

(Print) (ISSN 2537-0286)

(Online) (ISSN 2537-0294) https://nrmj.journals.ekb.eg/ DOI: 10.21608/NRMJ.2018.8152

Novel Research in Microbiology Journal (2018), 2(3): 48-52 *Editorial*

Efficacy of Pseudomonads as biocontrol agents of phytopathogens

Adel K. Madbouly*

Microbiology Department, Faculty of Science, University of Ain Shams, Cairo, Egypt

*Author E-mail: <u>adelkamelmadbouly@yahoo.com</u>

Received: 4 May, 2018; Accepted: 6 June, 2018; Published online: 24 June, 2018



Introduction

Microbial plant pathogens infecting different crops cause great economic losses worldwide. Thus plant diseases need to be controlled to maintain the quality and abundance of these crops. Different approaches have been used to control plant diseases, mainly chemical fertilizers and pesticides. However, environmental pollution and pathogens induced resistance due to excessive use of these agrochemicals; exclude their may successful application. Consequently, researchers have focused on developing alternatives to control plant diseases; among these alternatives is biological control. The term biological control or biocontrol applies to the use of living antagonists to suppress or kill deleterious plant pathogens. The saprophytic microbe which antagonises the pathogen is called the biological control agent (BCA) or bioagent.

Microbe-mediated biological control is considered safe as being close to the natural ecosystems (Coleman-Derr and Tringe, 2014). There are two general ways of applying biocontrol agents against phytopathogens; through addition of large amounts of bioagents to the soil or stimulating the activities of the existing bioagents using various amendments (Conrath, 2011). Biocontrol mechanisms involve production of antibiotics, volatile organic compounds (VOC), hydrolytic enzymes, competition for space, nutrients and induction of systemic resistance in plants (Faheem *et al.*, 2015; Yunus *et al.*, 2016). Senthilraja *et al.*, (2010) pointed that combination of more than one biocontrol agent which occupy different sites and exert different promoting effects on their host plants is more advantageous than their individual application.

Pseudomonas spp. are soil-borne, Gram-negative bacteria which possess characteristics that make them advantageous for plant disease biocontrol (Palleroni, 2008). Several species of these bacteria produce antibiotics which suppress phytopathogens growth (Yang and Cao, 2012). Moreover, Duke *et al.*, (2017) added that surface colonization by *Pseudomonas* spp. is accompanied with induction of host plant defence.

Fluorescent Pseudomonads belong to Plant Growth Promoting Rhizobacteria (PGPR), which play major role in the plant growth promotion, induced systemic resistance and biocontrol of several phytopathogens etc. These effects are attributed to the production of different secondary metabolites including; antibiotics, siderophores and hydrogen cvanide (HCN) (O'Sullivan and O'Gara, 1992). Siderophores are low molecular weight chelating agents of ferric ions produced by bacteria and fungi under conditions of low iron stress (Ngamau et al., 2014). According to Decheng et al., (2005) most of the plants are able to use bacterial iron siderophore complexes from the soil as a competitive advantage under iron stress, thus restrict proliferation and colonization of root by different phytopathogens. Siderophore-based bioagents have commercial importance as they are safer and do not lead to biomagnification, in addition, they also provide iron nutrition to the plants thereby promoting their growth (Sayyed et al., 2007).

Among various biocontrol agents, *P. fluorescens* strains inhabit different environments such as; plant, soil and water surfaces and have been used successfully to control different soil-borne pathogens (Ganeshani and Kumar, 2005). *P. fluorescens* BE8 showed 55% control of *Fusarium oxysporum* f. sp. *cucumerinum* in an *in vivo* pot experiment (Szentes *et al.*, 2013). Another strain of *P. fluorescens* inhibited *Xanthomonas oryzae* pv. *oryzae*, the bacterial causal agent of leaf blight in rice (Lingaiah and Umesha, 2013).

