The Role of Green Tea Extract and Vitamin C on Malathion-Induced Testicular Oxidative Damage in Rats

اعداد

Suzan A. Saad

Faculty of Home Economics, Al- Azhar University, Nawag, Tanta, Egypt. suzansaad@azhar.edu.eg



مجلة البحوث في مجالات التربية النوعية

معرف البحث الرقمي DOI: 10.21608/jedu.2020.40535.1054

المجلد السادس العدد 26 . يناير 2020

الترقيم الدولى

P-ISSN: 1687-3424 E- ISSN: 2735-3346

موقع المجلة عبر بنك المعرفة المصري <u>/https://jedu.journals.ekb.eg</u>

موقع المجلة http://jrfse.minia.edu.eg/Hom

العنوان: كلية التربية النوعية . جامعة المنيا . جمهورية مصر العربية



دور مستخلص الشاي الأخضر وفيتامين ج في الضرر التأكسدي للخصية التي يسببها الملاثيون في الجرذان

د. سوزان سعد

المستخلص

الملاثيون هو أحد مبيدات الفوسفات العضوى التي تستخدم على نطاق واسع في الزراعة لمكافحة الحشرات. يؤثر الملاثيون على أعضاء الجسم كالجهاز التتاسلي عن طريق تثبيط الأستيل كولينستريز وإحداث الإجهاد التأكسدى. بحثت هذه الدراسة تأثير مستخلص الشاى الأخضر وفيتامين ج في الضرر التأكسدي (للخصية) الناجم عن الملاثيون في الجرزان. لذلك تم تصميم التجربة على النحو التالي: تم تقسيم ٤٨ من ذكور الجرذان البالغة إلى مجموعتين رئيسيتين (ستة جرذان لكل مجموعة .(مج1: مجموعة ضابطة سلبية. مج2 (٤٢ جرذ) تحقن بالملاثيون (١٥٠ مجم / كجم من وزن الجسم) في محلول ملحى لإحداث الضرر التأكسدي للخصية. تم تقسيم المجموعة الثانية إلى سبع مجموعات فرعية ، مجموعة ضابطة موجبة ، ومجموعات تم معالجتها بفيتامين ج (۲۰۰، ۲۰۰، مجم / كجم من وزن الجسم) ومجموعات تم معالجتها مستخلص الشاي الأخضرالكحولي والمائي (١٥٠ و ٣٠٠ مجم / كجم من وزن الجسم). بعد ٢٨ يوم ، تم تخدير كل المجموعات وقياس مؤشرات الحيوانات المنوية. تم فحص الخصية لقياس الإنزيمات المضادة للأكسدة و جمع عينات من الدم لتقييم مستويات هرمونات المصل ، وهي الهرمون المنشط للحويصلة (FSH) والهرمون المنشط للجسم الاصفر (LH) والتستوستيرون. أظهرت النتائج أن المجموعات المعالجة اثرت معنويا في حركية الحيوانات المنوية التقدمية ،الشكل الطبيعي ، إجمالي عدد الحيوانات المنوية ، هرمون التستوستيرون ، CAT. ،SOD ،GPX في المقابل ، انخفض مستوى FSH ومستوى . ($p \le 0.05$) مقارنة مع المجموعة الضابطة الموجبة. ($p \le 0.05$) مقارنة مع خلصت الدراسة الى أن أكسدة الدهون التي يسببها الملاثيون والإجهاد التأكسدي في خصية الجرذان .يبدو أن حمض الأسكوربيك ومستخلص الشاى الأخضر بتأثيرهم المضاد للأكسدة ، يمكن أن يحسن التغيرات السامة التي يسببها الملاثيون.

الكلمات الرئيسة: التحطم التأكسدي للخصية، الملاثيون، فيتامبن سي، الشاي الأخضر، مضادات الأكسدة، الهرمون المنشط للحويصلة، الهرمون المنشط للحسم الاصفر.

The Role of Green Tea Extract and Vitamin C on Malathion-Induced Testicular Oxidative Damage in Rats

Dr.Suzan A. Saad

Abstract

Malathion is one of organophosphate pesticides that is widely used in agriculture to control insects. This study investigated the role of green tea extract and Vitamin C on malathion-induced testicular oxidative damage in rats. Forty eight adult male rats were divided into 2 main groups. The first group (6 rats) considered as a negative control. The second group (42 rats) administered with Malathion (150 mg/kg body weight) in saline solution to induce testicular oxidative damage. The second group were divided into seven subgroups as a following: control group (+ve), two groups treated with vitamin C (100 and 200mg/ Kg B.W.) and groups treated with alcohol or aqueous green tea extract (150 and 300 mg/ Kg BW). After 28 days, all groups were anesthetized then sperm parameters were measured. Results showed that treated groups were associated with significant increase in sperm motility (%), Progressive motility (%), Normal form, total sperm count (%), luteinizing hormone (LH) and testosterone antioxidative enzymes i.e. GP_x, SOD and CAT. In contrast, follicle-stimulating hormone (FSH) and level of MDA were significantly ($p \le 0.05$) decreased as compared with (+ve) control. Malathion-induced lipid peroxidation and oxidative stress in the testis of rats. It seems that antioxidant capabilities ascorbic acid and green tea extract improved malathion-induced poisonous changes.

