Dept. of Poultry Dis. Fac. Vet. Med. Assiut Univ.

CLOSTRIDIAL INFECTION IN CHICKENS "STUDYING THE PATHOGENICITY AND EVALUATION OF THE EFFECT OF SOME GROWTH PROMOTORS ON BROILER PERFORMANCE"

(With 7 Tables)

By R.S. IBRAHIM; IBTIHAL, M. MONAZI* and A.M. SOLIMAN,

*Animal Health Res. Inst., El- Menia (Received at 18/2/2001)

عدوى الكلوستريديم فى الدجاج " دراسة العدوى الصناعية وتقييم تأثير بعض منشطات النمو على اداء بدارى التسمين"

رجب سيد ابراهيم، ابتهال محمد منازع، عادل محمد سليمان

أجريت العدوى الصناعية في بدارى التسمين باستخدام أنواع الكلوستريديم الشائعة العرزل. كانت معدلات النفوق أعلى عند أجراء العدوى باستخدام ميكروب الكلوستريديم متحددا مسع الكوكسيديا عنها عند عمل العدوى بالكلوستريديم منفردا وخصوصا الكلوستريديم برفرنجينز. أزدات شدة الأعراض و الأفات التشريحية عند محاولة الاعداء باستخدام الأنواع المفرزة للسموم من ميكروب الكلوستريديم برفرنجينز عنها عند العدوي بميكرويات الكلوسستريديم برفرنجينز الغير مفرزة للسموم وكلوستريديم كولينم وكلوستريديم سبوروجينز واصبحت أكثر حدة عند العدوى المختلطة مع الكوكسيديا. عند أضافة ميثيلين داى سلساليت الباسيتر اسسين الفلافوميسين ، الانر اميسين ، اللنكوميسين كل على حده الى العلائق التي تم تعذيـــة الطيــور عليها لوحظ ان معدلات النفوق قد تقلصت عند العدوى بالانواع المفررزة السموم أ و ج وكذلك الانوع الغير مفرز للسموم من ميكروب الكلوستريديم برفرنجينز، وعلم ي الْناحيــة الاخرى تحسنت الاوزان بشكل معنوى حيث كان مركب فلاقوميسين هو الاكثر فاعلية عـن باقى المركبات كما قلت الافات التشريحية بشكل واضح. أظلهرت مركبات الانر اميسين واللنكوميسين انخفاض في معدل النفوق من ٤٥% الى ٢٥% وتلاه مركبي الباسينر اسين والفلافوميسين بمعدلي ٣٠ و ٣٥%. تأثرت جميع الميكروبات تحت الدراسة تأثراً تاما فــــــي الحتبار الحساسية عند أستخدام الاموكسيسيللين، ينسيللين، سيروفلوكساسين، انروفلوكساسين، فليموكوين، دوكسي سيكالين، وسلفات الكولستين بينما تأثرت بشكل متوسط عند أستخدام الارثروميسين والكلورامفينيكول والاوكسىتتر اسيكلين وكان هناك مقاومة كاملة ضد مركبات نيوميسين، استريتوميسين والجنتاميسين.

SUMMARY

Experimental infection of broiler chickens with the most commonly isolated clostridium species was done. Mortality rates were higher in birds subjected to combined infection with clostridium and coccidia than those infected with clostridium alone especially in birds infected with Cl perfringens. Symptoms and lesions were more severe in case of infection with toxigenic types of Cl perfringens than in case of experimental infection with non-toxigenic Cl perfringens, Cl colinum and Cl sporogenes. The intensity became more severe after mixed infection with clostridium and coccidia. In case of birds received bacitracine methyl disalicylate (BMD), flavomycin, enramycin and lincomycin, mortality rates due to infection with toxigenic Cl perfringens types A&C and non-toxigenic type were reduced. A significant difference between lesions in birds receiving growth promotors mixed ration was observed. Enramycin and lincomycin reduced the mortality rates from 45% to 25% more than BMD and flyomycin (30% and 35%). On the other hand, a significant improvment in weight gain was observed especially with flavomycin followed by BMD, enramycin and lincomycin respectively. Most of the tested clostridial isolates were highly sensitive to amoxycillin, pencillin, ciprofloxacin, enrofloxacin, flumequin, doxycyclin and colistin sulphate, while they were moderately sensitive to erythromycin, chloramphenicol, oxytetracycline. Complete resistance to neomycin, streptomycin and gentamicin was noticed.

