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# ISOLATION AND IDENTIFICATION OF LUMPY SKIN DISEASE VIRUS FROM UPPER EGYPT

(With 1 Table and 8 Figures)

By

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# عزل وتصنيف الفيروس المسبب لمرض التماب الجلد العقدى في مصــر العليــــا

طه العللوي ، مخدار الطرابيلي ، مراط اسماعيل ، صحيف رشوان

لقد تم عزل ٢٠ عترة من فيروس التهاب الجلد العقدى من بين ثلاث أماكن مختلفة من مصر العليا وهي : نجع حمادى، الوادى الجديد ، ( موشا والبدارى ) محافظة أسيوط ، وقد تم التعرف عليها بتأثيرها على الغشاء السلي السجقي وعلى خلايا خصية الأغنام وخلايا كلية المواشى ، وتم التعرف عليها أيضا باستخدام الاختبارات السيرولوجيه مثل اختبار الاليزا واختبار المصل التعادلي.

## SUMMARY

Twenty local isolates had been obtained from skin nodules and skin scapes from 3 different localities Nag-Hamadi, New Vally and Mousha, El-Badary (Assiut in Upper Egypt). These isolates were identified by their cytopathogenic effect on CAM, MDBK and LT. As well as serologically by solid phase Elisa Technique and surm neutralization test.

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## INTRODUCTION

Lumpy skin disease (LSD) is a highly infectious disease of cattle caused by a member of pox virida and exsists in various areas in Africa (MAC-DONALD, 1931; VON BAKSTROM, 1945; HAIG, 1957; MaCOWEN, 1959; AWAD, 1966 and EL-KANAWATY, 1989).

VANDENENDE et al. (1949) used a filtrate from an emulsion of skin nodes and milk duct nodules from a dead calf by bovine LSD to isolate the agent by serial passage in chick embryo. The agent appears to be filterable virus approximately 10-25 mu in diameter.

ALEXANDER et al. (1957) in the union of South Africa isolated 3 cytopathic virus from skin nodule of cattle suffering from LSD virus type one was Orpha the second Allerton and the 3rd was the Neethling type virus. The last one was the true causative agent of LSD. They added that the true causative agent (Neethling type virus) can be propagated in cell culture of calf and lamb kidney, calf and lamb testis with clear CPE change and on chorioallantoic membrane of chicken embryo (9-11) days old with production of macroscopic pock lesion.

LOSOS (1986) proved that LSD virus can be propagated in a number of mammalian cell culture as tissue from lamb, calves, rabbits, hamster and embryonated chicken egg (9-11) day old and incubated at 33-35.5 C for 6 days.

EL-KANAWATY (1989) isolated LSD virus on CAM from skin nodules and internal organs obtained from 5 cattle in El-Tal-Kabeer and 2 cattle from El-Noberia dairy farm.

WOODS (1988) reported that isolated virus of LSD from infected cattle could be confirmed by SNT with specific antisera. VERSTEEG (1985) mentioned that the Elisa was a widened scope for detection of viruses and their antigen. This work was carried out as a trial to isolate and identify the causative virus of lumpy skin disease in cattle in Upper Egypt.

## MATERIAL and METHODS

1) Skin samples:

Skin nodules or scales were taken aseptically from 58 clinically infected cattle ageing between 4 months to 8 years for viral isolation. These

samples were homogenised then kept in tubes containing 4 ml phosphate buffer saline containing 100 Iu of pencillin, 100 ug of streptomycine and 250 Iu of mycostatin. These emulsions, either left at room temperature or incubated at 37 C for 1 hour, were centrifuged at 3.000 r.p.m. for 30 minutes in a cold centrifuge to remove any dehris. The collected supernatant fluid were stored in deep freeze at-70 C until used (ALI and OBIED, 1977).

2) Embryonated hens eggs:

9-11 day-old embryonated hens egg were obtained from agriculture college of Assiut University and examined by candling before used for chorioallantoic (CAM) inoculation to GRIST et al. (1977).

3) Tissue culture:

MDBK as well as primary and secondary (LT) were obtained as cell line from the Institute of Veterinary Serum and Vaccine Research and Production Abbasia. These cell lines were propagated in Eagles MEM suplemented with 10% horse serum or new born calf serum and used for virus isolation according to PLOWRIGHT and FERRIS (1958) and for SNT according to DARCEL (1975).

4) Reference sera:

Specific antisers for LSD virus were obtained from Faculty of Veterinary Medicine, Cairo University, Department of Microbiology.

