause of the drastic decrease in albumin & total protein of cadmium-toxicated groups as clarified on our study and this may be due to damage of the endoplasmic reticulum, DNA & mitochondria by excessive ROS production, which has an impact on how proteins are formed [40]. So, cadmium toxicity changed the liver's ability to synthesize, which may have an impact on how much fluid is retained in the tissue space. On other hand the current study revealed increase in globulin level in cadmium group than control one as defense mechanism against toxicity which led to decrease of A/G ratio [4]. Significantly raised by vitamin C due to the ability to decrease damage of oxidation [41]. Due to an increase in oxidant substances, a decrease in glutathione, stimulation of the heme oxygenase enzyme, and an increase in red blood cell lysis, direct & total bilirubin of cadmium-intoxicated rats is higher than in normal control groups [42]. With the co-treatment of vitamin C led to significant decrease in total and direct bilirubin because red blood cells' ability to counteract oxidative stress is reduced, which can enhance red cell lysis [43]. In the current investigation, cadmium chloride

markedly decreased CAT, SOD, and GST as well as amount of GSH, showing that oxidative stress exists. Endogenous antioxidant enzymes SOD and CAT are considered the initial band of resistance because their potent capacity to neutralize ROS [44]. CAT facilitates the transformation of H<sub>2</sub>O<sub>2</sub> into H<sub>2</sub>O & oxygen by avoiding creation to OH [45]. SOD was produced as a by-product of oxidative stress into H<sub>2</sub>O<sub>2</sub> [46]. GST plays a crucial part in the detoxification process in the hepatic tissues by facilitating binding of any toxic substance with GSH [47]. According to research, exposure to Cd causes the production of ROS, which causes the lipids to oxidize either by decreasing the GSH concentration or by depleting antioxidant enzymes [48]. Due to lipid peroxidation, Cd increased the amount of MDA [49]. The delivery of vitamin C successfully increased GSH, CAT, GST, and SOD as Due to its reducing properties, vitamin C can diminish ROS like H<sub>2</sub>O<sub>2</sub> and neutralize them. MDA and ROS levels, however, were reduced [50]. This study demonstrated a considerable rise in TNF- $\alpha$  expression of cadmium intoxicated groups [51]. Increased reactive oxidative species brought on by decreased anti-oxidant enzyme activity and more oxidation of lipids in Cd toxicity may result in significant alterations to membrane functions, damage to cellular integrity, DNA fragmentation, and caspase-3 activation. The prevention of cell death is connected to vitamin C's impact on oxidative stress, and decrease level of caspase3 [52].

### **Conclusion**

The findings of our study demonstrated vitamin C's value in lowering the toxicity of cadmium chloride on hematological, biochemical, antioxidant, immunosuppressive, and histopathological effects.

### **Funding statement:**

There was no external support for this study.

### **Conflict of Interest:**

The writers say they have no competing of concern.

