## DIAGNOSTIC AND GENETIC STUDIES OF *THEILERIA ANNULATA* WITH SPECIAL REFERENCE TO GENETIC POLYMORPHISM OF *THEILERIA ANNULATA* MEROZOITE SURFACE (TAMS-1) ANTIGEN

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#### **ABSTRACT**

Received at: 23/12/2014

Accepted: 17/1/2015

Diagnosis of bovine theileriosis was carried out by blood film and polymerase chain reaction-PCR methods. In the current study, Theileria annulata merozoite piroplasm Surface Antigen (Tams-1) target based PCR was used for specific detection of T. annulata infection in cattle from different localities in Upper Egypt then followed by cloning and sequencing of this gene then alignment of all obtained sequences and their translated amino acids to studying its polymorphism among different local strains. The results of this study concluded that blood film is still important for diagnosis. (Tams-1) target based PCR test was more sensitive than blood film. The infection rates with T. annulata in the examined cattle were 46.19% and 28.57% by using Tams-1 target based PCR and thin blood film, respectively. The (Tams-1) gene sequencing, alignment and translation into amino acids concluded that Tams-1 sequences and its translated amino acids were highly variable. This makes it not recommended for use in vaccination and/or serological diagnostic tests. The sequences and their translated amino acids were deposited in the GenBankTM databases and available under accession numbers as following (GenBank<sup>TM</sup>: KJ021626, GenBank<sup>TM</sup>: KJ021627, GenBank<sup>TM</sup>: KJ021628, GenBank<sup>TM</sup>: KJ021629).

Keywords: Diagnosis; Blood film, PCR, Tams-1; Theileria annulata.

#### INTRODUCTION

Theileriosis is the most economically important disease of cattle; it causes severe losses in livestock production in Egypt Al-Gaabary, (1995) and AL-Hosary, (2009). (Tams-1) is an antigen of T. annulata, it is one of the merozoite piroplasm surface antigen (MPSA) (Katzer et al., 2002). Tams-1, polypeptide characterized by variable molecular mass it is between 30 to 32 kDa (Katzer et al., 1998). This gene encoded for protein of 281 amino acids (aa) containing a putative hydrophobic N-terminal signal peptide, which is highly variable. Several studies reported using of (Tams-1) target based PCR for sensitive and specific amplification of T. annulata DNA from blood of cattle and ticks D'Oliveira et al. (1995), Gubbels et al. (2000), Shiels et al. (1994, 2000) and Kirvar et al. (2000). The present study

aimed to compare between blood films and (Tams-1) target based PCR as a diagnostic tools for diagnosis of bovine theileriosis. Genetic polymorphism of Tams-1 gene in different isolates of *T. annulata* from different localities in Upper Egypt was monitored.

#### **MATERIALS and METHODS**

Since August 2010 till August 2013, a total number of 210 cattle belong to different localities in Upper Egypt were randomly selected to this study. Out of them, 11 positive samples were subjected to cloning, sequencing, amino acids translation and multiple sequences alignment. Two types of samples were collected from each animal; one blood sample was collected directly from the ear vein and used for preparation of blood films, another whole blood sample was collected on Ethylene Diamine Tetra-

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Acetic acid (EDTA) as anticoagulant (1mg/1ml) by direct puncture of the jugular vein and marked with numbered labels then stored at (-20°c) till use in DNA-extraction Charles, (2002).

#### **Blood film:**

Thin blood films were prepared immediately post ear vein puncture then fixed and stained by Giemsa stain 8%. Three thin blood films from each animal were examined for detection of piroplasms or trophozoite (Coles, 1986).

#### **Polymerase chain reaction (PCR):**

(Tams-1) target based polymerase chain reaction was used for molecular diagnosis of *T. annulata* infection in examined blood samples.