Key words: Clostridial infection, chickens, growth promotors.

INTRODUCTION

Enteritis is considered as one of the major problems affecting severely the broiler production stocks. One of the major enteritis producing bacteria is clostridium species which become more disastrous after coccidiosis resulting in ulcerative and necrotic enteritis. These conditions were reproduced by several workers (Balauca, 1976 & Shan et al, 1985 and Das et al, 1997 a,b). The use of antibiotics as feed additives (growth promotors) in poultry ration is wide spread, which act indirectly by retaining the natural balance of the bacterial flora of the digestive tract, and preventing the bacterial detrimental toxin formation and thus facilitate the absorption of nutrients and consequently depressing the activity of clostidial species.

In formerly published work, different species of genus clostridium were isolated and classified by same authors (Ibrahim et al, 2001). In this study, further work was done to investigate the pathogenicity of isolated bacteria and evaluation of the effect of feed additives on experimental infection with the same organisms as well as testing of their antimicrobial susceptibility.

MATERIALS and METHODS

Preparation of cultures for experimental infection:

A forty eight hours culture of clostridial isolates in cooked meat medium was prepared. The culture suspensions were centrifuged at 4000 rpm for 1 hr. Purity was checked through Gram staining from the sediments. The sediment was washed three times in saline and resuspended in thioglycollate medium. Colony count technique was done according to Cruickshank et al, (1975).

Collection and sporulation of oocysts:

The intestinal and caecal contents of chickens infested with coccidia were collected, suspended with normal saline and filtered through muslin. The filterates were mixed with 2.5% aqueous solution of potassium dichromate and placed as thin layer in petri dishes followed by incubation for a week at room temperature. Daily microscopic examination was carried out to detect sporocysts and sporozoites. The mixtures were washed 3X by normal saline. The sediments containing sporulated oocysts was resuspended in saline solution. Oocysts count were determined according to Parfitt (1958).

Experimental infection:

a) Experimental infection with isolated clostridial species only:

This was done according to Das et al., (1997c). A total of one hundred and twenty 3-week-old chickens (Balady breed) were divided into 6 groups, each of twenty birds.

First group was infected orally with 2 ml broth culture containing 1.8x10⁹ CFU/ml of Cl. perfringens type A. Second group was infected orally with 2 ml broth culture containing 1.8x10⁹ CFU/ml of Cl. perfringens type C. Third group was infected orally with 2 ml broth culture containing 1.8x10⁹ CFU/ml of non-toxigenic Cl. perfringens. Fourth group was

infected orally with 2 ml broth culture containing 10^7 CFU/ml of Cl. colinum. Fifth group was infected orally with 2 ml broth culture containing 1.5×10^9 CFU/ml of Cl. sporogens. Sixth group was inoculated orally with 2 ml sterile broth, and kept as control. All inoculated groups received 3 successive doses at 2 days intervals and kept under observation for 4 weeks postinfection. Clinical signs, PM findings, moralities, average body weights and microbial recovery rates were recorded.

b) Combined experimental infection with clostridium and coccidia:

A total of hundred and twenty 2-week-old chickens inoculated orally with 5×10^4 sporulated oocysts of coccidia. One week lattar, birds were grouped into 6 groups each of 20 birds. five groups were inoculated orally with clostridia as described in former experiment, while 6 th group was left as control.

Evaluation of the effect of feed additives on clostridial infection:

A total of 240 one-day-old chicks were divided into 4 groups (60 birds each). All groups received ration mixed with feed additives along the experiment time. Group I received granulated bacitracin methyene disalicylate (BMD) 10% (Alpharma Animal Health Divission) as 440 gm/ton. Group II received Flavomycin 40 (Hoechst) as 125 gm/ton. Group III received Enramycin F40 (Takeda, LTD, Tokyo, Japan) as 125 Group IV received Lincomix (Upjohn Company, Kalamazoo, Mi) as 200 gm/ton. After three weeks, each group was divided into three subgroups (each of 20 birds) A,B,and C. Subgroup A were inoculated orally with 2 ml broth culture containing 1.8x109 CFU/ml of toxigenic Cl perfringens types A and C. Subgroup B were inoculated orally with 2 ml broth culture containing 1.8x10⁹ CFU/ml of non-toxigenic Cl perfringens types. Subgroup C was left as control. All inoculated groups received 3 successive doses at 2 days intervals. Birds were kept separately and received the treated ration for 4 weeks. Clinical signs, PM lesions, mortalities, average body weights and microbial recovery rates were recorded.