Key word: Testicular oxidative damage, malathion, vitamins C, green tea, antioxidants, sperm, LH, FSH, testosterone.

Introduction

Diethyl methoxy thio-phosphoryl thio-succinat is a chemical pesticide Malathion organophosphate pesticide family which is widely used in industry, agriculture to control insects on crops, produce ornamental plants, grasses, fruits, and vegetables as well as in the medical sector for disease vector control in many countries. [Dorri et al., 2015] These pesticides are absorbed through the skin and mucous membranes [Sarabia and Bustos, 2009] as affect various organs of the body including liver, kidneys, pancreas, and testis. Shah and Iqbal, 2010 1 It inhibits acetylcholinesterase activity of most eukaryotes. The major metabolites of malathion are mono- and di-carboxylic acid derivatives, and malaoxon is a minor metabolite. However, it is malaoxon that is the strongest cholinesterase inhibitor. [Bonner et al, 2007] oxidative stress has been lately planned as a major toxicity mechanism for organophosphorus insecticide both in acute and chronic poisoning cases [Ranjbar et al, 2005]. Malathion affects the reproductive system. Humans, birds, and other animals are in contact to increased levels of this insecticide due to its common utilization and the rising rates of food contamination [Babu et al., 2006]. Testis, by producing steroids and possessing an unfortunate antioxidant group may possibly become a strong goal for the chronic oxidative stress produced during ageing [Turner and Lysiak, 2008]. It recommended that testicular oxidative stress dysfunction of the organ may result in infertility[Tremellen, 2008].

Tea is a pleasant, common, communally accepted, and safe drink that was originally used as a medicine and is now recognized as a significant industrial and pharmaceutical raw material ^[] Bansal *et al.*,2012 ^[]. Green tea polyphenols, especially epigallocatechin gallate (EGCG) have several beneficial properties, including anticancer [Wang et al.,2018. Posadino et al.,2017 [] antioxidant, antidiabetic,antihypertensive, antimicrobial, and anti-metabolic syndrome effects [Zhang, 2016] as well as improving fertility in humans and animals [Jin et al., 2015]. Regular consumption of green tea is associated with a decreased risk of ovarian cancer in women [Lee et al., 2013] Green tea is considered as a dietary source of antioxidant compounds, mainly

comprising polyphenolic components like catechins and gallic acid. Green tea also contains numerous other factors, such as vitamin C, carotenoids, and tocopherols; minerals, such as Cr, Mn, Se, or Zn; and certain phytochemical compounds [Hashim et al., 2016]. These compounds might enhance the Green tea polyphenols antioxidant activity [Kim et al., 2003].

The ascorbic acid is a known antioxidant present in the testis with the precise role of protecting the latter from the oxidative damage [Nayanatara et al., 2003]. It also contributes to the support of spermatogenesis at least in part through its capacity to maintain this antioxidant in an active state. Vitamin C is itself maintained in a reduced state by a GSH-dependent dehydroascorbate reductase. which is abundant in the testes. Vitamin C has been shown to improve sperm motility and enhances semen quality and fertility of rats [Rekha et al.,2009]. Deficiencies of vitamins C or E leads to a state of oxidative stress in the testes that disrupts both spermatogenesis and the production of testosterone. In recent years, vitamin C supplements have been widely used in rats diets and the levels for enhancing production and reproductive performance have been increased several fold. Supplementation with Vitamin C has also been shown to increase total sperm output and sperm concentration [Nayanatara et al., 2003]. Therefore, the present study aims to investigate the role of green tea extract and Vitamin C on malathion-induced testicular oxidative damage in rats.

Materials and Methods

Materials

Malathion was obtained from Kafr El- Zayat pesticide and Chemicals Company, Kafr El- Zayat, , ElGharbia Governorate, Egypt. Casein (85% protein), choline chloride, DL-methionine, vitamins and salt mixture were obtained from El-Sharqiya Company, Vitamin C was obtained from ElGomhorya Company for Trading Drugs, Chemicals and Medical Instruments, Cairo, Egypt. The leaves of green tea, sunflower oil and corn starch were purchased from local market Tanta City, ElGharbia Governorate, Egypt.

Animal

Forty- eight normal male albino rats of Sprague Dawley Strain weighing (150 \pm 10g) were obtained from the laboratory Animal Colony. Ministry of Health and Population, Helwan, Cairo, Egypt.

Methods

Preparation of alcohol green tea extract

Eight hundred grams of shade dried leaf powder were immersed in 4 L of 95% ethanol and left for 24 h under constant stirring and filtered. This was repeated twice with 2 L of 95% ethanol. Thus, a total of 6.5 L filtrate were collected and concentrated by rotary vapor at 40°C. The yield of ethanol extract (EE) was been 42 g (5.25%). The extraction of selected plants was done by the method of Villasenor et al., [Villaseñor *et al*] 2002.

