



EGYPTIAN ACADEMIC JOURNAL OF  
**BIOLOGICAL SCIENCES**  
TOXICOLOGY & PEST CONTROL

F



ISSN  
2090-0791

WWW.EAJBS.EG.NET

**Vol. 15 No. 1 (2023)**

[www.eajbs.eg.net](http://www.eajbs.eg.net)



## Efficacy of Common Synthetic Insecticides for Management of Fall Armyworm, *Spodoptera frugiperda* (Lepidoptera: Noctuidae) in Egypt

Sherehan A. R. Salem<sup>1</sup>, Hassan F. Dahi<sup>2</sup>, Farouk A. Abdel-Galil<sup>3</sup>, and Mervat A. B. Mahmoud<sup>1</sup>

<sup>1</sup>Zoology Dep., Fac. of Science, South Valley University, Qena, Egypt.

<sup>2</sup>Plant Protection Research Institute, Dokki, Giza, Egypt.

<sup>3</sup>Plant Protection Department, Faculty of Agriculture, Assiut University, Assiut, Egypt.

\*E-mail: [sherehan.abdelkareem@sci.svu.edu.eg](mailto:sherehan.abdelkareem@sci.svu.edu.eg) ; [hassandahi@yahoo.com](mailto:hassandahi@yahoo.com) ;  
[faagalil@aun.edu.eg](mailto:faagalil@aun.edu.eg) ; [mervat.mahmoud@sci.svu.edu.eg](mailto:mervat.mahmoud@sci.svu.edu.eg)

### ARTICLE INFO

Article History  
Received:29/2/2023  
Accepted:19/5/2023  
Available:26/5/2023

#### Keywords:

Chemicals,  
control, pest,  
insecticide, FAW,  
methomyl,  
chlorpyrifos, and  
spinosad.

### ABSTRACT

Being the most invasive and destructive pests in maize fields, the promising insect; is the fall armyworm (FAW), *Spodoptera frugiperda* (J. E. Smith) (Lepidoptera: Noctuidae). The first onset of fall armyworm in Egypt started in 2019 in the south of Egypt among maize plants in Aswan Governorate. Heading north, the pest spread quickly to Luxor, Qena, Sohag and Assiut governorates in the south. Many synthetic insecticides were recommended to control this insect pest. For this purpose, this research was to evaluate the potential effectiveness and toxicity of the following insecticides, Methomyl (Lannate<sup>®</sup> 90%), Chlorpyrifos (Dofos 48%) and Spinosad (Tracer 24%) on the biological aspects of *S. frugiperda* in the laboratory. Bioassays were conducted on the newly molted 4<sup>th</sup> instar of *S. frugiperda* larvae under  $25 \pm 2$  °C room temperature,  $70 \pm 10\%$  relative humidity. The LC<sub>50</sub> values were 105.5, 470, and 2.5 ppm for the compounds methomyl, chlorpyrifos, and spinosad, respectively. Distilled water was served to the control group. Accumulative larval mortality percentages were 42, 55 and 44 for methomyl, chlorpyrifos, and spinosad, respectively. A significant increase in the total duration of the consecutive larval instars and pupae was recorded post-treatment at the 4<sup>th</sup> instar larvae with the LC<sub>50</sub> concentrations of tested insecticides. Accordingly, the differences were significant in the toxicity to *S. frugiperda*; hence, the variation in LC<sub>50</sub> is a reference to the varying actions of the tested insecticides. These data indicate that these recommended synthetic insecticides are effective in controlling fall armyworm instars.

### INTRODUCTION

In early 2016, the fall armyworm *Spodoptera frugiperda* recorded the first outbreak in West and Central Africa (Goergen *et al.*, 2016). Whilst in Egypt, particulate in Upper Egypt; the Agricultural Pesticide Committee (APC) of the Ministry of Agriculture announced the first case of *S. frugiperda* in May 2019 among a maize yield in Kom Ombo City located in Aswan Governorate (FAO 2019; Dahi *et al.*, 2020). As well, on the 6<sup>th</sup> of

August of 2021, fall armyworm began to invade the maize crop in Assuit Governorate, Upper Egypt; where the damage was more severe on sorghum plants (Mohamed *et al.*, 2022).

As a result of the significant damaging influence of *Spodoptera frugiperda* in Africa, 300 million people could be threatened by hunger. *S. frugiperda* has a very broad host extent, and 353 plant species from 76 different plant families, including significant hosts from the Poaceae, Asteraceae, and Fabaceae, have been identified as larval hosts (Casmuz Augusto *et al.*, 2010; CABI 2018; Montezano *et al.*, 2018). This insect can significantly harm crops by decreasing productivity yield and resulting in considerable financial losses (Idrees *et al.*, 2022).

Under optimal conditions, in a single life cycle, a female *S. frugiperda* lays 1500 eggs that overlap on the adaxial face of a corn leaf. Four days after oviposition, the first instar larvae emerge and scrape the leaf, whereas subsequent instars completely consume the leaf, causing severe damage and plant death. The last larval instar is typically found within a leaf sheath (Cruz 1995; Galo *et al.*, 2002; Capinera 2008; Valicente & Tuelher 2009) where it is protected from insecticide applications (Gassen, 1996).

Chemical insecticides have been most commonly used to control pests in plants since they are the most effective, easy to use, and provide satisfactory results. Even with some disadvantages, chemical inputs are vital to the maintenance of high yields in modern agriculture (EPA, 2021).

Neurotoxic insecticides for example chlorfenapyr and methomyl (Viana & Costa 1998) are often applied to control *S. frugiperda* larvae. Nevertheless, these insecticides also can be used for eggs (Travares *et al.*, 2011) and the mature stage of *S. frugiperda* (Pratissoli *et al.*, 2004). Application of insecticides to the larvae of *S. frugiperda* consequently is widen control efficiency.

Methomyl, an oxime carbamate, was the first insecticide has been synthesized in 1968 for the management of many insect classes at the broad range, involving Hemiptera, Lepidoptera, Diptera, Homoptera, and Coleoptera. Similar to various carbamates, Methomyl is able to suppress the acetylcholine (AChE) activity, accompanied by nerve and/or tissue insufficiency and probably death occur. Owing to significant toxicity to varying stages of insects as larva and adult, methomyl is assumed to be highly degraded metabolically via mixed-function oxidase (Van Scoy *et al.*, 2013). Despite of own acute toxicity, rare incidents rise from labeled usage of methomyl, perhaps because of its weak stability, low bioaccumulative activity, and more reversible with speedy dispersal from almost environmental matrices (Mortensen and Serex, 2014).

