

CANDIDA PARAPSILOSIS COMPLEX SPECIES AND ANTIFUNGAL SUSCEPTIBILITY PROFILE IN PATIENTS OF INTENSIVE CARE UNITS OF MANSOURA UNIVERSITY HOSPITALS.

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ABSTRACT

Candida parapsilosis is an important non-albican species responsible for invasive fungal infections and hospital acquired infections especially in critical care patients. *C. parapsilosis* complex has been renamed according to genetic bases into 3 different species *C. parapsilosis sensu stricto*, *C. metapsilosis* and *C. orthopsilosis*. This study was designed to describe the distribution and antifungal susceptibility profile of the three members of *Candida parapsilosis* complex among patients of intensive care units (ICUs) in Mansoura University Hospitals. *Candida parapsilosis* was identified by Analytic Profile Index (API) 20 C. *C. parapsilosis sensu stricto*, *C. orthopsilosis* and *C. metapsilosis* were recog-

nized according to the secondary alcohol dehydrogenase (SADH) restriction pattern using *BanI* restriction enzyme. Antifungal susceptibility testing was performed by E test. A total of 68 *C. parapsilosis* isolates were included in this study. Sixty-two isolates (91.2%) were identified as *C. parapsilosis sensu stricto*, 4 (5.9%) were identified as *C. orthopsilosis*, and 2 isolates were identified as *C. metapsilosis* (2.9%). All isolates of the *C. parapsilosis* complex species were sensitive to amphotericin B. Fifty isolates (80.6%) of *C. parapsilosis sensu stricto* were susceptible to fluconazole; 7 isolates (11.3%) were susceptible-dose dependent (SDD) to fluconazole, and 5 isolates (8.1%) were resistant to fluconazole. Most of *C. parapsilosis*

sensu stricto isolates were sensitive to itraconazole 59 (95.2%). No itraconazole or fluconazole resistance were found among the *C. metapsilosis* and *C. orthopsilosis* isolates; there was single *C. orthopsilosis* isolate SDD to both itraconazole and fluconazole.

Keywords: *Candida*, *C. parapsilosis*, Fluconazole, Resistance, Anti-fungal.

INTRODUCTION

Candidiasis is a serious infection in hospitals worldwide, especially in intensive care units (ICUs) patients (1-2). Candidiasis can result from an endogenous colonization; however, hospital transmission and emergence of resistance to antifungal agents represent new and remarkable problems (3).

Although the main candidal species causing infections worldwide is still *Candida albicans*, there is an alarm from the increase of invasive infections caused by non-*albicans* species. *Candida parapsilosis* has emerged as the second most common causative agents of candidemia in Latin America, Asia (4-5) and in

many European surveys (6-8). *C. parapsilosis* is considered one of the main causes of invasive fungal infections in USA especially in transplant patients (9).

Isolates of *C. parapsilosis* cannot be distinguished phenotypically. However, genetic analysis by randomly amplified polymorphic DNA revealed that *C. parapsilosis* complex is composed of three different species, originally they were designed group (I, II, and III). This designation is replaced later by *C. parapsilosis sensu stricto*, *C. orthopsilosis* and *C. metapsilosis*, for group (I, II, and II) respectively (10). This three genetically different species can be identified by restriction analysis of secondary alcohol dehydrogenase (*SADH*) gene which is present in all groups (11).

Candida infections are mostly treated with amphotericin B (AMB) and its lipid formulations (12-14). However, *C. parapsilosis* resistance to amphotericin B has been reported (15).

Fluconazole (FLU) is an effective and safe alternative option for treat-

ment of patients with candidemia (16-17) and in particular for candidemia caused by *Candida parapsilosis* complex (18). Several studies reported resistance in *C. parapsilosis* to *fluconazole* (19-20).

This study aimed at giving insight into the prevalence of the different *C. parapsilosis* complex species; *C. parapsilosis sensu stricto*, *C. metapsilosis*, and *C. orthopsilosis* and their distribution among patients of ICUs of Mansoura University Hospitals. Moreover, this study describes the susceptibility profile of these species to antifungal agents commonly used for treatment of candidal infections namely, AMB, FLU and Itraconazole (ITC).

