Dept. of Animal Medicine, Fac. Vet. Med., KaferEl-Sheikh. Tanta University.

PREGNANCY TOXAEMIA IN SHEEP AND GOATS (CLINICAL AND BIOCHEMICAL ASPECTS)

(With 4 Tables)

By
M.H. NASSER; DAWLAT M.AMIN*; SH.M. MOUSA*;
OMIMA M. KANDIAL**; FATMA M. KAME*
and B.I. AGAG

*Animal health Research Institute, Dokki.,

**: National Research Center, Doki.

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تسمم الحمل في الأغنام والماعز الأعراض الإكلينيكية والصورة البيوكيميانية

محمد حسن ناصر ، دولت أمين ، موسى ، أميميه قنديل ، فاطمه كامل ، بدير عجاج

إشتمل البحث على فحص عدد ٢٠ من الأغنام والماعز والتابعة لمزرعة برج العرب بكفر الشيخ للأغنام والماعز وكان الهدف من البحث دراسة الأعراض الإكلينيكية والبيوكيميائية للحالات التي تعاني من تسمم الحمل. وقد سجلت أهم الأعراض الإكلينيكية متمثلة في فقدان الشهية والخمول الذي أتبعه الأعراض العصبية متمثلة في العمى والرقاد والإغماء والموت في بعض الحالات. وأوضحت التحاليل البيوكيميائية لمصل الدم على إنخفاض معنوي في معدلات الجلوكوز والدهون الثلاثية والكولسترول والماغنسيوم والألبيومين في الحيوانات المريضة بمقارنتها بالسليمة - بينما حدثت زيادة معنوية في معدلات الكورتيزون والأحماض المريضة بمقارنتها بالسليمة.

SUMMARY

This work was carried out to study clinical and biochemical aspects of field cases of ewes and does suffering from pregnancy toxaemia. The first signs observed were anorexia and depression which may be followed by locomotor disturbances (incoordination and star gazing), recumbency, blindness, coma and death. Laboratory findings of present data showed ketonuria, lower serum values of glucose, triglycerides, cholesterol, magnesium and albumin. Higher serum cortisol, free fatty acids, urea, liver enzymes, alpha and gamma globulins were observed. The obtained data were tabulated and discussed.

Key words: Sheep & Goats - Pregnancy Toxaemia

INTRODUCTION

Pregnancy toxaemia is a potentially total metabolic condition of sheep and goats which can cause substantial losses. It occurs in ewes and does during the last 2 to 4 weeks of gestation caused by a negative energy balance resulting from increased energy demands for rapid fetal growth in late gestation and insufficient intake (Smith, 1990). Adult ruminant obtains very little glucose from its diet and its metabolic requirements for glucose are supplied by gluconeogensis in the liver and kidney (Bergman, 1973). During pregnancy and l, the requirements for glucose increase where considerably, in the pregnant animal the fetus and uterus utilize glucose as a major energy source (Lindsay,1973) and during lactation large quantities of glucose are removed by the mammary glands for lactose synthesis (Annison and Linzell, 1964). As a result of the energy deficit, some mobilization of lipid reserves appear to occur which results in a doubling of the plasma free fatty acids, giving rise to a fatty liver and increased ketone bodies in blood and urine (Chaiyabuter, et al., 1982 and kimberling, 1988)

Main clinical manifestations in early pregnancy toxemia disease in ewes and does are reduced appetite, dullness, hypoglycaemia, ketonaemia and ketonuria. In a more advanced stage a severe ketoacidosis, haemoconcentration hyperglycaemia and uraemia often accompanied by dyspnea, recumbency and blindness (Kronfeld, 1972) and El-Sebaie (1995). Excessive salivation and fine muscle fasciculation were occasionally observed in the head region causing movements of the overlying skin and twitching of the ears (Sargison, et al., 1994 and Scott, et al., 1995). The condition can be

diagnosed provisionally by a clinical examination of animals and confirmed biochemically by a marked increase in the concentration of 3 - hydroxybutyrate and a corresponding decrease in the plasma glucose concentration (Scott and Woodman, 1993). The aim of this work was to investigate serum biochemical alterations in field cases of pregnancy toxaemia in ewes and does which may provide a diagnostic aid in such conditions.

