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Risk Assessment of Heavy Metal Pollution in *Oreochromis niloticus* from the Rosetta Branch of the River Nile, Egypt

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ABSTRACT

During the last three decades, an annual proliferation in the levels of heavy metals was detected in the River Nile due to sewage, industrial and agricultural wastewater discharged into the river waters. Thus, the current study was set with the aim of investigating the possible health risks associated with water use and the consumption of Oreochromis niloticus via comparing and determining various exposure pathways and potential health risks corresponding with heavy metals in surface water and fish off the River Nile using a number of indices. S1: El-Qanater El-Khayria site (S1; reference clean site) prior to the bifurcation, S2: El-Rahawy drain discharge point into the Rosetta Branch, and S3: El-Qatta site (about 7 km from S2) were the three locations covered by the present investigation throughout winter and summer of 2023. Heavy metals' levels were addressed (Fe, Cr, Cd, Zn, and Cu). Among the metals under research, Fe had the highest concentration, followed by Zn> Cu> Cr> Cd, and S2 had the highest heavy metal load over the course of the study. In contrast to the FAO guidelines for irrigation water and the USEPA guidelines for drinking water, water samples from S1 of the River Nile were suitable for both uses during the study, despite the fact that the heavy metal evaluation index (HMEI) of water samples taken from S2 and S3 of the Nile indicated that both sites are unsuitable for irrigation and drinking. O. niloticus off site S3 was monitored for non-carcinogenic health concerns in accordance with USEPA recommendations. However, chromium in fish at S3 exhibited higher lifetime potency of cocarcinogenic risks, particularly for habitual consumers. The current findings confirm the necessity of rapid water quality management planning, especially in heavily polluted areas, to secure safe water usage and maintain ecological balance and human well-being. These results also emphasize the importance of understanding the site-specific cancerogenic risks related to exposure to heavy metals in different locations and consumption patterns to appraise aquatic food safety and health risk assessments.

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INTRODUCTION

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Environmental pollution has been ranked as a critical problem in the 20th century, which requires national and international incorporated efforts to be encountered (**Azab** *et al.*, **2019**). Most environmental pollutants threaten the welfare of humans and the environment, as well as ecological productivity and integrity (**Said** *et al.*, **2021**). The previous authors attributed the

increase in chemical contamination to excessive anthropogenic activities. Unfortunately, this kind of pollution has a great impact on both aquatic and terrestrial environments.

Globally, water pollution has become a problem of high concern, which negatively impacts water's quality and ceases its benefit (Ali *et al.*, 2016). There are seven primary groups of contaminants that can contaminate water: inorganic, organic, radioactive materials, sediments, suspended solids, heat, and heavy metals (Botkin & Keller, 2008).

Water bodies, specifically rivers are markedly polluted since they receive urban, rural, and industrial wastes that usually contain high concentrations of hazardous pollutants (**Zyadah**, **1999; Azab** *et al.*, **2019**). The Nile is the principal body of fresh water in Egypt (**Hashem** *et al.*, **2020**). It runs through the narrow valley, then bifurcates forming two branches, the Damietta and the Rosetta, forming the Delta (**Abdel-Satar** *et al.*, **2017**). Currently, various challenges have encountered the Rosetta Branch. Three essential sources of pollutants have been detected releasing their effluents into the branch under estimation. El-Rahawy drain, where agricultural and domestic discharges are being released from the Giza City, forms the 1st source. Effluents discharged are estimated to be more than 1,900,000m³ /day. Another source of pollutants originated from the industrial region of Kafr El-Zayat, while pollution spotted from numerous tiny agricultural drains, with their wastes discharged into the Branch, has been assessed to be the third source. Furthermore, the sewage released into the branch of several cities and villages has added to the effluents received through the third source (**El Bouraie** *et al.*, **2011**).

The Rossetta Branch and its aquatic organisms are facing serious problems, among which is the heavy metal pollution (Darweesh et al., 2019). The previous authors elucidated that the heavy metal levels' annual ratio has experienced an increase in the River Nile. The high ratio of heavy metals in the habitat is caused by anthropogenic and natural activities, among which are sewage, agricultural and industrial wastewater, microplastics, organic fuel combustion, pesticides, and phosphate fertilizers (Said et al., 2021). Heavy metals are common harmful pollutants that damage people, plants, water, and the air these days (Huang et al., 2020; Khattab et al., 2021, Gouda & Taha, 2023; Taha & Gouda, 2023). Once in the water, these toxic, bioaccumulating and non-biodegradable materials precipitate on sediment particles and build up in the food chain by damaging the organs and tissues of aquatic life (Dural et al., 2006; Qiu, 2015; Mohamad et al., 2020; Younis et al., 2024b). It is significant to mention that the main sources of heavy metal accumulation are food and water, followed by muscles and gills (Awad et al., 2024). It was discovered that, in addition to aquatic animals' feeding patterns, the pH, temperature, and heavy metals concentrations all affect the buildup of heavy metals (Ali et al., 2016). The two categories of heavy metals are essential and non-essential. The vital role that critical heavy metals like Cr, Zn, Fe, Mn, and Cu play in biofunctions makes them vital for living things. Due to the harmful effects of both excess and deficiency of the important heavy metals, they are advised for this purpose at low quantities (Santos et al., 2014). Heavy metals that are not necessary, like Pb, Hg, and Cd, have no biological purpose in living things; they are only needed in minimal amounts and turn poisonous at high concentrations (Gati et al., 2016). The estimated 23.3% of the nation's total natural source production is derived from fish produced in the River Nile (GAFRD, **2015**). Being at the top of the aquatic food chain, eating fish would have a harmful impact on people since fish organs and tissues accumulate heavy metals (**Abdel-Mohsien & Mahmoud**, **2015**). According to **Mohamed** *et al.* (**2020**), fish can therefore be employed as bioindicators for the identification of aquatic contamination.

Because of their low cholesterol, sufficient omega-3 fatty acids in addition to other beneficial nutrients, fish are good for reducing the rate of cholesterol, the threat of heart disease as well as stroke in people. Thus, an increase has been detected in the consumption of fish (Knuth *et al.*, **2003; Abdel-Mohsien & Mahmoud, 2015**). Accordingly, the accumulation of heavy metals in fish and other aquatic animals can be thought of as a closed circuit, with the resultant negative health effects on humans, including kidney diseases, cancer, mental illness, developmental abnormalities, and neuromuscular issues (Zhuzzhassarova *et al.*, **2024**).

O. niloticus is the fish species most frequently found in the River Nile. For the Egyptians, it is highly popular, being the cheapest compared to other species and available all the year- round (Abdel-Mohsien & Mahmoud, 2015; El-Naggar *et al.*, 2021). *O. niloticus* is capable of tolerating deteriorated environmental conditions due to its high physical resistance and slight respiration demand, allowing it to withstand ammonia with high amounts in addition to low oxygen (Zhou *et al.*, 1998).

With the aim of addressing heavy metal pollution in the Rosetta Branch of the River Nile and the accumulation of these metals in *O. niloticus*, the present investigation was conducted. In addition, the study focused on determining the validity of water for both drinking and irrigation, as well as assessing the potential health risks in relation with oral and dermal contact of water and the ingestion of the Nile tilapia from the River Nile.

MATERIALS AND METHODS

1. Study area

The Rosetta Branch of the Nile River is roughly 225km long, 150–200 m wide, with an average depth of 2.0– 3.5m. Every day, the branch receives and discharges drainage water from agricultural areas, as well as partially treated and untreated domestic and industrial wastewaters with cubic meters of 3 million or more (**Gaber** *et al.*, **2013**). It terminates at the Rosetta Estuary after beginning at El-Qanater El-Khayria. El-Rahawy, the Zawyet El-Bahr, El-Tahrer, the Tala, and the Sabal, which are the five agricultural drains that discharge into the branch (**Donia**, **2005**). The present study focused on El-Rahawy drain, which is an extremely big drain receiving industrial, agricultural, and domestic wastes of Giza City. It discharges more than 1,9 million m^3/day as sewage waste directly into the Rosetta Branch.

The inquiry study area spans over 20 kilometers from the El-Qanater El-Khayria site to the El-Qatta site. Three locations were the subject of the current study: S1: El-Qanater El-Khayria site (reference clean site) before to the bifurcation; S2: El-Rahawy drain discharge point into the

Rosetta Branch (about 12.6km from S1); and S3: El-Qatta site (approximately 7km from Site 2) (Fig. 1). The latitudes and longitudes of the sites under study are displayed in Table (1).

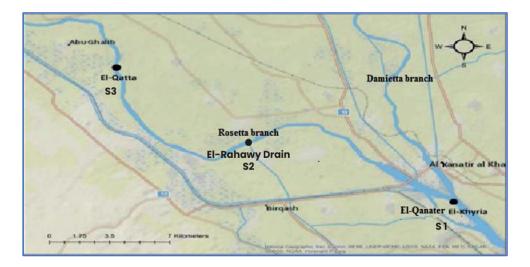


Fig. 1. The Nile Delta map showing sites under investigation (Mohamed et al., 2020)

Site	Location	Location Latitude				
S1	El-Qanater El-Khayria (before the	30° 10' 19.0344" N	31° 8' 24.0252" E			
	bifurcation)					
S2	El-Rahawy drain (discharge point)	30° 12' 20.0988" N	31° 1' 40.962" E			
S3	Al-Qatta site (downstream EL-Rahawy	30° 13' 27.6636" N	30° 58' 26.1444" E			
	drain)					

Table 1. Longitudes and latitudes of sites under study (GPS)

2. Samples collection

2.1. Water sampling

Under the code NIOF-FW3-F-23-R-003, the current study was conducted in compliance with the ethical requirements of the National Institute of Oceanography and Fisheries' (NIOF, Egypt) committee for the ethical care and use of animals/aquatic creatures. Throughout two seasons in 2023 (winter & summer), samples from surface water were taken from the three study locations, down to a depth of 30cm.

2.2.Fish sampling

Thirty *O. niloticus* specimens, regardless of their sex, with a mean total length of 20.03 ± 1.62 cm and mean total weight of 163.19 ± 24.08 g were sampled during the study period, with the help of fishermen available at two sites: S1 S3 (affected by El-Rahawy drain) with the help of fishermen. No fish were found alive during winter at S2, El-Rahawy drain discharge point.

3. Determination of HM

3.1. HMs analysis in water

For heavy metals analysis, the flame atomic absorption spectrophotometer (Savant AA, GBC Scientific Equipment) was utilized in the Central Laboratory, Faculty of Science, Ain Shams University in accordance with the American Public Health Association's Standard Method (APHA-3111B, 2017). The findings of the analysis of five heavy metals (Zn, Cu, Cd, Fe, and Cr) were expressed in µg/L.

3.2. HMs analysis in fish samples

According to **El-Dakar** *et al.* (2021), with minor modifications, fish were anesthetized with 40 mg/L clove oil upon capture. The muscles, liver, and gills of the fish were then removed, chopped into tiny pieces, and kept in an ice box until they were sent to the Central Laboratory, Faculty of Science, Ain Shams University, for the measurement of the aforementioned heavy metal levels.

According to **El-Naggar** *et al.* (2022), the subsequent procedure was implemented: The CEM microwave sample preparation equipment (MDS-2000, USA) was employed to digest the samples before analysis. Nitric acid (HNO₃) in a concentrated form was added to containers containing one gram of ground material.

For an adequate reaction, the vessels were left overnight and later, a turntable, where vessels were placed, was attached to the system and a heating program was implemented. Vessels were set to run till digestion was completed. Afterward, samples cooled down for 5min, followed by separating the turntable from the system.