METHODS

This cross sectional study was carried out including all patients aged ≥ 18 years with candidal infection during their hospital stay in ICUs during period extending from February 2013 to December, 2013 (11 months period). Our local ethical committee approved the protocol. Urine, respiratory samples, blood, and oral swabs were collected from cases with suspected candidal infections clinically.

Samples were collected and processed at the Medical Mycology unit and Microbiology Diagnostic and Infection Control unit in Medical Microbiology and Immunology department, Faculty of medicine, Mansoura University.

All media were prepared according to the manufacturer's instructions. Processing of specimens was performed according to *Koneman et al.* (21).

Candida was identified according to colonial morphology on Sabouraud Dextrose Agar (SDA), Gram stained film, and non *albicans Candida* were differentiated from *Candida albicans* by germ tube test.

Candida parapsilosis was identified using API 20 C AUX (bioMérieux), according to the manufacturer's instructions.

DNA extraction.

DNA Extraction Kit QIAamp was used to extract genomic DNA from *Candida parapsilosis* strains according to the manufacturer's instructions. The DNA obtained was finally suspended in 100 μ l TE buffer and stored at -20° C until use.

PCR amplification and SADH gene restriction analysis.

SADH gene was amplified by PCR for confirmation of *C. parapsilosis*, the reaction was done as described previously by Tavanti et al. (11) using the following primers Fwd, 5'- GTTGATGCTGTTGGATTGT-3' and Rev, 5'-CAATGCCAAATCTCCCAA-3'. PCR reaction was done in a PTC-100 TM instrument.

Isolates displaying *SADH* fragment sized of 716 bp were confirmed to be *Candida parapsilopsis* complex and used in the study.

The products of PCR reaction were treated with the *BanI* enzyme (Thermo Fisher Scientific) in a tube containing 10 µl of the amplification products and 2 µL of *BanI*. The products of restriction reaction were detected by agarose gel electrophoresis. *Candida parapsilopsis* species were distinguished as *C. parapsilosis sensu stricto*, *C. metapsilosis* and *C. orthopsilosis* according to the *SADH* restriction pattern. DNA bands were visualized using a UV transilluminator.

Antifungal susceptibility testing: was performed by E test (Liofill-

chem, Italy), and MIC results were interpreted according to the CLSI (22) guidelines.

RESULTS

This study enrolled 68 isolates of *Candida parapsilopsis* complex as identified by API 20 C AUX (bioMérieux) and confirmed by PCR amplification of *SADH* gene.

For the *C. parapsilosis* complex, the amplified *SADH* fragment (716 bp) was cut by *BanI* restriction enzyme. According to the *BanI* restriction profile described before (11), isolates with single *BanI* restriction site (at position 196) were identified as *C. parapsilosis sensu stricto*, isolated with no restriction site were classified as *C. orthopsilosis* and isolates with three *BanI* restriction sites (at positions 96, 469, and 529) were identified as *C. metapsilosis*.

Distribution of *C. parapsilosis* complex species: The sex distribution and age groups of patients are presented in table (1). Prevalence of the *C. parapsilosis* complex species and their distribution in different clinical samples are described in table (2). About ninety percent (91.2%) of the isolates (62 isolates) were identi-

fied as *C. parapsilosis sensu stricto*. Four isolates representing (5.9%) were identified as *C. orthopsilosis*. Only two isolates representing (2.9%) were identified as *C. metapsilosis*. *C. parapsilosis sensu stricto* were detected in all types of collected clinical samples including blood. However, *C. orthopsilosis* and *C. metapsilosis* were isolated only from urine and mucosal samples.

Antifungal susceptibility pattern: vSusceptibility profile to azole

agents (fluconazole, itraconazole) and AMB are described in table (3). All isolates were sensitive to AMB. Regarding fluconazole sensitivity, fifty isolates (80.6%) of *C. parapsilosis sensu stricto* were sensitive to FLU; 7 isolates (11.3%) were SDD to FLU, and 5 isolates (8.1%) were resistant to FLU. No azoles (fluconazole and itraconazole) resistance were detected among *C. metapsilosis* and *C. orthopsilosis* isolates; there was single *C. orthopsilosis* isolate SDD to both ITC and FLU.