#### **DNA Extraction and polymerase chain reaction:**

DNA extraction from whole blood was carried out according to manufacturer's instructions of commercial kits (according to QIA amp blood kit, Qiagen, Ltd, UK)

### DNA amplification by polymerase chain reaction (PCR) using Tams-1 target-based primer:

For the standard PCR, primer **Tams1 F** (5\ ATG CTG CAA ATG AGG AT) and **Tspms1 R** (5\GGA CTG ATG AGA AGA CGA TGA G), Amplifying a (785 bp) fragment of *T. annulata* 30 KDa major merozoite surface antigen gene Kirvar *et al.* (2000), Murat *et al.* (2008) and AL-Hosary, (2009).

**Cycling conditions:** 94°C for 5 min., followed by 37 cycles consisting of 1 min at 94°C, 1 min. at 55°C, 2 min. at 72°C and final extension step at 72°C for 10 min. longer then, the samples were stored at 4°C until use. The cycling condition carried out in thermocycler TECHNE TC–312. In addition to the samples positive control sample contain "DNA from *T. annulata* infected lymph" and negative control sample (DNase free water), then all were included in the amplification step.

Gel Electrophoresis: The chamber was connected to 75 volt power supply for 90 min., 10μl of each PCR product were separated by electrophoresis on 1.8% agarose gel (GX 040.90, Gen AGarose, L.E., Standard DNA /RNA agarose, Molecular Biology Grade, Inno-Train Diagnostic, D-61476, Kronberg/Taunus) Containing Ethidium bromide as 1 μl /ml electrophoresis buffer. The result obtained through high performance ultraviolet transilluminator,

the specific band were detected by using DOC-It ® LS, Image acquisition-software, (UVP, INC, UK) at 785 bp.

## Cloning of PCR product, DNA sequencing and data analysis

#### **Cloning:**

Ligation of the PCR product was carried out using pDrive Cloning vector according to manufacturer's instructions of QIAGEN Kit (Cat. NO.231224), Qiagen, Ltd, UK. Transformation by using QIAGEN Kit was carried out according to the manufacturer's instructions of Qiagen, Ltd, UK then Platting/Culture and the selection of clones was depended on (Blue / White Colony Selection).

**Preparation of the samples for sequencing:** The samples measured for detection the amount of dDNA by using eppendorf-Biophotometer, NO.6131 then adjust the sample concentration to  $1\mu g/\mu l$  in total volume 15  $\mu l$  in two tubes one for forward primer and the second for reverse primer.

Sequencing and data analysis: The amplicons were sequenced in both directions in the research center, Borstel, Germany by using Sanger 3730xl, Applied Biosystems (Life Technologies) as previously described by Holman, *et al.* (2005) and Schoelkopf *et al.* (2005). Tams-1 sequences were determined from alignments of 3 to 12 clones from each isolate and then the sequence results were subjected to Basic Local Alignment Search Tool (BLAST) similarity searches according to Altschul *et al.* (1990).

#### Sequences alignment and translation:

All the sequences were alignment together by using Clustal W, also translated into its amino acids (aa) by using Expasy.translate software and then alignment the obtained amino acids (aa) together by the same alignment program.

#### **RESULTS**

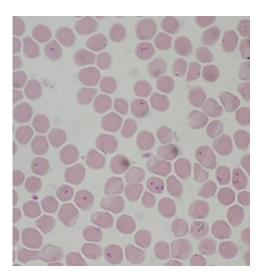
In the current study 210 cattle were randomly selected and subjected to examination for diagnosis of *T. annulata* infection.

#### Blood film and PCR:

The results revealed that the infection rate was 28.57% while the infection rate according Tams-1 target based PCR was 46.19% (Table 1 and Fig. 1, 2).

