

## **EFFECT OF RUMEN-PROTECTED-GLUCOSE SUPPLEMENTATION DURING TRANSITION PERIOD ON BLOOD METABOLITES AND REPRODUCTIVE PERFORMANCE IN DAIRY COWS**

By

**Howayda S. Belal\* and Dohreig R.M.A\*\***

\*Biology of Reproduction Dept. and \*\*Artificial insemination and Embryo Transfer Dept.,  
Animal Reproduction Research Institute (ARRI)

### **ABSTRACT**

Supplementation of rumen protected glucose (RPG) may be a useful dietary strategy to improve energy balance in transition dairy cows. The present study was performed to investigate whether supplementation with RPG could improve metabolic status, milk production and reduce infection during the transition period in cows. Fifty five healthy Holstein dairy cows (4–5 years old;  $515 \pm 42$  kg body weight; 32.1 kg milk production per day) with the second lactation were enrolled in the study. Cows were randomly assigned into two groups: group (1) control group which fed on control diet, group (2) RPG group which fed on control diet supplemented with RPG at rate of 250 g/cow/day. Supplementation was applied from 20 day before parturition to 60 days postpartum. Blood was sampled on 21 postpartum to examine some serum biochemical parameters [glucose (GLU), non-esterified fatty acid (NEFA) -  $\beta$ -hydroxybutyric acid ( $\beta$ -HBA) and insulin (INS)]. Results showed that serum glucose level was insignificantly different between RPG supplemented and control cows. On the contrary, a rapid decrease was observed in concentration of NEFA and  $\beta$ -HBA and activity of aspartate aminotransferase (AST) in response to RPG supplementation. Meanwhile, dietary RPG supplementation increased plasma concentration of insulin (INS) relative to control cows.

### **Keyword:**

Dairy cattle - Rumen protected glucose - Blood metabolites -Transition period –Reproductive performance.

## INTRODUCTION

There is a variety of situations during lactation in which insufficient glucose justify the time availability may limit productivity mainly in ruminants (**Kvidera, 2017**). Multiple studies on how to increase available glucose to relieve negative energy balance (NEB) in early lactating dairy cows have been operated. Some studies revealed that milk production increased by administering intravenous injection of exogenous glucose to lactating dairy cows (**Brown and Allen, 2013 and Curtis et al., 2018**). Plasma glucose concentration was elevated by infusing glucose into the abomasum (**Nichols et al., 2016 and Gualdrón-Duarte and Allen, 2018**). However, glucose supplementation by intravenous injection or gastrointestinal infusion is not feasible in large commercial dairy farm and providing glucose by feeding additional carbohydrates have a negative effect (**Colman et al., 2013**). Meanwhile, the direct absorption of glucose by the small intestine is more energy efficient (**Moran et al., 2014**). Therefore, it may be beneficial to find a way to deliver directly glucose in the small intestine, where it can be absorbed and used for milk production (**Zhang et al., 2019**) through feeding rumen protected glucose.

Rumen protected glucose (RPG) is a good source of glucose for early lactation dairy cows to relieve negative energy balance (NEB), encapsulated by hydrogenated fat to escape rumen digestion and it can be released completely after entering the intestine (**Wang et al., 2020a**). More glucose is thus delivered to the small intestine then absorbed directly by the small intestine epithelium, rather than relying on the gluconeogenesis of the liver independently (**Zhang et al., 2019**). The form of glucose supplementation can block fat mobilization and gluconeogenesis in dairy cows in the NEB state (**Nichols et al., 2016**) and also large amounts of  $\beta$ -hydroxybutyric acid (BHBA), non-esterified fatty acid (NEFA), acetone and acetoacetic acid, etc. were no longer produced in the liver. Meanwhile, the original high concentration of these substances in the blood are also rapidly metabolized in the body and the fat accumulated in the liver would be transported out of the liver in the form of lipoproteins in time, or be oxidized in liver cells, and the body gradually deposits protein and fat (**Omphalius et al., 2020**). The life-weight therefore returns to normal, so as to achieve the purpose of alleviating negative energy balance (NEB) (**Li et al., 2019**). In addition, due to the blocking of gluconeogenesis, the blood ammonia production is reduced, which no longer affects the reproduction of dairy cows, and the conception rate during estrus increases (**Wang et al.,**

2020a). Studies had found that RPG can increase the milk production (Li *et al.*, 2019), relieve the inflammatory response (Wang *et al.*, 2020b), participate in ileal epithelium metabolism and regulate genes related to immune homeostasis (Zhang *et al.*, 2019) in postpartum cows. The present study was performed to investigate whether supplementation with RPG could improve metabolic status, milk production and reduce infection during the transition period in cows.