Furthermore, chlorpyrifos is an insecticide of wide spectrum use that can destroy insects by means of disruption of the nervous system via block of the breakdown of a neurotransmitter AChE (Smegal, 2000). Chlorpyrifos inhibit the breakdown of ACh through binding to the active site of the cholinesterase (ChE) in the synaptic cleft. As a result, excess ACh accumulates in synaptic clefts, which results in neurotoxicity and eventual death for the neurons (Karanth *et al.*, 2000).

Ultimately, spinosad is an insecticide obtained naturally from an actinomycete bacterium species, *Saccharopolyspora spinosa* (Mertz and Yao 1990) that confer the efficiency of a synthetic insecticide. It is composed of spinosyn A and D which are designated the two most active metabolites. Both spinosyns are readily degraded in moist aerobic soil, and field dissipation, which is quite rapid (half-life, 0.3-0.5 d) can be attributed to photolysis or a combination of metabolism and photolysis. Spinosad induces neurologic impacts in insects that are appropriate to the general stimulation of nicotinic acetylcholine receptors but by a novel mechanism among familiar insecticide compounds. Spinosad is considered more efficient for lepidopteran larvae, in addition to some Diptera, Thysanoptera,

Coleoptera, and Hymenoptera, however, it exhibits limited toxicity to other insects and reveals less activity to mammals and wildlife.

Wherefore, the aim of this work was to estimate the efficiency of the known used synthetic insecticides methomyl, chlorpyrifos and spinosad against new invasive pests in Egypt and study their biological effects on the pest like larval mortality percentage, larval duration, pupation percentage, pupal weights of both sexes, pupal duration of male and female, pupal mortality percentage, adult emergence, sex ratio, male and female longevity, fecundity and fertility % for developing an emergency-based approach by novel insecticides for reducing crop wastes by repressing this notorious pest in Egypt and other influenced countries.

## MATERIALS AND METHODS

The current design was established for studying the effect of methomyl, chlorpyrifos and spinosad insecticides toward newly moulted 4<sup>th</sup> of *S. frugiperda* larvae.

### Test Insect:

The culture of FAW was initiated with larvae gathered from maize cultivation at Qena province existed in Upper Egypt. Where they were kept at the laboratory for several generations and reared in laboratory environments under controlled  $25 \pm 2$  °C,  $70 \pm 10$  % RH), and fed on castor leaves according to Dahi (1997). Larvae have daily provided and fed on castor bean leaves. The resultant pupae were placed in clean moist sawdust jars to supply the pupation site. But adults were supplied 10 % sugar solution.

### Insecticides:

Commercial formulations of the most applied insecticides belonging to various chemical groups were used. The insecticide concentrations assigned in this study were Methomyl (Lannate® 90), Chlorpyrifos (Dofos 48%), and Spinosad (Tracer 24%) (Table 1). The concentrations used for each insecticide were 180, 90, 45 and 22.5 ppm for methomyl, 1000 ppm, 600 ppm, 500 ppm, 250 ppm, 200 ppm and 125 ppm for chlorpyrifos and 5 ppm, 3 ppm, 2 ppm, 1 ppm, 1.5 ppm, and 0.5 ppm for spinosad.

### Bioassay of Tested Compounds on *S. frugiperda* (Smith):

The larvicidal efficiency of the studied compounds was estimated on newly moulted 4<sup>th</sup> of *S. frugiperda* larvae. Each of the prepared concentrations of tested compounds was immersed in fresh castor oil leaves; then allowed to dry at room temperature prior to being offered to the 4<sup>th</sup> instar larvae preserved in ice cube packs. Larvae were awarded contaminant leaves for a period of 24 hours. Each handling involved 50 larvae which were replicated five times. The control comprised similar numbers of larvae, and given castor oil leaves immersed in distilled water. The mortality percent of the larvae was calculated post-exposure by 24 h. and corrected according to Abbott's formula (1925). The obtained results were expressive graphically and LC<sub>50</sub> values were calculated using computerizing LDP line plots.

**Table 1.** Insecticide formulations tested against *S. frugiperda* larvae including active ingredients, formulation type, manufacturer, chemical group and respective mode of action.

Common name	Trade name	Manufacturer	Chemical group	Mode of action
Methomyl	Lannate 90% S P	DuPont de Nemours, USA	Carbamate	Acetylcholinesterase (AChE) inhibitors
Chlorpyrifos	Dofos 48% EC	chema industries -Elisra Industries	Organophosphorous	Acetylcholinesterase (AChE) inhibitors
Spinosad	Tracer 24% SC	UK Dow Agrosiences	5, Spinosyns	Nicotinic acetylcholine receptor (nachr) allosteric modulators - site i

### Biological Studies:

Fresh castor oil leaves were immersed in the LC<sub>50</sub> for each insecticide and then left to dry at room temperature before being offered contaminated leaves as before for each compound. 50 larvae were comprised in each treatment enclosed by five times replication (10 larvae/ice cube packs). The same number of larvae were considered a control; these larvae were offered castor oil leaves immersed in distilled water. The following parameters were recorded: larval mortality percentage, larval duration, pupation percentage, pupal weight, male pupal weight, female pupal weight, pupal duration, male pupal duration, female pupal duration, pupal mortality percentage, adult emergence, sex ratio, male and female longevity, fecundity (Number of eggs laid per female) and fertility percentage (Egg hatchability).

### Statistical Analysis:

The current data were subjected to analysis of variance (ANOVA) using the software package SPSS Statistics version 25 (New York, USA). The significance of variable treatments was determined by Duncan test ( $p < 0.05$ ). Lethal concentrations at LC<sub>50</sub>, 95% confidence limits (CLs) and slope were calculated using the Ldp Line software according to Finney (1971).