5) Solid phase Eliza:

Indirect microplate Eliza according to KENDAL et al. (1983).

#### RESULTS

Table (1) indicates that twenty viral isolates were obtained from skin nodules and skin scales using CAM inoculation and then on MDBK and LT cell cultures, 15 isolates were from the aluminum farm, Nagh-Hamadi, 2 isolates from Mousha and EL-Badary (Assiut) as well as 3 isolates from New Valley.

Table (1) and Fig. (1,2) showed that most of the twenty isolates produce pin point pock lesion in the form of streks or strips. In some cases thickness and congestion of membrane without any macroscopic lesion were observed.

Fig. (3, 4 & 5) demonsterate the cytopathic effect of the twenty isolates on MDBK, cell culture, which start from the third day post inoculation untile complete destruction of the cell sheet on the eight day.

Fig. (6, 7 & 8) showed the CPE of these isolates on primary and secondary lamb testis (LT) cell culture which start from the third day post inoculation till detachment and complete destruction of cell sheet after about 6-8 days post inoculation.

The identification and confirmatory test to performed on the twenty local isolates using SNT and Elisa in which specific LSD antisera with a titre (1/64) was used. The results revealed that these twenty local isolates were positive LSD virus.

#### DISCUSSION

Lumpy skin disease is a highly infectious skin disease of cattle caused by a virus, characterized by fever, skin nodules all over the body, oedema especially in hind limb and abortion in some cases (BLOOD and RADOSTITS, 1989).

The results, as shown in table (1) indicated the isolation of LSD virus on CAM, MDBK cell culture and LT cells from skin nodules and skin scales obtained from infected animals from 3 different localities in Upper Egypt. The same methods of isolation were applied by (VANDENENDE, 1949; ALEXANDER et al., 1957; MADIN & DARBY; 1959 and ALI & OBEID, 1977); WAADS, 1988 and EL-KANAWATY, 1989). In addition PROZESKY and BARNARD (1982) and EL-KANAWATY isolated LSD virus from lymph node, internal organs as liver, lung and kidney.

Table (1), on the otherhand, demonstrated that identification of LSD virus in this investigation depends on the cytopathic effect of thevirus on CAM and tissue culture (MDBK and LT). These CPE were also used by VAN-ROOYEN et al. (1949), ALEXANDER et al. (1957); PLOWRIGHT and WITCOMB (1959); VAN-ROOYEN et al. (1969); ALI & OBEID (1977); NAWATHE et al. (1980); WOODS (1988) and EL-KANEWATY (1989). Table (1) revealed also, that the 20 local isolates of LSD were identified and confirmed by S.N.T. AND Elisa using specific LSD antisera. These tests were previously used by (MARTIN & MAUREEN, 1968; VAN-ROOYEN et al., 1965; ALI 7 OBEICL, 1977; HEDGER 7 HAMBLIN, 1983; WOODS, 1988; EL-KANAWATY, 1989; CHO & BAHAC, 1985; LITTLE et al., 1985 and VERSTEEG, 1985). They also stated that Elisa was superior in viriological diagnosis than other methods.