For the heavy metals analysis, the digested samples were merged into 25ml of distilled water and were placed in the flame atomic absorption spectrophotometer. Each heavy metal concentration was measured at a specific slit width and wavelength. The resultant concentration was expressed in mg/kg of the tissue.

4. Risk assessment

4.1. Health hazard assessment for water uses

4.1.1 Water samples' heavy metal assessment index (HMEI)

Heavy metal (HM) concentrations are evaluated using HMEI to determine the water quality's numeric value, adhering to the permissible standard of the same HM for irrigation (FAO, 2019) and drinking (USEPA, 2018). This step was accomplished using Singh *et al.* (2017)'s equation. When HMEI is less than 1.0, water samples are considered felicitous for human use; if HMEI is greater than 1.0, they are considered unsuitable.

4.1.2 The daily chronic intake (CDI)

CDI was estimated as stated by **USEPA** (2004) for dermal adsorption and oral ingestion of HM detected in water samples of the River Nile.

4.1.3 Target hazard quotient (THQ)

The THQ is considered to compute the interplay of HM exposures through water and identify the safety threshold, which doesn't persuade antagonistic health impacts on humans, conforming the equations of **USEPA (2004)** and the reference dose guidelines of each metal (mg/Kg Day) for the dermal contact and ingestion of water established by **USEPA (2011)**.

4.1.4 Hazard index (HI)

The HI is the accumulation of total THQ; it reveals the potential non-carcinogenic health hazards encouraged by HM in water following **USEPA**'s (2004) equations. HI \leq 1.0 reflects the anticipation of no harmful healthiness consequences.

4.2. Assessment of Health hazard for fish consumption

4.2.1 Estimated daily intake (EDI) of HM

EDI was estimated for adults (70Kg) using the outline ingestion rates of fish (habitual & normal) covering a lifespan of 70 years, as detailed in the research works of **Song** *et al.* (2009) and **Salaah** *et al.* (2022).

4.2.2 Target hazard quotient (THQ)

The THQ is generally conducted to assess the non-carcinogenic hazards of every single consumed metal by means of the metal's reference dose stated by **USEPA** (2011, 2012).

4.2.3 Hazard index (HI)

HI is the metals' collective THQ. It presents the comprehensive health risk owing to vulnerability to several metals (USEPA, 2011).

4.2.4 Target cancer risk (TR)

According to USEPA (2011), TR is obtained to evaluate the likely carcinogenic risk in correspondence with the utilization of specific HM (Cd Cr) from fish. At $\leq 1.0 \times 10$ -6, Σ TR and TR are considered insignificant (USEPA, 2010).

5. Statistical analysis

The season and site efficacy on the heavy metal bioaccumulation in the tissues of *O. niloticus* were ascertained by employing Jeffreys's Amazing Statistics Program (JASP 0.16.4) to perform a two-way analysis of variance (ANOVA) on the data of the heavy metal levels in the tissues. Three separate analyses were conducted. Tukey's test (**Abdi & Williams, 2010**) was used to elucidate the differences between the specimens under study.

RESULTS AND DISCUSSION

1. Heavy metals concentrations in water

Heavy metals' average concentrations detected in the samples of surface water at the studied sites throughout the study period are shown in Fig. (2). Generally, they were determined in a descending order, as follows: Fe, Zn, Cu, Cr, and Cd.

As shown in Fig. (2A), the three sites' iron concentrations ranged from a maximum average value of 1200µg/ L during winter at S2 to a minimum mean value of 200µg/ L during summer at S1. According to the CCME (2017), the iron levels in the water at the three sites exceeded the threshold limit of $300 \mu g/L$, except for the Fe level that was recorded during summer at S1. The aforementioned findings coincide with the outcomes of Hashem et al. (2020) and Mohamed et al. (2020), who illustrated that the concentrations of Fe at Sites 2 and 3 surpassed the allowable levels. The phytoplankton's utilization of iron (Canli & Kalay, 1998), oxidation of Fe2+ to Fe3+, and subsequent precipitation of hydroxide an elevated dissolved oxygen content may be the reasons why the lowest concentration of Fe was detected at S1 during summer (Ghallab, 2000). S2 and S3 had the greatest mean Fe concentrations in both seasons (1200 and 950µg/L, respectively), with S3 having the highest concentration in winter (900µg/ L). The outflow of effluents from El-Rahawy drain, being filled with both domestic and agricultural sewage, may be the cause of the high Fe content at sites 2 and 3 (Haggag, 2017). Furthermore, according to El-Sayed and Salem (2015), this high value can be a result of the release of this heavy metal into the water through the breakdown of organic matter and dead microbes. Abdelsalam et al. (2024) added that the lowest concentration of iron during summer is due to the suspending of iron particles, surface microorganisms, presence of metal oxides owing to high temperature.

At the current three sites under study, zinc concentrations varied from a minimum average concentration of $80\mu g/$ L during the summer at S1 to a maximum mean level of $200\mu g/$ L during winter at S2 (Fig. 2B). Notably, the **CCME (2017)** found that the levels of zinc in the water of the three research locations were higher than the standards. The results presented above are consistent with those of **Hashem** *et al.* (2020), who hypothesized that zinc amount at sites 2 and 3 was higher than the allowed. Prior to S3 ($120\mu g/$ L) in winter, S2 under investigation had the highest zinc level (200 and $130\mu g/$ L, respectively). Abdel-Satar *et al.* (2017) suggest that the impact of anthropogenic activities could be the cause of this elevated concentration at S2. One of the most common contaminants found in waste and water, both in liquid and solid form, is zinc. Furthermore, due to its acute toxicity and lack of biodegradability, zinc is classified as a hazardous waste (Younis *et al.*, 2024 a). Accordingly, Abdelsalam *et al.* (2024) ascribed the increasing Zn concentration in winter to the drought period, when the water level drops, which in turn increases the microbiota activity and causes organic drips to bioaccumulate. These organic

drips may act as humic acids that contain a variety of heavy metals, such as Cu, Zn, Fe, Cd, Pb, and Mn. **Ezzat** *et al.* (2024) ascribed the elevated Zn concentration in aquaculture to the drainage of agricultural and industrial wastes into the water.

Data denoted in Fig. (2C) show that the highest wintertime concentration of Cu was recorded at S2 (55 μ g/L), followed by S3 (40 μ g/L). It was noted that during the summertime, the Cu content at S1 was less than the standards of the **Egyptian Governmental Law no. 48/1982-Decision 92 (2013)** (6 μ g/L)). The tendency of Cu to form complexes with humic matter and organic ligands, eliminating free ions' penetration into the water, is thought to be the cause of the lowest Cu level in water during summer at the studied sites (Mantoura *et al.*, 1978; Moustafa, 2017). For Cu levels, they surpassed the acceptable levels during winter at S2 & S3, whereas they were within the thresholds of the **Egyptian Governmental Law no. 48/1982- Decision 92** (2013) at S1 in summertime (10 μ g/L). Accordingly, the previous findings coincide with those of **Hashem** *et al.* **(2020)** and **Mohamed** *et al.* **(2020), who revealed that the Cu levels at Sites 2 and 3 exceeded the standard limits. The aforementioned data may be ascribed to the decline in the level of water in the River Nile during the rainy season as well as the rise in domestic waste at El-Rahawy drain, whose discharge will affect the metal's level at S3.**

Notably, the Cr concentration in water at S1 was within the quality standard values of the CCME (2017) (10 μ g/L). Cr concentrations at the investigated sites ranged between a maximum mean value of 26 μ g/L during winter at S2 and a minimum average value of 7 μ g/L during the dry season at S1 (Fig. 2D). It was noticed that Cr concentration exceeded the thresholds of the CCME (2017) at both sites (2 & 3) during both seasons (Fig. 2D). It was annotated that heavy metals' levels in the investigated samples showed season-specific variations, being the highest during the winter months (Abdel-Moati & El-Sammak, 1997). This may be due to the increase in the amount of industrial, agricultural, and sewage wastes that are discharged into the River Nile during winter (Khalil *et al.*, 2017). Conversely, the decrease of the HM contents in summer may be interpreted as an outcome of the phytoplankton's development, which can absorb significant amounts of HM from water, in addition to the rise in the River Nile's water level throughout the summer season (Tayel *et al.*, 2018).

Cd concentrations at the sites under investigation ranged between a maximum average value of $12\mu g/L$ during winter at S2 and a minimum mean concentration of $0.5\mu g/L$ during summer at S1 (Fig. 2E). All Cd values at the studied sites exceeded the standards recommended by the **CCME (2017)** and the **Egyptian Standards of the Environmental Laws no. 48/1982 decree 92/2013-Decision 92 (2013)** (1 $\mu g/L$), except for those observed during summer at S1. The previously mentioned findings may account for the effect of pollution sources in the locations under study, such as sewage and landfill at S3, residential wastes at S2 and anthropogenic activities; mainly agricultural at S1 that are especially amplified during the rainy season.

In their study (**El-Degwy** *et al.*, **2023**), an association was recorded between the elevated levels of Zn and Cd in the water of Lake Mariout and the high temperatures and fermentation during summer, with the former subsequently releasing from the bottom sediment into the water column above.

They concluded that all the investigated heavy metals' concentrations are greater in the rainy season compared to the dry one. Furthermore, the fluctuations in the levels of the heavy metals (Fe, Zn, Cu, Cr & Cd) at the studied sites may be ascribed to the dynamic variation of wastewater flow in the area or the time of water sample collection. Such seasonal variation of heavy metal levels in water is traced back to the variation of domestic, industrial and agricultural activities, Moreover, such differences may be related to the hazard impacts of varied pollutants, viz. residential, agricultural, and industrial ones (**El-Degwy** *et al.*, **2023**).

2. Heavy metals' bioaccumulation in various tissues of O. niloticus

Fish are commonly used as biological indicators to assess the level of metal contamination in aquatic environments (Al-Sayegh Petkovšek *et al.*, 2012). This is confirmed since they can accumulate high concentrations of certain metals found in the water (Daviglus *et al.*, 2002). When contaminated fish are consumed, toxic elemental pollutants are introduced into the human body, which can lead to a significant decline in health (Alinnor & Obiji, 2010). Therefore, consuming fish in amounts exceeding the recommended safety levels can be extremely harmful to humans (Basiony, 2014).

Heavy metals are known to aggregate in the tissues of fish. Through direct ingestion from the sediment and water or indirect ingestion throughout the food chain, they may acquire metals. These metals are subsequently built up in fish tissues and organs in amounts greater than those in their environment, which might have negative consequences (Agah *et al.*, 2009). Based on data in the study of Neima *et al.* (2016), heavy metals usually form cationic complexes built up in the internal organs of fish. According to Neima *et al.* (2016), the accumulation of heavy metals is often impacted by the concentration of the metal, the mode of metal uptake, the duration of exposure, intrinsic characteristics (like fish age and eating habits), and environmental factors (viz. salinity, temperature, and pH). Due to variations in permeability, metabolic rates, and the types and quantities of metal-binding ligands on the organism's surface, this buildup differs with the species, the type of heavy metal, and even the individual (Campenhout *et al.*, 2004; Javed & Usmani, 2019).

An understanding of the relationship between fish and their external environment can be gained from the accumulated metals in key organs, mirroring the concentrations of these metals in the fish species' surrounding environment (Monroy *et al.*, 2014).

2.1. Heavy metal contents in the gills tissues of O. niloticus

The mean concentrations of HMs in the gills samples of O. *niloticus* off different sites in the River Nile covering the period of work are depicted in Table (1). At both sites, the average levels of heavy metals in the gills were in the descending order of Fe> Zn> Cu> Cr> Cd.

