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ROLE OF ENVIRONMENT IN OCCURRENCE OF CALF DIARRHOEA

(With 4 Tables)

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دور البيئة في حدوث اسهال العجول

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تم جمع عدد ١٤٨ عينة براز من عجول سليمة ظاهريا (٨٠) و عجول مصابـة بالإسـهال (٦٨)، هواء (٤٥)، مياه (٣٠)، تربة (٣١)، مسحات جلدية (١٤٨) ومسحات من الحلمات (١٤٨) وتم فحصها بكتير يولوجيا. أظهرت النتائج تواجد ميكروب إشيريشيا كولاي والسالمونيلا (٧٠ و ٣٠٧ على التوالي) ومن العجول المصابة بالإسهال (٤٩٧٤ و ٣٠٠٣ على التوالي) بالإضافة الى انه تم عزل ميكروب الإشريشيا كولاى والسالمونيلا من عينات الهواء (١٣.٣ و ٤,٤% على التوالي)، مياه (٧٠ و ٢,٦% على التوالي)، تربسة (٧١) و ٢٥,٨ على التوالي)، مسحات جلاية (٧٠,٧ و ١٠,١% على التوالي) ومسحات الحلمات (٢٦,٣ و ٤,٧% على التوالي). هذا وقد تم التعرف على انواع السالمونيلا التي تـم عزلها من العجول والبينه المحيطه سيرولوجيا. كذلك تم عـزل بعـض البكتيريا السالبة و الموحية الحرام ينسب مختلفه من العجول والبيئة. هذا وقد لوحظ أن مطهر + TH4 لـــه تأثير قاتل على ميكروب اشيرشيا كولاي عند تركيز ٥٠، و ١ % بعد ٢٠ و ١٠ دقائق عند التوالي وكذلك بالنسبة لميكروب السالمونيلا تيفيميوريم بعد ١٠ و٥ دقائق علي التوالي. بالإضافة إلى ذلك فأن الأيودوفور ٢% له تأثير قاتل على ميكروب الأشيرشيا كولاى عند تركيز ١% بعد ٢٠ دقيقة وبالنسبة لميكروب السالمونيلا تيفيميوريم عند تركيز ٥,٠ و ١% بعد ٢٠ و ١٠ وقائق على التوالي. وقد وجد أن البيئة تلعب دورا كبيرا في الاحتفاظ ونقل المبكر ويات المسينة للاسهال في العجول.

SUMMARY

A total of 148 faecal samples (80 from apparently healthy and 68 from diarrheic calves), air (45), water (30), soil (31), dam coat (148) and teat swabs (148) were collected from a dairy farm and subjected for

bacteriological examination. The obtained results revealed that E. coli and Salmonella serotypes could be isolated from apparently healthy (70 and 3.7%) and diarrheic calves (79.4 and 10.3%). In addition, E. coli and Salmonellae were recovered from air (13.3 and 4.4%), water (70 and 6.6%), soil (71 and 25.8), dam coat (50.7 and 10.1%) and teat swabs (26.3 and 4.7%). Moreover, serotyping of isolated Salmonellae from calves and environment was carried out. Also, other Gram-negative and Gram-positive bacteria isolated from calves and environment were recorded. On the other hand, it was found that TH4+ is effective against E. coli (at concentrations of 0.5 and 1% after contact period of 20 minutes and 10 minutes respectively) and Sal typhimurium (at concentration of 0.5 and 1% after 10 and 5 minutes respectively). In addition, iodophor 2% is effective against E. coli (at a concentration of 1% after 20 minutes) and Sal. typhimurium (at concentrations of 0.5 and 1% after 20 and 10 minutes respectively). It can be concluded that the environment plays a dangerous role in maintaining the infectious agents responsible for diarrhoea in calves.

Keywords: Occurrence, Calf, Diarrhoea.

INTRODUCTION

Environment surrounding calves constitutes the main source of pathogenic and potentially pathogenic microorganisms. Such pathogens are involved as causative agents in calf diarrhea.

Environmental conditions in the intensively confined houses can markedly influence the incidence of the disease that may be higher if large number of animals are allowed to house in unhygienic conditions (Quigley, 1995).

Roy and Twenouth (1972) recorded that the enteric diseases was basically a result of alteration of gastric and intestinal function to less than the optimum and its clinical manifestation as an infectious disease depend on the age of the calf and the balance between its immunological and microbiological environment.