## RESULTS

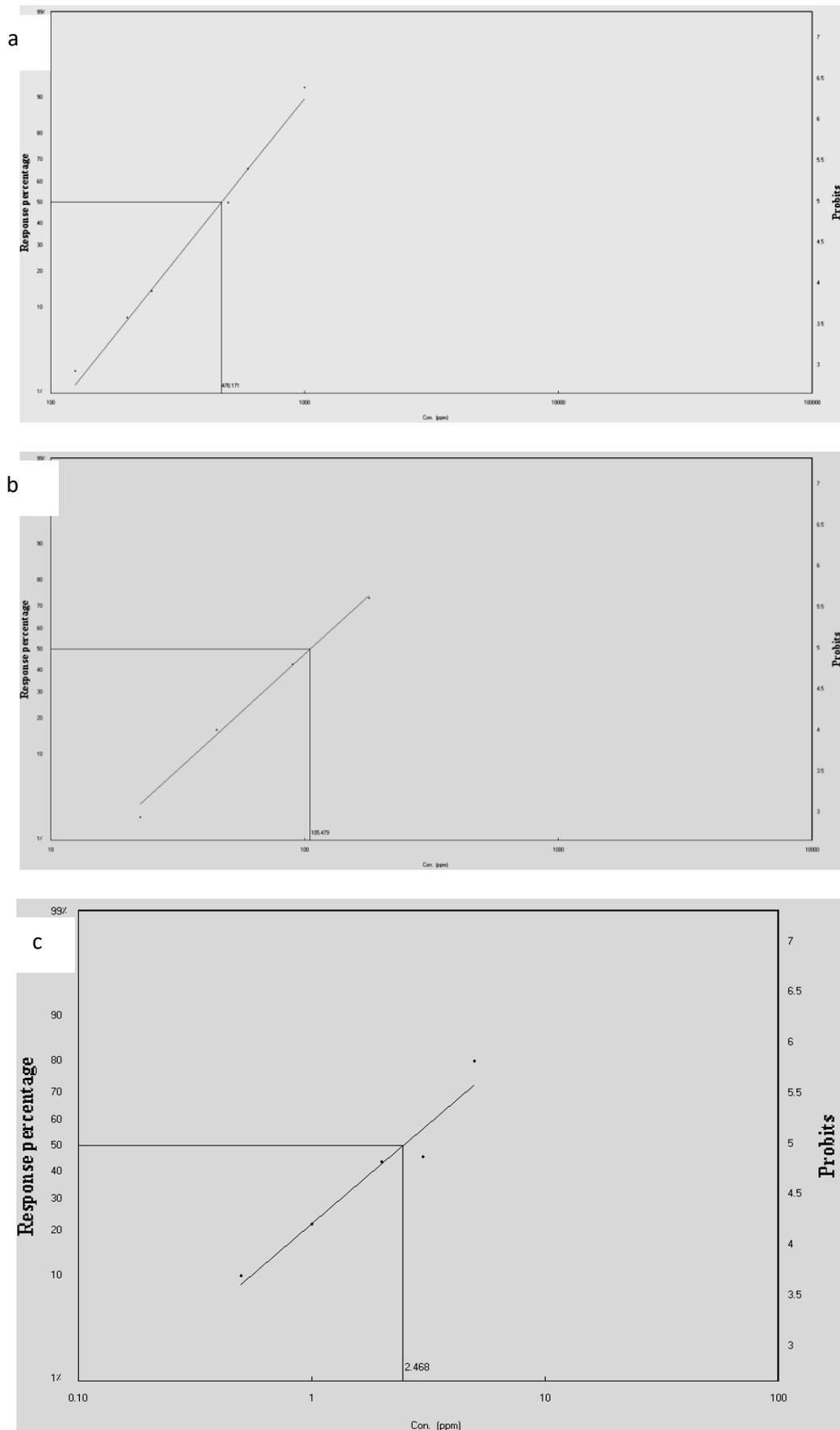
### Effect of Toxicity by Insecticides on 4<sup>th</sup> Instars Larvae of *S. frugiperda*:

As represented in Table 2, the toxicity by methomyl, chlorpyrifos, and spinosad on the 4<sup>th</sup> larval instar of *S. frugiperda* varied according to their concentrations and chemical structure. Overall, the high concentration attended by a rise in mortality rate, and conversely.

The LC<sub>50</sub> values were 105.5, 470 and 2.5 ppm for methomyl, chlorpyrifos, and spinosad, respectively. Correspondingly, the most effective chemical was spinosad, followed by methomyl, then chlorpyrifos. The results proved the toxicity effect on *S. frugiperda* was different. Moreover, variations in LC<sub>50</sub> values perhaps reflected the different modes of action of insecticides.

**Table 2.** The toxicity of the insecticides on the 4<sup>th</sup> larval instar of fall armyworm, *S. frugiperda*.

Insecticide	Concentrations (ppm)	Mortality %	Corrected mortality %	LC <sub>50</sub> (ppm)	Slope
Methomyl	180	74	73.45	105.5	2.816
	90	44	42.86		
	45	18	16.33		
	22.5	4	2.04		
	Control	2			
Chlorpyrifos	1000	92	92	470	3.87
	600	66	66		
	500	50	46		
	250	14	14		
	200	8	8		
	125	2	2		
	Control	0.0			
Spinosad	5	80	80	2.5	1.938
	3	46	46		
	2	44	44		
	1	40	40		
	1.5	22	22		
	0.5	10	10		
	Control	0.0			



**Fig. 1.** The toxicity LDP line of the insecticides used on the 4<sup>th</sup> larval instar of fall armyworm, *S. frugiperda*, a) Chlorpyrifos b) Methomyl c) Spinosad

## **1- Latent Effect of LC<sub>50</sub> of Methomyl on Some Biological Aspects of 4<sup>th</sup> Instars *S. frugiperda* Larvae:**

### **1.1-Effect on the Larvae Development:**

Under conditions of the present work, the duration of the initially treated instar (i.e., newly ecdysed 4<sup>th</sup> instar larvae) with LC<sub>50</sub> of methomyl, plus the subsequent 5<sup>th</sup> and 6<sup>th</sup> larval instars and up to pupation, was a total of 12.93±0.21 days. Meanwhile, control data expressed that the development of untreated larvae from the start of the 4<sup>th</sup> instar up to the initiation of the pupal stage was 11.09±0.28 days. This shows that as a result of the treatment the larval stage was extended by 1.84 days compared to the equivalent control (Table 3).

The accumulative larval mortality percentage was 42 % for the 4<sup>th</sup> instar larvae exposed to LC<sub>50</sub> methomyl at the termination of the larval stage and prior to pupation. This percentage was 3% in untreated larvae (Table 3).

### **1.2- Effect of LC<sub>50</sub> Methomyl on *S. frugiperda* Pupae Treated As 4<sup>th</sup> Instar Larvae:**

As illustrated in Table 3, the percentage of pupation was 58% among the recently produced 4<sup>th</sup> instar larvae following subjected to LC<sub>50</sub> methomyl. The percentage of pupation was 97% in untreated larvae.

The mean weight of a pupa in the control was 0.1824±0.01 g; a female pupa weighed slightly less than a male pupa, 0.1737±0.004 g and 0.1923±0.004 g, respectively. The mean weight of pupa following its treatment as 4<sup>th</sup> instar larvae with LC<sub>50</sub> of methomyl was 0.1998 g, which was not significantly different from their control. As depicted in Table 4, the weight of the female and the male pupae were comparable, being 0.1927±0.004 g and 0.2152±0.006 g, respectively. The pupal mortality reached 6% in treatment as well as in the control.