#### REFERENCES

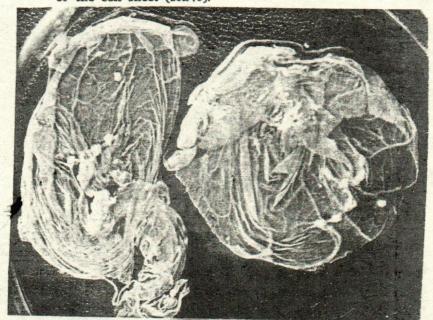
- Alexander, R.A.; Plowright, W. and Haig, D.A. (1957): Cytopathogenic agents associated with Lumgpy skin disease of cattle. Bull. Epiz. Dis. Afr., 5: 489-492.
- Ali, B.H. and Obeid, H.M. (1977): Investigation of the first outbreak of Lumy skin disease in sudan. Br. Vet. J., 133: 184-189.
- Awad, F.I. (1966): Studies on so called oedematous skin disease of buffaloes in URA. Vet. Rec.; Vol. 78 No. 23: 776-778.
- Blood, D.C. and Radostits, O.M. (1989): A test book of the disease of cattle, sheep, pigs, goat and horses. Bailliere Tindall, London.
- Cho, H. and Bohac, J.G. (1985): Sensitivity and specificity of an enzyme linked dmmunosorbent assay for detection of Ibr viral antibody in cattle. Candian J. of Comparative medicine 49(2): 189-194.
- Darcel, C.L.E.Q. (1975): Some factors influencing the micro test method for neutralizating antibodies to the virus of infectious bovine rhinotracheitis. Cand. Vet. J. 16(2): 59-62.
- El-Kanawaty, Z.R. (1989): Some study on Lumpy skin disease. Thesis M.V.Sc. Faculty Vet. Med. Zagazig Univ. Benha branch.
- Grist, N.R.; Bell, E.J.; Follett, E.A.G. and Mrquhart (1977): Diagnostic Methods in clinical virology. 3rd Ed. p. 84-85.
- Haig, D.A. (1957): Lumy skin disease. Bull. Epiz. Dis. Afr. 5: 420-430.
- Hedger, R.S> and Hamblin, C. (1983): Neutrulizing antibodies to Lumpy skin disease virus in African wild life. Comp. Nmmun. Microbial infect Dis. 6(3): 204-213.
- Kendal, G.I.; Ionescu, M. and Dreesman (1983): Utilization of the biotin/avidin systems to amplify the sensitivity of enzyme linked immunosorbent assay (ELISA). J. Immunol. Methods, 56: 329-339.
- Littel, R.; Thoren-Tolling, K. and Sjoquist, J. (1985): Binding of immunoglobulin levels in an mammalian sera. J. immunol, Method. 62: 1-13.
- Losos, G.J. (1986): Infectious tropical disease of domestic animal. First Edition the press Avon.
- MacDonald, R.A.S. (1931): Pseudo-urticaria of cattle. Northern Rhodesia Department of animal health. Annual Report 1930, 20-21.
- MaCowen, K.D.S. (1959): Observation on the epizootiology of Lumpy skin disease during first year of its occurrence in kenya Bull epz. Ds Afri., 7: 7-20.

- Madin, S.H. and Darby, N.B.(1959): Established kidney cell lines of normal adult bovine and ovine origin (2411). Proc-Soc. Exp. Biol. (N.Y) 98: 5744-5745.
- Martin, W.B. and Maureen, G. (1968): Antibodies rothe group II Lumpy skin disease viruses in the sera of cattle in Kenya. Bull. Epizoot. Dis. Afr. 16: 217-222.
- Nawathe, D.R.; Asagba, M.O.; Abegunde, A.; Ajayi, S.A. and Durkwa, L. (1982): Some observation on the occurrence of Lumpy skin disease in Nigeria. Zbl. Vet. Med. B, 29: 31-36.
- Plowright, W. and Ferris, R.D. (1958): The growth and cypathogenicity of sheep pox virus in tissue cultures. Brit. J. Exp. Path., 39: 424.
- Plowright, W. and Witcomb, M. (1959): The growth in tissue cultures of virus derived from lumpy skin disease of cattle. J. Patho. Bacteriol 78: 397-402.
- Prozesky, L. and Barnard, B.H. (1982): A study of the pathology of lumpyskin disease cattle. Onder Stepoort J. Vet. Res. 99: 167-175.
- Vandenende, M.R.; Don, P.A. and Kipps, A. (1949): The isolation in eggs of a new filtrable agent which may be the cause of bovine lumpy skin disease. J. Gen. Microbial., 3: 174.
- Van-Rooyen, P.J.; Munz, E.K. and Wiss, K.E. (1969): The optimal conditions for the multiplication of Nerrthling type lumpy skin disease virus in embryonated eggs. Onder Stepport J. Vet. Res. 36(2): 165-174.
- Versteeg, J. (1985): Acolour of Atlos virology. Published by Woffe Medicial Publication LTD, 1985.
- Von Backstrom, U. (1945): Preliminary report on a new disease, the infectious noture. J. S. Afr. Vet. Med. Ass., 16: 29-32.
- Weiss, K.E. (1963): Lumpy skin disease. Emerging diseases of animals. FAO Agriculture studies No. 61: 179-201.
- Woods (1988): Lumpy skin disease, a review. Trop. Anim. Hlth. Prod. 20: 11-17.