Boylan (1982) reported that under most farm conditions eliminating infectious agents from the environment was difficult, but good management procedures could aid in maintaining a level of environmental contamination that was less than critical.

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Infection may be transmitted directly from dams or infected calves or indirectly through contaminated food and water, feeding and watering utensils or a build up of infection in the calf pen (Williams et al., 1975).

In Egypt, several studies have been carried out on environmental pathogens (Abd-El-Karim, 1971; Hamoud et al., 1978; Marzouk et al. 1980; Mowafi et al. 1980; Bashandy et al., 1983; Mostafa, 1984; Soliman, 1984 and El-Masry, 1989 & 1996).

The aim of this work is to determine the environmental sources with certain pathogens as causative agents for diarrhea in calves and to study the efficacy of two disinfectants on the viability of some bacterial isolates recovered from diarrheic calves to assess preventive measure to avoid transmission of infection.

MATERIAL and METHODS

This study was carried out in a farm belonging to Beheira province. Two hundred and forty cows with 151 calves were included in this farm. Cows were housed in open yard system with dirty floor supplied with a parlour. Calves were kept in a calf pen and grouped by age in three pens. Disposal of manure was carried out mechanically.

2.1. Faecal samples:

A total of 148 faecal samples were collected from apparently healthy (80) and diarrheic calves (68). Each sample was divided into 2 parts, the first part was inoculated into nutrient broth and the second into selenite F broth. After incubation, plating was carried out on the selective media.

2.2. Environmental samples:

2.2.1. Air samples:

Forty five air samples were collected from the investigated cow house using liquid impinger (Brachman et al., 1964). Each sample was inoculated into Selenite F broth and nutrient broth. After incubation, plating was carried out using blood agar, Salmonella Shigella agar and Salt mannitol agar and the plates were incubated at 37 °C for 48 hours.

2.2.2. Water samples:

Thirty drinking water samples each of 200 ml were filtered through $0.45~\mu m$ membrane filter. Each membrane was cultured on the previously mentioned media.

2.2.3. Soil samples:

Thirty one soil samples each of 20 gm were collected aseptically by a sterile spatula and transferred to a sterile brown glass bottle and placed in 100 ml sterile distilled water and mixed on a mechanical shaker for one hour and left undisturbed for 10 minutes to allow the large particles to settle. Ten ml of the supernatant were added to 90 ml of nutrient broth and Selenite F broth. After incubation, 0.1 ml from each was taken immediately and plated on previously mentioned agar media.

2.2.4. Dam coat swabs:

One hundred and forty eight skin swabs were collected from dams of apparently healthy (80) and diarrheic calves (68). Sterile swabs moistened with sterile broth were rubbed on the skin of hind quarters and perineal region.

2.2.5. Teat swabs:

One hundred and forty eight teat swabs were collected from dams of apparently healthy (80) and diarrhea calves (68) according to Rendos et al. (1975). Coat and teat swabs were inoculated into sterile test tubes containing nutrient broth and plated on previously mentioned agar media.

- 2.3. Identification of bacteria was carried out according to Edward and Ewing (1972) and Murry et al. (1984).
- 2.4. Serological identification of isolated Salmonella spp. was carried out according to Kauffmann (1978).
- **2.5.** Efficacy of TH4+ and Iodophor on the viability of *E. coli* and *Salmonella typhimurium*:

The bactericidal effects of TH4+ (composed of Quaternary ammonium compounds and Glutaraldehyde as well as plant extracts) and Iodophor 2 % were assessed at on *E. coli* and *Salmonilla* typhimurium concentrations of 0.5 and 1 % after contact periods of 5, 10 and 20 minutes.

RESULTS and DISCUSSION

Table (1) revealed that E. coli was isolated from faecal matter of apparently healthy calves at percentage of 70 %. This indicates that E. coli is considered to be a normal inhabitant of the intestinal tract. Under stress factors the organims becomes pathogenic causing calf diarrhea. A finding is supported by Amstutz (1965) who reported that feeding and management errors were frequently associated with calf diarrhea.

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In addition, *E. coli* could be isolated from diarrheic calves at percentage of 79.4% (Table 1). This result is supported by that reported by Anon (1964), Soliman (1984), El-Masry (1989), Hozain (1990), Hafiz et al. (1994), Salib (1995) and El-Masry (1996).