MATERIAL and METHODS

Both pregnant ewes and does were included in this study. Fifty-two native breed ewes ranging from 2 to 4 years -old at Meseer, Kafr El- sheikh, production farm were transferred from concentrate to drawa feeding during the last month of pregnancy. Three mortality and 20 abortion cases beside severe to moderate clinical signs characteristics of pregnancy toxaemia had occurred a few days later. These signs include inappetance, dullness, incoordination, grinding on the teeth, sternal recumbency, drowsiness, salivation and blindness.

The diseased does were allotted into two groups (A and B) according to the locality and nutrition regimen. In a flock of 60 pregnant Barkey and Demashky does (2.5-5 years old) at Borg El-Arab sheep and goat production farm diseased cases had recorded (group A). They exhibited one or more of the following symptoms, anorexia, depression, disinclination to move, recumbency and drowsiness. Three cases died and necropsy findings showed paleness of the liver and kidney. The history of the flock indicated that these animals were inadequately fed (about 100 gm concentrate/ head/ day plus rice straw add lib.).

Several cases of suspected naturally occurring pregnancy toxaemia in pregnant native breed does were observed in small private farms at El- wahat Al- Daqhlia (group B). These were seen especially during bad weather days. Diseased cases showed loss of appetite, weakness, muscle tremors of the head and sternal recumbency. The post-mortem examination of the emergency slaughtered case revealed greasy friable pale liver and abdominal fat necrosis. In this locality goats fed mainly grasses beside dates, concentrate or barley and housed in open yards without shelter in winter.

Blood and urine samples were collected from 7 and 11 clinically affected does from groups A and B, respectively as well as 8 affected ewes. Blood and urine Samples were also collected from apparently healthy pregnant does and ewes, 10 samples from each samples for detection of

different causes of abortion (vaginal swabs) and serum were also taken from aborted cases and examined at the Animal Health Research Institute. Urine samples were immediately used for detection of ketone bodies using coumbour-9 test-strips (Boehringer Monnheim, Germany). Blood serum samples were subjected to glucose determination on the day of sampling according to Siest et at (1981), cortisol after Schlaghedee et al. (1992) and free fatty acids as described by Schuster, and Pilz, (1979). Serum alkaline phosphatase (ALP) and inorganic phosphorus after Kilchling and Freiburg (1951). Serum total lipids, phospholipids, triglyceride, cholesterol, total protein, urea, alanine amino-transferase (ALT), aspartate amino-transferase (AST), calcium and magnesium were estimated using reagent kits (Biomerieux, Marcy-L Etoile, France). Serum protein electrophoretic pattern was assayed according to Davis (1964) and Ornstein (1964). Statistical analysis was carried out after Snedecor and Cochran (1974).

RESULTS

Blood serum concentrations of glucose, phospholipids, triglyceride, calcium and magnesium showed a marked decrease in pregnancy toxaemic ewes and does compared to non-toxaemic ones (Tables, 1 and 3). Serum cholesterol levels was only decreased in toxemic ewes.

Serum cortisol, free fatty acids, urea, ALT, AST, and ALP were generally increased in toxaemic ewes and does than in non-toxaemic ones-In significant difference were seen between toxaemic and non toxaemic animals for: total lipids and total protein. While serum albumin concentrations were dropped in all toxaemic animals, alpha and beta globulins as well as total globulins were elevated especially in toxaemic ewes compared to apparently healthy ones (Tables, 2 and 4).

DISCUSSION

Anorexia and depression were the first signs observed in both pregnancy toxaemic ewes and does. Sternal recumbency and locomotor disturbances including incoordination in ewes and muscle tremors in goats were previously recorded by El-Sherif et al. (1978), Smith (1990) and El-Sebaie et al. (1992) and El-Sebaie(1995). Forbes and Singleton (1964) described that the nervous symptoms may be due to an inability of the nerve cells to utilize sugar, perhaps as a result of high cortisol levels. Moreover,

excess salivation and blindness showed in severe toxemic cases of ewes were in accordance with the previous reports of Buswell et al. (1986) and Sargison et al (1994).