Remarkably, site- as a factor- was recorded with a significant (P < 0.05) effect for HMs under investigation, monitoring high significant mean values at S3 compared to those recorded at S1. This means that the gills, due to their anatomical position, showed high levels of some metals' accumulation patterns, since they are continuously and directly affected by the surrounding contaminants (**Abdel-Khalek** *et al.*, **2016**).

Moreover, the levels of Fe, Zn, Cu, and Cr were recorded to be significantly (P < 0.05) impacted by site and season. Their highest significant values were recorded at S3 during winter (133.97, 45.12, 4.31, and 2.76µg/ g, respectively), while their lowest significant levels were found at S1, during summer (46.00, 23.79, 2.09 and 1.11µg/ g, respectively). On the other hand, neither the season nor the interaction between the two factors exhibited a crucial effect on the values of Cd, though they numerically differed. Furthermore, it was noted that the average values of Fe and Zn at S3 during winter (133.97 and 45.12µg/ g, respectively) exceeded the threshold of the **WHO** (**1993**) (100 and 40µg/ g). Whereas, the mean concentrations of Cr at both sites during both seasons surpassed the permissible level recommended by the **FAO** (**1983**) (1µg/ g). Whereas, Cd levels at S3 during both seasons (winter & summer) outreached the permissible dose reported by the **WHO** (**1993**) (0.5µg/ g).

In general, the increment of the addressed heavy metals in gills is associated with their elevated concentrations in water originated from the flow of huge quantities of sewage waste into the studied area. These results may be attributed to the following reasons: 1) It is well known that gills are the primary site that create a pathway for heavy metals to go into the fish's body (**Bols** *et al.*, **2001**); 2) Gills, being directly exposed to the environment, are known for their potential to excrete certain metals viz. lead (**Matthiessen & Brafield**, **1977**); 3) Wepener *et al.* (**2001**) noted that gills are the first site where waterborne metals accumulate owing to their bonding with gill cytosolic chemicals, and 4) the metals absorbed onto the gill surface, which serves as the initial point of contact for contaminants in the water, is another reason. Furthermore, according to **Ezzat** *et al.* (**2024**), Zn is highly accumulated in the body of *O. niloticus* in Burullus Lake due to the highly effective uptake and storage through fish gills and internal organs. In contrast with our observations, the previous authors detected a relation between the summer season and the increase in pathological changes and weakness of gills. They deduced that the following pathological changes are traced back to the elevated concentrations of heavy metals throughout summer: gill damage, gas exchange disruption and increasing gills inflammatory responses causing pathogen growth and activation within the gills.

Risk Assessment of Heavy Metals Pollution in *Oreochromis niloticus* from the Rosetta Branch of the River Nile

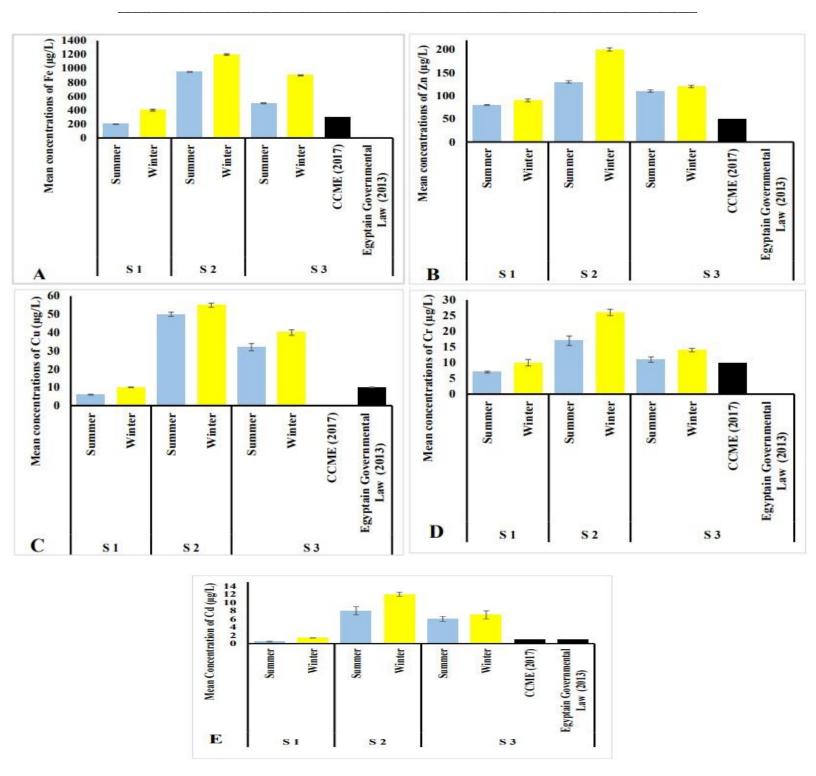


Fig. 2. Mean concentrations of HMs in samples of surface water at the investigated sites throughout the study period including (A) Fe, (B) Zn, (C) Cu, (D) Cr. and (E) Cd S1: El-Qanater El-Khayria (non-polluted site, reference site). S2: El-Rahawy drain (discharge point). S3: Al-Qatta site (polluted site)

2.2 Levels of heavy metals in O. niloticus liver tissues

Table (2) lists the average levels of heavy metals detected in the Nile tilapias' liver samples, with specimens from several locations. In general, liver was recorded with the largest amounts of most heavy metals. This can be confirmed because the liver substantially induces metallothionein and stores redistributes, detoxifies while converting contaminants (Yılmaz *et al.*, 2007). The liver is a highly active organ that is specialized in absorbing and storing a variety of heavy metals (Dural *et al.*, 2006; Yılmaz *et al.*, 2007). Furthermore, the liver tissue's metabolic activity makes it quite likely that most metals will be concentrated in large quantities within its tissues (Omar *et al.* 2014). The average heavy metal levels in the 1 iver at both sites are sequenced as follows: Fe> Zn> Cu> Cr> Cd. It is worth mentioning that all heavy metals were significantly affected by site and season (P < 0.05), in which their highest significant values were recorded at S3 during winter, while their lowest were noted during summer at S1.

Table 1. Mean concentrations of heavy metals $(\mu g/g)$ in *O. niloticus* gills from different sites throughout the study period

		Site per	: season				
	S1	l		53	Site effect (P-value)	Season effect (P-value)	Permissible dose for O. niloticus
Heavy metal	Summer	Winter	Summer	Winter			
Fe	46.00±4.17 ^d	61.04 ±1.8 ^c	96.50± 3.16 ^b	133.97± 0.551 ^a	< .001	< .001	100
Zn	$\begin{array}{c} 23.79 \pm \\ 0.42^{\ d} \end{array}$	30.60± 0.23 °	37.54± 1.21 ^b	45.12± 1.42 ^a	< .001	< .001	40
Cu	$\begin{array}{c} 2.09 \pm \\ 0.008^{\ d} \end{array}$	2.59± 0.169 [°]	3.82± 0.05 ^b	4.31±0.04 ^a	< .001	< .001	30
Cr	1.11± 0.018 ^c	1.59± 0.02 ^b	1.35± 0.14 ^{bc}	2.76± 0.092 ^a	< .001	< .001	1
Cd	0.09±0.007	0.15± 0.004	0.52±0.32	0.73±0.010	0.013	0.447	0.5
Each value	e illustrates the m	ean level± SE,	SE: Standard e	error (n=3).		•	•
Values in	the same row wit	h varied supers	scripts are signi	ficantly differen	t (<i>P</i> < 0.05) by 7	Fukey's test.	
-	rmissible doses a	-		-	which is in ac	cordance with I	FAO (1983).
	anater El-Khayria		site, reference	site).			
S3: Al-Qa	tta site (polluted a	site).					

Moreover, it was noticed that the mean concentrations of Fe at S3 during both seasons were 140.65 and 277.67µg/ g, respectively, whereas at S1 during winter (138.39µg/ g), values surpassed the permissible limits reported by the **WHO** (1993) (100µg/ g). On the other hand, the average values of Zn at S3 and S1 during winter (65.89 and 42.00µg/ g, respectively) exceeded the threshold of the **WHO** (1993) (40µg/ g). However, the mean value of Cu at S3 during winter (53.33µg/ g) exceeded the threshold determined by the **WHO** (1993) (30µg/ g). Whereas Cr

levels outreached the threshold reported by the FAO (1983) $(1\mu g/g)$ at both sites during both seasons. Nevertheless, Cd values at S3 during both seasons (0.64 and 0.84 $\mu g/g$, respectively) exceeded the recommended limits assigned by the WHO (1993) (0.5 $\mu g/g$). Cd is a serious environmental pollutant that can cause detrimental effects on fish, such as growth deficits (Peterson *et al.*, 1983), biochemical effects (Haux & Larasson, 1984), and osmoregulatory problems (Reid & McDonald, 1988) at sublethal doses or slightly high doses (Abdel-Mohsien & Mahmoud, 2015). The elevated levels of Cd may be ascribed to wastes discharged from mining and industrial operations, phosphate fertilizer, and sewage at El-Rahawy drain, where almost all Cd concentrations are accumulated in the habitat (Mohamed *et al.*, 2020).

Major histopathological alterations were induced in the liver of the Nile tilapia by the biomagnification of heavy metals (Cd, Fe, Cu, Zn, and Pb) in Burullus Lake, such as necrosis, pyknosis, and vascular damage of hepatocytes (**Ezzat** *et al.*, **2024**). The aforementioned pathological changes are assigned to the seasonal changes, i.e. during summer, heavy metal content and algae bloom increase in the water, and accordingly the energy requirement increases, causing oxidative stress and liver damage, which in turn reflects on fish health.

		Site per se	eason				
	S	51	S	53	Site effect (P-value)	Season effect (P-value)	Permissible dose for O. niloticus
Heavy metal	Summer	Winter	Summer	Winter			
Fe	57.07± 7.78°	138.39± 4.53 ^b	140.65 ± 0.325^{b}	299.67± 1.35 ^a	< .001	< .001	100
Zn	23.75± 0.42 °	42.00±0.07 ^b	38.72± 0.59 ^b	65.98±1.4 7 ^a	< .001	< .001	40
Cu	3.44±0.55 ^b	6.96±1.67 ^b	25.70± 9.4 ^b	53.33± 3.78 ^a	< .001	0.017	30
Cr	1.13± 0.014 ^d	1.78±0.034 °	1.94± 0.026 ^b	2.85 ± 0.05^{a}	< .001	< .001	1
Cd	0.13 ± 0.003^{d}	0.16±0.005 °	0.64 ± 0.005^{b}	0.84 ± 0.005^{a}	< .001	< .001	0.5
Values in All the per S1: El-Qa	the same row wit rmissible doses a	verage value± SE h different supers re according to W (non-polluted sit	cripts are sigr HO (1993) e	ificantly differ			FAO (1983).

Table 2. The average concentrations of heavy metals $(\mu g/g)$ in the liver of the Nile tilapia obtained from the sites under study during summer and winter of 2023

2.3 Metal contents in O. niloticus muscle tissues

The average levels of heavy metals in the muscles in both locations are as follows: Fe > Zn > Cu > Cd > Cr. All of the heavy metal concentrations in the muscles were shown to be significantly impacted by site and season (P < 0.05); the greatest significant values were found at

S3 in winter, while the lowest significant levels were found at S1 in summer. However, neither the Zn, Cu, and Cr values across the two seasons nor the Cd value at S3 during winter (Table 3) showed any discernible changes for S1.

The levels of Zn and Fe at S3 during winter exceeded the permissible doses for *O. niloticus*. While, the levels of Cd surpassed the recommended limits (**WHO**, **1993**) for the fish at both sites during both seasons. Whereas, for the safety of human consumption, all heavy metals were found within the allowable levels of the **FAO/WHO** (**2011**) at both sites during both seasons, except for Cd and Cr.