## **LEGENDS**

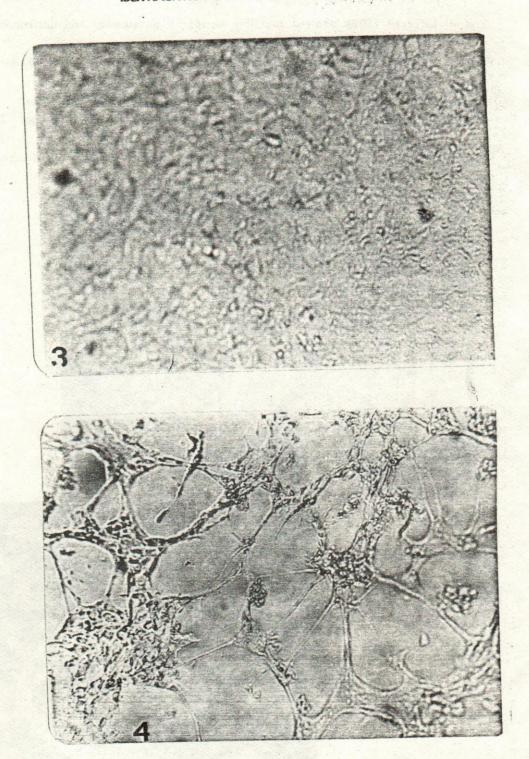
- Fig. 1: Chorioallantoic allantoic membrane showed pook lesions (6-7 days post inoculation).
- Fig. 2: Chorioallantoic allantoic membrane showed pock lesions after (6-7 days post inoculation).
- Fig. 3: Normal non infected MDBK showed a complete confluent sheet (5 days post inoculation) (10x40).

- Fig. 4: Infected MDBK showed rounding shrinking anastomoss and destruction of cells (5 days post inoculation) (10x40).
- Fig. 5: Infected MDBK showed complete destruction of cells sheets (7 days post inoculation) (10x40).
- Fig. 6: Normal non infected LT 4-day-old showed complete confluent sheet.
- Fig. 7: Infected LT alls showed rounding, shrinkage, clumping in grapes like formation (4 days post inoculation).
- Fig. 8: Infected LT cells 6 days post inoculation showed complete destruction of the cell sheet (10x40).



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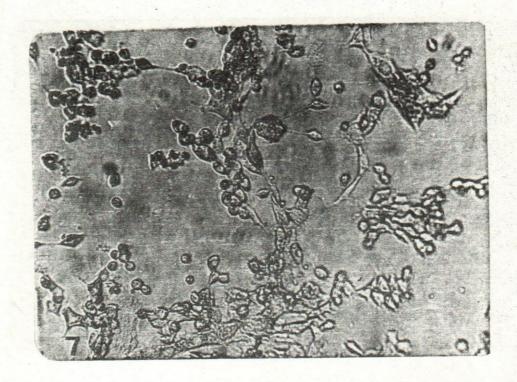


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Table, (1): Isolation and identification, of 20 local isolates of LSD wins.

CAM NOW LI CAM,9-11 Days MORK,4 days LI,4 days local isolates gal	+	+	p- + + + tulation night of the propertion and propertion and propertion and propertion as and propertion as the properties and properties are the properties and properties are the properties and properties are the properti
LT,4 days 1	Sound cell, Sound cell, shrinkage, granulation, shrinkage, granulation, clumping in cell detach- grapes like ment,3-8 day formation, cell detachment,2-6 day	Sound cell, Round cell, shrinkage, granulation, shrinkage, granulation, clumping in cell detach grapes like ment,4-8 day formation, cell detachment,3-6 day	Round cell, Angular sep- + shrinkage, aration, round anastomosis, cell, granulation gramulation, clumping in grapes gramulation, clumping in grapes ment, 4-7 day shrinking and cell detach- ment, 3-5 day
cytopathogen MDBK,4 days	Round cell, shrinkage, anastomosis, granulation, cell detach- ment,3-8 day	Round cell, shrinkage, arastomosis, granulation, cell detach- ment, 4-8 day	Round cell, shrinkage, anastonosis, gramulation, cell detach ment,4-7 day
NDEK LI CAM,9-11 Days MDEK,4 days LI,4 days local isolates	Pock lesion 6-7 days one case gave congestion & thicking of CAM,	Pock leston 6-7 days	Fork leston 6-7 days
LT	•Congression	+	+
MOR			+
CAM	+	+ E	+
Type or samples	Scrabs	Skin	Skin nodules
lifty Breed	Preizian cattle	Freizlan	Freizian Skin Native nodu rate)
Locality	Negh- Hamadd	Vally	Mousha Frei Badary Nati (Assiut Governorate)
			2