## MATERIAL AND METHODS

### Animals:

This study was conducted in lactating dairy farm in Kilo 59 Alexandria, Cairo desert, all cows were housed in free stalls, fed twice daily with a total mixed ration meeting the requirement for milk productions with ad-Librium access to water. Fifty five healthy Holstein dairy cows (4-5 years old;  $515 \pm 42$  kg body weight; 32.1 kg milk production per day) with the second lactation were enrolled in the study. Cows were randomly assigned into two groups: group (1) (n=25) control group which fed on control diet, group (2) (n=30) RPG group which fed on control diet supplemented with RPG at rate of 250g/cow/day (Gluco-Go-60) as follow in Table (1). The RPG used in this study adopted the latest international rumen evolving technology, and the rumen protection level of the product was 54.03 % (Wang *et al.*, 2020c). It was prepared with 45 % glucose as the core material, 45 % hydrogenated fat by mass as the coating, and 10 % water to make it (Li *et al.*, 2019). The RPG addition is granular and the diameters of the particles were 0.6-0.85 mm. All cows were fed from (20day before parturition to 60 day postpartum).

**Table (1):** Ingredients and nutrient composition of the diets.

<b>Ingredient ( % of DM)</b>	<b>20 day before parturition</b>	<b>60 day Postpartum</b>
<b>Corn</b>	<b>2.200</b>	<b>6.200</b>
<b>Soybean meal (46%)</b>	<b>2.200</b>	<b>6.00</b>
<b>Max pro.</b>	<b>0.500</b>	<b>2.00</b>
<b>Fat</b>	<b>0.050</b>	<b>0.200</b>
<b>Easy buffer</b>	<b>0.00</b>	<b>0.180</b>
<b>Magnesium oxide</b>	<b>0.00</b>	<b>0.040</b>
<b>Selenium</b>	<b>0.002</b>	<b>0.003</b>
<b>Cu, Zn, Mn, Co.</b>	<b>0.004</b>	<b>0.006</b>
<b>Biological antitoxic</b>	<b>0.012</b>	<b>0.005</b>
<b>Chemical antitoxic</b>	<b>0.006</b>	<b>0.040</b>
<b>Common salt</b>	<b>0.040</b>	<b>0.040</b>
<b>Mineral mix</b>	<b>0.030</b>	<b>0.050</b>
<b>Vitamin</b>	<b>0.015</b>	<b>0.025</b>
<b>Gluco-Go-60</b>	<b>0.250</b>	<b>0.250</b>
<b>Corn silage 30%</b>	<b>22.500</b>	<b>40.00</b>

**Nutrient composition of the diet:**

<b>Chemical analysis, (% of DM)</b>	<b>20 day before parturition</b>	<b>60 day postpartum</b>
<b>CP</b>	<b>11.6</b>	<b>14.6</b>
<b>Fat</b>	<b>2.0</b>	<b>2.1</b>
<b>Starch</b>	<b>10.3</b>	<b>14.8</b>
<b>NDF</b>	<b>53.6</b>	<b>45.2</b>
<b>ADF</b>	<b>31.3</b>	<b>25.5</b>
<b>Ash</b>	<b>6.8</b>	<b>6.0</b>
<b>NE<sub>L</sub><sup>2</sup> Mcal/kg</b>	<b>1.30</b>	<b>1.37</b>

**Blood Sampling:**

Blood samples were collected from the jugular vein of each cow on the +twenty-first day postpartum. Blood samples were centrifuged at 3000 rpm for 15 min to obtain serum; then it was stored at (-20 °C) until analysis.