The summer and wintertime witnessed iron levels in fish muscles ranging from 40.83 to $54.15\mu g/kg$ at S1 and from 90.80 to $124.52\mu g/kg$ at S3. Remarkably, iron was the highly prevailing metal in the tissues under investigation. Iron elevation in water is likely the cause of Fe, being densely accumulated in fish muscles, especially at S3 (**Khalil** *et al.*, **2017**). Data in Table (3) reveal that the iron contents in fish muscles are greater than the recommended dose of the **WHO** (**1993**) (100 $\mu g/kg$ at S3 during winter). Besides, the largest concentration of iron detected in fish muscles may be found in the River Nile waters and in household sewage at S2. On the other hand, the lowest values at S1 might be the result of Fe²⁺ being oxidized to Fe³⁺, which then remains as Fe (OH)₃ in the sediment of the oxygenated water.

Zinc was recorded with the maximum value in fish muscles at the polluted site (S3 during winter (41.66 μ g/kg), while the minimum value (22.96 μ g/kg) was noticed at S1 during summer. The value of Zn surpassed the permissible level of 40.0 μ g/kg (**WHO**, **1993**) at S3 during winter. A relation was assessed between the high uptake of heavy metals, the increase in zinc, and the great metabolic rates, previously spotted in the study of **Sorensen (1991)**. Another perception may relate the elevated levels of Zn in water at S3 to the decline in the water discharged throughout the months of winter, which subsequently increases the domestic waste in S2.

The summer and winter cadmium concentrations in fish muscles varied from 1.05 to $1.10\mu g/kg$ at S1 and from 1.07 to $2.13\mu g/kg$ at S3. The cadmium concentrations found in fish muscles surpassed the allowable threshold of **WHO** (1993) ($0.5\mu g/kg$). A significant bond between cadmium and the cysteine residue of metalothionein was previously detected in the studies of **Abu El-Ella** (1996) and **Tayel** *et al.* (2018) leading to high levels of cadmium found in fish muscles. Furthermore, the phosphate fertilizer used at S2, where most of the cadmium accumulates in the environment, along with waste from industrial, mining, and sewage activities, is probably associated with the elevated cadmium levels (**Dimari** *et al.*, 2008).

The results of **El-Degwy** *et al.* (2023) are contradictory with the current results, where the high heavy metals' levels (Fe, Mn, Cu, Zn, Cd and Ni) in the edible parts of *O. niloticus* were distinguished in summer owing to the untreated waste disposal, urban development, runoff from municipal sewage, and industrial wastes.

The present findings showed that copper, iron and zinc were the metals most dominating in fish organs. **Kumar** *et al.* (2011) analyzed these results and contended that these metals are necessary because they are regulated to preserve a particular homeostatic state in fish and act as co-factors for the activation of several enzymes. Conversely, non-essential metals are usually present in fish in trace concentrations and serve no biological purpose. For instance, the heavy metals cadmium and chromium are toxic, non-essential, and non-biodegradable. They are not involved in biological processes in living things, but they can be dangerous in small amounts. Consequently, these metals have the potential to harm fish even at low doses (**Badr** *et al.*, 2014).

Relative to other tissues, the lowest concentrations of all the heavy metals under study were detected in fish muscles. This result coincides with the finding of **Darweesh** *et al.* (2019). Moreover, **Bahnasawy** *et al.* (2010) reported that muscles are non-biologically active organs in heavy metals accumulation.

The levels of heavy metal residues in the liver and gills of *O. niloticus* were considerably greater than those in the muscles, according to the findings of the study on heavy metals in fish tissues. Numerous authors' data (Saeed & Shaker, 2008; Farouk, 2009; Yosef & Goma, 2011; Abd-El-Khalek *et al.*, 2012; EL-Shaer & Alabssawy, 2019; Aly *et al.*, 2020) support this result. This outcome can be ascribed to fish's gill tissues and liver creating metallothionein proteins to help with detoxification when they are vulnerable to heavy metals. According to Jobling (1995) and Yacoub (2007), these proteins are essential for shielding tissues from the detrimental influence of heavy metals.

In comparison with fish muscles, the higher levels of heavy metals in gills can be related to the muscles challenging to completely remove the heavy metals from the platelets (Heath, 1995). Hence, muscles do not actively accumulate heavy metals (Karadede & Ünlü, 2000). Various studies involving different fish species (Guerrin *et al.*, 1990; Alam *et al.*, 2002) reported similar findings, possibly due to the low binding affinity of the proteins in the muscles (El-Shaer & Alabssawy, 2019). It may also be the cause of the trace amounts of heavy metals in fish muscles because of the scales on the fish's skin, mucous secretions, and skin barrier, which all decrease the amount of water that carries heavy metals to the body through the muscles and skin.

Fish liver is a reliable indicator of contamination in the aquatic environment since it has a vital role in metabolizing and excreting xenobiotic compounds (**Rocha & Monteiro, 1999**). In addition, the liver filters the contaminated blood that is transported through the intestine. Additionally, **Habib** *et al.* (2023) and **Lawan & Akawu** (2024), in their studies on *O. niloticus*, attributed the highest quantities of heavy metals to the liver, being the main organ responsible for heavy metal accumulation for its function in the detoxification and removal of heavy metals from the body, due to its high metallothionein protein content, which has a great ability to bind with heavy metals and to prevent their circulation.

The current results indicate that fish living in aquatic habitats with high metal ion concentrations tend to absorb these ions into their tissues in various ways. Metal concentrations in natural aquatic environments are typically low, ranging from nanograms to micrograms per liter. Fish living in these environments primarily accumulate metals through their diet and water (**Bury** *et al.*, 2003). In this context, metal accumulation was observed in *O. niloticus* under investigation at significantly higher concentrations compared to those recorded in the surrounding water. The increased metal accumulation in the muscles of the species under investigation at S3 occurred during winter, and this elevation can be ascribed to the low water levels during the drought and the high levels of domestic sewage discharged into S2.

3. Health risk assessment

3.1. Oral and dermal exposure of the River Nile

In this study, the HMEI was assessed for both drinking and irrigation water quality standard of water sampled from three localities throughout the period of investigation (winter and summer of 2023). S1 recorded had appropriate HMEI values for both irrigation and drinking uses, with values that ranged between 0.79 and 0.24 in winter, and 0.62 and 0.21 in summer, respectively. These values categorized the water at S1 as 'Fit' for both uses during the study, as demonstrated in Table (4).

On the contrary, S2 and S3 exhibited significantly higher HMEI values, indicating 'Unfit' conditions for both drinking and irrigation. In winter, the HMEI values for drinking and irrigation were higher than summer, confirming the persistent and severe contamination at these sites, especially in winter.

Table (5) shows seasonal and site-based variations of the daily intake (EDI) of heavy metals evaluated for *O. niloticus* adults through both oral ingestion and dermal absorption at various stations. For oral ingestion, Fe displayed the highest EDI among estimated metals, with values reaching 3.77E-05 and 2.98E-05 mg/kg/day in winter and summer at S2, respectively. Conversely, Cd had the least metal EDI, recording 2.20E-08 and 1.57E-08 mg/kg/day in winter and summer at S1, respectively. Likewise, for dermal absorption, Fe presented the highest EDI, with 1.85E-07 and 1.47E-07 mg/kg/day in winter and summer at site S2, correspondingly. Cd had the lowest EDI, measured at 1.08E-10 and 7.71E-11 mg/kg/day in summer and winter at site S1. Basically, Fe had the highest EDI among metals for both paths, while Cd had the lowest EDI, with substantial differences among sites across seasons. Throughout the different pathways and seasons, S2 exhibited the highest EDI for heavy metals, and the lowest displayed at S1. Generally, the EDI values for most heavy metals were lower in summer than winter, signifying a potential seasonal effect on pollutant levels.

		Sites p	er season]			
	s	1	S	53	Site effect (P-value)	Season effect (P-value)	Permissible dose for O. niloticus (µg/ g)	Permissible dose for humans (µg/ g)
Heavy	Summer	Winter	Summer	Winter				
metal								
Fe	40.83±	54.15±	90.80±	$124.52 \pm$	< .001	< .001	100	425
	0.3 ^d	1.5 °	0.793 ^b	1.095 ^a				
Zn	22.96±	24.89±	34.35±	41.66±	< .001	< .001	40	99.4
	0.58 °	0.76 °	1.088^{b}	0.63 ^a				
Cu	1.22±	1.30±	2.26±	3.36±	< .001	0.001	30	50
	0.072 °	0.15 °	0.14 ^b	0.088^{a}				
Cr	0.09±	0.13±	0.50±	0.66±	< .001	< .001	1	0.5
	0.026 ^c	0.05 °	0.06 ^b	0.085^{a}				
Cd	1.05±	1.10±	$1.07 \pm$	2.13±	< .001	< .001	0.5	1.3
	0.002 ^b	0.0005^{b}	0.008^{b}	0.029 ^a				
Each value	e designates the	e mean value±	SE, SE: Standa	rd error (n=3).	·			

Table 3. The heavy metals mean levels $(\mu g/g)$ in the Nile tilapia's muscles gathered from the studied sites during summer and winter of 2023

Values in the same row with different superscripts are significantly different (P < 0.05) by Tukey's test.

The permissible doses for the fish are according to WHO (1993), except Cr which is in accordance with FAO (1983).

The permissible limits for human consumption are in accordance with FAO/WHO (2011).

S1: El-Qanater El-Khayria (non-polluted site, reference site).

S3: Al-Qatta site (polluted site).

The non-carcinogenic health risks of HM via different pathways of exposure of the Nile water samples covering the investigated seasons are demonstrated in Table (6). Across the studied seasons, the greatest oral THQ values were observed at S2 for Cd followed by Cr; whereas, the lowest THQ values were documented at S1 for Fe and Cu. Concerning the dermal pathway, the highest THQ values were recorded in winter for Cd at S1 followed by Cr at S2, while the lowest THQ values were noted at S1 for Fe succeeded by Cu at S1. In summer, the THQ highest values were found at S2 for Cr at S2 followed by Cd, with the lowest THQ values at S1 for Cu and Fe.

The HI for oral pathway displayed the highest value in winter at S2 and in summer at S2. While, both winter and summer witnessed the most elevated HI values at S2. The HI values across seasons exhibited no non-carcinogenic health risks related to different exposure pathways of heavy metals from the water of the Rive Nile.