The mean duration of the pupal stage in the control group averaged 11.29±0.47 days, this period was 11.28±0.54 days when larvae were treated as 4<sup>th</sup> instar. It is noteworthy that the duration of the pupal stage, whether male or female, in treated 4<sup>th</sup> instar larvae, didn't significantly change from that in the untreated insects (Table 4).

### **1.3- Effect on Adult Emergence and Reproductive Potential of Moths from Treated 4<sup>th</sup> Instar *S. frugiperda* Larvae with LC<sub>50</sub> Methomyl.**

Adult emergence was impaired following treatment of *S. frugiperda* larvae with LC<sub>50</sub> of methomyl. This effect was 94 %, as it was in their control (Table 4).

As seen in Table 5, normally, the sex ratio of emerging moths is 1:1 (male: female); however, in moths protruding from 4<sup>th</sup> larvae dealing with LC<sub>50</sub> of methomyl, the sex ratio was 1:1.4 (male: female). The average life span of adult females for the control moth was 11.25 days, represented by a mean of 3.0±0.71, 6.0±0.41 and 1.0±0.71 days for pre-oviposition, oviposition and post oviposition period, respectively. The life span of the emerged moths from 4<sup>th</sup> instar larvae treated with the LC<sub>50</sub> methomyl was 10.5±0.29 days. Whereas, the pre-oviposition, oviposition, and post-oviposition periods were 3.0±0.41, 6.75±0.63 and 0.0 days, respectively. The reproduction potential of moths emerging from treated larvae was also highly significantly reduced (Table 5). The totality of eggs deposited per female was 1991.25±45.0 eggs per female with 90 percent egg hatchability. In comparison, females in their respective control group deposited 2202±105.17 eggs/ female with 98 % egg viability. The incubation periods for the second generation eggs were 2.75±0.25 and 2.75±0.25 days for (treated and control) *S. frugiperda* eggs when treated as 4<sup>th</sup> instar larvae treated with LC<sub>50</sub> of methomyl, as shown in Table (6).

## **2- Effect of LC<sub>50</sub> of Chlorpyrifos on Some Biological Aspects of 4<sup>th</sup> Instar *S. frugiperda* Larvae:**

### **2.1-Effect on the Development of Treated Larvae:**

Following treatment of the 4<sup>th</sup> instar with LC<sub>50</sub> of chlorpyrifos, the mean development period of the subsequent larval instars was significantly extended. As seen in Table 3, larval development took 13.89±0.07 days following the initial treatment (4<sup>th</sup> instar) up to pupation,

as compared to  $11.09 \pm 0.28$  days in their respective control group. The accumulative mortality of the larvae was 55% at the end of the larval stage for 4<sup>th</sup> instar. Of the surviving larvae; 45% were successfully pupated in the respective treated instar, as compared to 97% of the untreated larvae (Table 3).

### **2.2- Effect of LC<sub>50</sub> Chlorpyrifos on *S. frugiperda* Pupae Treated as 4<sup>th</sup> Instar Larvae.**

Data presented in Table 4 mean weight of pupa developing from 4<sup>th</sup> instar larvae treated with LC<sub>50</sub> chlorpyrifos was  $0.2292 \pm 0.01$  g, respectively, which significantly exceeded the weight of control pupa. This was obvious in both female and male pupae.

It is noteworthy, that the duration of the pupal stage, whether male or female developed from treated 4<sup>th</sup> instar larvae was comparable and insignificantly similar to untreated insects (Table 4). Furthermore, the mortality of pupae was relatively low reaching 7 % in pupae treated as 4<sup>th</sup> instar, compared to 6% in the respective control. The percent of malformed pupae was 6.0 % as compared to zero % in the control.

### **2.3- Effect on Adult Emergence and Reproductive Potential of Moths from Treated 4<sup>th</sup> Instar *S. frugiperda* Larvae with LC<sub>50</sub> Chlorpyrifos.**

Table 4 exhibited that percentage of adult eclosion was relatively unaffected as a result of the treatment of 4<sup>th</sup> *S. frugiperda* larvae by LC<sub>50</sub> chlorpyrifos. In this respective mentioned instar, adult emergence was at a high of 94%, which was only slightly decreased from their control by 1 %. The sex ratio in emerged moths from untreated larvae was equal to 1:1 (male: female) however, this ratio was slightly in favor of an increase in female moths (1: 1.3) emerging from 4<sup>th</sup> instar larvae treated with LC<sub>50</sub> chlorpyrifos.

Under the laboratory conditions of the current work, the life span of female *S. frugiperda* treated with LC<sub>50</sub> chlorpyrifos averaged  $11.83 \pm 2.48$  days, giving a pre-oviposition, oviposition, and post-oviposition period of  $2.25 \pm 0.25$ ,  $7.5 \pm 0.29$  and  $0.75 \pm 0.48$  days, respectively Table 5. A female moth deposited  $2166 \pm 105.53$  eggs with 85% hatchability Table 6.

## **3-Effect of LC<sub>50</sub> of Spinosad on Some Biological Aspects of 4<sup>th</sup> instar *S. frugiperda* larvae**

### **3.1-Effect on the Development of Treated Larvae:**

As exhibited in Table 3, 4<sup>th</sup> instar *S. frugiperda* larvae when treated with LC<sub>50</sub> of spinosad caused a significant extension of 2.62 days in the duration of the larval stage, i.e.,  $13.71 \pm 0.1$  days versus  $11.09 \pm 0.28$  days in the control.

Larval mortality percentages were 44 % and 3 % at the termination of the larval stage in treated and untreated larvae, respectively, giving a respective pupation percentage of 56% and 97 % (Table 3).

### **3.2- Effect of LC<sub>50</sub> Spinosad on *S. frugiperda* Pupae Treated as 4<sup>th</sup> Instar Larvae.**

As seen in Table 4, the mean weight of a newly formed pupa was 0.2261 g, following their treatment as 4<sup>th</sup> instar larvae with LC<sub>50</sub> of spinosad was significantly more than the control. This was evident in both female and male pupae.

The duration of the pupal stage, being male or female in treated 4<sup>th</sup> instar larvae was a non-significant difference from that in untreated insects (Table 4).

The percent of malformed pupae was 6.0 % as compared to zero % in the control.

### **3.3- Effect on Adult Emergence and Reproductive Potential of Moths from Treated 4<sup>th</sup> Instar *S. frugiperda* Larvae with LC<sub>50</sub> Spinosad.**

As observed in Table 4, in the treatment of 4<sup>th</sup> instar *S. frugiperda* larvae with LC<sub>50</sub> spinosad, the percentage of adult eclosion was slightly reduced to 79% as compared to 94 % in the control. The Percentages of malformed moths were much higher in moths developed from treated 4<sup>th</sup> instar larvae (7.0%). The sex ratio in moths that emerged from the treated 4<sup>th</sup> instar was 1:1.7 (male: female) Table 5.