Table (1) Epidemiological features of patients

Sex	NO (%)
Male	29 (42.6)
Female	39 (57.4)
Age	
Mean \pm SD (min-max)	47.1 \pm 13.7 (18-68 years)
Age groups	NO (%)
≥ 18 - ≤ 29	8 (11.8)
> 29 - ≤ 39	12 (17.6)
> 39 - ≤ 49	10 (14.7)
> 49 - ≤ 60	17 (25)
> 60	21 (30.9)

Table (2) Distribution of *C. parapsilosis*, *C. orthopsilosis*, and *C. metapsilosis* in different clinical samples

Clinical Sample	No. (%) of isolates			
	<i>C. parapsilosis sensu stricto</i>	<i>C. orthopsilosis</i>	<i>C. metapsilosis</i>	Total
Urine	29	3	1	33
Respiratory tract	17	0	0	17
Blood	6	0	0	6
Mucosal surface	10	1	1	12
Total	62 (91.4)	4 (2.3)	2 (2.9)	68 (100)

Table (3): Susceptibility profile of the three *Candida parapsilosis* spp. and their antifungal MIC range.

Species (no. of isolates)	Antifungal agent	MIC (mg/ml) Range	Mean	MIC 90	MIC 50	No (%) of isolates		
						S	SDD	R
<i>C. parapsilosis sensu stricto</i> (62)	AMB	0.031-1	0.21	0.5	0.125	62 (100%)	0	0
	FLC	2-64	1.3	32	8	50 (80.6)	7 (11.3%)	5 (8.1%)
	ITC	0.031-0.25	0.15	0.25	0.125	59 (95.2%)	3 (4.8%)	0
<i>C. orthopsilosis</i> (4)	AMB	0.062-0.5	0.19	0.5	0.09	4 (100%)	0	0
	FLC	2-32	1.2	3.2	6	3 (75%)	1 (25%)	0
	ITC	0.125-0.5	0.34	0.5	0.375	3 (75%)	1 (25%)	0
<i>C. metapsilosis</i> (2)	AMB	0.25	0.25	-	-	2 (100%)	0	-
	FLC	0.5-8	4.3	-	-	2 (100%)	0	0
	ITC	0.25-1	0.63	-	-	2 (100%)	0	0

S: susceptible
 SDD: susceptible dose dependant
 R: resistant

DISCUSSION

Three different species of *C. parapsilosis complex* have previously been recognized according to genetic background namely; *C. parapsilosis sensu stricto*, *C. metapsilosis* and *C. orthopsilosis* (11, 23).

In this study, *C. parapsilosis sensu stricto* represents (91.2%) of all isolated *C. parapsilosis* strains. *C. orthopsilosis* and *C. metapsilosis* represent (5.9% and 1.5 %) respectively. *C. parapsilosis sensu stricto* was the only member of the complex that was isolated from blood samples of the ICUs patients.

This result agrees with results of other studies like Silva et al. and GE et al. (24-25). This augments the assumption that main member of *C. parapsilosis complex* responsible for hematogenous infections is *C. parapsilosis sensu stricto*. The other two members (*C. orthopsilosis* and *C. metapsilosis*) are responsible for other infections like urinary tract infections and mucosal infections.

The higher prevalence of *C. parapsilosis sensu stricto* may be due to its higher capacity for persistence in

the hospital environment which may help its transmission to patients. (26-27) And/or may be explained the its capacity to express virulence determinants more than the other two species (28-29) (e.g) adherence to host cells, the ability to form biofilm, and several hydrolytic enzymes production, such as phospholipases, lipases, and proteases (30).

C. parapsilosis sensu stricto was the only species of the complex can that can form biofilms (31,32). Tavanti et al. (33) found that most of *C. parapsilosis sensu stricto* strains are proteinase producers, the higher producers being recovered from blood and mucosal specimens.

The isolation frequency of the three species of *C. parapsilosis complex* is variable throughout the world. In almost all studies, *C. parapsilosis sensu stricto* is the most common isolated species. However, the prevalence and distribution of the three species is variable. This distribution may vary according to socioeconomic conditions of the affected patients population in different countries and cities throughout the world. For example, *C. parapsilosis sensu stricto*

incidence varies from (95.6%) in Kuwait (34) to (64.5%) in China (35). Also elevated incidence of *C. metapsilosis* (10–35.5%) was found in studies performed in hospitals from China (35-36) and in Hungary (37) compared to other countries. On the other hand, *C. orthopsilosis* has higher incidence about (9%) in other studies like Bonfietti et al. (38) in Brazil.