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## References

1. Ali, H. and Khan, E. What are heavy metals? Long-standing controversy over the scientific use of the term 'heavy metals'—proposal of a comprehensive definition. *Toxicological & Environmental Chemistry*, **100**, 6-19(2018).
2. Tang, J., Zhang, L., Zhang, J., Ren, L., Zhou, Y., Zheng, Y., Luo, L., Yang, Y., Huang, H. and Chen, A. Physicochemical features, metal availability and enzyme activity in heavy metal-polluted soil remediated by biochar and compost. *Science of the Total Environment*, **701**, 1-9 (2020).
3. Branca, J.J.V., Pacini, A., Gulisano, M., Taddei, N., Fiorillo, C. and Becatti, M. Cadmium-induced cytotoxicity: effects on mitochondrial electron transport chain. *Frontiers in Cell and Developmental Biology*, **8**, 1-7(2020).
4. Poli, V., Madduru, R., Aparna, Y., Kandukuri, V. and Motireddy, S.R. Amelioration of Cadmium-Induced Oxidative Damage in Wistar Rats by Vitamin C, Zinc and N-Acetylcysteine. *Medical Sciences*, **10**, 1-7 (2022).
5. Renuka, M., Aparna, Y., Venkataramanaiah, P. and Srinivasulu Reddy, M. Vitamin, C, E and Zinc ameliorates Cadmium toxicity induced biochemical changes in male albino rats. *Toxicol. Forensic Med.*, **7**, 13-19 (2021).
6. Treviño, S., Pulido, G., Fuentes, E., Handal-Silva, A., Moreno-Rodríguez, A., Venegas, B., Flores, G., Guevara, J. and Díaz, A. Effect of cadmium administration on the antioxidant system and neuronal death in the hippocampus of rats. *Synapse*, **76**, 22242-22246 (2022).
7. Demir, H., Kanter, M., Coskun, O., Uz, Y.H., Koc, A. and Yildiz, A. Effect of black cumin (*Nigella sativa*) on heart rate, some hematological values, and pancreatic  $\beta$ -cell damage in cadmium-treated rats. *Biological Trace Element Research*, **110**, 151-162 (2006).
8. El-Boshy, M.E., Risha, E.F., Abdelhamid, F.M., Mubarak, M.S. and Hadda, T.B. Protective effects of selenium against cadmium induced hematological disturbances, immunosuppressive, oxidative stress and hepatorenal damage in rats. *Journal of Trace Elements in Medicine and Biology*, **29**, 104-110 (2015).
9. Wang, L., Cao, J., Chen, D., Liu, X., Lu, H. and Liu, Z. Role of oxidative stress, apoptosis, and intracellular homeostasis in primary cultures of rat proximal tubular cells exposed to cadmium. *Biological Trace Element Research*, **127**, 53-68 (2009).
10. Shuh, M., Bohorquez, H., Loss, G.E. and Cohen, A.J. Tumor necrosis factor- $\alpha$ : life and death of hepatocytes during liver ischemia/reperfusion injury. *Ochsner Journal*, **13**, 119-130 (2013).
11. Xu, D.P., Li, Y., Meng, X., Zhou, T., Zhou, Y., Zheng, J., Zhang, J.J. and Li, H.B. Natural antioxidants in foods and medicinal plants: Extraction, assessment and resources. *International Journal of Molecular Sciences*, **18**, 96-102 (2017).
12. Adikwu, E. and Deo, O. Hepatoprotective Effect of Vitamin C (Ascorbic Acid). *Pharmacology & Pharmacy*, **4**, 84-92 (2013).
13. Nayak, S.K., Swain, P. and Mukherjee, S.C. Effect of dietary supplementation of probiotic and vitamin C on the immune response of Indian major carp, *Labeo rohita* (Ham.). *Fish & Shellfish Immunology*, **23**, 892-896 (2007).
14. Al Sammak, M., Ahmed, R.M. and Alazzo, N. The Role of Vitamin C in Amelioration of Hepatorenal Toxicity of Cefotaxime in Adult Albino Rats (Histological Study). *Journal of Medical Sciences*, **9**, 845-848 (2021).
15. Yuan, G., Dai, S., Yin, Z., Lu, H., Jia, R., Xu, J., Song, X., Li, L., Shu, Y. and Zhao, X. Toxicological assessment of combined lead and cadmium: acute and sub-chronic toxicity study in rats. *Food and Chemical Toxicology*, **65**, 260-268 (2014).
16. Hussen Ali, T. Determination of the lethal dose 50%(LD50) of cadmium chloride and the histopathological changes in male mice liver and kidneys. *Journal of Education and Science*, **25**, 27-38 (2012).
17. Dankbar, B., Padró, T., Leo, R., Feldmann, B., Kropff, M., Mesters, R.M., Serve, H., Berdel, W.E. and Kienast, J. Vascular endothelial growth factor and interleukin-6 in paracrine tumor-stromal cell interactions in multiple myeloma. *The Journal of the American Society of Hematology*, **95**, 2630-2636 (2000).
18. Kaneko, Y., Kimura, T., Kishishita, M., Noda, Y. and Fujita, J. Cloning of apg-2 encoding a novel member of heat shock protein 110 family. *Gene*, **189**, 19-24 (1997).

