

SINAI Journal of Applied Sciences



USING MORINGA LEAVES POWDER IN PRODUCTION OF PROBIOTIC YOGHURT

Ola F. El-Sayed¹, H.S.M. El-Taweel¹, A.A. El-Shibiny² and M.M.K. Metwally^{*2}

- 1. Dept. Dairy Sci., Food Technol. Res. Inst., Ministry Agri. and Land Reclamation, Egypt.
- 2. Dept. Dairy and Food Sci., Fac. Environ. Agric. Sci., Arish Univ., Egypt.

ABSTRACT

Moringa leaves powder was added during the manufacture of probiotic yoghurt at level (0.5%) before pasteurization. Three probiotic yoghurt treatments were prepared as follows: voghurt without moringa served as control (T1), yoghurt + moringa (T2) and yoghurt + moringa + 10% mango pulp (T₃). Yoghurt were inoculated with 2% lactic acid bacteria (LAB) and 2% L. acidophilus and incubated at 42°C until complete coagulation (pH 4.8), then stored at 5°C up to 14 days. Chemical, microbiological and sensory properties of the produced yoghurt were carried out. The level of 0.5% moringa was found to be the best ratio. The results showed that pH values and moisture content (%) decreased during storage period for all treatments, while the values of titratable acidity, total solids (%), protein content (%), fat content (%), antioxidant activity and total phenolic content were increased and the treatments (T₂ and T₃) had values greater than control yoghurt (T₁). Microbiological analysis indicated that the addition of 0.5% moringa leaves powder and 0.5% moringa leaves powder +10% mango pulp stimulate the growth of LAB and probiotic culture (L. acidophilus). On the other hand Yeasts and moulds, Coliform and Sporeforming bacteria were not detected in all treatments up to the end of storage period. Moreover the addition of 0.5% moringa leaves powder + 10% mango pulp increased the acceptability of product more than the addition of moringa alone up to the 14th day of storage at 5°C.

Kew words: Moringa, production, yoghurt.

INTRODUCTION

Yoghurt is a coagulated dairy product obtained by the lactic acid fermentation of milk by bacteria *i.e. Streptococcus thermopiles* (ST), *Lactobacillus delbrueckii* ssp. *bulgaricus* (LB) (Fadela *et al.*, 2009). Addition of these two cultures resulted in acidification of milk and produce of aromatic compounds (Sahan *et al.*, 2008). Although these microflora have been found to be valuable for human as they help in maintaining health and nutrition. Also efforts have been placed on developing yoghurt containing probiotic cultures like

Lactobacillus acidophilus (LA) and B. bifidus (BB) (Vinderola and Reinheimer, 2000). Probiotic cultures are live microbial food ingredients that are beneficial for human health (Salminen et al., 1999), which includes improvement of intestinal microbial balance which results in the inhibition of bacterial pathogens, reducing the risk of colon cancer, in the inhibition of bacterial pathogens, reducing the risk of colon cancer, improving the risk of colon cancer, improving the immune system, lowering serum cholesterol levels (Saarela et al., 2002), alleviation of lactose intolerance and nutritional enhancement (Alizadeh and Ehsani, 2008).

* Corresponding author: Tel.: +201227252488 E-mail address: mamdouhmk1@hotmail.com Mango (*Mangifera indica* L.) is a seasonal fruit grows in tropical regions and is regarded as one of the most important fruits of Asia.

The nutritional importance of mango is mainly due to its high amounts of bcarotene, a carotenoid which provides various health benefits, including pro-Α and antioxidant activity vitamin (Harnkarnsujarit and Charoenrein, 2011). Mango contains a variety of phytochemicals and nutrients. The fruit pulp is high in prebiotic dietary fiber, polyphenols vitamin C, diverse provitamin a carotenoids (Ajila and Prasada Rao, 2008).

Moringa oleifera is referred to as a "Miracle tree" or "Wonder tree" (Kasolo et al., 2010) of significant socio economic importance because of its several nutritional, pharmacological (Caceres et al., 1991) and industrial application (Makkar and Becker, 1996).

The leaves of this plant contain high amount of vitamin B complex, calcium, potassium, iron and protein. Also, they contain all of the essential amino acids in good proportion (Mishra et al., 2012).

Moringa oleifera leaves are active against the growth of bacteria such as: E. coli, S. arous, P. aeruginosa and B. cereus as these organisms range from pathogenic and oxygenic organism liable to cause food borne illnesses and food spoilage. It can be used as evaluable drug in the treatment of infections caused by E. coli and P. aeruginosa (Abalaka et al., 2012).