Sensitivity to antimicrobial agents:

The clostridial types used in this work were tested for their in vitro sensitivity to different antimicrobial agents using

blood agar plates under anaerobic conditions (Gas-Pack Anaerobic Jar, Baker Platinum LTD, London). The sensitivity was judged according to diameter and clearance of inhibition zone (Perclman et al, 1991).

RESULTS

Experimental infection:

Results are recorded in tables 1&2. Deaths occurred at first and second week postinfection. Mortality rates were 40%, 50%, 25%, 5% and 5% in chicks infected with Cl. perfringens toxigenic types A and C, non-toxigenic type, Cl colinum and Cl sporogens respectively, while in case of combined infection with clostridium and coccidia, mortality rates were much higher and recorded as 70%, 75%, 40%, 10% and 5% respectively. Clinical signs recorded as depression, ruffled feathers, decreased appetite and bloody or whitish diarrhea. Observed PM lesions were catarrhal to hemorrhagic,ulccerative and necrotic enteritis, together with mucosal thickening of small intestine (dudenum, jujunum, ileum and cecum). spleen and liver may be congested and enlarged with necrosis. Symptoms and lesions were more prominent in birds infected with toxigenic types of Cl perfringens with or without coccidia, while other types demonstrated mild signs and lesions. Clostridia were reisolated from intestinal tract of experimentally infected birds on cooked meat media and blood agar plates.

Evaluation of feed additives effect on chicks infected with clostridium:

Results are shown in Tables (3,4,5 and 6). All feed additives used in this study were effective in minimizing the mortality due to infection with toxigenic and non-toxigenic Cl perfringens. The average mortality rate due to infection with toxigenic Cl perfringens types A&C is 45%, while in case of non-toxigenic type is 25%. In case of birds received BMD, flavomycin, enramycin and lincomix, mortality rates due to infection with toxigenic Cl perfringens types A&C and non-toxigenic type were decreased from 45% & 25% to 30% & 5%, 35% & 5%, 25% & 0% and 25% & 5% respectively. A significant difference between lesions in birds receiving feed additives mixed ration and those received normal ration was

noticed on experimental infection, where intensity of lesion became much less severe. On the other hand, a significant improvement in weight gain was observed especially with flavomycin followed by BMD, enramycin and lincomix respectively.

Susceptibility of clostridial isolates to antimicrobial agents:

Results are illustrated in Table 7. Most of the tested clostridial isolates were highly sensitive to amoxycillin, pencillin, ciprofloxacin, enrofloxacin, flumequin, doxycycline and colistin sulphate. on the other hand, the isolates were moderately sensitive to crythromycin, chloramphenicol, oxytetracycline while it was resistant to neomycin, streptomycin and gentamicin.

DISCUSSION

Mortality rates were higher in birds subjected to combined infection with Clostridium and coccidia than those infected with Clostridium alone especially in birds infected with Cl perfringens. The observed clinical signs were manifested by whitish or bloody diarrhea and decreased body weight. On PM examination, different degrees of enteritis were demonstrated together with mucosal thickening and liver and spleen affections. Symptoms and lesions were more severe in case of toxigenic types of Cl perfringens than in case of experimental infection with non-toxigenic Cl perfringens, Cl colinum and Cl sporogenes, and getting more exaggerated after mixed infection with clostridium and coccidia. Similar results were described by Balauca et al (1976), Shane et al (1985), Fukata et al (1988) and Das et al (1997 b).

Mortality rates were 40%, 50%, 25%, 5% and 5% in chicks infected with *Cl. perfringens* toxigenic types A and C, non-toxigenic type, *Cl colinum* and *Cl sporogens* respectively, while in case of combined infection with clostridium and coccidia, mortality rates were much higher and recorded as 70%, 75%, 40%, 10% and 5% respectively. The formerly mentioned results were confirmed by Parish (1961), Kattich et al (1966), Hein and Timmus (1972), Bradley and Radharishnan (1973), Kondo et al (1988) and Baba et al (1997). Contrasting to our results, Long (1974) and Awad et al (1976) could not reproduce

the disease directly or indirectly with Cl perfringens or its toxins.