19. Nadeem, M., Abdullah, M., Hussain, I., Inayat, S., Javid, A. and Zahoor, Y. Antioxidant potential of *Moringa oleifera* leaf extract for the stabilisation of butter at refrigeration temperature. *Czech Journal of Food Sciences*, **31**, 332-339 (2013).
20. Bancroft, J.D. and Gamble, M. eds. Theory and practice of histological techniques. *Elsevier Health Science*, **3**, 1-11 (2008).
21. Elshopakey, G.E. and Elazab, S.T. Cinnamon aqueous extract attenuates diclofenac sodium and oxytetracycline mediated hepato-renal toxicity and modulates oxidative stress, cell apoptosis, and inflammation in male albino rats. *Veterinary Sciences*, **8**, 1-9(2021).
22. Thiex, N., Novotny, L. and Crawford, A. Determination of ash in animal feed: AOAC official method 942.05 revisited. *Journal of AOAC International*, **95**, 1392-1397 (2012).
23. Bax, L., Ikeda, N., Fukui, N., Yaju, Y., Tsuruta, H. and Moons, K.G. More than numbers: the power of graphs in meta-analysis. *American Journal of Epidemiology*, **169**, 249-255(2009).
24. Hyder, O., Chung, M., Cosgrove, D., Herman, J.M., Li, Z., Firoozmand, A., Gurakar, A., Koteish, A. and Pawlik, T.M. Cadmium exposure and liver disease among US adults. *Journal of Gastrointestinal Surgery*, **17**, 1265-1273 (2013).
25. Shi, T., Zhang, Y., Gong, Y., Ma, J., Wei, H., Wu, X., Zhao, L. and Hou, H. Status of cadmium accumulation in agricultural soils across China (1975–2016): From temporal and spatial variations to risk assessment. *Chemosphere Journal*, **230**, 136-143(2019).
26. Hong, D., Min, J.Y. and Min, K.B. Association between cadmium exposure and liver function in adults in the United States: a cross-sectional study. *Journal of Preventive Medicine and Public Health*, **54**, 471-480 (2021).
27. Zamani, M.M., Mortazavi, S.H., Monajjemzadeh, M., Piranfar, V., Aalidaeijavadi, Z. and Bakhtiarian, A. Protective Effect of Combined Long Time Administration of Selenium and Vitamin C on Liver and Kidney Toxicity of Cadmium in Rats. *Iranian Journal of Pathology*, **16**, 174-180 (2021).
28. Thévenod, F., Schreiber, T. and Lee, W.K. Renal hypoxia–HIF–PHD–EPO signaling in transition metal nephrotoxicity: *Archives of Toxicology*, **96**, 1573-1607 (2022).
29. Hounkpatin, A.S.Y., Johnson, R.C., Guédénou, P., Domingo, E., Alimba, C.G., Boko, M. and Edorh, P.A. Protective effects of vitamin C on haematological parameters in intoxicated Wistar rats with cadmium, mercury and combined cadmium and mercury. *International Research Journal of Biological Sciences*, **1**, 76-81 (2012).
30. Horiguchi, H., Oguma, E. and Kayama, F. Cadmium induces anemia through interdependent progress of hemolysis, body iron accumulation, and insufficient erythropoietin production in rats. *Toxicological Sciences*, **122**, 198-210 (2011).
31. Kataranovski, M., Janković, S., Kataranovski, D., Stošić, J. and Bogojević, D. Gender differences in acute cadmium-induced systemic inflammation in rats. *Biomedical and Environmental Sciences*, **22**, 1-7(2009).
32. Hossein-Khannazer, N., Azizi, G., Eslami, S., AlhassanMohammed, H., Fayyaz, F., Hosseinzadeh, R., Usman, A.B., Kamali, A.N., Mohammadi, H., Jadidi-Niaragh, F. and Dehghanifard, E. The effects of cadmium exposure in the induction of inflammation. *Immunopharmacology and Immunotoxicology*, **42**, 1-8(2020).
33. Piskin, E., Cianciosi, D., Gulec, S., Tomas, M. and Capanoglu, E. Iron absorption: factors, limitations, and improvement methods. *ACS omega*, **7**, 20441-20456(2022).
34. Samantaray, B.P. and Parida, S.P. Vitamin-C treated haematology assessment of Swiss albino mice (*Mus musculus*). *Biointerface Research in Applied Chemistry*, **11**, 9044-9050 (2021).
35. Gu, X. and Manautou, J.E. Molecular mechanisms underlying chemical liver injury. *Expert Reviews in Molecular Medicine*, **14**, 1-4(2012).
36. Hall, P. and Cash, J. What is the real function of the liver ‘function’ tests?. *The Ulster Medical Journal*, **81**, 30-36(2012).