Preparation of aqueous green tea extract:

The green tea extract was made by soaking amount of green tea powder which equivalent alcohol green tea extract (12 g) in boiling distilled water (100 m) five mints then solution filtered to make an effective dose [Maity et al., 1988].

Chemical analysis

Leaves of green tea were subjected to chemical analysis in order to determine **the t**otal phenols: Phenolic compounds were determined by HPLC according to the method of [Goupy *et al.*, 1999] Central lab. of Food Technology Research Institute Agricultural Research Center, Cairo, Egypt.

Experimental design and animal groups

Rats of were kept in single wire cages with wire bottoms under hygienic conditions. The diet was introduced to the rats in special food containers to avoid scattering of food. Also water supply was given *ad-libitum* and check daily. Rats fed on basal diet for one week for adaptation. After this week, the rats were divided into two main groups as a following: The first main group (**G1**= 6 rats) fed on basil diet as a negative control. The second main group administered with malathion (150 mg/kg body. weight.) to induced testicular oxidative damage and divided into

divided into seven subgroups (SG) as the following: SG (1): was fed on basal diet as a positive control group (G+). SG (3&4) administrated orally with (100 and 200 mg vitamin C /kg body weight), respectively. SG (5&6) administrated orally with (150 and 300 mg alcohol green tea extract / kg body weight), respectively. SG (7&8) administrated orally with (150 and 300 mg aqueous green tea extract /kg body weight), respectively.

At the end of the experiment the rats were fasted overnight before sacrificed and the blood samples were collected from each rat and centrifuged to obtain the serum. Testis were collected and removed cleaned in saline solution, dried by filter paper and weighted.

Biological evaluation

During the experimental period (28day), the consumed diet was recorded everyday, feed intake (FI) and body weight was recorded every week. Biological evaluation of the different diets was carried out by determination of body weight gain % (BWG %) ¹ Chapman *et al.*,1986 ¹.

Sperm parameters

Sperm count, Sperm motility, progressive motility and normal form were calculated according to the methods of [Ekaluo *et al.*,2005] Ekaluo *et al.*,2013 ^{1.}

Biochemical analysis

Hormonal assay

The levels of hormones were measured in serum according to the principle highlighted by [[] Tietz, 1995 []] for testosterone while the method of Uotila [[] **Uotila** *et al* **1981** []] was

used for luteinizing and follicle stimulating hormones.

Antioxidant enzymes

Glutathione peroxidase (GPx), Malondialdehyde (MDA), (Super Oxide Dismutase (SOD) and Catalase (CAT) were determined according to the methods of Paglia, D.E. and Valentine, Ohkawa *et al.*, Nishikimi *et al.*, and Aebi, [[] Paglia and Valentine, Aebi., 1984 ^{].}

Statistical analysis

Statistical analysis was carried out using the program of Statistical Package for the Social Sciences (SPSS), PC statistical software (Version 20; Untitled–SPSS Data Editor). The results were expressed as mean ± Standard deviation (mean ± SD). Data were analyzed using one way classification, analysis of variance (ANOVA) [Armitage and Berry (1987).].

Results and Discussion

Chemical analysis

Total phenolic compounds in green tea

High-performance liquid chromatography (HPLC) analysis of green tea leaves are reported in Figure (1). HPLC analysis of leaves of green tea revealed the presence of twenty compounds in leaves of green tea. The major components are found to be catechein and chlorogenic while the lowest compounds are Alpha –coumaric and Cinnamic.

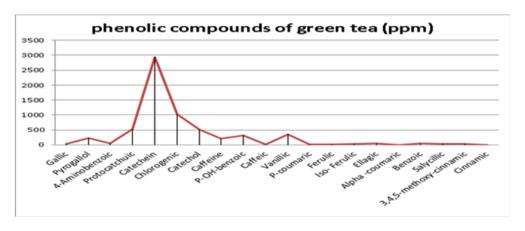


Figure (1): Phenolic compounds of green tea (ppm) by HPLC analysis

Biological evaluation

Effect of green tea extracts (alcoholic & aqueous) and vitamins C on feed intake (FI), body weight gain % (BWG) and testis weight in testicular oxidative damage rats

Data presented in Table (1) showed the effect of (alcoholic and aqueous) green tea extracts and vitamins C on feed intake

(FI), body weight gain (BWG, %) and testis weight in testicular oxidative damage rats. All treated groups showed increase in FI and the best results were found in vitamin C (200 mg/kg) and aqueous green tea extract (300 mg/kg) which recorded nonsignificant with negative control. All treated groups recorded significant increase (P≤0.05) in BWG% as compared to positive control group. The best results recorded in treated group with vitamin C (200 mg/kg). Results of testis weight recorded significant decrease in mean value of (+ve) control group as compared to all treated groups which showed improvement in testis weight when compared with (-) control group. The best results were recorded for the groups treated with Vitamin C (100 and 200 mg) and alcohol green tea extracts (300 mg/kg). Administration of aqueous extract of Green tea to pesticides cyromazine and chlorpyrifos treated groups has an ameliorated effect in the loss of body weight [Heilkal et al., 2013, Reddy et al., 2017] 1. Rats administered with aqueous green tea extract showed the observed testis weight loss in (+ve) control group may be due to reduced sex hormones [Nashwa et al., 2011