**Blood Biochemical Parameters:**

Measurement of serum glucose level according to **Trinder (1969)**,  $\beta$ -hydroxybutyric acid ( $\beta$ -BHBA) quantitative UV assay on serum with BS-300 analyzer according to **Young (2000)**, NEFA-HR (2) ACS-ACOD method for the quantitative determination of non-esterified fatty acids (NEFA) in serum according to **Rogiers (1978)**. The measurements were performed on a spectrophotometer type SPV-72, Federal Republic of Germany. The enzymatic activity of aspartate aminotransferase (AST) was carried out according to **Reitman and Frankel (1957)**.

**Some Hormonal Parameters:**

The insulin (INS) levels were measured using a commercial bovine ELISA kit (Moonblind Inc. Lakeforest, CA92630.USA) according to **Gerich (1988)**.

**Statistical Analysis:**

Data were presented as mean  $\pm$  SEM (standard error of the mean). Independent-samples T test was carried out for the obtained data using SPSS program version 16.0 and  $P \leq 0.05$  was considered as statistically significant.

**RESULTS**

Serum glucose level was insignificantly different between RPG supplemented cows and control cows. On the contrary, a decrease ( $P \leq 0.05$ ) was observed in NEFA, BHBA and AST activity in response to RPG supplementation. Dietary RPG supplementation significantly increases plasma concentrations of insulin ( $P \leq 0.05$ ) relative to control cows.

**Table (2):** Effect of rumen protected glucose supplementation (RPG) on some serum biochemical parameters of dairy cows during the transition period.

Items	Control group	RPG supplemented
Glucose mg/dl	63.24±2.32 <sup>a</sup>	66.27±3.34 <sup>a</sup>
NEFA mmol/L	1.83±0.01 <sup>a</sup>	0.95±0.02 <sup>b</sup>
B-HBA mmol/L	0.36±0.01 <sup>a</sup>	0.18±0.00 <sup>b</sup>
AST U/L	88.20±5.27 <sup>a</sup>	70.50±6.19 <sup>b</sup>
Insulin ng/ml	1.58±0.03 <sup>b</sup>	2.42±0.01 <sup>a</sup>

RPG= Rumen protected glucose, NEFA=non-esterified fatty acid, β-HBA=Beta-hydroxybutyric acid; Means within a row with different superscript letters differ at ( $P \leq 0.05$ ).

Dietary supplementation with RPG in the transition period showed significant increase in milk yield in supplemented group than in control dairy cows. Health disorders during transition period in RPG group were less than in control group which include retained fetal membranes and metritis.

**Table (3):** Effect of rumen-protected glucose supplementation (RPG) on some reproductive performance in dairy cows during the transition period.

Parameters	Control group	Supplemented group
First heat	38	36
Ins.no	4.5	4
Prev(-1) open day	121	123
open days	118	106
Prev (-1) lact. Yield	10145	10180
Lact. Avg	33.7	34.6
Preg %	88%	96%
R.F.M	4	2
Metritis	4	1

Ins.no: insemination number; prey (1) open day: previous open day; prey (1) lact. Yiel: previous lactation yield; preg: pregnancy; R.F.M.=retained fetal membrane; Difference between control and RPG supplementation in dairy cows.

274.5kg milk in lactation
1921.5 milk price
12 days in open days

## DISCUSSION

Glucose is a key nutrient for the maintenance of basic functions of body tissues and the synthesis of milk during lactation. Low blood glucose concentrations after calving are associated with infertility in postpartum dairy cows perhaps because glucose is a master regulator of hormones and metabolites that control reproductive processes (**Lucy et al., 2014**). Glucose metabolism imbalance exacerbates gluconeogenesis and lipid mobilization, which not only hinders milk quality and milk yield, but also limits the health and reproductive function in dairy cows (**Sauls-Hiesterman et al., 2018**). Concentrations of GLU,  $\beta$ -HBA, NEFA, and activity of AST are important indicators for evaluating the energy balance of cows (**Song et al., 2019**).