Table 1: Infection rate by using blood film and Tams-1 target based PCR

# Giemsa stained thin blood filmTams-1 target based PCRNo. of tested animalsPositive%No. of tested animalsPositive%2106028.572109746.19



**Figure 1:** Blood film stained with Giemsa stain demonstrating intracellular trophozoit "signet ring" of *T. annulata* (X100)

**Figure 2:** Agarose gel electrophoresis of (Tams-1) target based PCR amplified DNA from *T. annulata* infected and non-infected animals. M, 100 bp plus Marker DNA Lanes 1, 2, 4, 5 and 7 positive samples yielded 785 bp PCR product, lanes 3 and 6 Negative samples. Lanes 8 and 9 were the positive and negative control, respectively.

#### Sequences analysis:

The obtained results revealed that the local isolates of *T. annulata* are characterized by the polymorphism of Tams-1 gene. This finding obtained through alignment of the different obtained sequences together and with those in the gene bank by using (Clustal W) (Fig. 3). This was also confirmed after translation of the each sequence to its amino acids

(aa) by using Expasy.translate software and then alignment together (Fig. 4). The obtained sequences and their translated amino acids were deposited in the GenBankTM databases and available under accession numbers as following (GenBank<sup>TM</sup>: KJ021626, GenBank<sup>TM</sup>: KJ021627, GenBank<sup>TM</sup>: KJ021628, GenBank<sup>TM</sup>: KJ021629).