Table 1: Effect of green tea extracts (alcoholic & aqueous) and vitamins C on feed intake (FI), body weight gain % (BWG) and testis weight in testicular oxidative damage rats

Parameters Groups	FI (g)	BWG (%)	Testis weight (g)
Control –ve	252±2.0°	34.916±1.24 ^a	2.27 ± 0.02^{a}
Control +ve	240±2.0 ^d	19.84±1.16 ^f	$1.75\pm0.08^{\rm e}$
Vitamin C 100 mg	244 ± 2.0^{c}	25.19 ± 1.00^{c}	2.21 ± 0.02^{abc}
Vitamin C 200 mg	250±2.0 ^{ab}	29.63±1.10 ^b	2.25 ± 0.02^{ab}
Alcohol green tea extract 150 mg	244±1.0°	21.57±1.15 ^e	2.14±0.02 ^{cd}
Alcohol green tea extract 300 mg	247±1.0 ^{bc}	22.62±0.68 ^{de}	2.25±0.02 ^a
Aqueous green tea extract 150 mg	246±2.0°	22.68±0.53 ^{de}	2.11±0.04 ^d
Aqueous green tea extract 300 mg	250±2.0 ^{ab}	23.69±0.79 ^{cd}	2.18±0.04°

^{*}Values denote arithmetic means \pm SD. Means with different letters in the same column differ significantly at p \leq 0.05.

Effect of green tea extracts (alcoholic & aqueous) and vitamins C on the sperm parameters in testicular oxidative damage rat

Data listed in Table (2) declared that (+ve) control group showed a significant reduction in Sperm motility (%) as compared to all treated groups. The best result was recorded for the group treated with alcohol green tea extracts (150 and 300 mg/kg) and Vitamin C (200 mg) which recorded non- significant with negative control. As for Progressive motility (%) and Normal form (%) it could be observed that experimental groups treated with green tea extracts (alcoholic & aqueous) and vitamins C had noticed improvement which recorded increase in Progressive motility (%) and Normal form (%). The best results were recorded for the groups treated with Vitamin C (200 mg) and alcohol green tea extract (300 mg/kg).

The same table illustrate the change in total sperm count for controls and treated groups. Results of total sperm count showed that all treated groups had significant increase as compared to (+ve) control group. The best result recorded for the groups treated with alcohol green tea extract (300 mg/kg). Vitamin C can be found in high concentrations in seminal plasma [Larson et al., 2003 Jouanne et al. 1988] 'As vitamin C intake increases its concentration in seminal plasma rises and prohibits DNA damage [Douglas et al., 2005]. Administration of green tea improve sperm parameter because green tea significantly increases the antioxidant capacity in plasma as well as spermatozoa after consumption of 2-6 cups/day which may lead to decrease oxidative damage of lipids and DNA [Henning et al., 2003, Xu et al., 2004, Higdon and Frei, 2003]. Green tea increases the antioxidation level and protects against oxidative damage in humans. Therefore, green tea regulates defensive mechanisms against oxidative damage [Erba et al. 2005]. In addition to green tea antioxidant properties, it's may also decrease inflammation, reduce DNA fragmentation, and increase the motility and viability of semen [Asadi et al., 2017]. Other potential benefits of polyphenols of green tea include improved egg viability and reduced cellular damage of reproductive organs. In addition, its antioxidative concentration correlates with sperm levels and motility [Sreejaya and Nirmala 2016].

Table 2: Effect of green tea extracts (alcoholic & aqueous) and vitamins C on the sperm parameters in testicular oxidative damage rat

Parameters Groups	Sperm motility (%)	Progressive motility (%)	Normal form (%)	Total sperm count (10 ⁶)
Control –ve	66.67 ± 2.89^{a}	56.67±3.51 ^a	68.33±2.89 ^a	32.52 ± 0.80^{a}
Control +ve	38.67 ± 3.06^{d}	33.33 ± 2.89^{c}	$36.00\pm3.61^{\text{f}}$	17.49±1.25 ^e
Vitamin C 100 mg	56.67±2.89 ^{bc}	46.33±3.21 ^b	57.33±2.52 ^{cd}	23.25±0.73 ^d
Vitamin C 200 mg	61.00±3.61 ^{ab}	48.33±2.89 ^b	66.00±3.61 ^{ab}	26.34±1.34°
Alcohol green tea extract 150 mg	61.67±3.51 ^{ab}	44.67±2.52 ^b	56.33±1.53 ^d	25.66±0.69°
Alcohol green tea extract 300 mg	64.67±2.52 ^a	54.00±3.61 ^a	63.33±2.89 ^{ab}	29.64±0.69 ^b
Aqueous green tea extract 150mg	51.67±2.89°	43.33±3.51 ^b	50.33±2.52 ^e	24.52±1.26 ^{cd}
Aqueous Green tea extract 300mg	58.00±3.46 ^b	46.00±3.61 ^b	61.67±2.89 ^{bc}	25.65±0.93°

^{*}Values denote arithmetic means \pm SD. Means with different letters in the same column differ significantly at p \leq 0.05.