Under the conditions of the present work, the life span of a female moth in control averaged 11.0 days, giving  $3.0 \pm 0.71$ ,  $6.0 \pm 0.41$  and  $1.0 \pm 0.71$  days for the pre-oviposition, oviposition and post oviposition periods, respectively. The average of deposited eggs was 2202.5 eggs per female, with 98% egg viability.

Emerged moths from 4<sup>th</sup> instar larvae treated with LC<sub>50</sub> spinosad had a life span of 11.0 days. In the treated larval instar, the periods for pre-oviposition, oviposition and the post oviposition stages were not affected. The reproductive potential of these moths was significantly reduced. The mean number of deposited eggs was 2006.75 eggs per female in emerged moths from the treated 4<sup>th</sup> instar larvae, with respective egg viability of 90%.

**Table 3:** Effects of LC<sub>50</sub> concentrations of the tested insecticides on the larval stage of *S. frugiperda*

Biological aspects	Control	Methomyl	Chlorpyrifos	Spinosad
Larval duration (days)	$11.09 \pm 0.28$ C	$12.93 \pm 0.21$ b	$13.89 \pm 0.07$ a	$13.71 \pm 0.1$ a
Pupation %	97	58.0	45.0	56.0
Larval mortality %	3	42.0	55.0	44.0
Normal larvae %	100	100	100	100
Malformed larvae %	0.0	0.0	0.0	0-0

➤ The means that have the same letter horizontally are insignificant.

**Table 4:** Effects of LC<sub>50</sub> concentrations of the tested insecticides on pupal stage of *S. frugiperda*.

Biological aspects	Control	Methomyl	Chlorpyrifos	Spinosad
Pupal durations (days)	$11.29 \pm 0.47$ a	$11.028 \pm 0.54$ a	$10.74 \pm 0.01$ a	$10.58 \pm 0.33$ a
Female pupal durations (days)	$10.44 \pm 0.26$ a	$10.8 \pm 0.27$ a	$10.0 \pm 0.2$ a	$10.3 \pm 0.43$ a
Male pupal durations (days)	$12.17 \pm 0.23$ a	$12.38 \pm 0.43$ a	$11.50 \pm 0.2$ ab	$11.09 \pm 0.31$ b
Normal pupae %	100	100	94.0	94.0
Malformed pupae %	0.0	0.0	6.00	6.00
Pupal weight (gm.)	$0.1824 \pm 0.01$ c	$0.1998 \pm 0.01$ bc	$0.2292 \pm 0.01$ a	$0.2261 \pm 0.01$ ab
Female pupal weight (gm.)	$0.1773 \pm 0.004$ c	$0.1927 \pm 0.004$ b	$0.2137 \pm 0.001$ a	$0.2126 \pm 0.008$ a
Male pupal weight (gm.)	$0.1923 \pm 0.004$ a	$0.2152 \pm 0.006$ a	$0.2495 \pm 0.011$ b	$0.2451 \pm 0.001$ b
Pupal mortality %	6.0	6.00	7.0	21.0
Emergence %	94.0	94.0	93.0	79.0

➤ The means that have the same letter horizontally are insignificant.

**Table 5:** Effects of LC<sub>50</sub> concentrations of the tested insecticides on adult stage of *S. frugiperda*.

Biological aspects	Control	Methomyl	Chlorpyrifos	Spinosad
Adult malformation %	0.0	0.0	0.0	7.0
Sex ratio % (♂ : ♀)	1:1	1:1.8	1:1.3	1:1.7
Adult longevity (days)	$12.0 \pm 1.08$ a	$11.75 \pm 1.11$ a	$11.83 \pm 2.48$ a	$12.25 \pm 0.75$ a
Female longevity (days)	$11.25 \pm 0.85$ a	$10.5 \pm 0.29$ a	$11.0 \pm 0.58$ a	$11.0 \pm 0.0$ a
Male longevity (days)	$13.75 \pm 1.25$ a	$11.75 \pm 1.11$ a	$14.0 \pm 0.41$ a	$13.5 \pm 0.5$ a
Pre-oviposition period (days)	$3.0 \pm 0.71$ a	$3.0 \pm 0.41$ a	$2.25 \pm 0.25$ a	$3.0 \pm 0.29$ a
Oviposition period (days)	$6.0 \pm 0.41$ b	$6.75 \pm 0.63$ ab	$7.5 \pm 0.29$ a	$6.0 \pm 0.41$ b
Post-oviposition period (days)	$1.0 \pm 0.71$	0.0	$0.75 \pm 0.48$	0.0
Fecundity (No. eggs / female)	$2202.5 \pm 105.17$ a	$1991.25 \pm 45.0$ a	$2166.5 \pm 105.53$ a	$2006.75 \pm 157.22$ a

➤ The means that have the same letter horizontally are insignificant.

**Table 6:** Effects of LC<sub>50</sub> concentrations of the tested insecticides on egg stage of *S. frugiperda*.

Biological aspects	Control	Methomyl	Chlorpyrifos	Spinosad
Hatchability %	98	90	85	90
Incubation period (days)	2.75 ± 0.25 a	2.75 ± 0.25 a	3.0 ± 0.0 a	2.75 ± 0.25 a

➤ The means that have the same letter horizontally are insignificant.

## DISCUSSION

In the existing work, known-applicable three insecticides; methomyl, chlorpyrifos, and spinosad have been chosen against 4<sup>th</sup> instar larvae of *S. frugiperda* to evaluate their toxicity and their latent effect on the larvae, pupae and emerged adults