		Winter (HMEI)			Summe	r (HMEI)	
	(Drinking quality)		(Irrigation quality)		(Drinking quality)		(Irrigation quality)	
	Value	Description	Value	Description	Value	Description	Value	Description
S1	0.79	Fit	0.24	Fit	0.62	Fit	0.21	Fit
S 2	6.74	Unfit	2.08	Unfit	5.00	Unfit	1.47	Unfit
S 3	4.59	Unfit	1.28	Unfit	3.02	Unfit	1.03	Unfit

Table 4. Heavy metal evaluation index (HMEI) values for water sampled from the River Nile during winter and summer of 2023

Table 5. Estimated daily intake (EDI) of heavy metals for adults via dermal and ingestion
absorption of water sampled from the Nile River during the study

					EDI	Oral					
	Winter						Summer				
	S1	S2	S3	Mean	S. D	S1	S2	S3	Mean	S. D	
Fe	5.34E-06	3.77E-05	2.82E-05	2.3E-05	1.66E-05	4.08E-06	2.98E-05	1.57E-05	1.66E-05	1.29E-05	
Cu	2.51E-07	1.72E-06	1.25E-06	1.07E-06	7.54E-07	1.88571E-07	1.57E-06	1.03E-06	9.32E-07	6.97E-07	
Zn	1.88E-06	6.28E-06	3.77E-06	3.98E-06	2.20E-06	2.51429E-06	4.08E-06	3.45E-06	3.35E-06	7.90E-07	
Cd	2.20E-08	3.77E-07	2.20E-07	2.06E-07	1.77E-07	1.57E-08	2.51E-07	1.88E-07	1.51E-07	1.22E-07	
Cr	2.20E-07	8.17E-07	4.40E-07	4.92E-07	3.01E-07	2.20E-07	5.34E-07	3.46E-07	3.66E-07	1.58E-07	
					EDI D	ermal					
Fe	2.62E-08	1.85E-07	1.38E-07	1.16E-07	8.17E-08	2.01E-08	1.47E-07	7.71E-08	8.13E-08	6.34E-08	
Cu	1.23E-09	8.48E-09	6.17E-09	5.29E-09	3.70E-09	9.26E-10	7.71E-09	5.09E-09	4.58E-09	3.42E-09	
Zn	5.55E-09	1.85E-08	1.11E-08	1.17E-08	6.50E-09	7.41E-09	1.20E-08	1.02E-08	9.87E-09	2.33E-09	
Cd	1.08E-10	1.85E-09	1.08E-09	1.01E-09	8.73E-10	7.71E-11	1.23E-09	9.26E-10	7.46E-10	5.99E-10	
Cr	1.08E-09	4.01E-09	2.16E-09	2.42E-09	1.48E-09	1.08E-09	2.62E-09	1.70E-09	1.80E-09	7.77E-10	
				SI) = Standar	d deviation					

Across both seasons, the TR of HM for adults through both ingestion and dermal absorption of the River Nile's water samples consistently were within the "negligible" range for all sampled sites. In the present study, TR values of oral exposure pathway for Cd and Cr ranged from 5.97E-09 to 1.43E-07 and from 1.10E-07 to 4.09E-07, respectively, all classified as negligible. Likewise, the dermal exposure risks for Cd and Cr also were under the negligible rank. Table (6) reveals higher Σ TR from oral exposure compared to dermal adsorption. Moreover, the recorded Σ TR_(Oral=Dermal) was consistently below the threshold for any significant cancer risk. Thus, the study indicates that there is no cancerogenic risk from both Cd and Cr through both pathways of exposure, across all sites and seasons studied.

More than 15 million people in the Cairo depend on the River Nile for their daily needs, making it Egypt's main supply of freshwater. A variety of commercial and industrial operations that are essential to the region are also supported by it. But the constant inflow of drainage water from many human activities has been causing significant and harmful alterations to the River Nile. The amount and quality of the river's water are generally affected by various factors, viz. dams, weather patterns, and human activity (**Khalil** *et al.*, **2017; Salaah** *et al.*, **2018**). The largest health hazards were found in water samples from S2, the site of the El-Rahawy drain, although

the drain's detrimental effects were also seen at S3, the following location. El-Rahawy drain, identified as a primary contamination source in the River Nile, discharges runoff from agricultural, industrial, and household activities into the river. Hence, S2 receives a wide contaminants' range such as fertilizers, pesticides, and industrial effluents from El-Rahawy drain (**Khalil** *et al.*, **2017**). Consequently, these pollutants can limit the water usage in this area. Although both the dermal and oral contact with water from the Nile recorded no health hazards at any sampling site, the present study highlights the potential risks associated with using water from S2 and S3 for drinking or irrigation. It is advisable to avoid using this water to prevent potential negative health consequences.

			TR Oral				
		Winter		Summer			
	S1	S2	S 3	S1	S2	S 3	
Cd	8.36E-09	1.43E-07	8.36E-08	5.97E-09	9.55E-08	7.161E-08	
Cr	1.10E-07	4.09E-07	2.20E-07	1.10E-07	2.67E-07	1.73E-07	
Mean	5.91E-08	2.75E-07	1.518E-07	5.79E-08	1.81E-07	1.22E-07	
S. D	7.18E-08	1.87E-07	9.64E-08	7.35E-08	1.21E-07	7.15E-08	
$\sum TR_{Oral}$	1.18E-07	5.51E-07	3.03E-07	1.15E-07	3.62E-07	2.44E-07	
			TR Dermal				
Cd	4.10E-11	7.04E-10	4.10E-10	2.93E-11	4.69E-10	3.51E-10	
Cr	5.40E-10	2.01E-09	1.08E-09	5.4E-10	1.31E-09	8.48E-10	
Mean	2.90E-10	1.35E-09	7.45E-10	2.84E-10	8.90E-10	6.00E-10	
S. D	3.52E-10	9.20E-10	4.73E-10	3.61E-10	5.95E-10	3.51E-10	
∑TR _{Dermal}	5.69E-10	1.78E-09	1.20E-09	5.69E-10	1.78E-09	1.20E-09	
∑TR _(Oral=Dermal)	1.18E-07	5.53E-07	3.048E-07	1.16E-07	3.64E-07	2.45E-07	
SD = Standard d	leviation. Cancer	risk considered n	egligible at <10 ⁻⁶ : lo	w at <10 ⁻⁵ : medium	at <10 ⁻⁴ : high at	<10 ⁻³ and very	
			high >10 ⁻³				

Table 6. Total target cancer risk (TR) f of heavy metals for adults through ingestion and dermal absorption of the Nile water samples throughout the study period

3.2 Ingestion and the Nile tilapia

The assessment of hazard associated with heavy metals for adults *via* normal and habitual consumptions of *O. niloticus* of the River Nile during the study is represented in Tables (7-9). Mainly, habitual consumption recorded higher EDI values than normal consumption, while winter recorded higher EDI, compared to summer among all the studied sites. The highest EDI values were detected at site S3, as Fe intake recorded a peak for habitual consumption, followed by Zn> Cu> Cr> Cd during winter and summer. S1 represented the lowest EDI values among sites, while winter revealed higher EDI values compared to summer (Table 7).

			EDI			
			Summer			
		S1	S 3	S1	S 3	
Fe —	Normal	2.41E-02	5.55E-02	1.82E-02	4.05E-02	
	Habitual	1.10E-01	2.53E-01	8.31E-02	1.85E-01	
7	Normal	1.11E-02	1.86E-02	1.02E-02	1.53E-02	
Zn	Habitual	5.06E-02	8.47E-02	4.67E-02	6.99E-02	
C	Normal	5.79E-04	1.01E-03	5.44E-04	1.01E-03	
Cu	Habitual	2.64E-03	4.60E-03	2.48E-03	4.60E-03	
C.I	Normal	5.93E-05	2.96E-04	3.92E-05	2.24E-04	
Cd	Habitual	2.71E-04	1.35E-03	1.79E-04	1.02E-03	
Cr –	Normal	4.90E-04	9.49E-04	4.67E-04	4.78E-04	
	Habitual	2.24E-03	4.33E-03	2.13E-03	2.18E-03	

Table 7. An inquiry of the evaluated daily intake (EDI) of heavy metals for adults *via* normal and habitual uses of the Nile tilapia

In winter, the highest THQ values were recorded at site S3 for Cr, with a value of 7.46E-01 for habitual consumers, followed by Cd> Zn> Fe> Zn. In summer, a different pattern was recorded for habitual consumption, as Cd exhibited the highest THQ values, followed by Cr>Fe> Zn> Cu at S3 (Table 8). The HI in the present study revealed a significant non-carcinogenic risk (HI>1) for habitual consumers associated with consuming *O. niloticus* from S3 during winter and summer (Table 8). Both TR and Σ TR associated with Cd and Cr during summer and winter are demonstrated in Table (9).

Winter recorded the highest TR and \sum TR for habitual consumers at S3, where Cr revealed higher risk than Cd. *O. niloticus* samples from S1 were reported negligible (TR<10⁶) to low (TR<10⁵) carcinogenic risk for both normal and habitual consumers, respectively. While, fish from S3 exhibited medium cancerogenic (TR<10⁴) risks for habitual consumers. Moreover, the \sum TR of *O. niloticus* from S3 recorded high cancerogenic risk (TR<10⁻³) for habitual consumers and medium cancerogenic risk (TR <10⁻⁴) for normal consumers.

Fish are rich in essential amino acids, minerals, vital polyunsaturated fatty acids, and vitamins that are crucial for human health (Kawarazuka & Béné, 2011; Reksten *et al.*, 2020). Fish are a significant rich and affordable source of nutritional animal protein, especially in low-and middle-income countries where hunger and undernutrition are major challenges; these nations count on fish to mitigate these burdens.

Furthermore, food security is an important multifaceted aspect because it is included in many of the Sustainable Development Goals of the United Nations Agenda 2030, including the second objective (Zero Hunger) and the third goal (Good Health and Well-Being) (WHO, 2019). The availability and affordability of fish make it a staple in the Egyptian diet. However, fish collect HM in their tissues at higher ranges than what is allowed, often by multiple times (Ahmed *et al.*, 2022). The main method through which humans are exposed to metals is through ingestion. Therefore, consuming seafood that has significant levels of heavy metal contamination

can pose several health risks to people, especially for the majority of coastal inhabitants who rely on fish as their main source of animal protein (Liu *et al.*, 2018).

Therefore, *O. niloticus* was selected for the present study due to its prevalence in the River Nile and generally, the HMs load in both fish tissues and surface water recorded a relatively high increase in winter to summer. **Hekal and Fahmy (2023)** revealed that during the winter months of December, January, and February, the water flow and levels of the River Nile were the lowest. Conversely, the highest water flow and levels of the River Nile were observed during the summer months of June, July, and August.

Table 8. The target hazard quotients (HQ) and hazard index (HI) of non-carcinogenic health risk of heavy metals for adults via ingestion and dermal absorption of the Nile water samples covering the study period

					THQ Oral	!					
		W	inter						Summer		
	Fe	Cu	Zn	Cd	Cr]	Fe	Cu	Zn	Cd	Cr
S1	7.63E-06	6.29E-06	6.29E-06	2.20E-05	7.33E-05	5.84	4E-06	4.71E-06	8.38E-06	1.57E-05	7.33E-05
S2	5.39E-05	4.32E-05	2.10E-05	3.77E-04	2.72E-04	4.27	7E-05	3.93E-05	1.36E-05	2.51E-04	1.78E-04
S3	4.04E-05	3.14E-05	1.26E-05	2.20E-04	1.47E-04	2.24	4E-05	2.59E-05	1.15E-05	1.89E-04	1.15E-04
Mean	3.40E-05	2.70E-05	1.33E-05	2.06E-04	1.64E-04	2.36	6E-05	2.33E-05	1.12E-05	1.52E-04	1.22E-04
SD	2.38E-05	1.89E-05	7.36E-06	1.78E-04	1.01E-04	1.84	4E-05	1.74E-05	2.64E-06	1.22E-04	5.27E-05
					THQ Derm	al					
		W	inter			Summer					
S1	8.74E-08	1.03E-07	9.26E-08	4.32E-06	7.20E-05	6.69	9E-08	7.71E-08	1.23E-07	3.09E-06	7.20E-05
S2	6.17E-07	7.07E-07	3.09E-07	7.41E-05	2.67E-04	4.89	9E-07	6.43E-07	2.01E-07	4.94E-05	1.75E-04
S 3	4.63E-07	5.14E-07	1.85E-07	4.32E-05	1.44E-04	2.57	7E-07	4.24E-07	1.70E-07	3.70E-05	1.13E-04
Mean	3.89E-07	4.41E-07	1.95E-07	4.05E-05	1.61E-04	2.71	IE-07	3.81E-07	1.65E-07	2.98E-05	1.20E-04
SD	2.72E-07	3.09E-07	1.08E-07	3.49E-05	9.88E-05	2.11	E-07	2.85E-07	3.88E-08	2.40E-05	5.18E-05
					HI						
			W	inter					Summe	r	
	S1 S2 S		S 3			S1	S2		S 3		
HI	Oral	1.16E-04	7.6	8E-04	4.51E-04		1.0	08E-04	5.25E-04	4	3.64E-04
	Dermal	7.66E-05	3.4	3E-04	1.88E-04		7.5	54E-05	2.26E-04	4	1.51E-04
				SD = Sta	andard devia	tion.	* HI≤	1.0		I	

According to earlier research by **Rajeshkumar and Li (2018)**, the species, location, and type of tissue were directly influential on the levels of HMs accumulated in fish tissues. The current research findings concur with the heavy metal levels assessed in the study of **Abdel-Halim** *et al.* (2022). They reported the same seasonal pattern of heavy metal in varying fish species and water off the River Nile, detecting a remarkable HM content increase in water and fish throughout winter, especially for Fe and Zn concentrations. This is the first comprehensive attempt to estimate the potential health risks interrelated with both water usage and consumption of *O. niloticus* from the River Nile.