MATERIALS AND METHODS

Materials

Fresh cow's milk was obtained from the herd of Badwy farm of El-Arish, Egypt. Average chemical composition of milk (3% fat, 3.35% protein, 12.6% T.S) were determined according to **AOAC** (2011).

Skim milk powder (96% TS, product of Dairy America TM) USA, was obtained from the local market.

Direct Vat Starter (DVS) yoghurt culture was obtained from CHR-Hansen's laboratorie, Denmark, under commercial name type (FD-DVS-YC-X11) containing *Streptococcus thermophiles* and *Lactobacillus delbrueckii* ssp. *Bulgaricus*.

Probiotic bacteria strain *Lactobacillus* acidophilus (DSM20384) was obtained from Egyptian Microbial Culture Collection (EMCC) at Cairo Microbiological Resources Center (Cairo MIRCEN), Faculty of Agriculture, Ain Shams University.

Mango (*Mangifera indica*) fruit and sugar were obtained from local market of El-Arish, Egypt.

Moringa oleifera leaves was obtained from Cautia farm of North Sinai, Egypt.

Methods

Preparation of Additions

Preparation of mango pulp

Mango pulp (0.27% fat, 0.51% protein, 81.3% T.S) was obtained manually after thorough washing and peeling of the skin and blended to get smooth and then pasteurized at 90°C for 10 min according to the procedure mentioned by **Vijayalakshmi** *et al.* (2009).

Preparation of moringa leaves powder

The collected leaves were spread on a clean curtain cloth and kept at room temp.

The selected room for shade drying was well ventilated by natural current of air. The leaves took about six to seven days to dry completely and became crispy and brittle to touch then blended in a blender to get powder according to the procedure mentioned by **Delong (2003)**.

Preparation of probiotic culture

Strain *Lactobacillus acidophilus* (DSM 20384) was activated in MRS broth according to **De Man et al.** (1960).

Manufacture of Yoghurt

Yoghurt was made from standardized cow's milk according to **Tamime and Robinson (1999)** as shown in diagram (A).

Diagram (A) – Manufacure of yoghurt

Fresh cow milk (3% fat, 3.35% protein, 12.6% TS).

Methods of Analysis

Yoghurt samples were analyzed chmically, microbiologicaly and organoleptically when fresh and after 7 and 14 days of storage at 5°C.

Chemical Analysis

pH values were measured using Jenway pH meter with Jenway spear electrode No: 29010 (Jenway limited Gransmore Green, Felsted, Dunmow, England).

Titratable acidity, total solids, total protein and fat were determined according to the method described by **AOAC (2011)** .Moisture content was calculated using the regular equation as follows:

Moisture (%) = 100 - Total solids

Measurement of antioxidant activity using 1,1-diphenyl-2-picrylhydrazyl radical (DPPH) inhibition **assay** was carried out according to methods described by **Li** *et al.* **(2009)**. Total phenolic content (TPC) of the previously prepared yoghurt samples were determined using the Folin- Ciocalteau by method described by **Li** *et al.* **(2009)**.

Microbiological analysis

Preparation of all samples for microbiological examination was carried out as described by **Frazier and Foster** (1961).

Lactobacillus acidophilus and Lactobacillus delbrueckii ssp. Bulgaricus were determined using MRS agar medium as described by **De Man et al.** (1960).

Streptococcus thermophiles was determined by using M17 selective medium as

described by **Krusch** *et al.* (1987). Plates were incubated at 37°C for 48hr.

Moulds and Yeasts Count

Were determined on oxytetracycline glucose yeast extract agar medium as suggested by **Harrigan and Mcconce**, (1966). Plates were incubated at 25°C for 3 days.

Coliform group

Were determined according to the **American Public Health Association** (1978). Appropriate dilutions of samples were plated on Mac Conk's agar medium and incubated at 37°C for 48hr.

Organoleptic properties

Organoleptic properties of yoghurt samples were evaluated according to **Tamime and Robinson (1999)**.

RESULTS AND DISCUSSION

Chemical Analysis of Yoghurt

Based on the results presented in Table 1. Generally, pH of all yoghurt samples decreased during storage up to 14 days. This phenomena was due to the growth of lactic acid bacteria and the production of lactic acid, which was due to the especial synergistic effect between *Lac*. spp and *Strep*. spp. (Yousef et al., 2013).