The findings of the current investigation demonstrated that all of the synthetic insecticides tested were significantly effective against *S. frugiperda* larvae in their 4<sup>th</sup> instar. Researchers from all over the world have recently extensively examined the different possible management tactics used to combat FAW in Africa and India, were among the regions where registered pesticides were tested in the field and laboratory as an emergency measure after this invasive insect severely harmed the maize crop (Kumela *et al.*, 2019; Kassie *et al.*, 2020), (Kumar, and Mohan 2022, Kumar *et al.*, 2023). However, this was the first investigation to assess the toxicity of these specific pesticides to *S. frugiperda* larvae in a laboratory setting in Egypt. The findings of the current investigation showed that *S. frugiperda* larvae in their fourth instar were susceptible to these synthetic pesticides; nonetheless, Chlorpyrifos was the most effective one on the 4<sup>th</sup> instar *S. frugiperda* larvae, followed by spinosad and then methomyl. Based on fiducial limits at 95%, there were significant differences in LC<sub>50</sub>'s among the tested insecticides, Insecticide toxicity regression lines showed homogeneity between tested populations and tested insecticide concentrations. The current study's findings are in accordance with (Viana & Costa 1998). Also, Ahissou *et al.*, (2021) mentioned that methomyl, abamectin benzoate and chlorpyrifos-ethyl are extremely effective insecticides from different seven insecticides used against the 3<sup>rd</sup> larvae instar fall armyworm in Burkina Faso. Spinosad provided excellent control of beet armyworm, meanwhile, pyridalyl was lower in efficiency (Mascarenhas *et al.*, 1997 and Torrey *et al.*, 1999). A notable increase was observed in the total duration of the sequent larval instars after the 4<sup>th</sup> instar *S. frugiperda* larvae treated with methomyl, chlorpyrifos and spinosad at LC<sub>50</sub> concentrations. Clinical manifestations due to toxicity were in the form of nervous tremors of the thoracic legs and mouthparts and the larva become was unable to feed. Hence, prolongation of the larval instars and a reduction in the pupation percentage and adult emergence were the results of impairment in the feeding of treated larvae. A study by Abdel-Rahim (2011) showed that second and fourth larval instars of *S. littoralis* deal with pyridalyl and methomyl prolonged the duration of the treated *S. littoralis* larva than their control. The results of this study indicated that the larval duration of the 4<sup>th</sup> instar treated with methomyl was significantly increased ( $p=0.01$ ) to an average of 17 days, compared to 12 days for the control group. Chlorpyrifos and Spinosad insecticides at sub-lethal concentration enhanced deterrent activity feeding of *S. litura* larvae (Singh and Sohi, 2007). Further, it was noticed that *P. gossypiella* of the larval stage treated by LC<sub>50</sub> Radiant (spinetoram) exhibited an extension in larval and pupal developmental periods (El-Barkey *et al.*, 2009). Abd El-Kareem (2016) found that mortality rates gradually increased at 2<sup>nd</sup> and 4<sup>th</sup> instar larvae of *S. littoralis* when fed on treated cotton leaves with LC<sub>50</sub> of tested bioinsecticide included Protecto® (*Bacillus thuringiensis* kurstaki), Viruset® (*Spodoptera littoralis* NPV), and Profect® (mixture of BTK & SLNPV) until reaching the 5<sup>th</sup>-day post application.

Interestingly, for unknown reasons, our results showed that the male and female pupal stage duration was not affected by the LC<sub>50</sub> concentration of the three tested insecticides for 4<sup>th</sup> instar *S. frugiperda* larvae. While exposure to chlorpyrifos or spinosad significantly increased the weight of the sequential pupal stage, this can be explained by the original weight of the larvae and its effect on the insecticides. These results are agreeing with Ismail *et al.* (2022), who treated *S. littoralis* with five different insecticides among them spinosad. They found that the larval duration increased and spinosad was superior in its residual effect on 2<sup>nd</sup> and 4<sup>th</sup> larvae.

Treatments 4<sup>th</sup> instar *S. frugiperda* larvae with LC<sub>50</sub> of spinosad led to a decrease in the percentage of adult emergence. Quite similar results were obtained by Abouelghar *et al.* (2013), When treated cotton leafworm with sublethal and lethal doses of spinosad, reductions in adult emergence, fecundity and fertility.

Treatment with LC<sub>50</sub> values of the three aforementioned insecticides was insignificantly reproductive potential for the tested pest. Meanwhile, the lowest Egg hatchability was recorded in 4<sup>th</sup> instar *S. frugiperda* larvae treated by chlorpyrifos followed by methomyl and spinosad was the same. Similarly, El-Barkey *et al.* (2009) revealed that spinetoram resulted in a significant lowering in the deposited *P. gossypiella* female eggs.

Thus, in addition to inadequate dietary intake and impaired, it's probable that lethal doses of chlorpyrifos fed to FAW had an impact on the insect metabolism after treatment with the insecticide. Besides, delays in insect development are a minor manifestation of endocrine processes and have hormonal effects as shown in Yang *et al.* (2023) who discovered that fall armyworm (FAW) larvae when exposed to LC<sub>30</sub> chlorantraniliprole induced a deceleration in the production of juvenile hormone esterase (*SfJHE*) which have implications for their development and survival, as JH plays a critical role in regulating insect growth and metamorphosis.

The obtained results of the present work show the extent of the importance of methomyl, chlorpyrifos, and spinosad for the invasive eradication of the pest *S. frugiperda* in Egypt. Each of the three insecticides exhibits a different and untraditional mode of action in treated insects. At the same time, it's important to have a wide range of insecticides that kill pests right away and effectively. The obtained results would be important in preventing or delaying the appearance of resistance in treated insects and therefore considered in an IPM program for the control of these economically important and invasive lepidopterous insects. Follow up physiological studies are ongoing to evaluate the effect of these three insecticides on the FAW body enzymes

**Funding** This research was extracted from a project titled "Evaluation Efficacy of some Recommend Insecticides on Fall Armyworm, *Spodoptera frugiperda*" funded by South Valley University, Qena, Egypt.

#### **The Ethical Approval:**

The experimental procedure concerning this work was conducted and approved by the Institutional Review Board for Animal Experiments of South Valley University according to the Ethical Guidelines for the Animals Handling in laboratory experiments of the Faculty of Science, South Valley University, Qena, Egypt (approval no. 010/11/22).

## REFERENCES

- Abbott, W. S. (1925). A method of computing the effectiveness of an insecticide. *Journal of Economic Entomology*, 18 (2), 256-267.
- Abd El-Kareem, S. M. I. (2016). Efficacy of three bioinsecticide and a methomyl insecticide against cotton leafworm larvae, *spodoptera littoralis* under controlled semi-field