Regarding to our results, birds which have been received BMD (440 gm/ton), flavomycin (125 gm/ton), enramycin (125 gm/ton) and lincomix (200 gm/ton), showed lower mortality rates when experimentally infected with toxigenic CI perfringens types A&C and non-toxigenic type. Mortality rates were decreased from 45% & 25% to 30% & 5%, 35% & 5%, 25% & 0% and 25% & 5% respectively, Stutz et al (1983) and Das et al (1997c) found that bacitracin showed 100% efficacy against CI perfringens isolates in vitro, while Williams (1972) reported that bacitracin was active against Cl perfringens both in vitro and in vivo. Our results disagree with Benno et al. (1988) and Devriese et al, (1993) who detected aquired resistance against bacitracin by some isolates.

Regarding to the effect of lincomix, several authors reported on its good activity on clostridium, body weight, reducing morbidity and mortality rates, and clostridial shedding by infected birds (Hamdy et al, 1983 a&b; Secasiu, 1995 and Shen Jian Zhong et al, 1997). On the other side Watkins et al (1997) found that lincomycin appeared to be resisted by most strains of Cl perfringens.

In case of enramycin and flavomycin, our results agree with those recorded by Benno ct al., (1988), Sheldon and Essary (1982), Palic et al (1998) regarding to improvement of body gain, suppressing the shedding and reducing the mortality rate. Devriese et al., (1993) reported on flavomycin resistance by Cl perfringens.

The recorded susceptibility and resistance patterns of in vitro sensitivity test in present work had been fully described by several workers (Trishkins and Rokhmanina, 1973; Ibrahim, 1979; Kondo et al., 1988 and Secasiu, 1995).

REFERENCES

Awad, F.I.; Bassiouni, A.A.; Gadalla, M.S.; El-Sisi, M.A. and Hussein, A.Z. (1976): Studies on poultry anaerobes in Egypt. 1. An attempt to isolate anaerobic bacteria from the intestinal tract of normal and dead chickens. 2. The effect of alpha and beta toxins of Clostridium perfringens type A and C introduced by different routes. 3. The effect

- of ration on chickens infected with *Clostridium* perfringens type C. Egyptian Journal of Veterinary Science, 13(1): 1–22.
- Baba, E.; Ikemoto, T.; Fukata, T.; Sasai, K.; Arakawa, A. and McDougald, L.R. (1997): Clostridial population and the intestinal lesions in chickens infected with Clostridium perfringens and Eimeria necatrix. Veterinary Microbiology, 54 (3/4) 301–308.
- Balauca, N. (1976): Experimental reproduction of necrotic enteritis in the fowl. 1. Single and mixed infections with Clostridium perfringens and coccidia in chickens in cages. Archiv Für Experimentelle Veterinermedizin, 30 (6): 903–912.
- Benno, Y.: Endo, K. and Mitsuoka, T. (1988): Isolation of fecal Clostridium perfringens from broiler chickens and their susceptibility to eight antimicrobial agents for growth promotion. Japanese Journal of Veterinary Science, 50 (3): 832-834.
- Bradley, R.E. and Radhakrishnan, C.V. (1973): The role of cecal microbial flora in cecal coccidiosis of chicken. Poultry Science, 52(5): 2001.
- Science, 52(5): 2001.

 Cruickshank, R.; Duguid, J.P.; Marion, B.P. and Swain, R.H.A. (1975): Medical Microbiology. 12th edn Volume II Churchill Livingstone, Edinburgh, London & New York.
- Das, B.C.; Dutta, G.N.; Devriese, L.A. and Phukan, A. (1997 a): Antimicrobial susceptibility of Clostridium perfringens isolated from necrotic enteritis cases of poultry. Indian Veterinary Journal, 74 (7): 553–555.
- Das, B.C.; Dutta, G.N.; Phukan, A. and Mukit, A. (1997 b):

 Necrotic enteritis in chickens due to field isolate of Clostridium perfringens type A. Indian Journal of Veterinary Pathology, 21(1): 27–29.
- Das, B.C.; Gupta, G.N. and Phukan, A. (1997 c): Experimental production and treatment of necrotic enteritis in fowl. Indian Journal of Poultry Science, 32(1): 59–66.
- Devriese, L.A.; Daube, G.; Hommez, J. and Haesebrouck, F. (1993): In vitro susceptibility of Clostridium perfringens isolated from farm animals to growth-enhancing antibiotics. Journal of Applied Bacteriology, 75(1):55–57.