37. Go, Y.M., Sutliff, R.L., Chandler, J.D., Khalidur, R., Kang, B.Y., Anania, F.A., Orr, M., Hao, L., Fowler, B.A. and Jones, D.P. Low-dose cadmium causes metabolic and genetic dysregulation associated with fatty liver disease in mice. *Toxicological Sciences*, **147**, 524-534 (2015).
38. Genchi, G., Sinicropi, M.S., Lauria, G., Carocci, A. and Catalano, A. The effects of cadmium toxicity. *International Journal of Environmental Research and Public Health*, **17**, 3782-3788 (2020).
39. Abdulrazzaq, A.M., Badr, M., Gammoh, O., Abu Khalil, A.A., Ghanim, B.Y., Alhussainy, T.M. and Qinna, N.A. Hepatoprotective actions of ascorbic acid, alpha lipoic acid and silymarin or their combination against acetaminophen-induced hepatotoxicity in rats. *Medicina*, **55**, 181-187 (2019).
40. De Feo, P. and Lucidi, P. Liver protein synthesis in physiology and in disease states. *Current Opinion in Clinical Nutrition & Metabolic Care*, **5**, 47-50 (2022).
41. Helal, E.G., Abdelaziz, M.A., Fadel, H.A. and El-Shenawe, N.S. The Therapeutic Effects of Vitamin C on Changes of Some Biochemical Parameters in Male Albino Rats Treated with Mixture of Food Additives (Sodium Benzoate+ Mono Sodium glutamate+ Chlorophyllin). *The Egyptian Journal of Hospital Medicine*, **75**, 3131-3138(2019).
42. Ibiam, A.U., Ugwuja, E.I., Ejeogo, C. and Ugwu, O. Cadmium-induced toxicity and the hepatoprotective potentials of aqueous extract of *Jessiaea nervosa* leaf. *Advanced Pharmaceutical Bulletin*, **3**, 309-313(2013).
43. Hussein, S.A., Omnia, M. and Fayed, A. MF. Protective effects of alpha-lipoic acid and melatonin against cadmium-induced oxidative stress in erythrocytes of rats. *Journal Pharmacology and Toxicology*, **9**, 1-24(2014).
44. Ighodaro, O.M. and Akinloye, O.A. First line defence antioxidants-superoxide dismutase (SOD), catalase (CAT) and glutathione peroxidase (GPX): Their fundamental role in the entire antioxidant defence grid. *Alexandria Journal of Medicine*, **54**, 287-293(2018).
45. Andrés, C.M.C., Pérez de la Lastra, J.M., Juan, C.A., Plou, F.J. and Pérez-Lebeña, E. Chemistry of Hydrogen Peroxide Formation and Elimination in Mammalian Cells, and Its Role in Various Pathologies. *Stresses*, **2**, 256-274(2022).
46. Glisic, B., Mihaljevic, I., Popovic, M., Zaja, R., Loncar, J., Fent, K., Kovacevic, R. and Smital, T. Characterization of glutathione-S-transferases in zebrafish (*Danio rerio*). *Aquatic Toxicology*, **158**, 50-62(2015).
47. Hayes, J.D., Flanagan, J.U. and Jowsey, I.R. Glutathione transferases. *Annual Review of Pharmacology and Toxicology*, **45**, 51-88(2005).
48. Rizwan, S., Naqshbandi, A., Farooqui, Z., Khan, A.A. and Khan, F. Protective effect of dietary flaxseed oil on arsenic-induced nephrotoxicity and oxidative damage in rat kidney. *Food and Chemical Toxicology*, **68**, 99-107(2014).
49. Ogunrinola, O.O. Lipid profile and malondialdehyde concentrations in cadmium-induced rats: a study with relation to doses. *MOJ Toxicology*, **1**, 1-6(2015).
50. Chandra, D., Tripathi, U.N., Srivastava, S. and Swaroop, A. Carbofuran induced biochemical toxicity in mice: Protective role of *Momordica charantia*. *European Journal of Experimental Biology*, **1**, 106-112(2011).
51. Schwabe, R.F. and Brenner, D.A. Mechanisms of liver injury. I. TNF- $\alpha$ -induced liver injury: role of IKK, JNK, and ROS pathways. *American Journal of Physiology-Gastrointestinal and Liver Physiology*, **290**, G583-G589(2006).
52. Ozmen, O. and Mor, F. Effects of vitamin C on pathology and caspase-3 activity of kidneys with subacute endosulfan toxicity. *Biotechnic & Histochemistry*, **90**, 25-30(2015).