- conditions at el-Behara governorate. *Egyptian Academic Journal of Biological Sciences (F. Toxicology & Pest control)*, 8(2), 13-18.
- Abdel-Rahim, E. F. M. (2011). Comparative bio-residual activity of pyridalyl and methomyl insecticides against larvae of the cotton leafworm, *Spodoptera littoralis* (Bosd.). *Egyptian Journal of Agricultural Research*, 89(1): 55-71.
- Abouelghar, G. E., Sakr, H., Ammar, H. A. Y., Nassar, M. (2013). Sublethal effects of spinosad (tracer®) on the cotton leaf worm (Lepidoptera: noctuidae). *Journal of plant protection research*, 53(3), 1-10
- Ahissou, B. R., Sawadogo, W. M., Bokonon-Ganta, A. H., Somda, I., Kestemont, M. P., and Verheggen, F. J., (2021). Baseline Toxicity Data of Different Insecticides against the Fall Armyworm *Spodoptera frugiperda* (J.E. Smith) (Lepidoptera: Noctuidae) and Control Failure Likelihood Estimation in Burkina Faso," *African Entomology*, 29(2), 435-444.
- Barros, E. M., Torres, J. B., and Bueno, A. F., (2010). Oviposição, desenvolvimento e reprodução de *Spodoptera frugiperda* (J.E. Smith) (Lepidoptera: Noctuidae) em diferentes hospedeiros de importância econômica. *Neotropical entomology* 36, 996-1001.
- CABI (2018). *Spodoptera frugiperda* (fall armyworm) Invasive species compendium. Wallingford, UK: CAB international. <https://www.cabi.org/isc/datasheet/29810>.
- Capinera, J. L. (2008). *Encyclopedia of Entomology*. 2<sup>nd</sup> Ed., Springer Science & Business Media, Heidelberg ,2061 p. <https://doi.org/10.1007/978-1-4020-6359-6>
- Casmuz Augusto, J. M. L., Socias, M. G., Murúa, M. G., Prieto, S. and Medina, S. (2010), Revision de los hospedero sdelgusa nocogollero del maiz, *Spodoptera frugiperda* (Lepidoptera: Noctuidae). *Revista de la Sociedad Entomológica Argentina*, 69, 209-231.
- Chimweta, M., Nyakudya, I. W., Jimu, L., and Bray, M. A. (2020). Fall Armyworm [*Spodoptera Frugiperda* (J.E. Smith)] Damage in Maize: Management Options for Flood-Recession Cropping Smallholder Farmers. *International Journal of Pest Management.*, 66, 142-154.
- Cruz, I. (1995). A lagarta-do-cartucho na cultura do milho. Circular Técnica 21, Embrapa Milho e Sorgo, Sete Lagoas, Brazil <https://www.infoteca.cnptia.embrapa.br/infoteca/bitstream/doc/475779/1/circ21.pdf> (last accessed 10 Mar 2022).
- Dahi, H. F. (1997). New approach for management the population of cotton leafworm *Spodoptera littoralis* (Boisd.) and pink bollworm *Pectinophora gossypiella* (Saund.) in Egypt. M. Sc. Thesis, Fac. Agric., Cairo Univ., 142 pp.
- Dahi, H. F., Salem, S. A. R., Gamil, W. E. and Mohamed, H. O. (2020). Heat Requirements for the Fall Armyworm *Spodoptera frugiperda* (J. E. Smith) (Lepidoptera: Noctuidae) as a New Invasive Pest in Egypt. *Egyptian Academic Journal of Biological Sciences (A. Entomology)*, 13(4), 73-85.
- El-Barkey, N. M., Amer, A. E. and Kandeel, M. A. (2009). Ovicidal activity and biological effects of radiant and hexaflumuron against eggs of pink bollworm, *Pectinophora gossypiella* (Saunders) (Lepidoptera: Gelechiidae) *Egyptian Academic Journal of Biological Sciences (A. Entomology)*, 2(1): 23 - 36
- El-Hassanin, A. S., Samak, M. R., Abdel-Rahman, G. N., Abu-Sree, Y. H., and Saleh, E. M. (2020). Risk assessment of human exposure to lead and cadmium in maize grains cultivated in soils irrigated either with low-quality water or freshwater. *Toxicology Reports*, 7, 10-15.
- EPA, (2021). <https://www.epa.gov/caddis-vol2/insecticides>

- FAOSTAT, (2019). World Food and Agriculture - Statistical Pocketbook 2019; Food and Agriculture Organization of the United Nations: Rome, Italy, 2019. <https://www.fao.org/documents/card/en/c/ca6463en>
- Farias, P. R. S., Barbosa, J. C., and Busoli, A. C. (2001). Spatial distribution of the fall armyworm, *Spodoptera frugiperda* (J.E. Smith) (Lepidoptera: Noctuidae), on corn crop. *Neotropical Entomology*, 30, 681-689.
- Finney, D. J. (1971). Probit Analysis. 3<sup>rd</sup> Edition, Cambridge University Press, Cambridge.
- Fitzpatrick, B. J.; Mascarenhas, R. N.; Boyd, M. L.; Boyd, D. J.; Burris, E. and Cook, D. (1996). Soybean looper and beet armyworm control on soybean in Louisiana, 1995. *Arthropod Management Tests*, 21:289-291
- Galo, D., Nakano, O., Silveira-Neto, S., Carvalho, R. P. L., Baptista, G. C., Berti-Filho, E., Parra, J. R. P., Zucchi, R. A., Alves, S. B., Vendramim, J. D., Marchini, L. C., Lopes, J. R. S. and Omoto, C. (2002). Entomologia agrícola. FEALQ, Piracicaba, Brazil. 220 p.
- Gamil, W. E., (2020). Fall Armyworm *Spodoptera frugiperda* (J. E. Smith) (Lepidoptera: Noctuidae) Biological aspects as a new alien invasive pest in Egypt. *Egyptian Academic Journal of Biological Sciences (A. Entomology)*, 13(3). 189-196.
- Gassen, D. N., (1996). Manejo de pragas associadas à cultura do milho. Aldeia Norte, Passo Fundo, Brazil.
- Goergen G., Lava K. P., Sankung, S. B, Togola A. and Tamò M., (2016). First report of outbreaks of the fall armyworm *Spodoptera frugiperda* (J E smith) (lepidoptera, noctuidae), a new alien invasive pest in west and Central Africa. *PLoS ONE*, 11(10). e0165632.
- Idrees, A., Qadir, Z. A., Afzal, A., Ranran, Q., Li, J. (2022). Laboratory efficacy of selected synthetic insecticides against second instar invasive fall armyworm, *Spodoptera frugiperda* (Lepidoptera: Noctuidae) larvae. *PLoS One*, 17(5). e0265265.
- IITA – Transforming African Agriculture. (2016). First report of out breaks of the “Fall Armyworm” on the African continent. IITA Headquarters, Ibadan, Nigeria. <http://bulletin.iita.org/index.php/2016/06/18/first-report-of-outbreaks-of-the-fall-armyworm-on-the-african-continent/> (Last accessed 10 Mar 2021).
- Ismail, S. M., Hafez, S. Sh., and Sleem, F. M. A. (2022). Bio-residual activity of novel insecticides in *Spodoptera littoralis* (boisadual, 1833) throughout its life cycle. *Egyptian Academic Journal of Biological Sciences (F. Toxicology & Pest control)*, 14(1).149-158
- Juarez, M. L., Schöfl, G., Vera, M. T., Vilardi, J. C., Murúa, M. G., Willink, E., Hanninger, S., Heckel, D. G., Groot, A. T. (2014). Population structure of *Spodoptera frugiperda* maize and rice host forms in South America: are they host strains? *Entomologia Experimentalis et Applicata*, 152. 182-199.
- Karanth, S., Pope, C. (2000). Carbosylesterase and a-esterase activities during maturation and aging: relationship to the toxicity of chlorpyrifos and parathion in rats. *Toxicological Sciences*, 58. 282-289.
- Kassie, M., Ndiritu, S. W., Stage, J. (2014). What determines gender inequality in household food security in Kenya? Application of exogenous switching treatment regression. *World Development*, 56.153-171.
- Kumar, N. T. D. and Mohan, K. M. (2022). Variations in the susceptibility of Indian populations of the fall armyworm, *Spodoptera frugiperda* (Lepidoptera: Noctuidae) to selected insecticides. *International Journal of Tropical Insect Science*. 42 .1707-1712.