- Fukata, T.; Hadate, Y.; Baba, E.; Uemwa, T. and Arakawa, A. (1988): Influence of Clostridium perfringens and its toxin in germ-free chickens. Research in Veterinary Science, 44(1): 68 70.
- Hamdy, A.H.; Thomas, R.W.; Kratzer, D.D. and Davis, R.B. (1983a): Lincomycin dose response for treatment of necrotic enteritis in broilers. Poultry Science, 62(4): 585– 588.
- Hamdy, A.H. Thomas, R.W.; Yancey, R.J. and Davis, R.B. (1983 b): Therapeutic effect of optimal Lincomycin concentration in drinking water on necrotic enteritis in broilers. Poultry Science, 62(4): 589–591.
- Hein, H. and Timmus, L. (1972): Bacterial flora in the alimentary tract of chickens infected with Eimeria brunetti and in chickens immunized with Eimeria maxima and cross infected with Eimeria brunetti. Exp. Parasitol., 31: 188– 193.
- Ibrahim, A.A. (1979): Anaerobic microflora of the small intestine of chickens. Ph.D. Thesis presented to Faculty of Veterinary Medicine, Assiut University.
- Ibrahim, R.S.; Ibtihal, M. Monazi and Soliman, A.M. (2001): Epidemiological studies on intestinal closteridial infection in broilers in Assiut and El-Menia governorates. Assiut Vet. Med. J. 44(88), 291-305.
- Katitch, R.; Chibalitch, S.; Kositch, L.J.; Voukitchevitch, Z.; Djoukitch, B. and Tomanovitich, B. (1966): Role of Eimeria tenella and E. necatrix in the epidemiology of enteritis caused by Clostridium perfringens type C in chicks. Acad. Vet. Fr., 39: 101–107.
- Kondo, F.; Tottori, J. and Soki, K. (1988): Ulcerative enteritis in broiler chickens caused by Clostridium colinum and in vitro activity of 19 antimicrobial agents in tests on isolates. Poultry Science, 67(10): 1424–1430.
- Long, J.R. (1974): Studies on necrotic enteritis in broiler chickens with emphasis on the role of Clostridium perfringens.
 Dissertation Abstracts International, 35 B, (5) 2503.
- Palie, T.; Stankovice, G. and Novakovic, Z. (1998): Effect of simultaneous application of Sacox and Flavomycin

- preparations on broiler performance and health. Zivinarstvo, 33(9/10): 211-214.
- Parfitt. (1958): Cited by Abdel-Rahman (1972). Thesis presented to Faculty of Veterinary Medicine, Assiut University.
- Parish, W.E. (1961): Necrotic enteritis in fowl. J. Comp. Path. Therap., 71: 377-413.
- Perelman, B.; Mints, S.; Zjut, M.; Kuttin, E. and Machny, S. (1991): An unusual Clostridium colinum infection in broiler chickens. Avian Pathology, 20: 475 480.
- Secasiu, V. (1995): Chemosensitivity of Clostridium perfringens strains isolated from poultry. Revista Român, de Medicina Veterinara, 5(1): 31–37.

 Shane, S.M.; Gyimah, J.E.; Harrington, K.S. and Snider, T.G.
- Shane, S.M.; Gyimah, J.E.; Harrington, K.S. and Snider, T.G. (1985): Etiology and pathogenesis of necrotic enteritis. Veterinary Research Communications, 9(4): 269-287.
- Sheldon B. W. and Essary, E.O. (1982): Effect of antibiotics on intestinal microflora and flavor of broiler meat. Poultry Science, 61: 280-287.
- Shen JianZhong; Xiao XiLong; Yang HanChun; Wang YanJun and Cha ZhenLin (1997): Pharmacodynamic study of lincomycin soluble powder for necrotic enteritis in chickens. Acta Veterinaria et Zootechnica Sinica, 28(5): 464-468
- Stutz, M.W.; Johnson, S.L. and Judith, F.R. (1983): Effects of diet and bacitracin on growth, feed efficiency, and populations of Clostridium perfringens in the intestine of broiler chicks. Poultry Science, 62 (8) 1619 1625.
- Trishkins, E.T. and Rakhmanina, I.A. (1973): Sensitivity to antibiotics of strains of Clostridium perfringens isolated from fowls. Truda. Vsesoyuznogo Instituts Eksperimental Noiveterinarii 41: 243 248.
- Watkins, K.L.; Shryock, T.R.; Dearth, R.N. and Saif, Y.M. (1997):

 In-vitro antimicrobial susceptibility of Clostridium perfringens from commercial turkey and broiler chicken origin. Veterinary 'Microbiology, 54(2): 195–200.