The study of Bahrouna et al., (2018)demonstrated that bacteria with high biocontrol potency such as; Rahnella aquatilis B16C, P. yamanorum B12 and P. fluorescens B8P produce siderophores, chlorinated phenylpyrrole antibiotics (PRN) and a volatile compound. These are involved in biocontrol potential against many soil-borne fungal pathogens (de Souza and Raaijmakers, 2003). However, Weidner et al., (2017) reported that these three bacterial strains did not produce auxins, as they showed no plant growth promoting (PGP) activities in non-infested soil. This is attributable to the fact that biocontrol potency is not always associated with PGP activities, accordingly it's quite difficult to detect a strain of bacteria which encompasses both traits simultaneously.

Volatile compounds (VOCs) produced by soil microbes promoted plant growth, showed high antimicrobial and nematicidal activities, and induced systemic resistance in several crops (Audrain et al., 2015). The genus Pseudomonas is well known of producing large arrays of antifungal and plant growthpromoting metabolites, such as VOCs (Yan et al., 2017). In the study of Raza et al., (2016), VOCs produced by a biocontrol strain P. fluorescens WR-1 inhibited the growth and virulence characters of Ralstonia solanacearum; the bacterial causal agent of tomato wilt. VOCs produced by this strain showed more growth inhibition of *R. solanacearum* in natural soil than in sterilized soil. This is attributed to the contribution of soil indigenous microbes which also produced antimicrobial VOCs (Hernández-León et al., 2015). The VOCs produced by P. fluorescens B-4117 inhibited the growth of several Agrobacteria such as; A. tumefaciens and A. vitis (Dandurishvili et al., 2011). Moreover, P. fluorescens strains UM16, UM240, UM256 and UM270 produced VOCs which inhibited the growth of phytopathogenic Botrytis cinerea (Hernández-León et al., 2015).

Recently, Singha et al., (2016) revealed the impact of bio-protective microbes isolated from rhizosphere soil on plant growth promotion and triggering defence responses in rice plant when inoculated with Rhizoctonia solani. Rice plants cowith Ρ. fluorescens **PF-08** inoculated and Trichoderma harzianum UBSTH-501 strains demonstrated significant increase in the shoot and root biomass, and defence related biomolecules. Direct promotion mechanisms of these bioagents were associated with the reduction in pathogen population in the rhizosphere through production of antimicrobial metabolites such as antibiotics (Harman et al., 2004), mycoparasitism, competition for space and nutrients (Wang et al., 2013). However, indirect promotion effects were attributable to the enhancement in population of different beneficial soil microbes, and triggering defence genes inside the plant leading to suppression of growth and reproduction of phytopathogens.

Bacteriocins are ribosomally encoded proteins which expressed high specificity by killing bacteria closely related to the producer strain (Ahmad et al., 2016). P. fluorescens strain SF4c isolated from wheat rhizosphere soil produced a bacteriocin similar to phage tail-like pyocins of P. aeruginosa. This tailocin antimicrobial activity has against manv phytopathogenic strains of Xanthomonads and Pseudomonads, without affecting the microbial community of the plant growth promoting rhizobacteria (PGPRs) such as P. protegens CHA0. Screening against number of phytopathogenic strains revealed that SF4c tailocin has inhibitory activity against X. axonopodis pv. vesicatoria (X. cv Bv5-4a). Therefore, this SF4c tailocin could be used as a biocontrol agent of bacterial leaf spot disease. The Atomic Force Microscopy (AFM) demonstrated that this tailocin adheres and causes damage to the cell envelope of X. cv Bv5-4a. This caused rapid leakage of intracellular materials with subsequent decrease of bacterial cell volume, leading finally to lysis of these cells (Fernandeza et al., 2017).

In saprophytic conclusion. Pseudomonads especially fluorescent ones are highly effective bioagents for control of most plant pathogenic microbes. They have multiple modes of action, are normal inhabitants of roots surfaces (rhizospheres) and leaves surfaces (phylloplanes), highly adapted to live as saprophytes and endophytes within plants. However, their potentials as bioagents could be enhanced by inoculating them into plants in association with suitable amendments, thus providing nutrients necessary for their growth, proliferation and colonization of plants surfaces, hence become more advantageous over deleterious phytopathogens. Moreover, it is better to apply more than one species of these Pseudomonads simultaneously to have several modes of action thus obtaining better biocontrol potency.