Antifungal susceptibility tests were performed by E test to itraconazole, fluconazole and amphotericin B. All *C. parapsilosis sensu stricto*, *C. metapsilosis* and *C. orthopsilosis* isolates were susceptible to amphotericin B. The *C. parapsilosis sensu stricto* MIC₅₀ and MIC₉₀ of AMB was 0.125 µg/ml and 0.5 µg/ml respectively. This result agrees with most of studies before that found the *C. parapsilosis* MIC₅₀ and MIC₉₀ average values range from 0.13 to 1 µg/ml and from 0.5 to 1 µg/ml, respectively (39-42).

Regarding azole antifungal agents, about eighty percent of *C. parapsilosis sensu stricto* isolates were susceptible to fluconazole, (11.3%) were SDD and (8.1%) were

resistant. About ninety five percent of the isolates were sensitive to Itraconazole and (4.8%) were SDD. No FLU-resistant or ITC resistance was detected among *C. metapsilosis* and *C. orthopsilosis* isolates. Only one isolate of *C. orthopsilosis* were SDD to Fluconazole and Itraconazole (MIC 32 µg/ml and 0.5 respectively).

Fluconazole-resistance has been reported in clinical isolates of *C. parapsilosis sensu stricto* around the world (43-48).

We observed only one *C. orthopsilosis* isolate was SDD to fluconazole and itraconazole (MIC 32 µg/ml and 0.5 respectively). However, because of the small number of isolates belonging to the new species, this study may not present a complete picture about the antifungal susceptibility pattern of *C. orthopsilosis*, and *C. metapsilosis*.

This study has some limitations. First, the current study did not investigate possible risk factors for *C. parapsilosis* infections. Furthermore, the study did not search the virulence factors of *C. parapsilosis* complex and the differences of virulence fac-

tors between members of the complex that may increase the prevalence of *C. parapsilosis sensu stricto* infections among these patients. So, further studies are required to discuss these factors.

Conclusion

C. parapsilosis sensu stricto represent majority of *C. parapsilosis complex* causing infections in ICUs patients. AMB retains its activity against the members of the *C. parapsilosis complex*. There is an alarming of azoles resistance in the members of the complex especially fluconazole.

REFERENCES

- 1- **Falagas ME, Roussos N, Vardakas KZ (2010)** : Relative frequency of albicans and the various non-albicans *Candida* spp among candidemia isolates from inpatients in various parts of the world: a systematic review. *Int J Infect Dis*.14: e954-66.
- 2- **Pfaller MA, Messer SA, Moet GJ, Jones RN, Castanheira M (2011)** : *Candida* bloodstream infections: comparison of species distribution and resistance to echinocandin and azole antifungal agents in Intensive Care Unit (ICU) and non-ICU settings in the SENTRY Antimicrobial Surveillance Program (2008-2009). *Int J Antimicrob Agents*. 38: 65-9.
- 3- **Pfaller M (1995)** : Epidemiology of candidiasis. *J Hosp Infect*. 30:329-38.
- 4- **Hinrichsen SL1, Falcão E, Viçela TA, Colombo AL, Nucci M, Moura L, Rêgo L, Lira C, Almeida L (2008)** : Candidemia in a tertiary hospital in northeastern Brazil. *Rev Soc Bras Med Trop* Agosto De. 41: 394-8.
- 5- **Ma CF, Li FQ, Shi LN, Hu YA, Wang Y, Huang M, Kong QQ (2013)** : Surveillance study of species distribution, antifungal susceptibility and mortality of nosocomial candidemia in a tertiary care hospital in Chi-