Laboratory findings revealed the presence of ketone bodies in the urine samples of clinically affected animals. The results obtained for serum glucose indicate that the ketotic ewes and does (group A) were hypoglycemic. These results could be confirmed with those obtained by Storry and Rook (1962) and Jonsson and Pehrson (1972) who found that the glucose levels are related to the animals energy status, values fulling with a negative energy balance. Affected does in group B, in spite of showing clinical signs of pregnancy toxemia, had serum glucose concentration near the normal range. These results indicated that exposure to cold resulted in an increase in mobilization of glucose as well as free fatty acids to be used in the thermogenic process (Terashima et al., 1982)

Serum cortisol concentrations showed a marked increase in diseased animals when compared to healthy ones. Similar result were recorded by Lindler (1959) who found that in ketotic sheep, the circulating cortisol levels were almost 3 times those of healthy sheep. Kimberling (1988) postulated that the stress and low caloric intake in pregnancy toxemic animals have a profound effect on the kidney and adrenal gland with a significant reduction in renal blood flow and glomerular filtration rate which raise plasma renin activities and elevate plasma cortisol levels.

Concerning serum lipid pattern, the observed decrease in phospholipids and triglyceride in ketotic animals was in agreement with Payne (1977) who mentioned that in ketosis, serum lipid values especially phospholipids and triglyceride were reduced. Henricson et al. (1977) concluded that the lower triglyceride content is the result of reduced appetite in animals with hyperketonaemia. The observed decrease in serum cholesterol in the ketotic ewes could be confirmed with the result obtained by kamel (1992). The sharp increase in the concentrations of free fatty acids in the sera of both affected ewes and does could be attributed to the increased mobilization of fatty acids from the adipose tissues in response to an increased requirement for endogenous substrate for energy production during pregnancy (Noble et al., 1971). Haughey (1973) found that fat catabolism is a major change in cold-stressed neonatal lambs. Russel et al. (1967) suggested that plasma free fatty acids would be the most useful index of the degree of under nourishment in pregnant ewes.

The elevated values of serum urea in the diseased animals can be fully explained by the observation of Parry and Tylor (1956) who found fatty

infiltration in the tubular epithelium of the kidneys of ketotic ewes. The increase in the activities of serum enzymes (ALT, AST& ALP) in diseased animals is a strong evidence for the degree of their liver damage. EL-Sebaie et al. (1992) observed severe hepatic changes in ketotic ewes and does.

The present decrease in serum calcium concentrations in diseased does (group A) tends to support the conclusion of Yoshida (1979) and Blood et al. (1983) where at any stage of ketosis calcium is probably lost through urine with consequent uremia of compensating acidosis. While the present decrease in serum magnesium concentrations in both ewes and does could be confirmed with the results obtained by Terashima et al. (1982).

The obtained results for serum protein electrophoretic pattern (Tables, 2 and 4) in ketotic animals showed a significant drop in the albumin concentration and a significant elevation in the concentrations of alpha and beta globulins and consequently total globulins. Vihan and Rai (1984) reported hypoalbuminaemia in pregnancy toxemic goats, while Ceron et al. (1994) observed non significant decrease in serum albumin, slight increase in alpha and beta globulins and a consistent increase in gamma globulins in ketotic goats. They postulated that the increase in alpha fraction in ketotic animals may be due to the combining of alpha globulin fractions with lipid compounds to facilitate their transport as lipoproteins (in ketosis there is a mobilization of reserve triglycerides to compensate the lack of blood glucose). Similarly, beta globulins are involved in lipid transport

Kimberling (1988) described that the total feed requirements for the single bearing ewe during the last 6 weeks is 1.5 x maintenance and 2 x maintenance for a twin bearing ewe. The additional energy for this period is best supplied by concentrate feeds as the rumen capacity is limited by fetal expansion. Accordingly the animals included in this study receive low feed requirements (less concentrates or only drawa) which led to occurrence of the disease. Finally it could be concluded that early diagnosis of ovine pregnancy toxaemia is vital for satisfactory treatment and pregnant ewes should receive its full requirements during the last stage of pregnancy toxoid and prevent its occurrence.

REFERENCES

Annison, E.F. and Linzell, J.L. (1964): J. Physiol. Lond. 175: 372. Cited by Chaiyabuter et al (1982) Bergman, E.N (1973)Glucose metabolism in ruminants as related to hypoglycemia and ketosis Cornell Vet., 63:341. Cited by Chaiyabutr et al (1982).