References

Ahmad, V.; Khan, M.S.; Jamal, Q.M.S.; Alzohairy, M.A.; Al Karaawi, M.A. and Siddiqui, M.U. (2016). Antimicrobial potential of bacteriocins: in therapy, agriculture and food preservation. International Journal of Antimicrobial Agents. 49(1): 1-11.

Audrain, B.; Frag, M.A.; Ryu, C.M. and Ghigo, J.M. (2015). Role of bacterial volatile compounds in bacterial biology. FEMS Microbiology Reviews. 39: 222-233.

Bahrouna, A.; Joussetb, A.; Mhamdia, R.; Mrabeta, M. and Mhadhbi, H. (2018). Anti-fungal activity of bacterial endophytes associated with legumes against *Fusarium solani*: Assessment of fungi soil suppressiveness and plant protection induction. Applied Soil Ecology. 124:131-140.

Coleman-Derr, D. and Tringe, S.G. (2014). Building the crops of tomorrow: advantages of symbiont-based approaches to improving abiotic stress tolerance. Frontiers in Microbiology. 5: 283.

Conrath, U. (2011). Molecular aspects of defence priming. Trends in Plant Science. 16: 523-524.

Dandurishvili, N.; Toklikishvili, N.; Ovadis, M.; Eliashvili, P.; Giorgobiani, N.; Keshelava, R.; Tediashvili, M.; Vainstein, A.; Khmel, I.; Szegedi, E. and Chernin, L. (2011). Broad-range antagonistic rhizobacteria *Pseudomonas fluorescens* and *Serratia plymuthica* suppress *Agrobacterium* crown gall tumours on tomato plants. Journal of Applied Microbiology. 110: 41-352.

Decheng, R.; Zuo, R. and Wood, T.K. (2005). Quorum-sensing antagonist (5Z)-4-bromo-5-(bromomethylene) 3-butyl-2(5H)-furanone influences siderophore biosynthesis in *Pseudomonas putida* and *Pseudomonas aeruginosa*. Applied Microbiology and Biotechnology. 66: 689-695. **de Souza, J.T. and Raaijmakers, J.M. (2003).** Polymorphisms within the prnD and pltC genes from pyrrolnitrin and pyoluteorin-producing *Pseudomonas* and *Burkholderia* spp. FEMS Microbiology Ecology. 43: 21-34.

Duke, K.A.; Becker, M.G.; Girard, I.J.; Millar, J.L.; Dilantha Fernando, W.G.; Belmonte, M.F. and de Kievit, T.R. (2017). The biocontrol agent *Pseudomonas chlororaphis* PA23 primes *Brassica napus* defenses through distinct gene networks. BMC Genomics. 18: 467.

Faheem, M.; Raza, W.; Jun, Z.; Shabbir, S. and Sultana, N. (2015). Characterization of the newly isolated antimicrobial strain *Streptomyces goshikiensis* YCXU. Science Letters. 3: 94-97.

Fernandeza, M.; Godinoa, A.; Príncipea, A.; Moralesb, G.M. and Fischer, S. (2017). Effect of a *Pseudomonas fluorescens* tailocin against phytopathogenic *Xanthomonas* observed by atomic force microscopy. Journal of Biotechnology. 256: 13-20.

Ganeshani, G. and Kumar, A.M. (2005). *Pseudomonas fluorescens*, a potential bacterial antagonist to control plant diseases. International Journal of Plant Sciences. 1: 123-134.

Harman, G.E.; Howell, C.R.; Viterbo, A.; Chet, I. and Lorito, M. (2004). *Trichoderma* speciesopportunistic, avirulent plant symbionts. Nature Reviews Microbiology. 2: 43-56.

Hernández-León, R.; Rojas-Solís, D.; Contreras-Pérez, M.; Orozco-Mosqueda, M.C.; Macías-Rodríguez, L.I.; Reyes-de la Cruz, H.; Valencia-Cantero, E. and Santoyo, G. (2015). Characterization of the antifungal and plant growthpromoting effects of diffusible and volatile organic compounds produced by *Pseudomonas fluorescens* strains. Biological Control. 81: 83-92. Lingaiah, S. and Umesha, S. (2013). *Pseudomonas fluorescens* inhibits the *Xanthomonas oryzae* pv. *oryzae*: the bacterial leaf blight pathogen in rice. Candian Journal of Plant Protection. 1: 147-153.