- na. BMC Infect Dis. 13: 337.
- 6- **Almirante B, Rodríguez D, Cuenca-Estrella M, Almeida M, Sanchez F, Ayats J, Alonso-Tarres C, Rodríguez-Tudela JL, Pahissa A (2006) :** Epidemiology, risk factors, and prognosis of Candida parapsilosis bloodstream infections: case-control population-based surveillance study of patients in Barcelona, Spain, from 2002 to 2003. J Clin Microbiol. 44 (5):1681-5.
- 7- **Pfaller MA, Diekema DJ (2007) :** Epidemiology of invasive candidiasis: a persistent public health problem. Clin Microbiol Rev. 20(1):133-63.
- 8- **Sandven P (2000) :** Epidemiology of candidemia. Rev Iberoam Micol. 17(3):73-81.
- 9- **Raghuram A, Restrepo A, Safadjou S, Cooley J, Orloff M, Hardy D, Butler S, Koval CE (2012) :** Invasive fungal infections following liver transplantation: incidence, risk factors, survival, and impact of fluconazole-resistant Candida parapsilosis (2003-2007). Liver Transpl. 18 (9):1100-9.
- 10- **Lehmann PF, Lin D, Lasker BA (1992) :** Genotypic identification and characterization of species and strains within the genus Candida by using random amplified polymorphic DNA. J Clin Microbiol. 30 (12):3249-54.
- 11- **Tavanti A, Davidson AD, Gow NA, Maiden MC, Odds FC (2005) :** Candida orthopsilosis and Candida metapsilosis sp. nov. to replace Candida parapsilosis groups II and III. J Clin Microbiol. 43(1):284-92.
- 12- **Adler-Shohet F, Waskin H, Lieberman JM (2001) :** Amphotericin B lipid complex for neonatal invasive candidiasis. Arch Dis Child Fetal Neonatal Ed. 84 (2):F131-3.

- 13- **Linder N, Klinger G, Shalit I, Levy I, Ashkenazi S, Haski G, Levit O, Sirota L (2003)** : Treatment of candidaemia in premature infants: comparison of three amphotericin B preparations J Antimicrob Chemother. 52(4):663-7.
- 14- **Oppenheim BA, Herbrecht R, Kusne S (1995)** : The safety and efficacy of amphotericin B colloidal dispersion in the treatment of invasive mycoses. Clin Infect Dis. 21(5):1145-53.
- 15- **Ostrosky-Zeichner L, Rex JH, Pappas PG, Hamill RJ, Larsen RA, Horowitz HW, Powderly WG, Hyslop N, Kauffman CA, Cleary J, Mangino JE, Lee J (2003)** : Antifungal susceptibility survey of 2,000 bloodstream Candida isolates in the United States. Antimicrob Agents Chemother. 47(10):3149-54.
- 16- **Cornely OA, Bassetti M, Cailandra T, Garbino J, Kullberg BJ, Lortholary O, Meersseman W, Akova M, Arendrup MC, Arikan-Akdagli S, Bille J, Castagnola E, Cuenca-Estrella M, Donnelly JP, Groll AH, Herbrecht R, Hope WW, Jensen HE, Lass-Flörl C, Petrikos G, Richardson MD, Roilides E, Verweij PE, Viscoli C, Ullmann AJ; ESCMID Fungal Infection Study Group.. ESCMID* (2012):** Guideline for the diagnosis and management of Candida diseases 2012: non-neutropenic adult patients. Clin Microbiol Infect. 18:19-37.
- 17- **Colombo AL, Guimarães T, Camargo LF, Richtmann R, Queiroz-Telles Fd, Salles MJ, Cunha CA, Yasuda MA, Moretti ML, Nucci M (2013)** : Brazilian guidelines for the management of candidiasis - a joint meeting report of three medical societies: Sociedade Brasileira de Infectologia, Sociedade Paulista de Infectologia and Sociedade Brasileira de Medicina Tropical. Braz J Infect