Also, there were slightly differences in pH values between control yoghurt and treated yoghurts during the storage period. These results were in agreement with those obtained by **Vijayalakshmi** *et al.* (2009).

It was clear from Table 1 that acidity values of all treatments increased during the progress of storage period.

Moreover, there were slightly differences in acidity values between control yoghurt and treated yoghurts during the storage period (Lamoureux et al., 2002) the increase in acidity attributed to the decrease in lactose content and post acidification

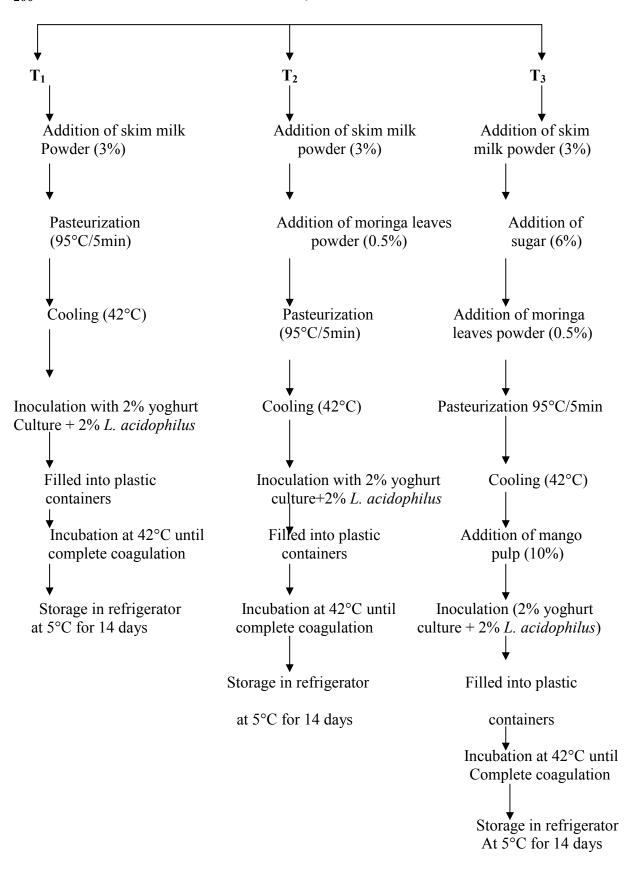


Table (1): pH and Acidity values of yoghurt fortified with moringa during storage up to 14 days at 5°C

Parameter	Storage (day)	Treatment			
		T_1	T_2	T_3	
	Fresh	4.60	4.50	4.50	
pН	7	4.18	4.13	4.13	
	14	4.14	4.07	4.06	
	Fresh	0.6	0.7	0.8	
Acidity (%)	7	0.8	0.9	0.9	
	14	0.9	0.9	1.0	

 T_1 : control (milk) T_2 : milk + moringa . T_3 : milk + moringa + mango + sugar.

especially by *L. delbreuckii* ssp. *bulgaricus* during the storage period. It was clear from Table 2 that the T.S of probiotic yoghurt slightly increased in all treatments during storage and the control treatment was lower than the others allover the storage period.

These results are similar to the values obtained by **Hashim** (2007). Moreover, the increase in total solids content during storage period attributed to the loss of moisture (**Tamime**, 1978). Also the protein content of all treatments gradually increased during storage and the control treatment was lower than the others allover the storage period. These results were in agreement with those obtained by **Salem** *et al.* (2013).

It was clear from Table 2 that the fat content of all treatments were slightly increased gradually during the progress of storage and the control treatment was lower than the others as storage period proceeded.

Increasing fat content in all treatments during storage may be due to the loss of moisture. These results are in agreement with those obtained by **Ismail** *et al.* (2006).

It is clear from this Table 2 that the moisture content of all treatments decreased gradually during the progress of storage. The values of control treatment was higher

than the other treatments during storage period.

The decrease in moisture contents of all treatments allover the storage period was probably due to the increase in total solid values.

Evaluation of Antioxidant activity (%) and Total phenolic content (TPC)

It is clear from Table 3 that the antioxidant activity in fresh yoghurt made with moringa (T_2) exhibited higher significant scavenging activity followed by yoghurt made with moringa + mango (T_3) while the plain yoghurt was found to have a lower scavenging effect. High potential of antioxidant activity of treatments may be due to that they are rich in photochemical contents, which possessed high antioxidant.