Basically, the evaluation of human health risks associative with the heavy metal load in edible fish tissues was the key purpose of the present study. The standards of human health estimation are determined by several factors, such as the contaminants ingested, exposure levels and duration, body weight in average, and consumption rates (**Salaah** *et al.*, **2022**).

In reference to USEPA hazard guidelines, consuming *O. niloticus* from S3 could induce a non-cancerogenic health risk. While Cr recorded higher lifetime cancer risk for both groups of the Nile tilapia's end- users throughout the study, particularly for habitual consumers. The Σ TR persuaded by consuming *O. niloticus* exhibited higher risk for normal and habitual *O. niloticus* consumers, especially in winter, based on the Σ TR data at S3, consuming *O. niloticus*.

Table 9. A survey illustrating the target cancer risk (TR) and total TR (Σ TR) of heavy metals for adults *via* normal and habitual ingestions of the Nile tilapia in the Nile River

	TR		S1	S3	
	W/ter 4 are	Normal	2.25E-06	1.12E-04	
Cd	Winter	Habitual	1.03E-05	5.13E-04	
	C	Normal	1.49E-06	8.50E-05	
	Summer	Habitual	6.80E-05	3.88E-04	
Cr	XX/:4	Normal	2.06E-05	3.99E-04	
	Winter	Habitual	9.40E-05	1.82E-03	
J r	Summer	Normal	1.96E-06	2.01E-04	
	Summer	Habitual	8.95E-05	9.16E-04	
		∑TR			
	XX /* 4	Normal	2.28E-06	5.11E-04	
	Winter	Habitual	1.04E-05	2.33E-03	
	C	Normal	2.11E-06	2.86E-04	
Summer		Habitual	9.63E-06	1.30E-03	

CONCLUSION

It is important to note that the El-Rahawy drain's effluents, which are influenced by sewage wastewater as well as industrial and agricultural operations, are causing serious environmental degradation in the Rosetta Branch. As demonstrated by the bioaccumulation of heavy metals in its tissues, the results indicated that *O. niloticus* is significantly impacted by this pollution. Because of its resilience to adverse environmental conditions, *O. niloticus* can thus be regarded as a good bioindicator of the contamination caused by heavy metals. The current study emphasizes the possible risks connected with using water from the El-Rahawy drain and El-Qatta area for drinking or irrigation, even though oral and dermal contact with water from the River Nile did not reveal any health problems at any sampling site. This water should not be used

in order to avoid any possible health risks. Furthermore, eating *O. niloticus* from the El-Qatta Site carries a significant non-carcinogenic health risk, whereas using Cr increases the risk of developing cancer in the future, especially for regular users, according to USEPA standards. *O. niloticus* consumption in this region carries a high overall cancer risk, which might result in two to five cancer cases per 10,000 normal consumers and one to two cases among thousand habitual consumers over the course of a lifetime. Given the rise in the number of possible cancer cases, this risk was noticeably higher in winter.

These results highlight the disparities in the quality of water across the Nile River and spot the grave consequences of human activity on public health. In order to guarantee safe water use, maintain ecological balance, and protect human health, the current study highlights the need for strong water quality management methods, particularly in areas with greater contamination levels. As a result, the government ought to exert an effort to increase public awareness about the Rosetta Branch. To simultaneously protect both people as end-users and fish from the harmful influences of these contaminants, it is also imperative to exercise greater caution when it comes to the El-Rahawy drain. This includes treating wastewater to remove pollutants before it is released into the Rosetta Branch or establishing regulations that prohibit the discharge of such pollutants into the River Nile.

REFERENCES

- Abdel-Halim, K. Y.; El-Monem, A.; Wanas, A. A. K. and Mostafa, N. A. (2022). Bioaccumulation profiles of heavy metals in fish collected from Rosetta branch of River Nile, Egypt: A case study for organ's responsibility. Egypt. J. Chem., 65(132):501-518. https://doi.org/10.21608/ejchem.2022.133006.5875.
- Abd-El-Khalek, D. E.; El -Gohary, S. El. and El -Zokm, G. M. (2012). Assessment of heavy metals pollution in *Oreochromis niloticus* in El-Max fish farm, Egypt. J. Exp. Biol., 8(2): 215-222.
- Abdel-Khalek A.A.; Elhaddad, E.; Mamdouh S. and Marie, M-A. S. (2016). Assessment of metal pollution around Sabal drainage in River Nile and its impacts on bioaccumulation level, metals correlation and human risk hazard using *Oreochromis niloticus* as a bioindicator. Turk. J. Fish. Aquat. Sci., 16: 227-239. <u>http://dx.doi.org/10.4194/1303-2712-v16_2_02</u>
- Abdel-Moati, M. A. R. and El-Sammak, A. A. (1997). Man-made impact on the geochemistry of the Nile delta lakes: a study of metals concentrations in sediments. Wat. Air Soil Poll. 97: 413-429. <u>https://doi:10.1007/bf02407476</u>
- Abdel-Mohsien, H.S. and Mahmoud, M.A.M. (2015). Accumulation of some heavy metals in Oreochromis niloticus from the Nile in Egypt: Potential hazards to fish and consumers. J. Environ. Prot. Sci., 6: /10.4236/jep.2015.69089

- Abdel-Satar, A.M.; Ali, M.H.H. and Goher, M.E. (2017). Indices of water quality and metal pollution of River Nile, Egypt. Egypt. J. Aquat. Res., 43: 21-29. <u>https://doi.org/10.1016/j.ejar.2016.12.006</u>
- Abdelsalam, K. M.; Tadros, H. R.; Moneer, A. A.; Khalil, M. K.; Hamdona, S. K.; Shakweer, L., Moawad, M.N.; El-Sayed, A. A. M.; El-Said G.F.; Ismail, M.M.; Shobier, A.H.; Hosny, S.; Dabbous, A.S.; Alzeny, A.M. and Khedawy, M. (2024). The Egyptian Nile estuarine habitats: a review. Aquat. Sci., 86(4), 95. https://doi.org/10.1007/s00027-024-01111-9
- Abdi, H. and Williams, L.J. (2010). Newman-Keuls Test and Tukey Test. In: Salkind, N. (Ed.), Encyclopedia of Research Design, Thousand Oaks, CA: Sage. <u>https://personal.utdallas.edu/~herve/abdi-NewmanKeuls2010-pretty.pdf</u>
- Abu El-Ella, S. M., (1996). Studies on the toxicity and bioconcentration of cadmium on grass carp *Ctenopharyngo donidella*. M. Sc. Thesis, Fac. of Sci., Helwan University, Egypt.
- Agah, H.; Leemakers, M.; Elskens, M.; Fatemi, S.M.R. and Baeyens, W. (2009). Accumulation of trace metals in the muscles and liver tissues of five fish species from the Persian Gulf. Environ. Monit. Assess., 157: 499-514. <u>https://doi.org/10.1007/s10661-008-0551-8</u>
- Ahmed, N.M.; Salaah, S.M. and Tayel, S. (2022). Accumulation and risk assessment of heavy metals-induced biochemical and histopathological alterations in O. niloticus from Lake Nasser, Egypt. Egypt. J. Aquat. Biol. Fish, 26(2):409-425. https://dx.doi.org/10.21608/ejabf.2022.234222
- Alam, M. G. M.; Tanaka, A.; Allinson, G.; Laurenson, L. J. B.; Stagnitti, F. and Snow, E. T. (2002). A comparison of trace element concentrations in cultured and wild carp (*Cyprinus carpio*) of Lake Kasumigaura, Japan. Ecotox. Environ. Safe, 53: 348–354. https://doi.org/10.1016/S0147-6513(02)00012-X
- Ali, N.A.; Mohamed, M.A. and Abd El-Hameed, E.A.A. (2016). Water quality and heavy metals monitoring in water and tissues of *O. niloticus* fish from different governorates "Egyptian Aquaculture farms". Egypt. J. Aquat. Biol. Fish., 20(1110-6131): 103-113. http://dx.doi.org/10.21608/ejabf.2016.10987
- Alinnor, I. J. and Obiji, I. A. (2010). Assessment of trace metal composition in fish samples from Nworie River. Pak. J. Nutr., 9(1): 81–85. <u>https://doi.org/10.3923/pjn.2010.81.85</u>
- Al-Sayegh Petkovšek, S.; MazejGrudnik, Z. and Pokorny, B. (2012). Heavy metals and arsenic concentrations in ten fish species from the šalek lakes (Slovenia): assessment of potential human health risk due to fish consumption. Environ. Monit. Assess., 184: 2647–2662. <u>https://doi.org/10.1007/s10661-011-2141-4</u>

- Aly, M. Y. M.; El-Gaar, D. M. K.; Salaah, S. M. and Abdo, M. H. (2020). Evaluation of Heavy Metals and Oxidative Stress with Biochemical Parameters as Bioindicators of Water Pollution and Fish in Lake Burullus, Egypt. Mari. Scie. Res. Ocean. J., 1(3): 30-34.
- American Public Health Association (APHA); American water work association (AWWA), and water environmental federation (3111B). ((2017). Standard methods for the examination of water and wastewater. 23rd edition. Rice, E.W.; Baird, R.B. and Eaton, A.D (eds.). APHA, Washington, . 209-216 Pp.
- **American Water Work Association (AWWA) (1998)**. Standard methods for the examination of water and wastewater. 20th edition. Denver, CO.
- Awad, S. T.; Hemeda, S. A.; El Nahas, A. F.; Abbas, E. M.; Abdel-Razek, M. A.; Ismail, M.; Mamoon, A. and Ali, F. S. (2024). Gender-specific responses in gene expression of Nile tilapia (*Oreochromis niloticus*) to heavy metal pollution in different aquatic habitats. Sci. Rep., 14(1):14671. <u>https://doi.org/10.1038/s41598-024-64300-4</u>
- Azab, A.M.; Aly-Eldeen, M.A.; Khalaf-Allah, H.M.M. and El-Battal, M.M.A. (2019). Effect of heavy metals on the ovary of *Tilapia zillii* in some canals of Nile Delta area, Egypt. Egypt. J. Aquat. Biol. Fish., 23(3): 329-345. https://dx.doi.org/10.21608/ejabf.2019.45670
- Badr, A. M.; Mahana, N. A. and Eissa, A. (2014). Assessment of Heavy Metal Levels in Water and Their Toxicity in Some Tissues of O. niloticus (Oreochromis niloticus) in River Nile Basin at Greater Cairo, Egypt. Glob. Vet., 13(4): 432-443. https://dx.doi.org/10.5829/idosi.gv.2014.13.04.8561
- Bahnasawy, M.; Khidr, A. and Dheina, N. (2010). Assessment of heavy metal concentrations in water, plankton, and fish of Lake Manzala, Egypt. Turk. J. Zool., 34: 1-10. https://dx.doi.org/10.3906/zoo-0810-6
- **Basiony, A. I. (2014).** Environmental studies on heavy metals pollution and management of lake Burullus, Egypt. M. Sc. Thesis, Fac. of Sci., .Port Said Uni., Egypt.
- Bols, N.C.; Brubacher JL; Ganassin, R.C. and Lee, L.E. (2001). Ecotoxicology and innate immunity in fish. Dev. Comp. Immunol., 25(8):853-873. <u>https://doi.org/10.1016/S0145-305X(01)00040-4</u>

