# Egyptian J. Anim. Prod., 35, Suppl. Issue, Dec. (1998):75-86. BUFFERING COMPONENTS IN RUMEN LIQUOR AND BLOOD OF BUFFALO HEIFERS

Neama A. Ashmawy

Department of Animal Production, Faculty of Agriculture, Cairo University

## SUMMARY

Five buffalo heifers aging 18-30 months were used to study buffering components in rumen liquor and blood. Samples of rumen liquor were taken by stomach tube, to determine the buffering components in the rumen fluid, mainly, total volatile fatty acids (TVFA's), bicarbonate (HCO<sub>3</sub>), phosphates and ammonia (NH<sub>3</sub>) concentrations with simultaneously measuring the pH, and buffering capacity value. Blood samples were withdrawn from the jugular vein to determine the relevant buffering components and pH value. Two hours postfeeding, rumen bicarbonate concentration showed high concentration, inorganic phosphorus concentration varied widely among animals, that may be due to different rate of saliva secretion and rumination rate of each animal. At nearly neutral pH, ammonia concentrations showed positive relationship between rumen and blood ammonia concentration. Total volatile fatty acids in rumen were appreciably high, that is attributed to the rapid fermentation of the ingested diets (concentrate, hay and rice straw). Rumen buffering capacity was approximately equal in all animals.

Keywords: Buffalo, buffers, bicarbonate, blood, rumen, ammonia, inorganic phosphorus, total volatile fatty acids

## INTRODUCTION

The buffering components of the reticulo-rumen contents results directly from the complex interrelationships among the amount and rate of saliva production, saliva composition and rate of microbial metabolism within reticulorumen (Kay, 1960 and Baily & Balch, 1961). Many factors affect buffering value of the rumen content such as the concentration of the end-products of fermentation, particularly, VFAs, carbon dioxide, bicarbonate (HCO<sub>3</sub>), phosphate and ammonia concentrations (Turner & Hodgetts, 1955). Within the

normal range of ruminal pH (5.5 - 7.5), bicarbonate concentration, phosphate and the volatile fatty acids are all important buffers. At pH levels below 5.5, the VFAs become the major components for preventing further reduction in pH. These authors concluded that, during fasting, rumen bicarbonate concentration was relatively more important than phosphate; when ruminal fermentation progresses, VFAs accumulate and pH decreases, bicarbonate levels decline and phosphate becomes relatively more important. During active fermentation, phosphate is relatively more important. With high concentrations of VFAs and pH below 6.0, VFAs contributed the major buffering action against further additions of acids. Briggs et al. (1957) stated that, rumen pH was closely related to VFA levels in the rumen.

This study was conducted to determine buffering components in rumen liquor and at the same time in blood serum, plus haemoglobin as functional factor in the stability of blood pH.

This work was designed as a preliminary investigation for the coincidentally relative concentrations of buffering components in rumen liquor and blood serum as related to general acid-base balance. The study is considered to provide parameters for subsequent studies of dynamic changes, in this components, in reaction to different thermal conditions and nutritional treatments.

## MATERIALS AND METHODS

#### Animals:

This work was carried out during summer (August). Ambient air temperature and relative humidity means were  $28.4\pm1.21$  °C &  $68.0\pm0.58\%$  in the Animal Physiology Laboratory, Animal Production Department, Faculty of Agriculture, Cairo University, Egypt.

Five buffalo-heifers 2.5 years old, weighing 240 - 380 kg were used in this study. The heifers were offered daily ration of hay (5 kg/head/day) plus concentrate (5 kg/head/day) and rice straw. The concentrate ration was composed of, undecorticated cotton seed cake, rice bran, yellow corn, limestone and common salt plus molasses. The nutrients contents were appoximalty 16% crude protein, 15% crude fiber and 3% crude fat.

The animals were deprived of food and water for 20 hours before feeding at 08:00 h. Samples of the rumen liquor and blood serum were collected simultaneously at 2 hours post feeding.

## Sampling of rumen liquor:

Samples of rumen liquor were obtained by a stomach tube at two hours postfeeding under a layer of liquid paraffin. The sample was centrifuged, the supernatant rumen liquid was transferred to another tube under liquid paraffin to determine rumen bicarbonate and buffering capacity value. Another rumen

liquid sample was drawn at the same time to determine pH value, TVFAs, ammonia concentration and inorganic phosphorus.