E. coli was recovered from air, water and soil of calf pen at percentage of 13.3, 70 and 71 % respectively (Table 2). Moreover, E. coli could be isolated from dam coat and teat swabs at percentage of 50.7 and 26.3 % respectively (Table 2). These results are supported by those Johansen (1972), Fiser and Svitasvsky (1973), Rendos et al. (1975), Eberhart (1977), Mowafi et al. (1980), Zakarya et al. (1980), Bashandy et al. (1983), Mostafa (1984), Soliman (1984), Asma et al. (1996), El-Masry (1989 & 1996).

The difference in isolation rate of E. coli may be attributed to environmental and managemental conditions (build up of infection in calf house, the farm dirts and the adverse climatic changes) as reported by Acres (1985) and Hinton et al. (1994).

Salmonellae were isolated from apparently healthy and diarrheic calves at percentage of 3.7 and 10.3% respectively (Table 1). In addition, Salmonellae could be recovered from air, water, soil, dam coat and teat swabs at percentage of 4.4, 6.6, 25.8, 10.1 and 4.7% respectively (Table 2). Similarly, Salmonellae could be isolated from calves and environment by Abd-El-Karim (1971), Osborne et al. (1977), Bulgin et al. (1982), El-Sayed (1987), Mettias (1987), Saad (1993), Asma et al. (1996) and El-Masry (1996).

Serological identification of Salmonellae isolated from calves and environment revealed that Sal. typhimurium could be isolated from apparently healthy and diarrheic calves at percentage of 2.9 and 9.7% respectively. Also, Sal. enteritidis was isolated from diarrheic calves at a percentage of 2.4% and Sal. dublin was recovered from apparently healthy and diarrheic calves at percentage of 1.4 and 4.8% respectively. In addition, Sal. typhimurium was isolated from air, water, dam coat and teat swabs at percentage of 2.2, 6.6, 12.9, 3.4 and 2% respectively. The respective values for Sal. entertidis were 0.0, 0.0, 6.4, 2.7 and 0.7%. Moreover, Sal. dublin and Sal. newport were recovered from dam coat (3.2 and 3.2%) and teat swabs (1.3 and 0.7%), Table (3).

It was found that Sal. typhimurium was the most common serotype isolated from calves and environment causing diarrhea in calves as reported by Hurd et al. (1994). The survival of infective agents, the presence of rats and mice and the management practices may change the

balance between the calf and its microbial flora especially Salmonella as recorded by Watts and Wall (1952), Abd-El-Karim (1971), Williams et al. (1975) and Roy (1980).

The results recorded in Table (1) revealed isolation of *Klebsiella pneumoniae* (5 and 11.8%), *Pseudomonas aeuroginosa* (6.2 and 10.3%), Shigella (1.2 and 0.0%), Proteus spp. (8.7 and 19.1%), Arizona (2.5 and 0.0%), Citrobacter (2.5 and 1.5%), Enterobacter (2.5 and 1.5%), *Staph. aureus* (6.24 and 13.2%) and *Strept. faecalis* (2.5 and 7.3%) from healthy and diarrheic calves respectively. These results are coinciding with those reported by El-Amrousi et al. (1972), Ahmed (1975), Farid et al. (1979), Mettias (1987), Mnatsakanov et al. (1991), Salib (1995) and El-Masry (1996).

Results presented in Table (2) showed that *Klebsiella pneumoniae*, *Pseudomonas aeruginosa*, Shigella, Proteus spp., Arizona, Citrobacter, Enterobacter, Providencia, *Staph aureus* and *Strept. faecalis* could be isolated from air, water, soil, dam coat and teat swabs of examined animals at various percentages. These results are supported by those obtained by Abd-El-Karim (1971), Brander (1973), Knittel (1975), Rendos et al. (1975), Mowafi et al. (1980), Marzouk et al. (1980), Zakarya et al. (1980), Soliman (1984) and El-Masry (1989 & 1996). Such organisms may have a role in inducing enteritis in calves under certain circumstances (Topley and Wilson, 1975; Buxton and Fraser, 1977).

The results indicated that TH4+ is effective against *E. coli* (at concentrations of 0.5 and 1% after contact period of 20 and 10 minutes respectively) and *Sal. typhimurium* (at concentrations of 0.5 and 1% after 10 and 5 minutes respectively). In addition, iodophor 2% is effective against *E. coli* (at concentration of 1% after 20 minutes) and *Sal. typhimurium* (at concentrations of 0.5 and 1% after 20 and 10 minutes respectively), Table (4).