The serum glucose concentration is the most important factor affecting the secretion of insulin. In the present study, there were no difference in blood glucose concentration between RPG supplemented group and the control group, and these results agree with **Sauls-Hiesterman et al., 2018; Zhang et al., 2019 and McCarthy et al., 2019; 2020**. The increased serum glucose concentration after RPG supplementation was consistent with previous studies (**Brown and Allen, 2013 and Wang et al., 2020a**). The increased glucose concentration in RPG supplemented group may be related to the dose of 0, 200, 350 and 500 g of RPG supplementation and its own metabolic adaptation (**Wang et al., 2020a**). Meanwhile, **Li et al., (2019)** found that, dietary RPG supplementation tended to decrease the plasma glucose concentration, lesser blood glucose may affect the reproduction through ovarian activity which restored via glucose itself or glucose-mediated actions on circulating concentrations of insulin, IGF1, and / or lipid metabolites (**Green et al., 2012 and Garverick et al., 2013**). In the current study, supplementation with dietary RPG increased the concentration of insulin in the blood and this result agrees with previous work **Wagner and Schimek, 2010 and McCarthy et al., 2020**. Meanwhile, **Sauls-Hiesterman et al., (2018)** said that, changes in insulin concentration were greater for the control cows compared with the cows received the dose of RPG (2.2, 4.4, 8.8 lb /head/day). Insulin decreases circulating NEFA concentration through its capacity to stimulate lipogenesis and inhibit lipolysis in adipose tissue (**Hayirli, 2006**). Insulin also has anti-ketogenic properties because it increases peripheral ketone utilization and decreases hepatic ketone production (**Hayirli, 2006**). The concentrations of  $\beta$ -HBA and NEFA were reduced after glucose supplementation. This is similar to a previous

report (McCarthy *et al.*, 2020 and Wang *et al.*, 2020a). On the contrary, Li *et al.*, (2019) found that plasma NEFA concentration increased in RPG supplemented group, this result agreed with supplement amount of RPG at 200 g/d may not be sufficient for increased milk production and therefore more body fat will be mobilized to meet the energy requirements for milk production.

AST activity in RPG supplemented group was significantly decreased than in control group. This is similar to a previous report (Wang *et al.*, 2020a). Meanwhile, Li *et al.*, 2019 found that no significant differences were observed in the plasma activity of AST between the RPG supplemented group and control group of cows during the transition period.

Dietary supplementation with RPG in the transition period showed significant increase in milk yield in supplemented group than in control dairy cows. Some studies found that duodenal infusion of glucose could increase milk yield and milk fat yield (Li *et al.*, 2019 and Zhang *et al.*, 2019). In contrast, some other studies showed that infusion of glucose through abdomen or rumen-protected did not affect milk yield (Larsen and Kristensen, 2009 and Sauls-Hiesterman *et al.*, 2018), but reduced milk fat yield (Larsen and Kristensen 2009). The opposite results may be due to the supplementary source, dosage and form of glucose, and the parity, basal diet composition, and physiological state of test animals.

Health disorders during transition period in RPG supplemented group were less than in control groups which include retained fetal membranes (4%, after a normal calving) and metritis. These result are similar to (Hooshman Dabbasi *et al.*, 2018) retained fetal membrane, metritis (Santos *et al.*, 2015), these disorders have adverse effects on animal welfare, milk production, reproduction, and farm profitability (Neves *et al.*, 2018).

## CONCLUSION

The key findings of the study were that supplementation with RPG in transition cows improved the postpartum lactation performance (increased milk yield), decreased the degree of negative energy balance (NEB) since it decreased NEFA,  $\beta$ -HBA and increased insulin concentrations, and presumably reduced the incidence of reproductive disorders.