CLUSTAL 2.1	multiple sequence alignment
KJ021627 KJ021629 KJ021628 KJ021626	ATGCTGCAAATGAGGATGAAAAGAAAAAGGA <mark>G</mark> GAAAA———————AAAAGATGTTG ATGCTGCAAATGAGGATGA <mark>T</mark> AAGAAAAAGGAAGAAAA <mark>AAAAGATGAAAA</mark> AAAAGATGTTG ATGCTGCAAATGAGGATGATAAGAAAAAGGAAGAAAA <mark>AAAAGATGAAAA</mark> AAAAGATGTTG ATGCTGCAAATGAGGATGAAAAAAAAAA
KJ021627 KJ021629 KJ021628 KJ021626	TTCTTGATGTTACTCTCACTTCATGTGATAATGTAACCTTTAAAGTCGTCGACTCTAACA TTCTTGATGTTCTTGCCACTTCATGTGAGAATGTAGTCTTTCATACTCCCGACTCTAACA TTCTTGATGTTCTTGCCACTTCATGTGAGAATGTAGTCTTTCATACTCGCGACTCTAACA TTCTTGATGTTACTGCCACTTCATGTGAGAATGTAGCCTTTTGATAACCGCGACCCTAACT ******************************
KJ021627 KJ021629 KJ021628 KJ021626 * * * * * * * *	CCACTGAGTTAACTGTCGCGGATGGCTACCGTTTCAAGACCCTTAAGGTCGGAGACAAGA CTACTGAATTACTTGTCAAAGACGGCTTCCGTTTCAAGACCCTTAAGGTCGGAGACAAGA CTACTGAATTACTTGTCAAAGACGGCTTCCGTTTCAAGACCCTTAAGGTCGGAGACAAGA CCGTGGTGTTTAACTGCGGGAAGGCTACCGTTTCAAGACCCTTAAGGTCGGAGACAAGA ** *** ****************************
KJ021627 KJ021629 KJ021628 KJ021626	CCTTGTTCAATGTTGACACCTCAAAACATACCCCAGTACAGGCATTCAAACTTAAGCATG CATTGTTCAATGTTGACACCTCAAAACATACCCCAGTACAGGCATTCAAACTTAAGCATG CATTGTTCAATGTTGACACCTCAAAACATACCCCAGTAAAGGCATTCAAACTTAAGCATG CCTTATTCAATGTTGACACAGAAACATACCCCAGTAAAGGCATTCAAACTTAAGCATG **** *******************************
KJ021627 KJ021629 KJ021628 KJ021626	AATCCGATGAGTGGTTCAGACTTAATCTTCA <mark>TG</mark> CTGCCCAGCCAAAGATGTTCAAGAAGA AATCCGATGAGTGGTTCAGACTTAATCTTCACCCTGCCCAGCCAAAGATGTTCAAGAAGA AATCCGATGAGTGGTTCAGACTTGATCTTCACCCTGCCCAGCCAAAGATGTTCAAGAAGA AATCCGAGGAGGTGGTCAGGCTTGATCTTCACCCGCCCAACCAA
KJ021627 KJ021629 KJ021628 KJ021626	AGGGAGACAAGGAATATTCTGAGG <mark>T</mark> CAAATTCGAGACCTACTACGATGAAGTCTTGTTCA AGGGAGACAAGGAATATTCTGAGG <mark>T</mark> CAAATTCGAGACCTACTACGATGATGTCTTGTTCA AGGGAGACAAGGAATATTCTGAGTCCAAATTCGAGACCTACTACGATGAAGTCTTGTTCA AGGGAGACAAGGAATATTCTGAGTCCAAATTCGAGACCTACCT
KJ021627 KJ021629 KJ021628 KJ021626 **** **** ****	AGGGAAAAT <mark>CA</mark> GCCAAGGAACTAGATGCTTCCAAGTTCGAAGATACA <mark>T</mark> CTTTGTTCACC <mark>T</mark> AGGGAAAAT <mark>CA</mark> GCCAAGGAA <mark>T</mark> TAGATGTTTCCAAGTTCGAAGATACAGCTTTGTTCACC <mark>T</mark> AGGGAAAATCCGCCAAGGAACTAGATGTTTCCAAGTTCGAAGATACAGCTTTGTTCACCG AGGGGAAATCCGCCAAGGAACTAGATGTTTCCAA <mark>A</mark> TTCGAAGATACAGCTTTGTTCACCG