- Bergman, E.N. (1973): Glucose metabolism in ruminants as related to hypoglycemia and ketosis Cornell Vet., 63: 341. Cited by Chaiyabutr et al. (1982).
- Blood, D.C., Henderson, J.A. and Radostits, O.M. (1983): Veterinary Medicine. 6th ed. London: Bailliere Tindall.
- Buswell, J.F.; Haddy, J.P. and Bywater, R.J. (1986): Treatment of pregnancy toxaemia in sheep using a concentrated rehydration solution. Vet, Rec., 118: 208-209
- Ceron, J.J.; Carcia Partida, P.; Sotillo, J.; Bayon, A. and Gutierrez Panizo, C. (1994): :Serum protein and protein electrophoretic pattern in goats with ketosis during various stages of reproduction. In XVIII World Buiatrics Congress. Proceedings XXVII Congress of Italian Association of Buiatrics. Augst 29 to 2 Sept. 99. 1309-1313.
- Chaiyabuter, N.; Faulkner, A.and Peaker, M.(1982): Glucose metabolism in vivo in fed and 48h starved goats during pregnancy and lactation.Br.J.Nutr., 47:87-94.
- Davis, B. G. (1964): Disc electrophoresis 11 Methods and application to human serum proteins. Ann. N.Y. Acad. Sci., 121: 404-427.
- El-Sebaie, A.H.; El-Shazly, M.A.; El-Gharram, M. and Alhindi, A.B. (1992): Pregnancy toxaemia in goats and sheep: Clinical updating. Proc. 5th Sci. cong., Fac. Vet. Med., Assiut Univ., Nov. 8-10, pp. 9-18.
- El-Sebaie, A.H. (1995): Caprine ketosis in does .3rd Sci Cong. Egyptian Soc. for Cattle Disease Vol. (368-371.
- El-sherif, M.M.T.; Mohamed, O.M.E. and Mostafa, M.A. (1978): Fermented berseen as a perdisposing cause of pregnancy toxaemia in ewes. Vet. Med. J., Fac. Vet. Med. Cairo Univ., 26: 157-164
- Forbes, T. J. and Singleton A. G. (1964): Ovine Pregnancy Toxaemia: A Review Br. Vet. J., 120:56-68.
- Haughey, K.G. (1973): Cold injury in newborn lambs. Aust. Vet. J., 49: 554-563.
- Henricson, B.; Jonsson, G. and Pehrson, B. (1977): "Lipid pattern", Glucose Concentration and Ketone level in the blood of cattle. Zbl. Vet. Med. A, 24: 89-102.
- Jonsson, G. and Pehrson, B. (1972): Some blood parameters in dairy cows at different feeding intensities. ({:::).International Meeting on Diseases of Cattle, Lond., 250-257.

- Kamel, F.M. (1992): Clinical and Biochemical Studies On Pregnancy toxaemia in three breeds of goats-Proc. 5th Sci., Cong., Fac. Vet. Med. Assiut Univ. Nov.8-10 pp. 1-8.
- Kilchling, H. and Freiburg, Br. (1951): Alkaline Phosphatase And Inorganic Phosphorus in serum. In Clinical Photometric, 3 rd ed. Verl Ges. Mph, Stutgart
- Kimberling, C.V. (1988): Jensen and Swifts Diseases of Sheep. 3rd ed. Lea& Febiger, Philadelphia.
- Kronfeld, D.S. (1972): Ketosis in pregnant sheep and lactating cows. A. review. Austr. Vet. J. 48, 680-687 hgsiol. Lond. 175: 372. Cited by chaiyabuter et al (1982).
- Lindler, H. R (1959): Nature (lond.), 184, Supp, 21: 1645. Cited by Forbes and Singleton (1964).
- Lindsay, D.B. (1973): In Production Disease in farm Animals. (1973), p.107 (Paync, K.G., Hibbitt, K.G.and Sansom, B.F. editors) London: Bailliere Tindal.
- Noble, R.C.; Steel, W. and Moore, J.H. (1971): The plasma lipids of the ewe during pregnancy and lactation. Res. Vet. Sci., 12: 47-53-
- Ornstein, L. C. (1964): Disc electrophoresis. I. Background and Theory AnnNY. Acad. Sci., 212: 221-235
- Payne, J.M. (1977): Metabolic Diseases in Farm Animals. Willam Heinemann Medical Books Ltd., London.
- Parry, H.B. and Tylor, W.H. (1956): Renal functions in sheep during normal and toxaemic pregnancy. J. Physiol., 131: 383-392.
- Russel, A.J.F, Doney, J.M. and Reid, R.L. (1967): The use of Biochemical Parameters in Controlling Nutritional State in Pregnant Ewes and the Effect of Undernorishment During pregnancy on lamb birth-weight. J Agric Sci Cambridge, 68: P359.
- Sargison, N. D. scott, P.R Penny, C. D., Pirie, R. S and kelly, JM (1994):
 Plasma Enzymes and Metabolites as Potential Prognostic Indices of
 Ovine Pregnancy Toxaemia A Preliminary Study Br. Vet. J., 150:271