From the previous results, it can be concluded that the environment plays a dangerous role in maintaining the infectious agents responsible for diarrhea in calves especially *E. coli* and Salmonella. So, to avoid infection by environmental pathogens, good hygienic measures should be applied including adequate ventilation, proper housing, hygienic disposal of sewage, thorough cleaning and disinfection of calf houses as well as proper management of calves and dams.

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Table (1): Bacteria isolated from feacal samples of examined calves.

Isolates	Appai healthy		Diarrh	eic calves
	+ ve	%	+ ve	%
Gram negative				
- E. coli	56	70	54	79.4
- Salmonella	3	3.7	7	10.3
- Klebsiella pneumoniae	4	5	8	11.8
- Pseudomonas aeruginosa	5	6.2	7	10.3
- Shigella spp.	1	1.2		0.0
- Proteus spp.	7	8.7	13	0.0
 Arizona spp. 	2	2.5		0.0
- Citrobacter	2	2.5	1	1.5
- Enterobacter	2	2.5	1	1.5
Gram positive				
- Staphylococcus aureus	5	6.2	9	13.2
- Streptococcus faecalis	2	2.5	5	7.3

No. of examined apparaently healthy calves = 80.

No. of examined diarrheic calves = 68.

Table (2): Bacteria isolated from environment of calves, dam coat and teat swabs:

	A	Air	Water	ter	Soil	lic	Dan	Dam coat	Teat	Teat swab
Isolates	+ ve	%	+ ve	%	+ ve	%	+ ve	%	+ ve	%
Gram negative										
- E. coli	9	13.3	21	70	22	7.1	75	50.7	39	26.3
- Salmonella	2	4.4	2	9.9	00	25.8	15	10.1	7	4.7
- Klebsiella pneumoniae	2	4.4	9	20	3	1.6	14	9.4	00	5.4
 Pseudomonas aeruginosa 	5	11.1	3	10	9	19.3	15	10.1	10	6.7
- Shigella spp.	1	0.0	-	3.3	_	3.2	ł	0.0	}	0.0
- Proteus spp.	8	17.8	9	20	6	29	2	1.3	11	7.4
- Arizona spp.	l	0.0	_	3.3	-	3.2	1	0.0	1	0.0
- Citrobacter	ł	0.0	4	13.3	_	3.2	9	4	2	1.3
- Enterobacter	_	2.2	3	10	2	6.4	2	3.4	4	2.7
- Providencia	_	2.2	3	10	7	6.4	1	0.0	4	2.7
Gram positive										
 Staphylococcus aureus 	I	0.0	3	10	6	29	4	2.7	7	1.3
 Streptococcus faecalis 	2	4.4	4	13.3	11	35.5	7	4.7	9.	4

No. of examined air samples = 45 No. of examined water samples = 30 No. of examined soil samples = 31

No. of examined dam coat samples = 148 No. of examined teat swab samples = 148

Table (3): Salmonella serotypes isolated from calves, environment, dam coat and teat swabs:

	Apparently	rently	Diarrheic	heic										
Salmonella	healthy	healthy calves	calves	es	Air	<u>.</u>	Water	ter	Soil	.E	Dam coat	coat	Teat swab	swab
serotypes	+ ve	%	+ ve	%	+ ve	%	+ ve	%	+ ve	%	+ ve	%	+ ve	%
Salmonella typhimurium	2	2.9	4	7.6	_	2.2	2	9.9	4	12.9	5	3.4	3	7
Salmonella enteritidis	1	0.0	_	2.4	;	0.0	1	0.0	2	6.4	4	2.7	1	0.7
Salmonella dublin	-	1.4	2	4.8	1	0.0	1	0.0		3.2	3	2	2	1.3
Salmonella newport	1	0.0	1	0.0	ŀ	0.0	;	0.0	_	3.2	3	7	1	0.7

Table (4): Efficacy of TH4+ and Iodophor against E. coli and Salmonella typhimurium:

			E. coli		Salı	Salmonella typhimurium	rium
Disinfectants	Disinfectants Concentration		Time of contact			Time of contact	
	of disinfectant	5 min.	10 min.	20 min.	5 min.	10 min.	20 min.
TH4+	0.5 %	+	+	1	+	ı	1
	1 %	+	i		i	ī	ī
Iodophor	0.5 %	+	+	+	+	+	, i
	1%	+	+		+	1	