- 18- **Pappas PG, Kauffman CA, Andes D, Benjamin DK Jr, Calandra TF, Edwards JE Jr, Filler SG, Fisher JF, Kullberg BJ, Ostrosky-Zeichner L, Reboli AC, Rex JH, Walsh TJ, Sobel JD; Infectious Diseases Society of America (2009) :** Clinical practice guidelines for the management of candidiasis: 2009 update by the Infectious Diseases Society of America. Clin Infect Dis. 48:503-35.
- 19- **Tumbarello M, Posteraro B, Trecarichi EM, Fiori B, Rossi M, Porta R, de Gaetano Donati K, La Sorda M, Spanu T, Fadda G, Cauda R, Sanguinetti M (2007) :** Biofilm production by Candida species and inadequate antifungal therapy as predictors of mortality for patients with candidemia. J J Clin Microbiol. 45(6):1843-50.
- 20- **Sarvikivi E, Lyytikäinen O, Soll DR, Pujol C, Pfaller MA, Richardson M, Koukila-Kähkölä P, Luukkainen P, Saxén H (2005) :** Emergence of fluconazole resistance in a Candida parapsilosis strain that caused infections in a neonatal intensive care unit. J Clin Microbiol. 43(6):2729-35.
- 21- **Koneman EW, Allen SD, Janda WM, Schreckenberger RC, Winn WC.** Color Atlas and Textbook of Diagnostic Microbiology, 5th ed., Lippincott-Raven, Philadelphia; 1997 (a). Introduction to microbiology. Part II: Guidelines for the collection transport, processing analysis and reporting of culture from specific specimen sources. p. 70-121.
- 22- **CLSI (2012) :** Reference method for broth dilution antifungal susceptibility testing of yeasts; 4th informational supplement. CLSI document M27-S4. Clinical and Laboratory Standards Insti-

tute, Wayne, PA

- 23- Rycovska A, Valach M, Tomaska L, Bolotin-Fukuhara M, Nosek J (2004) :** Linear versus circular mitochondrial genomes: intraspecies variability of mitochondrial genome architecture in *Candida parapsilosis*. *Microbiology*. 150 (Pt 5):1571-80.
- 24- Ge YP, Boekhout T, Zhan P, Lu GX, Shen YN, Li M, Shao HF, Liu WD (2012) :** Characterization of the *Candida parapsilosis* complex in East China: species distribution differs among cities. *Med Mycol*. 50 (1):56-66.
- 25- Silva AP, Miranda IM, Lisboa C, Pina-Vaz C, Rodrigues AG (2009) :** Prevalence, Distribution, and Antifungal Susceptibility Profiles of *Candida parapsilosis*, *C. orthopsilosis*, and *C. metapsilosis* in a Tertiary Care J Clin Microbiol. 47(8): 2392-7.
- 26- Clark TA, Slavinski SA, Morgan J, Lott T, Arthington-Skaggs BA, Brandt ME, Webb RM, Currier M, Flowers RH, Fridkin SK, Hajjeh RA (2004) :** Epidemiologic and molecular characterization of an outbreak of *Candida parapsilosis* bloodstream infections in a community hospital. *J Clin Microbiol*. 42 (10):4468-72.
- 27- Ge YP, Lu GX, Shen YN, Liu WD (2011) :** In vitro evaluation of phospholipase, proteinase, and esterase activities of *Candida parapsilosis* and *Candida metapsilosis*. *Mycopathologia*. 172(6):429-38.
- 28- Gácser A, Schäfer W, Nosanchuk JS, Salomon S, Nosanchuk JD (2007) :** Virulence of *Candida parapsilosis*, *Candida orthopsilosis*, and *Candida metapsilosis* in reconstituted human tissue models. *Fungal Genet Biol*. 44 (12):1336-41.

- 29- **Ruzicka F, Holá V, Votava M, Tejkalová R (2007)** : Importance of biofilm in *Candida parapsilosis* and evaluation of its susceptibility to antifungal agents by colorimetric method. *Folia Microbiol (Praha)*. 52 (3):209-14.
- 30- **Trofa D, Gácsér A, Nosan-chuk JD (2008)** : *Candida parapsilosis*, an emerging fungal pathogen. *Clin Microbiol Rev*. 21(4):606-25.
- 31- **de Toro M, Torres MJ, Maite R, Aznar J. (2011)** : Characterization of *Candida parapsilosis* complex isolates. *Clin Microbiol Infect*. 17(3):418-24.
- 32- **Song JW, Shin JH, Shint DH, Jung SI, Cho D, Kee SJ, Shin MG, Suh SP, Ryang DW. (2005)** : Differences in biofilm production by three genotypes of *Candida parapsilosis* from clinical sources. *Med Mycol*. 43(7):657-61.
- 33- **Tavanti A, Hensgens LA,**
- Mogavero S, Majoros L, Senesi S, Campa M. (2010)** : Genotypic and phenotypic properties of *Candida parapsilosis sensu strictu* strains isolated from different geographic regions and body sites. *BMC Microbiol*. 2010 Jul 28;10:203. doi: 10.1186/1471-2180-10-203.
- 34- **Asadzadeh M, Ahmad S, Al-Sweih N, Khan ZU (2009)** : Rapid molecular differentiation and genotypic heterogeneity among *Candida parapsilosis* and *Candida orthopsilosis* strains isolated from clinical specimens in Kuwait. *J Med Microbiol*. 58(Pt 6):745-52.
- 35- **Ge YP, Boekhout T, Zhan P, Lu GX, Shen YN, Li M, Shao HF, Liu WD (2012)** : Characterization of the *Candida parapsilosis* complex in East China: species distribution differs among cities. *Med Mycol*. 50 (1):56-66.
- 36- **Ge YP, Lu GX, Shen YN, Liu**