#### Blood sampling:

Blood samples were taken from jugular vien, one sample under a layer of liquid paraffin to determine serum bicarbonate concentration. Other heparinized blood samples were obtained to determine blood pH value, ammonia concentration, TVFAs, inorganic phosphorus and haemoglobin concentration.

## Technical procedure:

Rumen fluid and serum bicarbonate concentration was determined by the titration method of Van Slyke (1922). Rumen buffering value was determined by the method reported by Emmanuel *et al.* (1969). Rumen fluid and blood pH values were determined instantly at sampling time by glass electrode pH meter. Total VFAs concentrations were determined by steam distillation method (Barnett & Reid, 1957). The method of Fiske and Subbarrow (1925) was used to measure the inorganic phosphorus. Ammonia concentrations in blood and rumen liquid were determined by micro-diffusion technique of Conway (1950). Haemoglobin concentration was determined colourimeterically as described by Bauer (1970).

#### RESULTS AND DISCUSSION

#### 1- Bicarbonate

Two hours post-feeding, rumen bicarbonate concentration was high (Table 1), these concentrations were affected by the fairly high pH values ranging between 6.19 and 7.78. This high concentration might be due to the increase in saliva secretion rate at this time of active rumination post feeding period. Washburn and Brody (1937) observed that the concentration of carbon dioxide in the ruminal gas of normal cows varies from a maximum of 80 % at 1 - 2 hours post-feeding to a minimum of 10 % at 24 hours after feeding. Ash and Dobson (1963) concluded that hydration of CO2 in the sheep rumen provided a source of hydrogen ions for the production of undissociated VFA. It explains the accumulation of bicarbonate within the rumen contents observed in the present work. On the other hand, Masson and Phillipson (1951) and Dobson and Phillipson (1968) reported that substantial amounts of carbonic anhydrase have been found in rumen epithelial cells, and this could be involved in subsequent release of bicarbonate into the rumen. Most of the CO2 seems to be produced by intracellular epithelial metabolism, although CO2 also could be absorbed from the lumen side of the epithelium to provide the carbonic acid for the donation of hydrogen and bicarbonate ions (Stevens, 1973). In most cases the animals which had high values of rumen bicarbonate, showed high serum bicarbonate and vice versa (Table 1). These results agree with those reported

29.44 ± 4.09

28.16±4.02 6.37±0.56

39.04±14.92

5.73±0.45

17.76±3.56 5.94±0.32

31.68±7.79

30.56±11.47

5.60±0.66

Serum inorg. phos. (mg/100ml) Rumen inorg. phos. (mg/100ml)

Rumen NH<sub>3</sub> (mg/100ml)

Ammonia:

Blood NH<sub>3</sub> (mg/100ml)

6.10±1.08

7.60±0.67

Rumen TVFA (m.eq./100 ml)

Blood TVFA (m.eq./100 ml)

Inorganic phosphorous:

5.38±0.48

5.80 ± 0.22

 $1.44 \pm 0.09$  $6.51 \pm 1.30$ 

6.81±3.41 1.39±0.24

7.46±3.41 1.56±0.18

4.97±2.79 1.32±0.05

4.39±1.66 1.32±0.24

9.57±3.57 1.61±0.24

41.48±16.86 27.72±2.45 31.40±2.80 5 27.90±6.23 25.92±2.02 37.21±6.13 Table 1. Means ± S.E. of buffering agents in rumen fluid and blood serum of buffalo heifers 37.70±2.88 28.22±1.58 25.62±2.99 6.71±1.49 28.06±4.25 41.50±2.32 29.14±2.00 40.26±13.17 37.54±25.8 31.14±1.00 Rumen bicarbonate (m.mol/I) Serum bicarbonate (m.mol/I) Animal No. Bicarbonate: TVFA:

 $35.20 \pm 1.79$  $28.43 \pm 0.86$ 

 $34.53 \pm 4.38$ 

 $6.22 \pm 0.45$ 

5.80±0.75

4.88±0.75

Overall mean

Each mean represents 5 observations for each animal.

by Bödeker et al. (1992) in sheep and Remond et al. (1993) in Texel wethers. The first author recorded that, bicarbonate favors NH $_3$  absorption across the ruminal epithelium. The second author found that, if carbonic acid was in equilibrium with CO $_2$  in ruminal gas, and the pH was nearly neutral, the increase in carbonic acid (H $_2$ CO $_3$ ) may have led to an increase in bicarbonate ions (HCO $_3$ ).