Biochemical analysis

Hormonal assays

Data found in the Table (3) declared that LH (mIU/mL) and testosterone (ng/ml) in all treated groups with green tea extracts (alcoholic & aqueous) and vitamins C recorded significant increase as compared to positive control group. The best results in LH and testosterone reported in the group treated with Vitamin (C 200 mg) which recorded the nearest mean value from normal control group.

As for FSH (mIU/mL) it could be observed that positive control group achieved mean value higher than negative and all treated groups with significant difference between them. The best result founded in the group treated with Vitamin (C 200 mg) which recorded the nearest mean value from normal control group.

Testosterone is produced primarily in the gonads under the influence of the pituitary FSH and LH. The observations made from the results of this study therefore suggested that malathion the potential to interact and disrupt the functionality of the gonadal tissues in male rats. In our study, vitamin C administration to rats exposed to malathion produced an appreciable increase in the serum testosterone level to the level within the range of the control level and prevention of the alteration of the architectural integrity of the testicular tissues. These observations gave an indication that vitamin C counteracted the adverse effects of the constituents of malathion on the gonadal tissues in male rats. The result of this study agrees with the previous reports that vitamin C is an effective antioxidant in various biological systems [Ayo et al 2006 Ambali et al., 2007, Chen et al., 2000. Frei, 2004 J.,

Table 3: Effect of green tea extracts (alcoholic & aqueous) and vitamins C on Luteinizing hormone (LH), Follicle-stimulating hormone (FSH) and Testosterone in testicular oxidative damage rats

Parameters Groups	LH (mIU/mL)	FSH(mIU/mL)	Testosterone (ng/ml)
Control –ve	2.87 ± 0.08^{a}	2.79±0.03 ^g	2.91±0.03 ^a
Control +ve	$1.42\pm0.04^{\rm f}$	5.33 ± 0.06^{a}	$1.82\pm0.05^{\text{f}}$
Vitamin C 100 mg	$2.54\pm0.04d^{e}$	3.06 ± 0.05^{d}	2.52 ± 0.02^{e}
Vitamin C 200 mg	2.71 ± 0.05^{b}	$2.89\pm0.04^{\rm f}$	2.82 ± 0.02^{b}
Alcohol green tea extract 150 mg	2.45±0.05 ^e	3.18±0.04°	2.48±0.03 ^e
Alcohol green tea extract 300 mg	2.63±0.07 ^{bc}	2.98±0.03 ^e	2.70±0.04 ^c
Aqueous green tea extract 150 mg	2.45±0.04 ^e	3.30±0.03 ^b	2.59±0.04 ^d
Aqueous green tea extract 300 mg	2.60±0.03 ^{cd}	3.10±0.05 ^d	2.66±0.03°

*Values denote arithmetic means \pm SD. Means with different letters in the same column differ significantly at p \leq 0.05

Green tea extract indicate stimulation in testosterone synthesis in testes by components of green tea extract which increase the activity of enzymes which are responsible for synthesis and metabolism of lydig's cells steroids by stimulation of interstitial cells stimulating hormone (ICSH)which convert cholesterol to pregnonolone and then testosterone. Also spermatogenic stimulating hormone (FSH) or (SSH)) play a role in testosterone synthesis by increasing receptors sensitivity to (LH) or (ICSH)) which found on the surfaces of leydig's cells and then increase steroidogensis and testosterone release in the seminiferous tubules ¹ Pineda *et al.*, 2006 ¹.

Antioxidant enzymes

Glutathione peroxidase (GPx), Super Oxide Dismutase (SOD) and Catalase (CAT) were determined as important endogenous antioxidant enzymes in the testis. As can be seen from Table 4, the malathion-treated group has MDA levels that were substantially increased (p < 0.05), and their GPx, SOD and CAT levels were significantly reduced (p < 0.05). Results of GPx, SOD and CAT showed that all treated groups recorded significant increase when compared with positive control group. On the other hand, all treated groups recorded significant decrease in MDA when compared with positive control group.

Table 4: Effect of green tea extracts (alcoholic and aqueous) and vitamins C on antioxidant enzymes (GP_X, SOD, CAT and level of MDA) of testicular tissue in testicular oxidative damage rats

Parameters Groups	GPX ng/mg	SOD U/L	CATng/mg	MDA nmol/mg
Control –ve	0.40 ± 0.05^{a}	0.34 ± 0.03^{a}	0.31 ± 0.04^{a}	0.14 ± 0.01^{d}
Control +ve	0.18 ± 0.02^{d}	0.19 ± 0.02^{d}	0.19 ± 0.02^{c}	0.28 ± 0.02^{a}
Vitamin C 100	0.30 ± 0.01^{b}	0.28±0.01 ^{bc}	0.27 ± 0.02^{ab}	0.19 ± 0.01^{bc}
mg				
Vitamin C 200	0.37±0.02 ^a	0.32 ± 0.06^{ab}	0.30±0.04 ^a	0.16 ± 0.02^{d}
mg				