- WD (2011)** : In vitro evaluation of phospholipase, proteinase, and esterase activities of *Candida parapsilosis* and *Candida metapsilosis*. *Mycopathologia*. 172(6):429-38.
- 37- **Kocsubé S, Tóth M, Vágvölgyi C, Dóczy I, Pesti M, Pócsi I, Szabó J, Varga J (2007)** : Occurrence and genetic variability of *Candida parapsilosis* sensu lato in Hungary. *J Med Microbiol*. 56(Pt 2):190-5.
- 38- **Bonfietti LX, Martins Mdos A, Szeszs MW, Pukiskas SB, Purisco SU, Pimentel FC, Pereira GH, Silva DC, Oliveira L, Melhem Mde S (2012)** : Prevalence, distribution and antifungal susceptibility profiles of *Candida parapsilosis*, *Candida orthopsilosis* and *Candida metapsilosis* bloodstream isolates. *J Med Microbiol*. 61(Pt 7):1003-8.
- 39- **Fleck R, Dietz A, Hof H (2007)** : In vitro susceptibility of *Candida* species to five antifungal agents in a German university hospital assessed by the reference broth microdilution method and Etest. *J Antimicrob Chemother*. 59(4):767-71.
- 40- **Kuhn DM, George T, Chandra J, Mukherjee PK, Ghanoum MA (2002)** : Antifungal susceptibility of *Candida* biofilms: unique efficacy of amphotericin B lipid formulations and echinocandins. *Antimicrob Agents Chemother*. 46(6):1773-80.
- 41- **Marco F, Danés C, Almela M, Jurado A, Mensa J, de la Bellacasa JP, Espasa M, Martínez JA, Jiménez de Anta MT (2003)** : Trends in frequency and in vitro susceptibilities to antifungal agents, including voriconazole and anidulafungin, of *Candida* bloodstream isolates. Results from a six-year study (1996-2001). *Diagn Microbiol Infect Dis*. 46(4):259-64.

- 42- **Pfaller MA, Jones RN, Doern GV, Sader HS, Hollis RJ, Messer SA (1998)** : International surveillance of bloodstream infections due to Candida species: frequency of occurrence and antifungal susceptibilities of isolates collected in 1997 in the United States, Canada, and South America for the SENTRY Program. The SENTRY Participant Group. J Clin Microbiol. 36 (7):1886-9.
- 43- **Moudgal V, Little T, Boikov D, Vazquez JA (2005)** : Multiechinocandin- and multiazole-resistant Candida parapsilosis isolates serially obtained during therapy for prosthetic valve endocarditis. Antimicrob Agents Chemother. 49 (2):767-9.
- 44- **Lockhart SR, Messer SA, Pfaller MA, Diekema DJ (2008)** : Geographic distribution and antifungal susceptibility of the newly described species Candida orthopsilosis, in comparison to the closely related species Candida parapsilosis. J Clin Microbiol. 46 (8):2659-64.
- 45- **Silva AP, Miranda IM, Lisboa C, Pina-Vaz C, Rodrigues AG (2009)** : Prevalence, distribution, and antifungal susceptibility profiles of Candida parapsilosis, C. orthopsilosis and C. metapsilosis in a tertiary care hospital J Clin Microbiol. 47(8):2392-7.
- 46- **Cantón E, Pemán J, Quindós G, Eraso E, Miranda-Zapico I, Álvarez M, Merino P, Campos-Herrero I, Marco F, de la Pedrosa EG, Yagüe G, Guna R, Rubio C, Miranda C, Pazos C, Velasco D; FUNGEMYCA Study Group (2011)** : Prospective multicenter study of the epidemiology, molecular identification, and antifungal susceptibility of Candida parapsilosis, Candida orthopsilosis, and Candida metapsilosis isolated from

patients with candidemia.
Antimicrob Agents Chemo-
ther. 55(12):5590-6.