## REFERENCE

- Brown, W.E. and Allen, M. S. (2013):** Effects of intrajugular glucose infusion on feed intake, milk yield, and metabolic responses of early postpartum cows fed diets varying in protein and starch concentration. *J. Dairy Sci.*, 96: 7132–7142.
- Colman, E., Khafipour, B., Vlaeminck, B., De-Baets, J. and Plaizier, V. (2013):** Grain-based versus alfalfa-based subacute ruminal acidosis induction experiments: similarities and differences between changes in milk fatty acids. *J. Dairy Sci.*, 96: 4100 - 4111.
- Curtis, R.V., Kim, J.J.M., Doelman, J. and Cant, J.P. (2018):** Maintenance of plasma branched-chain amino acid concentrations during glucose infusion directs essential amino acids to extra-mammary tissues in lactating dairy cows. *J. Dairy Sci.*, 101: 4542-4553.
- Garverick, H.A., Harris, M.N., Vogel-Bluel, R., Sampson, J.D., Bader, J., Lamberson, W.R., Spain, J.N., Lucy, M.C. and Young Quist, R.S. (2013):** Concentrations of nonesterified fatty acids and glucose in blood of periparturient dairy cows are indicative of pregnancy success at first insemination. *J. Dairy Sci.*, 96: 181-188.
- Gerich, J.E. (1988):** Hormonal control of hemostasis In Goolowoy, J.A., Potvin, j.H., Shuman, C.R. (eds): *Diabetus Mellitus*, ninth edition. Eli-Lilly Co., Indiana Polis, 46-63.
- Green, J.C., Meyer, J.P., Williams, A.M., Newsom, E.M., Keisler, D.H. and Lucy M.C. (2012):** Pregnancy development from day 28 to 42 of gestation in postpartum Holstein cows that were either milked (lactating) or not milked (not lactating) after calving. *Reproduction*, 143: 699-711.
- Gualdrón-Duarte, L.B. and Allen, M.S., (2018):** Fuels derived from starch digestion have different effects on energy intake and metabolic responses of cows in the postpartum period. *J. Dairy Sci.*, 101: 5082–5091.
- Hayirli, A. (2006):** The role of exogenous insulin in the complex of hepatic lipidosis and ketosis associated with insulin resistance phenomenon in postpartum dairy cattle. *Vet. Res. Commun.*, 30: 749-774.
- Hooshmandabbasi, R., Zerbe, H., Bauersachs, S., de Sousa, N.M., Boos, A. and Klisch, K. (2018):** Pregnancy-associated glycoproteins in cows with retained fetal membranes. *Theriogenology*, 105:158–63.
- Kvidera, S.K., Horst, E.A., Abuajamieh, M., Mayorga, E.J., Fernandez, M.V. and Baumgard, L.H. (2017):** Glucose requirements of an activated immune system in lactating Holstein cows. *J. Dairy Sci.*, 100: 2360 -2374.
- Larsen, M. and Kristensen, N.B. (2009):** Effect of abomasal glucose infusion on splanchnic and whole-body glucose metabolism in periparturient dairy cows. *J. Dairy Sci.*, 92: 1071-1083.

- Li, X.P., Tan, Z.L., Jiao, J.Z., Long, D.L., Zhou, C.S., Yi, K.L., Liu, C.H., Kang, J.H., Wang, M. and Duan, F.H. (2019):** Supplementation with fat-coated rumen-protected glucose during the transition period enhances milk production and influences blood biochemical parameters of liver function and inflammation in dairy cows. *Anim. Feed Sci. Technol.*, 252: 92–102.
- Lucy, M.C., Butler, S.T. and Garverick, H.A. (2014):** Endocrine and metabolic mechanisms linking postpartum glucose with early embryonic and foetal development in dairy cows. *Animal*, 8: 82-90.
- Mayorga, E. J., Al-Qaisi, M., Abeyta, M.A., Perez-Hernandez, G., B. M. Goetz, B.M. and Castillo, A. R. (2019):** Effect of feeding rumen-protected glucose on lactation performance, energetic metabolism and inflammation in transitioning dairy cows. *Animals Sci.*, 253-272.
- McCarthy, C.S., Dooley B.C., Ramstad E.H., Appuharmy J.A., Ramirez H.A. and Baum Gard L.H. (2020):** Energetic metabolism, milk production, and inflammatory response of transition dairy cows fed rumen-protected glucose. *J. Dairy Sci.*, 103: 7451-7461.
- McCarthy, B. C., Dooley, E. H., Branstad, A.J., Kramer, E. A., Horst, E. J., Mayorga, M., Al-Qaisi, M. A., Abeyta, G., Perez-Hernandez, B. M., Goetz, A. R., Castillo, M. R., Knobbe, C. A., Macgregor, J. P., Russi, J. A., Appuhamy, H. A. and Ramirez-Ramirez, L. H. (2019):** Effects of feeding rumen protected glucose on lactation performance, energetic metabolism and inflammation in transitioning dairy cows. Iowa state university, 1-65.
- Moran, A.W., Al-Rammahi, M., Zhang, C., Bravo, D., Calsamiglia, S. and Shirazi-Beechey, S.P. (2014):** Sweet taste receptor expression in ruminant intestine and its activation by artificial sweeteners to regulate glucose absorption. *J. Dairy Sci.*, 97: 4955-4972.
- Neves, R.C., Leno, B.M., Bach, K.D. and Mcart J.A.A. (2018):** Epidemiology of subclinical hypocalcemia in early-lactation Holstein dairy cows: The temporal associations of plasma calcium concentration in the first 4 days in milk with disease and milk production. *J Dairy Sci.*:101:9321–31.
- Nichols, K., Kim, J.J.M., Carson, M., Metcalf, J.A., Cant, J.P. and Doelman, J. (2016):** Glucose supplementation stimulates peripheral branched-chain amino acid catabolism in lactating dairy cows during essential amino acid infusions *J. Dairy Sci.*, 99: 1145-1160.
- Omphalius, C., Lemosquet, S., Ouellet, D.R., Bahloul, L. and Lapierre, H. (2020):** Postruminal infusions of amino acids or glucose affect metabolisms of splanchnic, mammary, and other peripheral tissues and drive amino acid use in dairy cows. *J. Dairy Sci.*, 103: 2233-2254.
- Reitman, S. and Frankel, S. (1957):** A colorimetric method for the determination of serum glutamic oxalacetic and glutamic pyruvic transaminases. *American journal of clinical pathology*. 28 (1):56-63.