Alcohol green tea extract 150 mg	0.25±0.04°	0.28±0.01 ^{bc}	0.28±0.01 ^{ab}	0.19±0.01 ^{bc}
Alcohol green tea extract 300 mg	0.31±0.02 ^b	0.31±0.02 ^a	0.30±0.01 ^a	0.17±0.02 ^{cd}
Aqueous green tea extract 150 mg	0.30±0.01 ^b	0.25±0.02°	0.24±0.01 ^b	0.21±0.02 ^b
Aqueous green tea extract 300 mg	0.31±0.01 ^b	0.31±0.01 ^{ab}	0.28±0.01 ^a	0.16±0.01 ^d

^{*}Values denote arithmetic means \pm SD. Means with different letters in the same column differ significantly at p \leq 0.05.

Pesticides by making some changes in DNA or its binding protein can damage the testis tissues and cause mutations in spermatogonia cells, which ultimately lead to changes in the sperm. Ogutcu *et al.*, 2006 And also, it is proved that any increase in the levels of MDA and reducing the antioxidant immune system. Jahromi *et al.*, 2012 In such circumstances, along with avoiding exposure to organophosphate pesticides, use of an appropriate antioxidant such as ascorbic acid used in this research can reduce adverse effects of exposure to these pesticides. Vitamin C is available in many foods and easily intake an antitoxic effect by daily consuming Moron *et al 1979* Ascorbic Acid is a water-soluble vitamin that can decrease the amount of free radicals through its antioxidant properties Sutcu *et al 2006*.

Antioxidative enzymes are activated by green tea extracts intake ^[]. Frei and Higdon 2003^[], and the antioxidative strength of human plasma increases with continual ingestion of green tea ^[] Kimura *et al 2006*, ^[] These antioxidative defense systems might also prevent oxidative damage in the brain. Long-term intake of green tea extracts may be important because cells are constantly exposed to oxidative stress ^[] Khan *et al 1992*^[].

Conclusion

The present study data demonstrate that malathion can produce adverse effects on fertility, reproductive performance, and sperm parameters in male rats. In addition, it seems that ascorbic acid and green tea extract due to its antioxidant capabilities, can improve malathion-induced poisonous changes.

References

Aebi, H. Methods enzymol, 1984. 105: 121-126.

Ambali S, Akanbi D, Igbokwe N, Shittu M, Kawu M, Ayo J. Evaluation of subchronic chlorpyrifos poisoning on haematological and serum biochemical changes in mice and protective effect of vitamin C. The J Toxicol Sc 2007; 32 (2): 111-120.

Armitage, P. and Berry, G. (1987): Statistical Methods in Medical Research. Blackwell, Oxford, UK, pp. 93-213.

Asadi, N.; Bahmani, M.; Kheradmand, A.; Rafieiankopaei, M. The Impact of Oxidative Stress on Testicular Function and the Role of Antioxidants in Improving it: A Review. J. Clin. Diagn. Res. 2017, 11, IE01.

Ayo JO, Minka NS, Mamman M. M. Excitability scores of goats administered ascorbic acid and transported during hot-dry conditions. J Vet Sc 2006; 7(2):127-131...

B. Frei, J.V. Higdon. Antioxidant activity of tea polyphenols in vivo: evidence from animal studies. J Nutr, 133 (10) (2003), pp. 3275-3284.

Babu NS, Malik JK, Rao GS, Aggarwal M, Ranganathan V. Effects of subchronic malathion exposure on the pharmacokinetic disposition of pefloxacin. Environ Toxicol Pharmacol 2006, 22(2):167–171.

Bansal, S.; Syan, N.; Mathur, P.; Choudhary, S. Pharmacological profile of green tea and its polyphenols: A review. Med. Chem. Res. 2012, 21, 3347–3360.

Bonner M.R.; Coble, J.; Blair A.; Beane- Freeman L.E.; Hoppin J.A.; Sandler, D.P. and Alavanja, M.C. Malathion exposure and the incidence of cancer in the agricultural health study. Am J. Epidemiol. 2007; 1; 166(9):1023-1034.

Chapman, D.G.; Castilla, R. and Campbell, J.A. Evaluation of protein in food. LA. method for the determination of protein efficiency ratio. Can. J. Biochem. Physiol., 1959. 37: 679 – 686.

Chen K, Suh J, Carr AC, Morrow JD, Zeind J, Frei B. Vitamin C suppresses oxidative lipid damage in vivo, even in the presence of iron overload. Am J Physiol, Endocrinol Metab 2000; 279(6): 1406-1412.