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- Schlaghedee, R. E., Kornely, Santen, R. T. and Ridderskamp, P. (1992): The Effect of Long-term Glucocorticoid on Pituitary- Adrenal Responses to Exogenous Corticotrophic Releasing Hormone. New Eng. J. Med., 326: 226-230
- Schuster, V.H.G. and pilz, V.K. (1979): Kolorimetrische Mikromethode zur Bestimmung der Unveresterten Langkettigen Fettsauren Im Serum. Z.Med Labor. Diagn. 20: 212-217.

- Scott, P.R., Sargison, N.D., Penny, C.D., Pirie, R.S. and Kelly, J.M. (1995):
 Cerebrospinal fluid and plasma glucose concentrations of ovine
 pregnancy toxaemia cases, MZappetant ewes and normal ewes during
 late gestation.Br.Vet.J.,151: 39-44.
- Scott, P.R. and Woodman, M.P. (1993): An outbreak of Pregnancy Toxaemia In a Flock of Scottish Blackface Sheep. Vet. Rec., 133: 597-598.
- Siest, G., Henny, J. and Schiele, F. (1981): Interpretation Des Examens de Lboratoire, Karger ed, 206-223
- Smith, B.P (1990): Large Animal Internal Medicine. The C.V. Mosby Company. Philladel Phia, Torunto.
- Snedecor, G.W. and Cochran, W.G. (1974): Statistical Methods. 6th ed. Iowa State Univ. Press. Ames, Iowa USA.
- Storry, J. E. and Rook, A.F. (1962): The Effect of Level of Feeding Before and After Calving On the Concentration of Plasma Glucose in the Cow. Proc. Nutr. Soc., 21: 39-40.
- Terashima, Y., Tucker, R.E., Deetz, L.E., Degregorio, R.M., Muntifering, R.B. and Mitchell, C.E. (1982): Plasma Magnesium Levels as Influenced by Cold Exposure in Fed or Fasted Sheep. J. Nutr., 112: 1914-1920.
- Vihan, V.S. and Rai, P. (1984): Studies on Biochemical and Haematological Changes in Metabolic Derangements of Sheep and Goats. Ind. J.Vet. Med. (IV): 9.
- Yoshida, S. (1979): Blood serum Ca, Mg and P. Levels in Ketotic cows. Am. Dairy Sci. Association 74th Annual Meeting.

Table (1) some Laboratory findings in toxaemic pregnant and apparently healthy ewes.

Parameter	Diseased ewes	Healthy ewes	
Glucose (mg/dl)	26.96* ± 2.80	36.24 ± 3.14	
Cortisol (ng/m1)	48.87*** ± 3.11	30.32 ± 1.84	
Phospholipids(mg/dl)	64.13 ± 4.00	80.06 ± 5.20	
Triglyceride (mg/dl)	41.50* ± 3.06	51.43 ± 2.84	
Cholesterol (mg/dl)	65.40* ± 2.63	76.11 ± 3.56	
Free fatty acids(mg/dl)	49.83*** ± 2.88	23.33 ± 1.96	
Total protein (gm/dl)	6.45 ± 0.33	7.03 ± 0.24	
Urea (mg/dl)	34.75* ± 2.17	28.36 ± 2.08	
ALT (U/L)	30.77** ± 1.50	24.24 ± 0.63	
AST (U/L)	41.85*** ± 0.98	27.13 ± 0.75	
ALP (mmol U/L)	5.61* ± 0.60	3.58 ± 0.35	
Phosphorus (mg/dl)	5.26 ± 0.48	6.39 ± 0.41	
Calcium (mg/dl)	9.61 ± 0.43	10.52 ± 0.53	
Magnesium (mg/dl)	2.03* ± 0.16	2.69 ± 0.21	

^{*} Significant at:

^{*} P<0.05

^{*} P<0.01

^{*} P<0.001

Table (2) Serum protein electrophoretic pattern in toxaemic and healthy ewes.