crobiol Infect Dis. 68
(3):284-92

- 47- **Chen YC, Lin YH, Chen KW, Lii J, Teng HJ, Li SY (2010) :** Molecular epidemiology and antifungal susceptibility of *Candida parapsilosis* sensu stricto, *Candida orthopsilosis*, and *Candida metapsilosis* in Taiwan. *Diagn Mi-*
- 48- **Pfaller MA, Boyken L, Hollis RJ, Kroeger J, Messer SA, Tendolkar S, Diekema DJ (2008) :** In vitro susceptibility of invasive isolates of *Candida* spp. to anidulafungin, caspofungin, and micafungin: six years of global surveillance. *J Clin Microbiol.* 46(1):150-6

الملخص العربي

انواع وحساسية ميكروب الكانديدا باراسليوبسز لمضادات الفطريات

في مرضى العناية المركزة بمستشفيات جامعة المنصورة.

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ميكروب الكانديدا باراسليوبسز هو واحد من الأنواع الكانديدا الهامة المسؤولة عن العدوي الفطرية الغازية وعدوى المستشفيات المكتسبة وبخاصة في المرضى الرعاية الحرجة. وقد تمت إعادة تسمية مجمع ميكروب الكانديدا باراسليوبسز وفقا لقواعد وراثية الي ثلاثة انواع سانسيس تريكتو , اورثوبسليوبسز و ميتابسليوبسز.

وقد صممت هذه الدراسة لوصف توزيع وخصائص حساسية اعضاء مجمع الكانديدا باراسليوبسز لمضادات الفطريات بين المرضى من وحدات العناية المركزة في مستشفيات جامعة المنصورة. وقد تم التعرف علي اعضاء مجمع ميكروب الكانديدا باراسليوبسز بواسطة مؤشر الملف التحليلي API20 C ووفقا لخصائص قطع جين SADH باستخدام انزيم القطع و تم إجراء اختبار الحساسية لمضادات الفطريات عن طريق اختبار E.

وقد تم عزل ٦٨ عزلة لميكروب الكانديدا باراسليوبسز تتضمن ٦٢ عزلة من نوع سانسيس تريكتو تشكل نسبة (٩١,٢%) و اربع عزلات من نوع اورثوبسليوبسز بنسبة (5.9% و عزلتين من النوع ميتابسليوبسز بنسبة (٢,٩%) جميع العزلات كانت حساسة لعقار الامفوتريسين- ب. و ما يزيد عن ثمانين في المئة (٨٠,٦%) من العزلات من نوع سانسيس تريكتو كانت حساسة لعقار الفلوكونازول و (٨٠,٦%) العزلات (١١,٣%) ذو حساسية متغيرة علي حسب الجرعة و خمس عزلات (٨,١%) مقاومة لعقار الفلوكونازول. و معظم العزلات (٩٧,٩%) كانت حساسة لعقار الاتراكونازول. لم يتم عزل اي عزلات من الانواع اورثوبسليوبسز و ميتابسليوبسز مقاومة لعقارات الفلوكونازول و الاتراكونازول في هذه الدراسة. و كانت هناك عزلة واحدة من النوع اورثوبسليوبسز ذو حساسية متغيرة علي حسب الجرعة لعقارات الفلوكونازول و الاتراكونازول.

و نستخلص من هذا البحث ان نوع سانسستريكتو يشكل غالبية مجمع ميكروب الكانديدا بارابسليوبسز المسبب للعدوي في مرضي العناية المركزة و لازال عقار امفوتريسين-ب يحتفظ بفاعليته ضد جميع انواع مجمع ميكروب الكانديدا بارابسليوبسز و لكن هناك زيادة فى مقاومة الميكروب لعقارات الازولات خاصة عقار فلوكونازول.