- Botkin, D. B. and Keller, E. D. (2003). Environmental science earth as a living planet. 4th edition. John Wiley and Son Inc. New York, USA. 410-476 pp. <u>https://doi.org/10.1108/meq.2003.14.5.623.4</u>
- Bury, N.R., Walker, P.A. and Glover, C.N. (2003). Nutritive metal uptake in teleost fish. J. Exp. Biol., 206: 11- 23 Pp https://doi.org/10.1242/jeb.00068
- Canadian Council of Ministers of the Enironment (CCME) (2017). CCME water quality index; User's Manual 2017, Update. In: Canadian Water Quality, Guidelines for the protection of Aquatic life, Winnipeg.
- Canli, M.; Ay, Ö. and Kalay, M. (1998). Levels of heavy metals (Cd, Pb, Cu, Cr and Ni) in tissue of *Cyprinus Carpio, Barbus Capito* and *Chondrostoma Regium* from the Seyhan River. Turkey, Turk. J. Zool., 22 (2): 149-157.
- Campenhout, K. V.; Infante, H. G.; Adams, F. and Blust, R. (2004). Induction and binding of Cd, Cu, and Zn to metallothionein in carp (*Cyprinus carpio*) using HPLC-ICP-TOFMS. Toxicol. Sci. 80: 276–287. <u>https://doi.org/10.1093/toxsci/kfh149</u>
- Darweesh, M.M.; Gamal El-Dein, H.M.; Abou-Shleel, S.M. and El Shirbeny, M.A. (2019). Seasonal variation of heavy metals in water and organs of *Oreochromis niloticus* at Rosetta Branch, River Nile, Egypt. Egypt. J. Aquat. Biol. Fish., 23(3): 513-526. https://doi.org/10.21608/ejabf.2019.51648
- Daviglus, M.; Sheeshka, J. and Murkin, E. (2002). Health benefits from eating fish. Comments on Toxicology, 8: 345–374. <u>https://doi.org/10.1080/08865140215064</u>
- Dimari, G. A.; Abdulrahman, F. I.; Akan, J. C. and Garba, S. (2008). Metals concentration in tissues of Tilapia gallier, Crariaslazera and OsteoglossidaeCanght from Alau Dam, Maiduguri, Borno State, Nigeria. Am. J. Environ. Sci., 4(4), 373-379. <u>https://doi.org/10.3844/ajessp.2008.373.379</u>
- **Donia, N. (2005).** "Rosetta Branch Waste Load Allocation Model." Presented at the Ninth International Water Technology Conference, Sharm El-Sheikh, Egypt. 277Pp.
- Dural, M.; Göksu, M.L.; Özak, A.A. and Derici, B. (2006). Bioaccumulation of some heavy metals in different tissues of *Dicentrarchus labrax* L, 1758, *Sparus aurata* L, 1758 and *Mugil cephalus* L, 1758 from the Camlik lagoon of the eastern cost of Mediterranean (Turkey). Environ. Monit. Assess., 118(1): 65-74. <u>https://doi.org/10.1007/s10661-006-0987-7</u>

- Egyptian Governmental Law No. 48/ 1982- Decision 92 (2013): The implementer regulations for Law 48/ 1982, 92/ 2013 regarding the protection of the River Nile and water ways from pollution. Map Periodical Bull., 21-30.
- El Bouraie, M.M.; Motawea, E.A.; Mohamed, G.G. and Yehia, M.M. (2011). Water quality of Rosetta branch in Nile delta, Egypt. Suoseura, 62(1), 31:32 Pp..
- El-Dakar, A.Y.; Shalaby, S.M.; Abdelshafy, H.T. and Abdel-Aziz, M. (2021). Using clove and mint oils as natural sedatives to increase the transport quality of the Nile tilapia (*Oreochromis niloticus*) broodstock. Egypt. J. Aquat. Biol. Fish., 25(4): 437-446. https://dx.doi.org/10.21608/ejabf.2021.190302
- El-Degwy, A. A.; Negm, N. A.; El-Tabl, A. S. and Goher, M. E. (2023). Assessment of heavy metal pollution in water and its effect on Nile tilapia (*Oreochromis niloticus*) in Mediterranean Lakes: a case study at Mariout Lake. Appl. Water Sci., 13(2): 50. https://doi.org/10.1007/s13201-022-01858-2
- El-Naggar, M. M.; Salaah, S.; Ashraf, S.; Khalil, M. T. and Emam, W. W. M. (2022). Impact of chitosan and chitosan nanoparticles on reducing heavy metals within the O. niloticus, *Oreochromis niloticus*. Egypt. J. Aquat. Biol. Fish., 26(2): 859–874. <u>https://dx.doi.org/10.21608/ejabf.2022.239463</u>
- El-Naggar, M. M.; Salah, S.; El-Shabaka, H. A.; Abd El-Rahman, F. A. A.; Khalil, M. T. and Suloma, A. (2021). Efficacy of dietary chitosan and chitosan nanoparticles supplementation on health status of *O. niloticus*, *Oreochromis niloticus* (L.). Aquacult. Rep., 19: 100628. http://dx.doi.org/10.1016/j.aqrep.2021.100628
- El-Sayed, M. and Salem, W.M. (2015). Hydrochemical assessments of surface Nile water and ground water in an industry area South West Cairo, Egypt. J. Petrol., 24: 277-288. https://doi:10.1016/j.ejpe.2015.07.014.
- EL-Shaer, F. M. and ALabassawy, A. N. (2019). Assessment of heavy metals concentration in water and edible tissues of *O. niloticus* (*Oreochromis niloticus*) and (*Clarias garpinus*) from Burullus lake, Egypt with liver histopathological as pollution indicator. J. Egypt. Soc. Parasitol. (JESP), 49(1): 183-194. <u>https://doi.org/10.21608/jesp.2019.68301</u>
- Ezzat, N.; Al-Hawary, I.; Elshafey, A.; Althobaiti, N. and Elbialy, Z. I. (2024). Effect of Seasonal Changes in Heavy Metals on the Histomorphology of the Liver and Gills of Nile Tilapia (*Oreochromis niloticus* L.) in Burullus Lake, Egypt. J. Vet. Sci., 55 (6): 1705-1716. <u>http://dx.doi.org/10.21608/EJVS.2024.259471.1759</u>

- Farouk, A. A. (2009). Some studies on water pollution of Manzala Lake by heavy metals and others and effect of these on stock assessment of fish. M.SC. Thesis. Fac. of Sci. Al Azhar Univ. Chemistry Dept.
- Food and Agriculture Organization (FAO) (1983). Compilation of legal limits for hazardous substances in fish and fishery products. Food and Agriculture Organization of United Nations. Rome, Italy.
- Gaber, H. S., Midhat, A. E., Seham, A. I. and Mohammad, M. N. A. (2013). Effect of water pollution in El-Rahawy drainage canal on hematology and organs of freshwater fish *Clarias gariepinus*. World Appl. Sci. J., 21: 329-341. <u>http://doi.org/ 10.5829/idosi.wasj.2013.21.3.71192</u>
- Gati, G.; Pop, C.; Brudasca, F.; Gurzau, A.E.; Spinu, M. (2016). The ecological risk of heavy metals in sediment from the Danube Delta. Ecotoxicol., 25: 688-696. https://doi.org/10.1007/s10646-016-1627-9
- General Authority for Fish Resources Development (GAFRD) (2015). Yearly Book for fish production in Egypt, Agriculture Ministry.
- **Ghallab, M.H. (2000).** Some physical and chemical changes on the River Nile downstream of Delta barrage at El-Rahawy drain. M.Sc. Thesis. Fac. Sci. Ain Shams Univ., Egypt.
- Gouda S. A. and Taha A. (2023). Biosorption of Heavy Metals as a New Alternative Method for Wastewater Treatment: A Review. Egypt. J. Aquat. Biol. Fish., 27(2): 135-153. https://dx.doi.org/10.21608/ejabf.2023.291671
- Guerrin, F.; Burgat-Sacaze, V. and Saqui-Sames, P. (1990). Levels of Heavy Metals and Organochlorine Pesticides of Cyprinid Fish Reared Four Years in a Wastewater Treatment Pond . Bull. Environ. Contam. Toxicol. 44:461-467. https://doi.org/10.1007/BF01701230. Habib, S. S.; Batool, A. I.; Rehman, M. F. U. and Naz, S. (2023). Evaluation and association of heavy metals in commonly used fish feed with metals concentration in some tissues of *O. niloticus* cultured in biofloc technology and earthen pond system. Biol. Trace Elem. Res., 201(6), 3006-3016. https://doi.org/10.1007/s12011-022-03379-0
- Haggag, Y.H.M. (2017). Environmental and chemical studies on the River Nile Pollutants and their relation to water quality at Rosetta branch, Egypt. Ph. D. Thesis, Gen. Eng. Biotech. Inst.Sadat City Univ. Hashem, M. H.; Tayel, S. I.; Sabra, E. A. and Yacoub, A. M. (2020). Impact of the water quality of El-Rahawy Drain on some genetic and

histopathological aspects of *Oreochromis niloticus*. Egypt. J. Aquat. Biol. Fish., **24**(2): 19-38. <u>https://doi.org/10.21608/ejabf.2020.78272</u>

- Haux, C. and Larsson, A. (1984). Long-term sublethal physiological effects on rainbow trout, *Salmo gairdneri*, during exposure to cadmium and after subsequent recovery. Aquat. Toxicol., 5: 129-142. <u>https://doi.org/10.1016/0166-445X(84)90004-3</u> Hekal, N. and Fahmy, W. A. (2023). Assessment of water level fluctuation impacts along River Nile, Egypt. Water Sci., 37(1): 80-97. <u>http://dx.doi.org/10.1080/23570008.2023.2216040</u>
- Huang, L.; Rad, S.; Xu, L.; Gui, L.; Song, X.; Li, Y. and Chen, Z. (2020). Heavy metals distribution, sources, and ecological risk assessment in Huixian wetland. South China. Water, 12(2): 431. <u>https://doi.org/10.3390/w12020431</u>
- Javed, M., and Usmani, N. (2019). An overview of the adverse effects of heavy metal contamination on fish health. Proc.Nat. Acad. Sci., India Section B: Biol. Sci., 89(2): 389-403. https://doi.org/10.1007/s40011-017-0875-7
- **Jobling, M. (1995).** Environmental Biology of Fishes. 1sted. Printed in Great Britian. Chapman and Hall, London.
- Karadede, H. and Ünlü, E. (2000). Concentrations of some heavy metals in water, sediment and fish species from the Atatürk Dam Lake (Euphrates), Turkey. Chemosphere, 41(9): 1371-1376. https://doi.org/10.1016/S0045-6535(99)00563-9
- Kawarazuka, N. and Béné, C. (2011). The potential role of small fish species in improving micronutrient deficiencies in developing countries: building evidence. Public health nutr., 14(11): 1927-1938. <u>https://doi.org/10.1017/S1368980011000814</u>
- Khalil, M., T.; Nahed, S. G.;Nasr, A. M. A. and Sally, S. M. (2017). Antioxidant defense system alterations in fish as a bio-indicator of environmental pollution, Egypt. J. Aquat. Biol. Fish., 21(3): 11-28. <u>https://doi:10.21608/ejabf.2017.3536</u>.
- Khattab, N. M. A.; Saber, S. A.; El-Salkh, B. A. and Said, R. E. M. (2021). Genotoxicity and limbs asymmetry in the Egyptian toad (*Sclerophrys regularis*) as biomarkers for heavy metals toxicity. Egypt. J. Aquat. Biol. Fish., 25(4): 705-17. <u>https://dx.doi.org/10.21608/ejabf.2021.194098</u>
- Knuth, B. A.; Connelly, N. A.; Sheeshka, J. and Patterson, J. (2003). Weighing health benefits and health risk information when consuming sport-caught fish. Risk Anal., 23: 1185-1197. <u>https://doi.org/10.1111/j.0272-4332.2003.00392.x</u>
- Kumar, B.; Mukherjee, D. P. N.; Sanjay Kumar; Meenu Mishra; Dev Prakash; Singh, S. K. and Sharma, C. S. (2011). Bioaccumulation of heavy metals in muscle tissue of fishes from selected aquaculture ponds in east Kolkata wetlands. Ann. Biol. Res., 2(5):125-134.