 Williams, S.H. (1972): The antibacterial activity of nitrovin in
- Williams, S.H. (1972): The antibacterial activity of nitrovin in vitro; the effect of this and other agents against Clostridium welchii in the alimentary tract of chickens. Veterinary Record, 90(10): 310 312.

Table (1): Results of experimental infection of chicks with commonly isolated clostridial species.

Mean	body	240	238	271	279	275	- 00
	Mortality rate	40	20	2.5	ς.	w,	c
	Total no. of deaths	00	10	v.	-	-	0
ection	함	0	0	0	0	0	U
Weekly deaths post infection	51 51	0	0	0	0	0	0
y deaths	2110	5	(C)	ro.	0	0	0
Week	<u> 21</u>	9	7	CI	800	-	0
	Route of inoculation	Oral	Oral	Oral	Oral	Oral	Oral
No of	inoculated	20	20	20	20	20	30
	Dose (ml.)	1.8×109	1.8×10 ⁹	1.8×10°	103	1.5×10°	Sterile broth
	organisms	Cl. perfeingein, type A	('I. perfringens type C	Non-toxigenic (7. perfringens	CL colimum .	(Т. хрогоденк	Control
OH	Group	-	rı	re,	π	æ,	9
		2	63				

Table (2): Results of combined experimental infection of chicks^(a) with coccidia and commonly isolated clostridial species

- 1					Week	ly deaths	Weekly deaths post infection	ection			
Group no	Inoculated	Dose (mf.)	No. of inoculated birds	Route of inoculation	21	Dir.C	Pic C	#	Total no. of deaths	Mortality rate %	weight weight gen
and .	Cl. perfringens type A	1.8×10°	20	Oral	6	V)	0	0	7	70	229
CI	(7. perfringens type C	1.6×10°	20	Oral	-	4	0	0	15	7.5	230
60	Non-toxigenic (T. Perfringens	1.8×10°	30	Oral	40	m	0	0	∞	40	242
4	Cl. colinum	107	20	Oral		-	0	0	C1	01	253
us	Cl. sporogens	1.5×10°	20	Oral		0	0	0	-	\$	258
9	Control	Sterile broth	20	Oraf	0	0	0	0	0	0	272

N.B. (a) Previously infected with 5×104 sporulated oocysts of Eimeria at 14-day-old.

Table (3): Results of experimental infection of chicks received Bacitracine Methylene Disalicylate (BMD).

u dno.i	Inoculated		No of	Route of inoculation	7_	a	70,	B		Nonality	1000
0		Dose (ml.)	inoculated			ť	100	El	of deaths		body weight gm
2000	Cl. perfimpens type A. C	,01×81	20	Oral	5	-	0	0	9	30	297
6.11	Non-toxigenic CI. perfimgens	1.8×10"	20	Oral	-	0	0	0	-	ς.	308
m	Control	Sterile broth	20	Oral	0	0	0	0	0	0	31.5
(1)					Week	ly death:	Weekly deaths post infection	fection			Mean
a quorið -	Inoculated	Dose (ml.)	No or inoculated birds	Route of inoculation	71_	21 71	36	4	Total no of deaths	Monality rate %	body weight
-	(T. perfringens type A. C	1.8×10°	50	Oral	v.	CI	0	0	7	322	302
CI	Non-toxigenic (7. per/ringens	1 8×10 ⁴	50	Oral	-	0	0	0	-	v.	317
50	Control	Sterile broth	20	Oral	0	0	0	C	0	0	320
			STEEN.	200.00	1	1138	10		2	5	16.00

Table (5): Results of experimental infection of chicks received Enranycin.