## 2- Volatile fatty acids (VFAs)

In the present results, the total VFAs concentration was appreciably high at 2 hours postfeeding (Table 1). The increase in concentration of VFAs indicates rapid fermentation of ingested diets (concentrate & hay and rice straw). This result agrees with Topps et al. (1968) who indicated that, total VFAs in the rumen of young Friesian steers, aged 6 months, fed concentrate diets were 55.0 m.eq./l before feeding, 186.6 m.eq./l after feeding, and when steers fed hay, TVFAs concentration was 83.8 before vs. 132.0 m.eq./l after feeding. In heifers No. 2 & 3 (heavier and older than other animals), rumen TVFAs and ammonia concentrations were the least while bicarbonate concentration showed the highest values. These findings indicated that TVFAs and NH3 were absorbed from the rumen into blood and blood bicarbonate intered the rumen instead (Table 1). These results agree with Remond et al. (1993) who found that, at pH 6.7, NH<sub>3</sub> absorption increased with high NH<sub>3</sub> and butyrate concentrations in the rumen. These two heifers had the highest concentration, alongside heifer No. 1, in both VFAs and bicarbonat with the lowest concentration of NH3 in heifers No. 2 & 3.

## 3- Inorganic phosphorous

Rumen inorganic phosphorous concentration varied widely between the studied heifers (Table 1), this may be due to the differences in the rate of saliva secretion and rumination rate of each animal. Poutiainen (1968) indicated that, daily secretion rates of saliva for non-lactating cows varied from 90 to 190 I/day. Ternouth et al. (1985) noted that, the salivary secretion of inorganic phosphorous, generally, increase with salivary flow rate. The daily turnover of inorganic phosphorous in the saliva of sheep was 60 - 320 mg kg day 1 (Kay, 1960). Because of the relatively large addition of salivary inorganic phosphorous to the rumen contents there is no net absorption of inorganic phosphorous from this region. It is not until the small intestine that the bulk of the net absorption of Inorganic phosphorous occurs (Poppi & Ternouth, 1979) although there is also some net absorption of Inorganic phosphorous from the large intestine at higher phosphorous intake. Tomas et al. (1967) indicated that salivary phosphorus was the major source of phosphorus to the sheep rumen and was the principal determinant of rumen fluid inorganic phosphorus levels. The phosphorus level in blood was not affected by the phosphorus level in the rumen (Table 1). Tomas (1974 a & b) indicated that, the salivary

glands largely replace the kidneys as a means of removing excess Inorganic phosphorous from the circulation. The salivary Inorganic phosphorous helps to buffer the production of volatile fatty acids in the rumen and to satisfy the phosphorus requirements of the microbial population of the rumen. A relationship between kidney and salivary secretion of Inorganic phosphorous has been demonstrated.

#### 4- Ammonia

Lewis (1957) demonstrated that the portal blood ammonia concentration increased as a curvilinear function of rumen ammonia content. Data in the present study agree with the finding reported by Lewis (1957) since blood ammonia concentrations were parallel to that of rumen fluid (Table 1). Hogan (1961) and Bödeker et al. (1992) stated that at nearly neutral pH the passage of NH<sub>3</sub> across the ruminal epithelium increased with rise of NH<sub>3</sub> concentration in the ruminal fluid. Hogan suggested that, NH3 passage across the epithelium of the rumen mainly occurred by diffusion in the ionized form and that some NH<sub>3</sub> cations (NH<sub>4</sub><sup>+</sup>) may accompany the absorption of VFA anions. Heifers which had minimum values of NH3 and TVFA in the rumen, the HCO3 concentration in their rumen was higher. This low concentration in rumen NH<sub>3</sub> and TVFAs may be attributed to the release of these two buffering components from the rumen into blood, at the same time, blood bicarbonates pass across the rumen wall. The main factor that control NH3 transepithelial flux seemed to be the concentration of unionized NH3 in the ruminal contents and blood flow to the rumen (Remond et al., 1993).