## التأثير التحسيني لفيتامين سي ضد التأثير المثبط للمناعة والمتغيرات في الكيمياء الحيوية والدموية والأوكسدة لكلوريد الكادميوم على الجرزان

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كان الهدف الاساسي من هذا البحث هو التأكد مما إذا كان لفيتامين سي تأثيراً وقائياً آمناً في مواجهة الآثار الضارة للكادميوم الذي يعتبر من المعادن الثقيلة ذات السمية العالية والتلوث البيئي الكبير , تكونت المجموعة التجريبية من ٢٠ فأر مقسمة إلى ٤ مجموعات , أجريت الدراسة عليها لمدة ٣٠ يوم , قسمت على النحو التالي : مجموعة أساسية لم تعطى شيء , ( مجموعة كلوريد الكادميوم ) تم تجريعها بـ ١٥ مجم / كجم من كلوريد الكادميوم , ( مجموعة فيتامين سي ) تم تجريعها بـ ١٥٠ مجم / كجم من فيتامين سي , ( مجموعة كلوريد الكادميوم مع فيتامين سي ) تم تجريعها بـ ١٥ مجم/كجم من كلوريد الكادميوم و ١٥٠ مجم / كجم من فيتامين سي , وكانت جميع الجرعات عن طريق الفم , وبعد انقضاء ٤ أسابيع من التجريع تم أخذ عينات مختلفة من الدم وأنسجة الكبد لإجراء الفحوصات المختلفة , حيث كشفت النتائج عن انخفاض في عدد كريات الدم الحمراء , وتركيز الهيموجلوبين , وحجم الخلايا المكسدة بما يعادل (  $P < 0.05$  ) , وارتفاع عدد كريات الدم البيضاء بما يعادل (  $P > 0.05$  ) (من ناحية أخرى , كما كشفت النتائج عن انخفاض كبير في البروتين الكلي والألبومين وزيادة تركيز الجلوبيولين والبيروبين الكلي والمباشر , وزيادة تركيز إنزيمات الكبد (ALT,AST,ALP) بالإضافة لمضادة الأكسدة (GST,GSH,CAT,SOD) , وارتفاع MAD لمجموعة كلوريد الكادميوم مقارنة بالمجموعة الأساسية التي لم يتم اعطاءها شيء , ولكن عند دمج فيتامين سي مع كلوريد الكادميوم أشارت النتائج إلى تحسن ملحوظ في المعلمات قيد الدراسة , مما يشير إلى التأثير العلاجي لفيتامين سي ضد التسمم بكلوريد الكادميوم في الفئران .

**الكلمات الدالة :** كادميوم – فيتامين سي – إنزيمات مضادة للأكسدة – دلالات مضادات الأكسدة – إمكانية التحسين .