C.1. perfringens Dose (ml.) inoculated Route of 134 2 m² C.1. perfringens 1.8×10" 20 Oral 4 1 Von-toxigenic 1.8×10" 20 Oral 0 0 C.2. perfringens 0.8×10" 20 Oral 0 0 C.2. perfringens 0.8×10" 20 Oral 0 0 C.2. perfringens 0.8×10" 0.0 C.2. perfringens 0.9×10" 0.0 C.3. perfringens 0.9×10" 0.0	Oral collation 1 Oral collation Collad Coll	2 2 2 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0	हा _र 0 0 0	4 0 0 0	Total no of deaths of deaths	Mortality rate % 25 0 0 0	Medyll gm gm gm 312 315
Cl. perfringens 18×10° 20 Oral Non-toxigenic Cl. perfringens 18×10° 20 Oral Control Sterile broth 20 Oral Control Sterile broth 20 Oral Inoculated Dose (ml.) inoculated inoculation birds inoculation birds 18×10° 20 Oral Non-toxigenic Cl. perfringens 18×10° 20 Oral Oral Cl. perfringens 18×10° 20 Oral	Oral C Oral C Oral C Oral C Www.	t 1 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0	0 0 0	0 0 0	\$ 0 0	255	315 312
Non-toxigenic 18x10° 20 Oral Control Sterile broth 20 Oral Control Sterile broth 20 Oral Inoculated No. of Route of Organisms Dose (ml.) inoculated inoculation birds Cl. perffungens 1.8x10° 20 Oral Non-toxigenic Cl. perffungens 1.8x10° 20 Oral	Oral COral C	0 0 0 0 comix.	0 0	0 0	0 0	0 0	315
Locatrol Sterile broth 20 Oral Local Control Sterile broth 20 Oral Local Laber (6): Results of experimental infection of chicks received noculated organisms Lipserfringens	received lim We of Wellition 13	comix.	0	0	0	0	200
lnoculated bose (ml.) inoculated Route of crises received organisms Dose (ml.) inoculated inoculation birds type A, C 1.8×10° 20 Oral Non-toxigenic C. perfiningens 1.8×10° 20 Oral	received linu. W. ute of	comix.					
Inoculated Dose (ml.) inoculated Route of birds inoculation 1st 2ml birds (7. perffulgens 18×10° 20 Oral 3 2 Non-toxigenic (7. perfulgens 1.8×10° 20 Oral 0 1			ths post ini	fection			Adam
1.8×10° 20 1.8×10° 20			3104	40	Total no. of deaths	Mortality rate %	body weight
1.8×10° 20)ral 3	(~)	0	0	No.	25	162
	oral 0	-	0	0		S	307
Control Sterile broth 20 Oral 0 0 0		0	0	0	0	0	

Spent C. perfengent C. colinum C. spongenes	Antimicrobial	timicrobial Concentrations Isolated clostridial species	Isolated clostridial species	tial species					
15 yg 25 yg 4+ + + + + + + + + + + + + + + + + + +			C. parfringens Type A (5)		Ci perfringens Type D (3)	Non-toxigenic CLiperfingens (5)	Cl. colinum (\$)	Cl.sporogenes	Cl. spirafor
25 pg 25	Erythromycin	15 не	++	+	+	+ +	177		1
2.5 prog 2.5	Colistin sulphate	25 µg	+ +	+	+++	+ +	+ +	+ #	+
10 Mg +++ +++ +++ +++ +++ +++ +++ +++ +++	Neomycin	30 µg				19			+
10 pg	Amoxycillin	25 µg	+ +	+ + +	÷	4.4	H + +	1	
10 pg	Oxytetracycline	30 Hg	1	+	+	1	10000		+
### ### ### ### ### ### ### ### ### ##	Ampicillin	10 ид	+ + +	+ + +	++++	7 7	- 1		‡
5 pg + + + + + + + + + + + + + + + + + +	Streptomycin	10 µg	- 1					+	+
30 pg ++ ++ ++ ++ ++ ++ ++ ++ ++ ++ ++ ++ ++	Ciprofloxacin	S 48	+++	+++	+ +	. +			×
15 18 18 18 18 18 18 18 18 18 18 18 18 18	Dovycycline	30 µg	+ +	+++	++	H H		+	+
+++ ++ ++ ++ ++ ++	Enrofloxacm	5 µg	+ + +	+	+++	1 1	h -	++	+
## ## ## ## ## ## ## ## ## ## ## ## ##	Flumequin	15 429	+ +	++	+++++++++++++++++++++++++++++++++++++++		+ :	++++	+
	Chloramphenicol	30 µg	+ +		+		+ + +	+	+
	Gentamycin	10 µg		,	. ()		+		+

resistant +; weak + +; moderately susceptible
 Figures to parenthesis indicate number of examined strains.