- Kumar, S., Suby, S. B., Vasmatkar P., Nebapure, S. M., Kumar N. and Kumar G. M., (2023). Influence of temperature on insecticidal toxicity and detoxifying enzymes to *Spodoptera frugiperda*. *Phytoparasitica*, 1-13
- Kumela, T., Simiyu, J., Sisay, B., Likhayo, P., Mendesil, E., Gohole, L., and Tefera, T. (2019). Farmers' Knowledge, Perceptions, and Management Practices of the New Invasive Pest, Fall Armyworm (*Spodoptera frugiperda*) in Ethiopia and Kenya. *International Journal of Pest Management*, 65. 1-9.
- Mascarenhas, R. N., Fitzpatrick, B. J., Boyd, M. L., Clemens, C. G., Boethel, D. J., Vidrine, P. R., Moore, S. H., (1997). Evaluation of Selected Experimental and Standard Insecticides Against Soybean Looper, *Arthropod Management Tests*, 22, (1), 314.
- Mertz, E. P. and Yao, R. C. (1990). *Succharopolyspora spinosa* sp now isolated from soil collected in a sugar run still. *International Journal of Systematic Bacteriology*, 40:34-39.
- Mohamed, H. O., El-Heneidy, A. H., Dahi, H. F. and Awad, A. A. (2022). First Record of the Fall Armyworm, *Spodoptera frugiperda* (J. E. Smith) (Lepidoptera: Noctuidae) on Sorghum Plants, A new invasive pest in Upper Egypt. *Egyptian Acaemic Journal of Biological Sciences (A. Entomology)*, 15(1). 15-23
- Montezano, D. G., Sosa-Gómez, D. R., Specht, A., RoqueSpecht, V. F., Sousa-Silva, J. C., Paula-Moraes, S. D. (2018). Host plants of *Spodoptera frugiperda* (Lepidoptera: Noctuidae) in the Americas. *African Entomology*, 26(2). 286-300.
- Mortensen, S. R. and Serex, T. L. (2014). In *Encyclopedia of Toxicology* (Third Edition).
- Pratissoli, D., Thuler, R. T., Pereira, F. F., Reis, E. F., Ferreira, A. T. (2004). Ação transovariana de lufenuron (50 g/l) sobre adultos de *Spodoptera frugiperda* (J. E. Smith) (Lepidoptera: Noctuidae) e seu efeito sobre o parasitóide de ovos *Trichogramma pretiosum* Riley (Hymenoptera: Trichogrammatidae). *Ciência e Agrotecnologia*, 28. 9-14.
- Singh, I.; Sohi, A. S. (2007): Influence of sub-lethal doses of insecticides on the ovipositional pattern of *Spodoptera litura* (Fab.) on cotton. *Journal of Research, Punjab Agricultural University*. 44: 3, 214-215.
- Smegal, D. C. *Human Health Risk Assessment Chlorpyrifos*; U.S. Environmental Protection Agency, Office of Prevention, Pesticides and Toxic Substances, Office of Pesticide Programs, Health Effects Division, U.S. Government Printing Office: Washington, DC, 2000; pp 1-131.
- Tavares, W. S., Cruz, I., Petacci, F., Freitas, S. S., Serrão, J. E., Zanuncio, J. C., (2011). Insecticide activity of piperine: toxicity to eggs of *Spodoptera frugiperda* (Lepidoptera: Noctuidae) and *Diatraea saccharalis* (Lepidoptera: Pyralidae) and phytotoxicity on several vegetables. *Journal of Medicinal Plants Research*, 5. 5301-5306.
- Toepfer, S., Fallet, P., Kajuga, J., Bazagwira, D., Mukundwa, I. P, Szalai M., et al. (2021). Streamlining Leaf Damage Rating Scales for the Fall Armyworm on Maize. *Journal of Pest Science*, 94, 1075-1089.
- Torrey, K. D., Leonard, B. R. and Gore, J. (1999). Control of soybean looper and beet armyworm, 1998. *Arthropod Management Tests*, 24: F115, <http://www.entsoc.org/Protected/AMT/amt1999/F115.html>.
- Valicente, F. H., Tuelher, E. S., (2009). Controle biológico da Lagarta do Cartucho, *Spodoptera frugiperda*, com Baculovírus. Circular Técnica n° 144. Ministério da Agricultura Pecuária e Abastecimento, Sete Lagoas, Brazil. <https://ainfo.cnptia.embrapa.br/digital/bitstream/CNPMS-2010/22431/1/Circ-114.pdf> (last accessed 10 Mar 2021).

- Van Scoy, A. R., Yue, M., Deng, X., Tjeerdema, R. S. (2013). Environmental fate and toxicology of methomyl. *Rev Environ Contam Toxicol.*, 222:93-109.
- Viana, P. A., Costa, E. F., (1998). Controle da Lagarta-do-Cartucho, *Spodoptera frugiperda* (J.E. Smith) (Lepidoptera: Noctuidae) na cultura do milho com inseticidas aplicados via irrigação por aspersão. *Anais da Sociedade Entomológica do Brasil*, 27: 451-458.
- Viana, P. A., Costa, E. F. 1998. Controle da Lagarta-do-Cartucho, *Spodoptera frugiperda* (J.E. Smith) (Lepidoptera: Noctuidae) na cultura do milho com inseticidas aplicados via irrigação por aspersão. *Anais da Sociedade Entomológica do Brasil* 27: 451–458.
- Yang, J., Guan, D., Wei, J., Ge, H., Cao, X., Lv, S., Zhou, X., Zheng, Y., Meng, X., Wang, J., Qian K. (2023). Mechanisms underlying the effects of low concentrations of chlorantraniliprole on development and reproduction of the fall armyworm, *Spodoptera frugiperda*. *Pesticide Biochemistry and Physiology*, 191.105362.