- Rogiers, V. (1978):** Stability of the long chain non- esterified fatty acid pattern in plasma and blood during different storage conditions. Clin. Chem. Acta., 84: 49-54.
- Santos, J., Pinedo, P., Schuenemann, G.M., Bicalho, R.C., Chebel, R.C., Seabury, C., Fetrow, J. and Thatcher, W.W. (2015):** Improving fertility through genomic selection. Proc. Fairy Cattle Repro. Caincil, NY, DCRC, Champaign, IL, 3-9.
- Sauls-Hiesterman, J.A., Banuelos, S., Attanasov, B. and Bradford, B. (2018):** Physiologic responses to feeding rumen-protected glucose to lactating dairy cows. Kansas Agricultural Experiment Station Research Reports, 24-40.
- Song, Y., Bai, W., Qian, C., Xia, H., Zhang, C. and Xu E. (2019):** Establishment of early warning indicators for the risk of negative energy balance in beef cattle. Prog. Vet. Med., 40, 133-136.
- Trinder P. (1969):** Determination of glucose using glucose oxidase with an alternative oxygen acceptor. Ann. Clin. Biochem, 6, 24.
- Wagner, S.A. and Schimek, D.E. (2010):** Evaluation of the effect of bolus administration of 50% dextrose solution on measures of electrolyte and energy balance in postpartum dairy cows. Am. J. Vet. Res.; 71: 1074-1080.
- Wang, Y.P., CAI, M., Hua, D.K., Zhang, F., Jiang, L.S., Zhao, Y.G., Wang, H., Nan, X.M.N. and Xionga, B.H. (2020a):** Metabolomics reveals effects of rumen-protected glucose on metabolism of dairy cows in early lactation. Animal feed science and technology, 269: 235-141.
- Wang, Y.P., Han, X., Tan, Z., Kang, J. and Wang, Z.(2020b):** Rumen-protected glucose stimulates the insulin-like growth factor system and mTOR/AKT pathway in the endometrium of early postpartum dairy cows. Animals, 10: 357-371.
- Wang, Y.P., Zhang, F., Hua, D.,Jiang, L.and Xiong,B.(2020c):** Rumen stability of rumen-protected glucose and its effects of different doses on rumen degradation characteristics of oat hay Chin. J. Anim. Nutr., 32: 3428-3438.
- Young, D.S. (2000):** Effect of drug on Clinical Lab. Test, 5<sup>th</sup> Ed. AACC Press.
- Zhang, X., Wu, J., Han, X., Tan, Z. and Jiao, J. (2019):** Effects of rumen-protected glucose on ileal microbiota and genes involved in ileal epithelial metabolism and immune homeostasis in transition dairy cows. Anim. Feed Sci. Tech., 254: 0377-8401.