Coimbra, E. Castro, P. Rocha ereira, I. Rebelo, S. Rocha, A

- Dorri S.A; Hosseinzadeh H; Abnous K; Hasani F.V;, Robati R.Y and Razavi B.M. Involvement of brain-derived neurotrophic factor (BDNF) on malathion induced depressive-like behavior in subacute exposure and protective effects of crocin. Iran J Basic Med Sci 2015;958:18-66.
- Douglas-Hamilton DH, Smith NG, Kuster CE, Vermeiden JP, Althouse GC. Particle distribution in low-volume capillary-loaded chambers. J Androl. 2005;26:107–14.
- Ekaluo, U.B.; Udokpoh, A.E.; Udofia, U.U. and Ajang, R.O. Comparative toxicity of five commonly used analgesics on sperm count and sperm head abnormalities. Glob. J. Pure Applied Sci., 2005. 11: 81-84.
- Ekaluo, U.B., E.V. Ikpeme, Y.B. Ibiang and O.S. Amaechina, Attenuating role of vitamin C on sperm toxicity induced by monosodium glutamate in albino rats. J. Biol. Sci., 2013. 13: 298-301.
- Erba, D.; Riso, P.; Bordoni, A.; Foti, P.; Biagi, P.L.; Testolin, G. Effectiveness of moderate green tea consumption on antioxidative status and plasma lipid profile in humans. J. Nutr. Biochem. 2005, 16, 144–149.
- Frei B. Efficacy of dietary antioxidants to prevent oxidative damage and inhibit chronic disease. J Nutrition 2004; 134 (Suppl.): 3196-3198.
- Goupy, P.; Hugues, M.; Biovin, P. and Amiot, M. antioxidant composition and activity of barley (Hordeum vulgare) and malt extracts of isolated phenolic compounds. J.Sci. Food Agric., 1999.79:1625-1634.
- Hashim, F.; Tvrdá, E.; Massányi, P.; Stawarz, R.; Lukáč, N. Effects of biological active substances to the spermatozoa quality. *J. Microbial. Biotechnol. Food Sci.* 2016, *5*, 263–267.
- Heilkal, T.M.; Mossa, A.T.; Rasoul, M.A. and Mare, G.I. The ameliorating effects of green tea extract against cyromazine and chlorpyrifos induced liver toxicity in male rats. Asian J. Pharm .Clin. Res., 2013. 6 (1): 48-55.
- Henning, S.M.; Fajardo-Lira, C.; Lee, H.W.; Youssefian, A.A.; Go, V.L.; Heber, D. Catechin Content of 18 Teas and a Green Tea Extract Supplement Correlates With the Antioxidant Capacity. Nutr. Cancer 2003, 45, 226–235.
- Higdon, J.V.; Frei, B. Tea catechins and polyphenols: Health effects, metabolism, and antioxidant functions. Crit. Rev. Food Sci. Nutr. 2003, 43, 89–143.

- Jahromi VH, Koushkaki M, Kargar H. The effects of malathion insecticide on ovary in female rats. National park forschung in der schweiz. Switzerland Res Park J 2012;101:231-5.
- Jin, D.; Hui, W.; Zhen-Biao, W.; Jie, Z.; Shun, Z.; Wei, L. Protection of murine spermatogenesis against ionizing radiation-induced testicular injury by a green tea polyphenol. Biol. Reprod. 2015, 92, 1–13.
- Jouannet P, Ducot B, Spira A. Male factors and the likelihood of pregnancy in infertile couples. I. Study of sperm characteristics. Int J Androl. 1988;11:379–94.
- Kim, J.H.; Kang, B.H.; Jeong, J.M. Antioxidant, Antimutagenic and Chemopreventive Activities of a Phyto-extract Mixture Derived from Various Vegetables, Fruits, and Oriental Herbs. *Food Sci. Biotechnol.* 2003, *12*, 631–638.
- Larson KL, DeJonge CJ, Barnes AM, Jost LK, Evenson DP. Sperm chromatin structure assay parameters as predictors of failed pregnancy following assisted reproductive techniques. Hum Reprod. 2000;15:1717–22.
- Lee, A.H.; Su, D.; Pasalich, M.; Binns, C.W. Tea consumption reduces ovarian cancer risk. Cancer Epidemiol. 2013, 37, 54–59.
- M. Kimura, K. Umegaki, Y. Kasuya, A. Sugisawa, M. Higuchi.The relation between single/double or repeated tea catechin ingestions and plasma antioxidant activity in humans. Eur J Clin Nutr, 56 (12) (2002), p. 1186.
- Maity, S.; Vadasiromoni, J.; and Ganguly, D. Role of glutathione in the antiulcer effect of hot water extract of green tea. JPN J. Pharmacol., 1988, 78:285-292.
- Moron MS, Depierre JW, Mannervik B. Levels of glutathione, glutathione reductase and glutathione Stransferase activities in rat lung and liver. Biochimica et Biophysica Acta 1979; 582: 67-7.
- Nashwa A, Abu A, Kawkab A, Ahmed, Samar MM. The protective effect of Ginger and N-Acetyl cystine on ciprofloxacin induced reproductive toxicity in male rats. Journal of American Sciences. 2011;7(7):741–52.
- Nayanatara AK, Vinodini NA, Ahemed B, Ramaswamy CR, Bhat Shabarianth Ramesh. Role of ascorbic acid in monosodium glutamate mediated effect on testicular weight, sperm morphology and sperm count, in rat testis. Journal of Chinese Clinical Medicine. 2008;3 (1):1–5.
- Nishikimi, M.; Roa, N.a. and Yogi, K. Biochem. Bioph. Res. Common., 1972. 46,849-854.