On the other hand, DPPH radical scavenging activity of all treatments dropped at the 7th and the 14th days of storage period. Data presented in Table 3 show that the total phenolic content (TPC) of yoghurt fortified with moringa and moringa + mango were significant higher than plain yoghurt. Moreover, at the 7th day of storage period the TPC of all samples decreased significantly and also at the 14th day of storage this may be due to the decreased in pH values throughout storage period.

Table (2): Chemical analysis of yoghurt fortified with moringa during storage up to 14 days at $5^{\circ}\mathrm{C}$

Chemical analysis (%)	Storage (day)		t	
		T ₁	T ₂	T ₃
	Fresh	13.2	14.6	18.4
Total solids	7	14.3	15.7	19.1
	14	15.9	16.8	20.5
	Fresh	3.5	3.9	4.0
Protein	7	3.6	4.2	4.3
	14	3.8	4.5	4.8
	Fresh	3.2	3.3	3.4
Fat	7	3.3	3.5	3.6
	14	3.4	3.6	3.7
	Fresh	86.8	85.4	81.6
Moisture	7	85.7	84.3	80.9
	14	84.1	83.2	79.5

T₁: control (milk) T₂: milk + moringa . T₃: milk + moringa + mango + sugar.

Table (3): Antioxidant activity (%) and Total phenolic content (TPC) of yoghurt fortified with moringa during storage up to 14 days at 5°C

Parameter	Storage (day)	Treatment		
	_	T ₁	T ₂	T ₃
	Fresh	67.34	93.46	87.83
Antioxidant activity (%)	7	43.85	84.63	75.39
	14	37.64	77.56	63.84
	Fresh	9.86	15.89	14.71
TPC (mg Gallic acid/100 gm.sample)	7	8.73	15.04	14.13
	14	7.93	13.85	12.96

 T_1 : control (milk) T_2 : milk + moringa . T_3 : milk + moringa + mango + sugar.

Microbiological Analysis

Results cleared that there were an obvious differences between treatments of bio-yoghurt in the viable numbers of S. thermopiles and L. bulgaricus when fresh and during storage period. The counts were increased up to the 7th day then decreased during the progress of storage. The highest count obtained was of voghurt made with moringa (T₂) followed by yoghurt made with moringa + mango (T_3) while the lowest count was in control. Moringa alone stimulate the growth of both (Lactobacillus acidophilus and Lactobacillus bulgaricus) more than moringa + mango. The decline in bacterial counts may be due to the decreasing in the pH value of yoghurt (Yannawa et al., 2014).

These results are in agreement with those obtained by Vijayalakshmi et al. (2009), Sharareh et al. (2015), Salem et al. (2013) moreover Van Tienen et al. (2011) suggested that the growth of the probiotics in M. oleifera-supplemented yoghurt was found to have a growthenhancing effect.

It was clear from this Table 4 that the count of *S.thermophilus* increased gradually up to the 7th day of storage then decreased during the progress of storage. The highest count obtained was in yoghurt made with moringa (T₂) while the lowest count obtained was in control (T₁). These results are in agreement with those obtained by Vijayalakshmi *et al.* (2009), VanTienen *et al.* (2011), Salem *et al.* (2013) and Sharareh *et al.* (2015).

It was clear from the same Table that yeast and mould and coliform group were not detected in all treatments allover the storage period.

Organoleptic properties

Data in Table 5 shows that the total scores of sensory evaluation of all treatments were gradually decreased during storage. This may be due to the increase in the acidity which affect the rheological properties. In general, the values of total sensory evaluation were in the following desending order $T_3 > T_1 > T_2$. These results are in agreement with those obtained by Madhu *et al.* (2012) and Sharareh *et al.* (2015).

Table (4): Microbiological analysis of yoghurt fortified with moringa during storage up to 14 days at 5°C

Type of culture	Storage (day)	Treatments (log (cfu/ml)		
	-	T ₁	T ₂	T ₃
	Fresh	8.32	8.45	8.44
L. acidophilus, L. bulgaricus count	7	9.42	10.30	10.13
	14	9.30	9.93	9.03
	Fresh	8.10	8.12	8.11
S. thermophilus count	7	8.93	10.15	10.04
	14	7.90	9.01	8.93
	Fresh	ND	ND	ND
Yeast & Mould and Coliform count	7	ND	ND	ND
	14	ND	ND	ND

 T_1 : control (milk) T_2 : milk + moringa . T_3 : milk + moringa + mango + sugar.