KJ021627 KJ021629 KJ021628 KJ021626 *******	CCTCCGCCTTCGGCACTGGAAAGATGTACACCTTTAAAAAGGAATTTAAACCTTCCAAAG CCTCCGCCTTCGGCACTGGAAAGATGTACACCTTTAAAAAGGAATTTAAACCTTCCAAAG CCTCCGCCTTCGGCACTGGAAAGAAGTAGAACCTTTAAAAAGGATTTCAAACCTTCCAAAG CCTCCGCCTTCGGCACTGGAAAAAAGTAACACCTTTAAAAAGGATTTCAAACCTTCCAAAG
KJ021627 KJ021629 KJ021628 KJ021626	TCACCTTCGACAAGAAAGAAGTCGGAAAGCCAAACAATGCCAAGTATCTTGAAGTTGTTG TCACCTTCGA <mark>C</mark> AAGAAAGAAGTCGGAAAACCAAACAATGCCAAGTATCTTGAAGTTGTTG TCACCTTCGAAAAGAAAGAAGTCGGAAAACCAAACAATGCCAAGTATCTTGA <mark>T</mark> GTTTTCG TTCCTCCAAAAAGAAAGAAGTCGGAAAAACAAATAATGCCAAGTATCTTGATGTTTTCG *******************************
KJ021627 KJ021629 KJ021628 KJ021626	TCTTTGTTGGTTCTGATTCCAAGAAGTTAGTCAAACTCTACTACTTCTATACCGGTGACT TTTTTTGTTGGTTCTGATTCCAAGAAGTTCGTCAAACTCTACTACTTCTATACCGGTGACT TCTTTGTCAGTGCTGATTCCAAGAAGGTCGTCAGACTCGACTACTTCTATACCGGTGACT TCTTTGTCAGTGCTGATTCCAAGAAGGTCGTCAGACTCGACTACTTCTATACCGGTGACT ************************************
KJ021627 KJ021629 KJ021628 KJ021626	CAAGGTTGAAGGAGACCTACTTCGAGCTTAAGGACGATAAGTGGGTACAAATGTCACAGG CAAGGTTGAAGGAGACCTACTTCGAGCTTAAGGACGATAAGTGGGTTCAAATGACACAGG CAAGGTTGAAGGAGACCTACTTCGAGCTTAAAGGACGATAAGTGGGTGCAAATGTCACAGG CAAGGTTGAAGGAGACCTACTTCGAGCTTAAGGACGATAAGTGGGTGCAAATGTCACAGG
	CAGATGCAAACAAGGCCTTGAATGCCATGAACTCATCCTGGTCCACCGATTACAAACCAG CAGATGCAAACAAGGCCTTGAATGCCATGAACTCATCCTGGTCAACCGATTACAAACCAG CAGATGCAAACAAGGCCTTGAACGCCATGGACTCATCATGGTCGGCCGATTACAAACCAG CAGATGCAAACAAGGCCTTGAATGCCATGGACCCAACTGGTCATCAGATTCAGATTCAACCAG ********************************
KJ021626	TTGTCGACAAGTTCTCTCCCCTTGCAGTCTTCGCCTCAGTACTCATCGTCTTCTCATCAG TTGTCGACAAGTTCTCCCCCCTTGCAGTCTTCGCCTCAGTACTCATCGTCTTCTCATCAG TTGTCGACAAGTTCTCTCCCCCTTGCAGTCTTCGCCTCAGTACTCATCGTCTTCTCATCAG TTGTCGACAAGTTCTCCCCCCTTGCAGTCTTCGCCTCAGTACTCATCGTCTTCTCATCAG * ***********************************
KJ021627 KJ021629 KJ021628 KJ021626	TCCAATCAC TCCAATCAC TCCAATCAC TCCAATCAC