Rumen buffering capacity

Rumen buffering capacity at 2 hours postfeeding (Table 2) was approximately equal in all animals (buffering capacity per 2 ml rumen liquid ranged from 0.023 to 0.029 m.eq. at pH = 4.0 and it was 0.009 to 0.013 m.eq. at pH = 8.0). This denotes effective function of the buffering components in the rumen liquor.

pH values

The mean pH values in rumen liquor differed between animals within the range 6.19 - 7.98 that is 1.79 degrees of pH value (Table 2). This may be due to an increase in saliva bicarbonate concentration at the time of sampling. In spite of this wide variation of pH of rumen liquor the blood pH value was almost stable within the normal range in all animals (pH = 7.23 - 7.43). Upadhyay (1993) stated that maintenance of normal blood pH depends ultimately on the activity of blood buffers which effectively resist appreciable changes in the pH.

It is clear in table (2) that their was parallel trend in the values of haemoglobin concentrations and blood serum pH values. It has been proved

Table 2. Means ± S.E. of pH values in rumen fluid and blood, buffering capacity and haemoglobin in buffalo heifers

- <						
Item Animal No.	<del>-</del>	2	3	4	5	Overall mean
pH values: Rumen pH Blood pH Rumen buffering capacity:	7.24± 0.29 7.37± 0.13	6.88 ± 0.45 7.32 ± 0.17	7.78 ± 0.37 7.23 ± 0.25	6.19 ± 0.14 7.39 ± 0.07	7.01 ± 0.22 7.43 ± 0.15	7.02 ± 0.17 7.35 ± 0.07
at pH = 4.0 at pH = 8.0 Blood Hb (g/100ml)	0.029± 0.01 0.011± 0.00 9.88 ± 0.29	0.025 ± 0.00 0.013 ±0.00 8.02 ± 0.42	0.023 ± 0.00 0.009 ± 0.00 7.72 ± 0.36	$0.025 \pm 0.01$ $0.010 \pm 0.00$ $8.91 \pm 0.43$	$0.025 \pm 0.01$ $0.009 \pm 0.00$ 8.23 + 0.59	0.025 ± 0.00 0.010 ± 0.00 8 55 ± 0.24

Each mean represents 5 observations for each animal.

that haemoglobin has an important biological function in buffering for pH stabilization (Frandson, 1986 and Neama, 1987).

It is expected that there are integrated control on the exchange of buffering components between blood and rumen liquor to fulfill stability of the vital pH normal in blood. This concept needs extensive research work.

#### REFERENCES

- Ash, R.W. and A. Dobson, 1963. The effect of absorption on the acidity of rumen contents. J. Physiol. Lond. 169:39-61.
- Baily, C.B. and C. C. Balch, 1961. British J. Nutr., 15:371. cited by: Bartley, E. E., 1976. Bovine saliva production and function in buffers in ruminant physiology and metabolism. ed. by: Weirbera, M. S. and A. Sheffner. Church and Dwight Co. New Yourk.
- Barnett A.J. G. and R. L. Reid, 1957. Studies on the production of volatile fatty acids from grass and rumen liquor in an artificial rumen. J. Agric. Sci. Cambridge. 48:315-318.
- Bartley, E.E., 1976. Bovien saliva production and function in Buffers in ruminant physiology and metabolism. ed. by: Weirbera, M. S. and A. Sheffner. Church and Dwight Co. New Yourk.
- Bauer, J.D., 1970. Haemoglobin , part III. Haematology in, Gradwoll clinical laboratory methods and diagnosis, 17th ed. by Frankel's. Reitman's and Sonnen Wirth, A. C. the C. V. Mosby Co. Saint Louis.
- Briggs, P.K., J. P. Hogen and R. L. Reid, 1957. Aus. J. Agric. Res. 8:674. cited by: Darrell Van Campen. Effects of buffers on ruminal acids (1976) In Buffers in ruminant physiology and metabolism. ed. by Weirbera, M. S. and A. Sheffner. Church and Dwight Co. New Yourk.
- Bödeker, D. Y. Shen, J. Kemskowski and H. H. Holler, 1992. Influence of short-chain fatty acids on ammonia absorption across the rumen wall of sheep. Exp. Physiol. 77:369.
- Conway, E. J., 1950. Determination of ammonia. Microdiffusion analysis and volumetric error. cited by Oser, B. L., 1965. Hawk's physiological chemistry. Fourteenth Edition pp.1223 & 1224. New Yourk.
- Dobson, A. and A. T. Philipson, 1968. Absorption from the ruminant forestomach. In: Handbook of physiology. Alimentary canal. Bile, Digestion; Ruminal physiology. Washington, D C: Am. Physiol. Soc., Sect. 6, Vol. 5, Chapt. 132, pp.2761-2774.
- Duncan, J. R., 1986. Blood chemistry values in domestic animals. cited by : Rudolf Clarenburg, 1992. Physiological chmistry of domestic animals. Index.
- Emmanuel, M. J. Lawlor and D. M. Mcaleese, 1969. The rumen buffering system of sheep fed pelleted roughage-concentrate rations. Br. J. Nutr. 23:805.