Table (5): Organoleptic properties of yoghurt fortified with moringa during storage up to 14 days at 5°C

Sensory parameter	Storage (day)	Treatment		
		T ₁	T ₂	T ₃
	Fresh	4.95	4.87	4.80
Appearance (5 marks)	7	4.95	4.87	4.80
	14	4.90	4.85	4.75
	Fresh	4.93	4.90	4.85
Body & Texture (5 marks)	7	4.93	4.90	4.85
	14	4.90	4.85	4.80
	Fresh	8.85	8.0	9.5
Flavor (10 marks)	7	8.85	8.0	9.5
	14	8.80	7.5	9.3
	Fresh	18.7	17.77	19.15
Total acceptance (20 marks)	7	18.7	17.77	19.15
	14	18.6	17.20	18.85

T₁: control (milk) T₂: milk + moringa. T₃: milk + moringa + mango + sugar.

Conclusion

Finally it was concluded, from the previous data that, the addition of 0.5% moringa leaves powder and 0.5% moringa leaves powder +10% mango pulp in the manufacture of pro-bioticyoghurt stimulate the growth of LAB and probiotic culture (L. acidophilus) so increased the nutritional value of yoghurt. Also all treatments had a high positive effect on total phenolic contents and its antioxidant properties. Moreover the addition of 0.5% moringa leaves powder +10% mango pulp in the manufacture of voghurt increased the acceptability of product more than the addition of moringa alone up to the 14th day of storage at 5°C.

REFERENCES

Abalaka, M.E.; Daniyan, S.Y.; Oyeleke, S.B. and Adeyemo, S.O. (2012). The antibacterial evaluation of *Moringa*

oleifera leaf extracts on selected bacterial pathogens. J. Microbiol. Res., 2: 1-4.

Salem, A.S.; Salama, W.M.; Hassanein, A.M. and El-Ghandour, H.M.A. (2013). Enhancement of nutritional and biological values of labneh by adding dry leaves of *Moringa oleifera* as innovative dairy.

Salem, A.S.; Salama, W.M.; Hassanein, A.M. and El-Ghandour, H.M.A. (2013). Enhancement of nutritional and biological values of labneh by adding dry leaves of *Moringa oleifera* as innovative dairy of nutritional and biological values of labneh by adding dry leaves of *Moringa oleifera* as innovative dairy products. World Appl. Sci. J., 22 (11): 1594-1602.

Ajila, CM. and Prasada, R.U.J. (2008).

Protection against hydrogen peroxide induced oxidative damage in rat

- erythrocytes by *Mangifera indica* L. peel extract. Food Chem. Toxicol., 46: 303-309.
- Alizadeh, A. and Ehsani, M.R. (2008). Probiotic survival in yogurt made from ultrafiltered skim milk during refrigeration storage. Res. J. Biol. Sci., 3: 1163-1165.
- **American Public Health Association** (1978). Standard Method for the Examination of Dairy Products. 14th Ed. Washington, USA.
- AOAC (2011). Association of official Analytical Chemists. Official Methods of Analysis. 17th Ed., Gaitherburg, MD, USA.
- Caceres, A.; Cabrera, O.; Morales, O.; Mollinedo, P. and Mendia, P. (1991). Pharmacological properties of *Moringa oleifera*. 1: Preliminary screening for antimicrobial activity. J. Ethnopharmacol., 33: 213-216.
- **Delong, D. (2003)**. How to Dry Foods. H.P books.
- De Man, J.C.; Rogosa, M. and Sharp, M.E. (1960). A medium for the cultivation of lactobacilli. J. Appl. Bacteriol., 23:130-136.
- Fadela, C.; Abderrahim, C. and Ahmed, B. (2009). Physicochemical and rheological properties of yoghurt manufactured with ewe's milk and skim milk. Afr. J. Biotec., 8: 1938-1942.
- Frazier, W.C. and Foster, E.M. (1961).
 Laboratory Manual for Dairy
 Microbiology. Burgoss publishing Co.
 USA.
- Harnkarnsujarit, N. and Charoenrein, S. (2011). Effect of water activity on sugar crystallization and beta-carotene stability of freeze-dried mango powder. J. Food Eng., 105: 592-598.
- Harrigan, W.F. and Mcconce, M.E. (1966). Laboratory Methods in