**Figure 3:** Sequences' alignment for *T. annulata* Tams-1 gene.

CLUSTAL 2.1 multiple sequence alignment

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KJ021626
                      MFKKKGDKEYSE<mark>S</mark>KFETYYD<mark>D</mark>VLFKGKSAKELDVSKFEDT<mark>A</mark>LFT<mark>A</mark>SAFGTGKKYTFKK<mark>D</mark>F
KJ021628
                      MFKKKGDKEYSE<mark>S</mark>KFETYYDEVLFKGKSAKELD<mark>V</mark>SKFEDT<mark>A</mark>LFT<mark>A</mark>SAFGTGKKYTFKK<mark>D</mark>F
                      MFKKKGDKEYSEVKFETYYDEVI,FKGKSAKEI,D<mark>A</mark>SKFEDTSI,FTSSAFGTGKMYTFKKEF
кл021627
KJ021629
                      MFKKKGDKEYSEVKFETYYDDVLFKGKSAKELDVSKFEDTALFTSSAFGTGKMYTFKKEF
KJ021626
                      KPSKV<mark>L</mark>F<mark>E</mark>KKEVGKPNNAKYLDV<mark>F</mark>VFVS<mark>A</mark>DSKKVVRL<mark>D</mark>YFYTGDSRLKETYFELKDDKWV
KJ021628
                      KPSKVTF<mark>E</mark>KKEVGKPNNAKYLDV<mark>F</mark>VFVS<mark>A</mark>DSKKVVRL<mark>D</mark>YFYTGDSRLKETYFELKDDKWV
                      KPSKVTFDKKEVGKPNNAKYLEVVVVFVGSDSKK<mark>L</mark>VKLYYFYTGDSRLKETYFELKDDKWV
KPSKVTFDKKEVGKPNNAKYLEVVVFVGSDSKKFVKLYYFYTGDSRLKETYFELKDDKWV
KJ021627
KJ021629
                                             *****
KJ021626
                      QMSQADANKALNAMDPTWSSDYKPVVDKFSPLAVFASVLIVFSSVQS
KJ021628
                      QMSQADANKALNAMDSSW<mark>LA</mark>DYKPVVDKFSPLAVFASVLIVFSSVQS
KJ021627
                      QMSQADANKALNAM<mark>N</mark>SSWSTDYKPVVDKFSPLAVFASVLIVFSSVQS
         KJ021629
```

Figure 4: The alignment of translated (aa) of *T. annulata* Tams-1 gene

#### **DISCUSSION**

The infection rate with *T. annulata* of the examined cattle was 28.57% by thin blood film and 46.19% by Tams-1 target based PCR. These obtained results revealed that molecular technique was more sensitive it was in agreement with D'Oliveira *et al.* (1995), Aktas *et al.* (2002, 2006), Tabidi *et al.* (2006), Azizi *et al.* (2008), Durrani and Kamal, (2008) and AL-Hosary, (2009). They reported that blood smear is a quick and cheap for diagnosis of *T. annulata* infection in cattle, and PCR test was more sensitive and specific test.

Regarding to the genetic polymorphism of (Tams-1) gene that responsible for T. annulata Merozoite piroplasm surface antigen, the results revealed that Tams-1 gene from different T. annulata isolates are differ from each other. This agree with Manuja et al. (2006) who reported that (Tams-1) is a polymorphic gene and it was studied amongst four T. annulata isolates collected from three different parts of India. The four isolates of T. annulata were different from each other and may be express different antigenic determinants on their cell surface. Furthermore, Altay et al. (2007) and Esmaelizad et al. (2001) reported that the polypeptide (Tams1) is a highly variable immunodominant major merozoite piroplasm surface antigen of the protozoan parasite T. annulata and its associated with the evasion of the bovine immune system. Another possible explanation for the high levels of genetic diversity in Tams-1 gene among T. annulata isolates was reported by Gubbels et al. (2000) who concluded that it may be due to random mutation of nucleotides during asexual reproduction. This makes it difficult to use in vaccination or serological diagnostic assays. It concluded that Tams-1 gene merozoite surface antigens in T. annulata characterized by high level of genetic polymorphism making it unfit for serological diagnostic tools or vaccine production.

#### **ACKNOWLEDGMENT**

We would like to express sincere thanks for DFG project "Molecular epidemiology network for

promotion and support of delivery of life vaccines against *Theileria parva* and *Theileria annulata* infection in Eastern and Northern Africa" (DFG AH 41/7-1) for supporting this work.

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#### دراسات تشخيصيه وجينيه للثيليريا الحلقيه مع اشاره خاصه للتنوع الجيني في انتجين التامز-١ للثيليريا الحلقيه

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استهدفت الدراسه الي استخدام الطريقه التقليديه متمثله في المسحات الدمويه المصبوغه بصبغه الجيمسا بالاضافه الي تفاعل البلمره المتسلسل باستخدام بأدئ التامز - ١ لتشخيص الاصابه بمرض ثيليريا الماشيه في الابقار في صعيد مصر. واوضحت النتائج ان طريقه المسحه الدمويه مازالت تتمتع باهميه اكلينيكيه ولكن استخدام طريقه تفاعل البلمره المتسلسل باستخدام بادئ التامز- آهي الاكثر حساسيه. بادئ التامز-١ مستخلص من جين التامز-١ وهو جين مسئول عن الانتجين السطحي الخاص بالطفيل. وكان من أحد اهداف هذه الدراسه ايضا تتبع هذه الخاصيه في جين التامز-١ المعزول من طفيل الثيلريا الحلقيه المُعزوله من الابقار في صعيد مصر وتم تأكيد هذا الننوع الجيني وتم نشر المتواليات الجينيه وتتابعات الاحماض الامينيه (البروتين) الخاص بها في بنك الجينات تحت ارقام الانضمام التاليه: KJ021266, KJ021267, KJ021268, KJ021269 ان هذا الجين له القدره على التغير من نفسه بالشكل الذي يجعل البروتين الناتج عنه مختلف وبالتالي يعمل علي حمايه الطفيل من الاجسام المضاده التي ينتجها الجهاز المناعي الخاص بالحيوان المصاب ولذلك لا ينصح باستخدام هذا الجين في طرق التشخيص السير ولوجيه او التحصين ضد المرض.