Fraction percentage	Diseased ewes	Healthy ewes
Albumin	29.75* ± 1.12	33.15 ± 1.03
Alpha-globulins	20.73 ± 0.90	21.12 ± 0.84
Alpha-1	5.77 ± 0.23	5.92 ± 0.29
Alpha-2	4.49** ± 0.21	3.35 ± 0.26
Alpha-3	10.47 ± 0.60	11.85 ± 0.38
Beta-globulins:	$11.13* \pm 0.66$	9.12 ± 0.55
Beta-1	$6.16* \pm 0.30$	5.34 ± 0.24
Beta-2	4.97* ± 0.41	3.78 ± 0.28
Gamma-globulins:	38.39 ± 1.70	36.61 ± 0.89
Gamma-1	25.67 ± 1.24	24.26 ± 1.08
Gamma-2	12.72 ± 0.54	12.35 ± 0.41
Total globulins	70.25* ± 1.12	66.85 ± 1.03
A/G Ratio	$0.42* \pm 0.25$	0.50 ± 0.22

^{*} Significant at:

P < 0.05

P < 0.01

Table (3) Some laboratory findings in Toxaemic and apparently healthy does.

Parameter	Diseased does		Healthy does	
	Group A	Group B		
Glucose (mg/dl)	28.75* ± 2.18	31.55 ± 2.12	37.12 ± 2.06	
Cortisol ng/m1	46.22*** ± 2.42	n.d.	18.76 ± 1.05	
Total lipids (mg/dl)	339.87 ± 19.40	'n.d.	315.30 ± 20.86	
Phospholipids (mg/dl)	50.38*** ± 3.22	63.18* ± 4.36	77.16 ± 4.23	
Triglyceride (mg/dl)	37.56** ± 3.58	61.43 ± 2.84	54.20 ± 4.00	
Cholesterol (mg/dl)	69.14 ± 4.10	n.d.	62.05 ± 5.46	
Free fatty acids (mg/dl)	38.56*** ± 2.66	53.17*** ± 3.80	20.70 ± 1.56	
Total protein (gm/dl)	6.34 ± 0.22	6.65 ± 0.30	6.58 ± 0.27	
Urea (mg/dl)	35.17* ± 2.11	38.30** ± 3.24	29.28 ± 1.18	
ALT (U/L)	27.12*** ± 0.48	33.41*** ± 1.36	22.92 ± 0.37	
AST (U/L)	36.70*** ± 0.84	42.70*** ± 1.75	26.11 ± 0.57	
ALP (mmo1 U/L)	5.06*** ± 0.30	6.53*** ± 0.48	3.16 ± 0.19	
Phosphorus (mg/dl)	7.89* ± 0.48	6.82 ± 0.56	6.17 ± 0.38	
Calcium (mg/dl)	8.53 ± 0.22	9.18 ± 0.36	9.87 ± 0.34	
Magnesium (mg/dl)	1.94* ± 0.48	1.81* ± 0.11	2.36 ± 0.14	

n.d. = Not determined.

* Significant at:

P < 0.05

P < 0.01

P < 0.001

Table (4) serum protein elechophoretic pattern in toxaemic and healthy does.

Fraction	Disease	Healthy does	
	Group A	Group B	
Albumin	28.44* ± 1.08	29.25* ± 1.16	32.47 ± 0.85
Alpha-globulins	15.29* ± 0.87	14.78 ± 0.76	13.09 ± 0.39
Alpha-1	7.18 ± 0.52	6.88 ± 0.44	6.05 ± 0.56
Alpha-2	8.11 ± 0.64	7.90 ± 0.36	7.04 ± 0.47
Beta-globulins:	22.78** ± 0.48	21.14 ± 0.83	20.16 ± 0.68
Beta-1	11.64 ± 0.51	10.32 ± 0.49	11.08 ± 0.47
Beta-2	11.14* ± 0.48	10.82 ± 0.56	9.07 ± 0.60
Gamma-globulins:	33.49 ± 1.60	34.83 ± 1.08	34.28 ± 1.39
Gamma-1	24.00 ± 1.21	23.90 ± 0.95	24.29 ± 1.15
Gamma-2	9.49 ± 0.54	10.93 ± 0.78	9.99 ± 0.45
Total globulins	71.56* ± 1.08	70.75* ± 1.16	67.53 ± 0.85
A/G Ratio	0.40* ± 0.022	0.41* ± 0.026	0.48 ± 0.015

^{*} Significant at:

^{*}P < 0.05

^{**}P < 0.01