- Microbiology. Academic press London and New York.
- **Hashim, I.B. (2007).** Effect of cooling temperature and defrosting time on date quality during cold storage. The seventh annual U.A.E. Univ. Res. Conf., 1-10.
- Ismail, A.M.; Harby, S. and Salem, A.S. (2006). Production of flavored labneh with extended shelf life. Egypt. J. Dairy Sci., 34:59-68.
- Kasolo, J.N.; Bimenya, G.S.; Ojok, L.; Ochieng, J. and Ogwal-Okeng, J.W. (2010). Phytochemicals and uses of *Moringa oleifera* leaves in Ugandan rural communities. J. Med. Plants Res., 4: 753-757.
- Krusch, U.; Neve, H.; Luschei, B. and Teuber, M. (1987). Characterization of virulent bactreophages of *Streptococcus thermophilus* by host specificity and electron microscopy. Kieler Milschwirtschaftl. Forsch. Ber., 39: 155-167.
- **Lamoureux, L.; Roy, D. and Gauthier, S.F. (2002).** Production of oligosaccharides in yoghurt containing Bifidobacteria and yoghurt cultures. J. Dairy Sci., 85:1058.
- Li, W.; Hosseinian, F.S.; Tsopmo, A.; Friel, J.K. and Beta, T. (2009). Evaluation of antioxidant capacity and aroma quality of breast milk. Nutr., 25: 105-114.
- Madhu, S.; Prakash, M.; Anita, K.H. and Neetu (2012). An investigation of sensory evaluation of flavoured yoghurts made with different starter culture during storage. Int. J. Food and Nutr. Sci., 1:1.
- Makkar, H.P.S. and Becker, K. (1996).

 Nutritive value and antinutritive components of whole and ethanol extracted *Moringa oleifera* leaves.

 Animal Feed Sci. and Tech., 63 (1 -4): 211 218.

- Mishra, S.P.; Singh, P. and Singh, S. (2012). Processing of *Moringa oleifera* leaves for human consumption. Bull. Environ. Pharmacol. Life Sci., 2: 28-31.
- Saarela, M.; Lahteenmaki, L.; Crittenden, R.; Salminen, S. and Mattila-Sandholm, T. (2002). Gut bacteria and health foods-the European perspective. Int. J. Food Microbiol., 78: 99-117.
- Sahan, N.; Yasar, K. and Hayaloglu, AA. (2008). Physical, chemical and flavour quality of non-fat yoghurt as affected by a β-glucan hydrocolloidal composite during storage. Food Hydrocolloid, 22: 1291-1297.
- Salminen, S.; Ouwehand, A.; Benno, Y. and Lee, Y.K. (1999). Probiotics: how should they be defined? Trends Food Sci. Tech., 10: 107-110.
- Sharareh, H.; Kathryn, M.; Mohammad S. and Robert, G. (2015). Sensory evaluation of locally-grown fruit purees and inulin fibre on probiotic yoghurt in Mwanza, Tanzania and the microbial analysis of probiotic yoghurt fortified with *Moringa oleifera*. J. Health Popul Nutr., 33 (1): 60–67.
- **Tamime, A.Y. (1978).** The production of yoghurt and concentrated yoghurt from hydrolysed milk. Cult. Dairy. Prod. J., 13 (3): 16.

- Tamime, A.Y. and Robinson, R.K. (1999).
 Yogurt. Science and Technology.
 London, UK: Wood head Publishing.
- Van Tienen, A.; Hullegie, Y.; Hummelen, R.; Changalucha, J. and Reid, G. (2011). Development of a locally sustainable functional food for people living with HIV in Sub-Saharan Africa: laboratory testing and sensory evaluation. Published Online: October 10, 2011, DOI:http://dx.doi.org/10.3920/BM 2011. 0024, 2 (3): 193 198.
- Vijayalakshmi, R.; Nareshkumar, C. and Dhanalakshmi, B. (2009). Storage studies of cereal based low fat fruit yoghurt. Egypt. J. Dairy Sci., 38: 53-61.
- Vinderola, C.G. and Reinheimer, J.A. (2000). Enumeration of *Lactobacilii casei* in the presence of *L. acidophilus*, bifidobacteria and lactic starter bacteria in fermented dairy products. Int. Dairy J. 10: 271 275.
- Yannawa, S. and Bangkok (2014). Effect of that fruits on sensory properties of fruit yoghurt and survival of yoghurt starter culture added with probiotic strains in fruit yoghurt. Res. J. Pharmaceutical, Biol. and Chem. Sci., 5: 283-290.
- Yousef, M.; Nateghi, L. and Azadi, E. (2013). Effect of different concentration of fruit additives on some physicochemical properties of yoghurt during storage. Ann. Biol. Res., 4 (4): 244-249.