- Fiske, C. H. and Y. Subbarow, 1925. Determination of Inorganic phosphate.
  J. Biol. Chem. 66, 375. cited by Oser, B. L., 1965. Hawk's physiological chemistry. Fourteenth Edition pp.1113. New Yourk.
- Frandson, R. D., 1986. Anatomy and physiology of farm animals. p. 392. Lea and Febiger, Philadelphia, USA.
- Hogan, J. P., 1961. The absorption of ammonia through the rumen of the sheep. Aust. J. Biol. Sci., 14:448.
- Kaneko, J. J., 1989. Blood chemistry values in domestic animals. cited by: Rudolf Clarenburg, 1992. Physiological chmistry of domestic animals. Index.
- Kay, R. N. B., 1960. J. Physiol., 150-155. cited by: Bartley, E. E., 1976. Bovine saliva production and function in buffers in ruminant physiology and metabolism. ed. by: Weirbera, M. S. and A. Sheffner. Church and Dwight Co. New Yourk.
- Lewis, D., 1957. Blood-urea concentration in relation to protein utilization in the ruminant. J. Agric. Sci., 48:438.
- Masson, M. J. and A. T. Philipson, 1951. Absorption of acetate, propionate and butyrate from the rumen of sheep. J. Physiol. Lond. 113:189-206.
- Neama, A. Ashmawy, 1987. Acid-base balance by bicarbonate store in ruminant animals under environmental hot condition. M.Sc. Thesis, Fac. Agric., Cairo Unvi., Egypt.
- Oser, B.L., (ed.) 1965. Hawk's physiological chemistry (14th ed.) pp.952-955 the 13 Lakiston Divion Mc Graw Hill, London.
- Poppi, D. P. and J. H. Ternouth, 1979. Secretion and absorption of phosphorus in the gastro-intestinal tract of sheep fed on four diets. Aust. J. Agric. Res. 30, 503-512.
- Poutiainen, E., 1968. Factors influencing the flow of fluid, saliva and some cations through the reticulo-omasal orifice of the cow. Ann. Agric. Fenn, 7 (3):1.
- Remond, D., J. Pchaise, E. Delval and C. Poncet, 1993. Net transfer of urea and ammonia accross the ruminal wall of sheep. J. Anim. Sci., 71:2785-2792.
- Stevens, C. E., 1973. Transport across rumen epithelium. pp.404-426. In: Transport mechanism in epithelia, edited by H. H. Ussing and N. A. Thorn. Munksgaard, Copenhagen.
- Ternouth, J. H. Davies, H. M. S. Milton, J. T. B. Simpson, M. W. Morgan and N. E. Sands, 1985. Phosphorus metabolism in ruminants. 1- Technique for phosphorus depletion. Aus. J. Agric. Res. 36, 637-645.
- Tomas, F. M., 1974 a. Phosphorus homestasis in sheep. II- Influences of diet on the pathway of secretion of phosphorus. Aust. J. Agric. Res. 25, 485-495.
- Tomas, F. M., 1974 b. Phosphorus homestatis in sheep. III- Relationship between the amount of salivary phosphorus secretion and the quantities of