Ogutcu A, Uzunhisarcikli M, Kalender S, Durak D, Bayrakdar F, Kalender Y. The effects of organophosphate insecticide diazinon on malondialdehyde levels and myocardial cells in rat heart tissue and protective role of Vitamin E. Pestic Biochem Physiol 2006;86:93-8.

Ohkawa, H.; Ohishi, W. and yagi, k. Anal. Biochem., 1979. 95, 351.

Paglia, D.E. and Valentine, W. J. Lab. Clin. Med., 1967. 70:158-169.

Pineda, M.H.and Dooly, M.P. McDonald's Veterinary Endocrinology and Reproduction. 5 thed.lowa State Press.A.Blackwell publishing Company. 2003.pp.17-32;239-256.

Posadino, A.M.; Phu, H.T.; Cossu, A.; Giordo, R.; Fois, M.; Dtb, T.; Piga, A.; Sotgia, S.; Zinellu, A.; Carru, C.; et al. Oxidative stress-induced Akt downregulation mediates green tea toxicity towards prostate cancer cells. Toxicol in Vitro 2017, 42, 255–262.

Ranjbar A.; Solh H; Mashayekhi F.J; Susanabdi A. A. and Rezaie M. AbdollahiOxidative, stress in acute human poisoning with organophosphorus insecticides; a case control study Env Toxicol Pharmacol, 20 (1) (2005), pp. 88-91.

Reddy, M.A.; Kumar, B.K.; Boobalan, G.; Reddy, K.M.; kumar, C. S. V.; Reddy, K.M. and Lakshman, M. Hepatoprotective potential of green tea extract against experimental hepatotoxicity in rats. Indian J. Pharm. Sci, 2017.79 (1): 58-64.

Rekha DK, Nayanatara AK, Ramaswamy, Sheila RP, Bhat Ramesh, Venkappa SM. Infertility in male wistar rats induced by cadmium chloride: role of ascorbid acid. Journal of Chinese clinical medicine.2009;4 (11):616–21.

S.G. Khan, S.K. Katiyar, R. Agarwal, H. MukhtarEnhancement of antioxidant and phase II enzymes by oral feeding of green tea polyphenols in drinking water to SKH-1 hairless mice: possible role in cancer chemopreventionCancer Res, 52 (14) (1992), pp. 4050-4052.

Santos-Silva. The effect of green tea in oxidative stress. Clin Nutr, 25 (5) (2006), pp. 790-796.

Sarabia L; Maurer I and Bustos-Obregón E. Melatonin prevents damage elicited by the organophosphorous pesticide diazinon on the mouse testis. Ecotoxicol Environ Saf. 2009; 72:938-42.

Sendcor, G. and Cochran, W. Statistical Method 6th ed. Lowa State Collage U. S. A. 1979. 841.

Shah M.D and Iqbal M. Diazinon-induced oxidative stress and renal dysfunction in rats. Food Chem Toxicol 2010;48:3345-53.

Sreejaya, P.; Nirmala, P. Evaluation of anti-oxidant effect of green tea extract in cryo preserved human semen samples. Int. J. Curr. Res. 2016, 8, 27270–27274.

Sutcu R, Altuntas I, Yildirim B, Karahan N, Demirin H, Delibas N, et al. The effects of subchronic methidathion toxicity on rat liver: Role of antioxidant Vitamins C and E. Cell Biol Toxicol 2006;22:221-7.

Tietz, N.W. Clinical guide to laboratory tests. 3rd ed. Philadelphia: W.B. Saunders, Co.; 1995. p.405.

Tremellen K.. Oxidative stress and male infertility—a clinical perspective Hum Reprod Update, 14 (3) (2008), pp. 243-258.

Turner T.T. and Lysiak J.J. Oxidative stress: a common factor in testicular dysfunction J Androl, 29 (5) (2008), pp. 488-498.

Uotila, M., Ruoslathi, E., and Envall E. Two-site sandwich enzyme immunoassay with monoclonal antibodies to human alphafetoprotein. J Immunol Methods. 1981,42:11-15.

Villaseñor, I.M.; Echegoyen, D.E. and Angelada, J.S. A new antimutagen from Mentha cordifolia Opiz. Mutat Res.J., 2002. 515: 141-146.

Wang, X.; Li, X.; Liang, X.; Liang, J.; Zhang, C.; Yang, J.; Wang, C.; Kong, D.; Sun, H. ROS-Responsive Capsules Engineered from Green Tea Polyphenol-Metal Networks for Anticancer Drug Delivery. J. Mater. Chem. B 2018, 6, 1000–1010.

Xu, J.Z.; Yeung, S.Y.; Chang, Q.; Huang, Y.; Chen, Z.Y. Comparison of antioxidant activity and bioavailability of tea epicatechins with their epimers. Br. J. Nutr. 2004, 91, 873–881.

Zhang, L. Alleviation of Metabolic Syndrome by Green Tea Polyphenol EGCG. Master's Thesis, The State University of New Jersey, Rutgers, NJ, USA, 2016.