الملخص العربي

استخدام مسحوق أوراق المورنجا في إنتاج زبادي وظيف على المسورنجا في علا فتحي الشبيني ، ممدوح مصطفى كمال متولى المدالم ال

١. قسم بحوث الألبان، معهد بحوث تكنولولوجيا الأغذية، مركز البحوث الزراعية، وزارة الزراعة واستصلاح الأراضي، مصر

٢. قسم علوم وتكنولوجيا الأغذية والألبان، كلية العلوم الزراعية البيئية، جامعة العريش، مصر.

تم تصنيع الزبادي باستخدام لبن بقرى تم تقسيمه إلى ثلاثة أجزاء متساوية كل جزء تم معاملته كالاتي: المعاملة الأولى: تم تسخين اللبن على 90° م لمدة خمس دقائق ثم تم تبريده لدرجة حرارة 20° لتلقيح بكتريا البادئ المكونة من 20° لمن كلا من Lactobacillus delbrueckii ssp. bulgaricus and Streptococcus thermophiles من كلا من للمعاملة المعاملة التحضين على درجة حرارة 20° متى تمام التماسك وهذه المعاملة هي المعاملة الثانية تم إضافة 20° من مطحون اوراق المورنجا للبن قبل البسترة وباقى الخطوات كما في المعاملة الأولى وفي المعاملة الثالثة تم إضافة 20° سكر و 20° من مطحون أوراق المورنجا للبن قبل البسترة وأضافة 20° من مطحون أوراق المورنجا للبن قبل البسترة وأخلى النقيح بالبكتريا وباقي الخطوات كما في المعاملة الأولى النقيم النقوم الأولى وكذلك أثناء التخزين على 20° معد 20° بعد 20° بعد 20° من حيث تم عمل اختبارات كميائية وميكر وبيولوجية وحسية للمنتج ويمكن تلخيص النتائج كما يلى:

لوحظ انخفاض في قيم الpH لجميع المعاملات وكانت المعاملة الكنترول اعلى بنسبة بسيطة عن باقي المعاملات من اليوم الأول وحتى اليوم ٤٤ من فترة التخزين وازدادت قيم الحموضة اثناء فترة التخزين وكانت المعاملة الثالثة (زبادى بالْمَوٰرنجَا والْمانجو) أعلى من باقي المعاملات بينما كانت المعاملة الكنترول اقلّها من الّيوم الاول وحتى ١٤ يوم مُن فترة التخزين كما ازدادت قيم المادة الصلبة لجميع المعاملات اثناء فترة التخزين وكان الزبادي المدعم بالمانجو والمورنجا اعلى من باقى المعاملات بينما كانت المعاملة الكنترول اقلها من اليوم الأول وحتى نهاية فترة التخزين و ازدادت نسبة البروتين تدريجياً لجميع المعاملات اثناء فترة التخزين وكان الزبادي المدعِّم بالمورُّنجا والمأنجو اعْلَى من باقي المعاملات يليُّه الزبادي المدعم بالمورنجا بينما كانت المعاملة الكنترول اقلها بدءاً من اليوم الاول وحتى انتهاء فترة التخزين و أردادت نسبة الدهن معنويا لجميع المعاملات اثناء فترة التخزين وكانت المعاملة الكنترول أقل من باقى المعاملات خلال فترة التُخزين وانخفضت الرطوبة تدريجيا لجميع المعاملات اثناء فترة التخزين وكانت المعاملة الكنترول اعِلى في محتوى الرطوبَّة من باقى المعاملات بينماً كأن الزبادى المدعم بالمورنجا والمانجو اقلها في محتوى الرطوبة بدءاً مِن اليوم الاول وحتى نهاية فترة التخزين كما احتوى الزبادي المدعم بالمورنجا على قيم مرتفعة من النشاط المضاد للأكسدة ثم يعقبه الزبادي المدعم بالمورنَّجاً والمانجو وكانتُ المعاملة الكنترولُ اقل المعاملات في قيم النشاط المضاد للاكسدة وقد لوحظ الزبادي المعتلم بالمورك والمعاجو وقالت المعاملة التسرون الله المعاملات في نيم المعتلف المعتلف باليوم ؟ امن فترة الخفاض في قيم النشاط المضاد للاكسدة في اليوم السابع من التخزين وقد استمر هذا الانخفاض حتى اليوم الاول سجلت التخزين والمحتوى الكلى للفينولات في الزبادي المدعم بالمورنجا أم الزبادي المدعم بالمورنجا والمانجو بينما سجلت اقل قيمة للمعاملة الكنترول، أما خلال اليوم السابع من فترة التخزين حدث انخفاضا لقيم المحتوى الكلى للفينولات لجميع المعاملات واستمر هذا الانخفاض حتى اليوم ؟ امن فترة التخزين، إضافة كلا من المورنجا والمورنجا مع المانجو الى النبادة والمعاملات واستمر هذا الانخفاض حتى اليوم ؟ امن فترة التخزين، إضافة كلا من المورنجا والمورنجا مع المانجو الى النبادة والمعاملات واستمر هذا الانخفاض حتى اليوم ؟ امن فترة التخزين، إضافة كلا من المورنجا والمورنجا مع المانجو الى النبادة والمورنجا مع المانجو الى النبادة والمورنجا مع المانجو الم الزبادى أدى إلى تحفيز نمو كلا من L. bulgaricus, L. acidophilus and S. thermopiles حيث كان للمورنجاً بمفردها تأثيرا محفزا لنمو هذه البكتريا اكثر من المورنجا مع المانجو، وقد سجلت اعلي قيم لاعداد البكتريا في اليوم السابع من التخزين بينما حدث انخفاض لا عداد البكتريا في اليوم الرابع عشر من فترة التخزين بينما تلاشي تماما المحتوى الميكروبي من الخمائر والفطريات ومن بكتريا الكوليفورم في جميع المعاملات حتى نهاية فترة التخزين و حصل الزبادي الكنترول على اعلى درجة تحكيم في المظهر خلال اليوم الاول واليوم السابع من التخزين ثم يعقبه الزبادي المدعم بالمورنجا بينما حصل الزبادي المدعم بالمورنجا والمانجو على اقل درجة تحكيم في المظهر، وفي اليوم الرابع عشر من التخزين قلت درجات التحكيم لحميع المعاملات حصل الزبادي الكنترول على اعلى درجات تحكيم في القوام والتماسك في اليوم الاول واليوم السابع من التخرّين بينما حصِل الزبادي المدعم بالمورنجا والمانجو على اقلُ دِرجة تحكيم في القوآم والتماسك، وفي اليوم الرآبع عشر من التخزين قلت درجات التحكيم نسبيا وحصل الزبادي المدعم بالمورنجا والمانجو عل أعلى درجة تحكيم في النكهة ثم يعقبه الزبادي الكنترول في اليوم الأول واليوم السابع من فترة التخزين بينما حصل الزبادي المدعم بالمورنجا على اقل درجة تحكيم في النكهة، وفي اليوم الرابع عشر من فترة التخزين قلت درجات التحكيم نسبيا حصل الزبادي المدعم بالمورنجا والمانجو على أعلى درجات القبول العام ثم يعقبه الزبادي الكنترول من اليوم الاول وحتى ١٤ يوم من فترة التخزين بينما حصل الزبادي المدعم بالمورنجا على اقل درجة قبول عام عن باقي المعاملات خلال فترة التخزين ومن من النتائج المتحصل عليها يمكن القول أن تصنيع الزبادي وتدعيمه ب (٠,٥٪ مطحون أوراق مورنجا - ۰,۰% مطحون أوراق مورنجا + ۱۰% لب مانجو) أدى إلى تحفيز نمو بادئ الزبادي وايضا ال probiotic culture (المحتوى الكلى الفينولات والنشاط المضاد للاكسدة في الزبادي (L. acidophilus) النَّاتَجُ وَعُلُّوهُ عَلَى ذَلْكَ فَانَ تَدْعِيمُ الْزَّبَادَى بُ (٠,٠% مطحونَ أُورِ اقَ المُورُنَّجَا + ١٠% مانجو) أدى إلى تحسين الْقَبُولَ العام للزبادى اثناء فترة التخزين حتى ١٤ يوم على درجة حرارة ٥°م.

الكلمات الإسترشادية: المورنجا، إنتاج، زبادي.

المحكم___ون:

۱ ـ أ.د. سمير غنيم إبراهيم ۲ ـ أ.د. أميرة محمد الخولي