- phosphorus exceted via the urine and faeces. Aust. J. Agric. Res. 25, 495-507.
- Tomas, F. M., R. J. More and M. Somers, 1967. Phosphorus turnover in sheep. Aust. J. Agric. Res. 18, 635-645.
- Topps, J. H., R. N. B. Kay, E. D. Goodall, F. G. Whitelaw and R. S. Reid, 1968. Digestive of concentrate and hay diets in the stomach and intestines of ruminants. Br. J. Nutr. 22, 261.
- Turner, A. W. and W. E. Hodgetts, 1955. Aust. J. Agric. Res. 6:115, cited by: Emmanuel, B., M. J. Lawlor and D. M. Mcaleese, 1969. The rumen buffering system of sheep fed pelleted roughage concentrate rations. British J. Nutr., 23:805.
- Upadhyay, R. C., 1993. Physiological responses and acid-base status of rural working buffaloes. Indian J. Dairy Sci., 46, 5, pp.227-229.
- Van Slyke, 1922. J. Biol. Chem., 52:495 cited by Oser, B. L. (Plasma bicarbonate titration method) pp.952-955. In Hawk's Physiological Chemistry. the 13 Lakiston Divion Mc Graw Hill, London.
- Washburn, L. E. and S. Brody, 1937. Growth and development. Methan, hydrogen and carbon dioxide production in the digestive tract of ruminants in relation to the respiratory exchange. Missouri Univ. Agri. Expt. Sta. Res. Bull. No. 263.

# منظمات الحموضة والقلوية في سائل كرش عجلات الجاموس وعلاقتها بالمنظمات في الدم.

# نعمة أحمد عشماوى

قسم الإنتاج الحيواني - كلية الزراعة - جامعة القاهرة.

أجريت هذه الدراسة في معمل فسيولوجيا الحيوان بكلية الزراعة جامعة القاهرة لدراسة منظمات الحموضة والقلوية في الكرش وعلاقتها بالمنظمات الموجودة في الدم في خمسة عجلات جاموسي عمرها ١٨ - ٣٠ شهر.

- وقد تم سحب عينات سائل الكرش بواسطة اللى المعدى بعد ساعتين من التغذية على دريس ،
   عليقة مركزة ، قش أرز وقد تم تثييت العليقة طوال فترة التجربة.
- عينات الدم تم سحبها من الوريد الوداجي جزء منها تم وضعة في أنابيب معاملة بالهبارين وجزء أخر تحت طبقة من زيت البرافين لتقدير تركيز البيكربونات.
- وقد تم تقدير تركيز كل من الأحماض الدهنية الطيارة الكلية ، البيكربونات ، الفوسفات ، الأمونيا في كل من عينات الدم وعينات سائل الكرش وتم تقدير قيمة pH لكل العينات عند وقت أخذ العينة، وتقدير كفاءة هذة المنظمات بالكرش.

# وكان من أهم النتائج:-

- بعد ساعتين من التغذية تركيزات بيكربونات الكرش وصلت عند أعلى تركيز لها بينما تركيز الفوسفات تباينت بشدة مابين الحيوانات.
- أظهرت النتائج علاقة موجبة مابين تركيز الأمونيا في الدم والأمونيا في الكرش عند قيم PH القريبة من التعادل. بينما العجلات التي سجلت قيم منخفضة لتركيز الأمونيا والأحماض الدهنية الطيارة الكلية كان تركيز البيكربونات في كرشها عالية وهذا قد يكون راجع لمرور الأمونيا والأحماض الدهنية الطيارة من الكرش للدم في الوقت نفسة يتم عبور البيكربونات من الدم للكرش من خلال جدار الكرش.
- قيم pH سائل الكرش إختلفت مابين الحيوانات في مدى من PH = 6.19 7.98 بينما pH الدم تقريبا ثابتة في المدى الطبيعي لها 7.43 7.23 = pH حيث أن ثبات هذة الأخيرة يعتمد أساسا على المنظمات الموجودة بالدم.

- أما قيم كفاءة هذة المنظمات بالكرش كانت تقريبا متساوية في كل العجلات (كفاءة المنظم في ٢ مل من سائل الكرش) وهذا قد يرجع لتغذيتها على علائق ثابتة طوال فترة التجرية ويدل على كفاءة عمل هذه المنظمات في تنظيم درجة حموضة سائل الكرش.