Ngamau, C.N.; Matiru, V.N.; Tani, A. and Muthuri, C.W. (2014). Potential use of endophytic bacteria as biofertilizer for sustainable banana (*Musa* spp.) production. African Journal of Horticultural Science. 8: 1-11.

O'Sullivan, D.B. and O'Gara, F. (1992). Traits of fluorescent *Pseudomonas* spp. involved in suppression of plant root pathogens. Microbiological Reviews. 56: 662-676.

Palleroni, N.J. (2008). The road to the taxonomy of *Pseudomonas*. In: Cornelis, P. (Ed.), *Pseudomonas*. Genomics and Molecular Biology. Caister Academic, Norfolk, pp. 1-18.

Raza, W.; Ling, N.; Liu, D.; Wei, Z.; Huang, Q. and Shen, Q. (2016). Volatile organic compounds produced by *Pseudomonas fluorescens* WR-1 restrict the growth and virulence traits of *Ralstonia solanacearum*. Microbiological Research. 192: 103-113.

Sayyed, R.Z.; Naphade, B.S. and Chincholklar, S.B. (2007). Siderophore producing *A. feacalis* promoted the growth of Safed musali and Ashwagandha. Journal of Applied Research on Medicinal and Aromatic Plants. 29: 1-5.

Senthilraja, G.; Anand, T.; Durairaj, C.; Kennedy, Suresh, **S.**; Raguchander, J.S.; T. and Samiyappan, R. (2010). A new microbial consortia containing entomopathogenic fungus, Beauveria bassiana and plant growth promoting rhizobacteria, Pseudomonas fluorescens for simultaneous management of leaf miners and collar rot disease in groundnut. Biocontrol Science and Technology. 20: 449-464.

Singha, U.B.; Malviyaa, D.; Wasiullaha; Singha, S.; Pradhana, J.K.; Singh, B.P.; Roya, M.; Imram, M.; Pathak, N.; Baisyal, B.M.; Raie, J.P.; Sarma, B.K.; Singh, R.K.; Sharma, P.K.; Kaur, S.D.; Manna, M.C.; Sharma, S.K. and Sharma, A.K. (2016). Bio-protective microbial agents from rhizosphere eco-systems trigger plant defense responses provide protection against sheath blight disease in rice (*Oryza sativa* L.). Microbiological Research. 192: 300-312.

Szentes, S.; Gabriel-Lucian, R.; Laslo, É.; Lányi, S. and Mara, G. (2013). Selection and evaluation of potential biocontrol rhizobacteria from a raised bog environment. Crop Protection. 52: 116-124.

Wang, B.; Yuan, J.; Zhang, J.; Shen, Z.; Zhang, M. and Li, R., *et al.*, (2013). Effect of novel bioorganicfertilizer produced by *Bacillus amyloliquefaciens* W19 on antagonism of *Fusarium* wilt of banana. Biology and Fertility of Soils. 249; 435-446. Weidner, S.; Latz, E.; Agaras, B.; Valverde, C. and Jousset, A. (2017). Protozoa stimulate the plant beneficial activity of rhizospheric pseudomonads. Plant Soil. 410: 509-515.

Yan, Q.; Philmus, B.; Chang, J.H. and Loper, J.E. (2017). Novel mechanism of metabolic co-regulation coordinates the biosynthesis of secondary metabolites in *Pseudomonas protegens*. eLife. 6: e22835.

Yang, F. and Cao, Y.J. (2012). Biosynthesis of phloroglucinol compounds in microorganisms. Applied Microbiology and Biotechnology. 93: 487-495.

Yunus, F.; Iqbal, M.; Jabeen, K.; Kanwal, Z. and Rashid, F. (2016). Antagonistic activity of *Pseudomonas fluorescens* against fungal plant pathogen *Aspergillus niger*. Science Letters. 4: 66-70.