- Liu, M.; Chen, L.; He, Y.; Baumann, Z.; Mason, R. P.; Shen, H. and Wang, X. (2018). Impacts of farmed fish consumption and food trade on methylmercury exposure in China. Environ Int., 120: 333-344. <u>https://doi.org/10.1016/j.envint.2018.08.017</u>
- Mantoura, R.F.C.; Dickson, A. and Riley, J.P. (1978). The complexation of metals with humic materials in natural waters. Estuar. Coast. Mar. Sci., 6(4): 387-408. <u>https://doi:10.1016/0302-3524(78)90130-5</u>
- Matthiessen, P. and Brafield, A. E. (1977). Uptake and loss of dissolved zinc by stickle back *Gasterosteus aculeatus* (L). J. Fish. Biol., 10:399-410. <u>https://doi.org/10.1111/j.1095-8649.1977.tb04071.x</u>
- Mohamed, M.F.; Ahmed, N.M.; Fathy, Y.M. and Abdelhamid, I.A. (2020). Impact of heavy metals on *Oreochromis niloticus* fish and using electrophoresis as bio-indicator for environmental pollution of Rosetta branch, River Nile. Egypt. Eur. Chem. Bull., 9(2): 48-61. <u>https://doi:10.17628/ecb.2020.9.48-61</u>
- Monroy, M.; Maceda-Veiga, A. and de Sostoa, A. (2014). Metal concentration in water, sediment and four fish species from Lake Titicaca reveals a large-scale environmental concern. Sci. Tot. Environ., 487: 233-244. https://doi.org/10.1016/j.scitotenv.2014.03.134
- Lawan, M. M. and Akawu, A. (2024). Spatial and Tissue-Specific Accumulation of Heavy Metals in Tilapia (*Oreochromis niloticus*) Fish from Nguru River. BIJOS. (2536-6041), 8(2B), 187-193.
- Moustafa, S. S. (2017). Alterations in antioxidant defense system and biochemical metabolites in fish from Rosetta branch of the River Nile as a bio-indicator of environmental pollution. Ph.D. Thesis, Fac. of science, Ain ShamsUni
- Neima, A. A.; Mohamed, M. A. and Abd El-Hameed, E. A. A. (2016). Water quality and heavy metals monitoring in water and tissues of Nile tilapia fish from different governorates "Egyptian aquaculture farms". Egypt. J. Aquat. Biol. Fish., 20(3): 103-113. http://dx.doi.org/10.21608/ejabf.2016.10987
- Omar, W. O.; Saleh, Y. S. and Marie, M- A. S. (2014). Integrating multiple fish biomarkers and risk assessment as indicators of metal pollution along the Red Sea coast of Hodeida, Yemen Republic. Safe. Ecotoxicol. Environ. Safe., 110: 221-231. <u>https://doi.org/10.1016/j.ecoenv.2014.09.004</u>
 - Peterson, R. H.; Metcalfe, J. L. and Ray, S. (1983). Effects of cadmium on yolk utilization, growth, and survival of Atlantic Salmon alevins and newly feeding fry. Arch. Environ. Contam.Toxicol., 12: 37-44. <u>https://doi.org/10.1007/BF01054999</u>

- Qiu, Y.W. (2015). Bioaccumulation of heavy metals both in wild and mariculture food chains in Daya Bay, South China. Estuar. Coast. Shelf Sci., 163: 7-14. <u>https://doi.org/10.1016/j.ecss.2015.05.036</u>
- Rajeshkumar, S. and Li, X. (2018). Bioaccumulation of heavy metals in fish species from the Meiliang Bay, Taihu Lake, China. Toxicol., 5: 288-295. https://doi.org/10.1016/j.toxrep.2018.01.007

Reksten, A. M.; Somasundaram, T.; Kjellevold, M.; Nordhagen, A.; Bøkevoll, A.; Pincus, L.; Rizwan, A.A.M.; Mamun, A.; Thilsted, S.H.; Htut, T.; and Aakre, I., (2020). Nutrient composition of 19 fish species from Sri Lanka and potential contribution to food and nutrition security.

- J. Food Compos. Anal., 91: 103508. https://doi.org/10.1016/j.jfca.2020.103508
- Reid, S.D. and McDonald, D.G. (1988). Effects of cadmium, copper, and low pH on ion fluxes in the rainbow trout, *Salmo gairdneri*. Canad. J. Fisheries and Aquat. Sci.,45: 244-253. https://doi.org/10.1139/f88-029
- Rocha, E. and Monteiro, R. A. F. (1999). Histology and Cytology of Fish Liver: A Review. In: "Ichthyology: Recent Research Advances", Saksena D.N. (Ed.). Science Publishers, Enfield, New Hampshire. ISBN: 1-57808-053-3, : 321-344Pp.
- Saeed, S. M. and Shaker, I. M. (2008). Assessment of heavy metals pollution in water and sediment and their effect on *Oreochromis niloticus* in the northern delta Lakes, Egypt. 8th International Symposium on Tilapia in Aquaculture, 475-490 Pp.
- Said, R.E.M.; Ashry, M. and AbdAllah, E.M. (2021). The use of biomarkers in the O. niloticus (Oreochromis niloticus) as biological signals to track Nile contamination in Egypt. Egypt. J. Aquat. Biol. Fish., 25(5): 203-214. https://dx.doi.org/10.21608/ejabf.2021.198551
- Santos, I.; Diniz, M. S.; Carvalho, M. L.; Santos, J. P. (2014). Assessment of essential elements and heavy metals content on *Mytilus galloprovincialis* from River Tagus estuary. Biol. Trace Elem. Res., 159: 233-240. <u>https://doi.org/10.1007/s12011-014-9974-</u> y
- Salaah, S. M.; Khalil, M. T.; Gad, N. S. and Ahmed, N. A. M. (2018). Physico-chemical characteristics and physiological changes in *Oreochromis niloticus* from Rosetta branch of The River Nile. Eur. Chem. Bull.,7(2): 63-71. <u>http://dx.doi.org/10.17628/ecb.2018.7.63-71.</u>
- Shalloof, K. A. S. (2020). State of fisheries in lake Qarun, Egypt. EASEDJ-D.ES, Environmental Studies, 21(2), 1-10. <u>https://doi.org/10.21608/jades.2020.73188</u>
- Singh, R.; Venkatesh, A. S.; Syed, T. H.; Reddy, A. G. S.; Kumar, M. and Kurakalva, R. M. (2017). Assessment of potentially toxic trace elements contamination in groundwater

resources of the coal mining area of the Korba Coalfield, Central India. Enviro. Earth Sci., **76**(*16*): 1-17. https://doi.org/10.1007/ s126 65 -017-6899-8

- Song, B.; Mei, L. and Chen, T. (2009). Assessing the health risk of heavy metals in vegetables to the general population in Beijing, China. J. Environ. Sci., 21: 1702-1709. <u>https://doi.org/10.1016/s1001-0742(08)62476-6</u>
- Sorensen, E. M. (1991). Metal poisoning in fish, environmental and life sciences Associates. Austin. Texas. CRC Press Inc., Boston.
- Taha, A.; Hussien, W. and Gouda, S. A. (2023). Bioremediation of Heavy Metals in Wastewaters: A Concise Review. Egypt. J. Aquat. Biol. Fish., 27(1): 143-166. <u>https://doi.org/10.21608/ejabf.2023.284415</u>
- Tayel, S. I.; Mahmoud, S. A.; Ahmed, N. A. M. and Abdel Rahman A. A. S. (2018). Pathological impacts of environmental toxins on *Oreochromis niloticus* fish inhabiting the water of Damietta branch of the River Nile, Egypt. Egypt. J. Aquat. Biol. Fish., 22(5): 309-321. <u>https://doi:10.21608/ejabf.2018.25660</u>
- Wepener, V.; Van Vuren, J. H. J. and Du Preez, H. H. (2001). Uptake and distribution of a copper, iron and zinc mixture in gill, liver and plasma of a freshwater teleost, *Tilapia sparmanii*. Water SA., 27(1): 99-108. <u>https://doi.org/10.4314/wsa.v27i1.5016</u>
- World Health Organization (WHO) (1993). Evaluation of Certain Food Additives and Contaminants (Forty-first report of the joint FAO/ WHO Expert Committee on Food Additives). WHO, Geneva, Technical Report Series no 837.
- Yacoub, A. M. (2007). Study on some heavy metals accumulated in some organs of three river Nile fishes from Cairo and Kalubia Governorates. Afr. J. Biol. Sci., 3: 9-21.
- Yılmaz, F.; Ozdemir, N.; Demirak, A. and Levent Tuna, A. (2007). Heavy metal levels in two fish species *Leuciscus cephalus* and *Lepomis gibbosus*. Food Chem., 100: 830-835. <u>https://doi.org/10.1016/j.foodchem.2005.09.020</u>
- Yosef, T. A. and Gomaa, G. M. (2011). Assessment of Some Heavy Metal Contents in Fresh and Salted (Feseakh) Mullet Fish Collected from El-Burullus Lake, Egypt. J. Amer. Sci., 7(10): 137-144.
- Younis, A. M.; Hanafy, S.; Elkady, E. M.; Alluhayb, A. H. and Alminderej, F. M. (2024b). Assessment of health risks associated with heavy metal contamination in selected fish and crustacean species from Temsah Lake, Suez Canal. Sci. Rep., 14(1), 18706. https://doi.org/10.1038/s41598-024-69561-7
- Younis, M. L.; Rizk, E. S. T.; Elewa, S. E.; M. Abo-Elfotouh, O. and Mola, H. R. (2024a). Seasonal evaluation of heavy metals and zooplankton distribution and their co-

relationship in the Rosetta branch area of the Nile Delta in Egypt. Appl. Water Sci., 14(4), 73. <u>https://doi.org/10.1007/s13201-024-02121-6</u>

- Zhou, H. A.; Cheung, R. Y. H.; Chan, K. M. and Wong, M. H. (1998). Metal concentrations in sediments and Tilapia collected from inland waters of Hong Kong. Water Res., 32: 3331-3340. <u>https://doi.org/10.1016/S0043-1354(98)00115-8</u>
- Zhuzzhassarova, G; Azarbayjani, F. and Zamaratskaia, G. (2024). Fish and seafood safety: human exposure to toxic metals from the aquatic environment and fish in central Asia. Int. J. Mol. Sci., 25(3): 1590. <u>https://doi.org/10.3390/ijms25031590</u>
- Zyadah, M. (1999). Accumulation of some heavy metals in *Tilapia zillii* organs from Lake Manzalah, Egypt. Tr. J. Zool., 23: 365-372.