

Mycotoxins effect on male fertility Hormones

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Abstract

A total of 12 fungal species belonging to 8 genera were isolated from various collected and examined samples. *Fusarium graminearum* was grown on suitable medium for crude toxin production, 2ry metabolites were used for treating experimental animal models (not for control). After treatment period blood samples were collected for hormonal assays, Serum testosterone concentration was greatly reduced but follicle stimulating hormone values showed significant increase in treated samples. Testosterone and follicle-stimulating hormone are potential factors required to obtain male fertility and the process of spermatogenesis in the testis

Introduction

Mycotoxins are toxic secondary metabolites produced by fungi (molds) that cause an undesirable effect (mycotoxicosis) when human are exposed. Exposure is usually by consumption of contaminated foods, but may also by contact or inhalation. Biological effects include liver and kidney toxicity, central nervous system effects and estrogenic effects, to name a few. Only some molds produce mycotoxins and they are referred to as toxigenic. The fungal toxins are chemically diverse representing a variety of chemical families and range in molecular weight from about 200 to 500 kD. There are hundreds of known mycotoxins, but few have been extensively researched and even fewer have

good methods of analysis available. The primary classes of mycotoxins are aflatoxins, zearalenone, trichothecenes, fumonisins, ochratoxin A and the ergot alkaloids. (CAST, 2003).

Testosterone and follicle-stimulating hormone (FSH) are required to obtain full reproductive potential. In the tests, somatic Sertoli cells transduce signals of testosterone and FSH resulting the production of factors that are required by germ cells as they mature into spermatozoa (Walker, and Cheng, 2005)

The present study aimed to clarify the effect of crud toxins on male fertility using experimental model animals

Materials and Methods

Isolation of fungi

Three different culture media were used to isolate the fungi: Czapek-Dox agar, Malt extract-glucose agar and Potato-dextrose agar (PDA). The antibiotic, chloramphenicol, was added at a concentration of 0.5 mg/ml medium before autoclaving to suppress bacterial growth. Fungi in the samples were detected and examined using the direct plating method (Flannigan, 1977). Samples of 10 g of each grains (wheat, barley, rice, maize and sorghum) were surface-sterilized in 1% NaOCl for 1 minute and rinsed twice in sterile distilled water. The surface sterilized grains were aseptically transferred onto the solidified agar. A total of 9 plates were plated per sample. Ten grains were plated on each agar plate. Inoculated plates were incubated for seven days at 27°C prior to visual differentiation and counting of colonies (Czerwiecki et al, 2002). The different fungal colonies on the plates were subcultured on PDA media for identification of species (Raper and Fennell, 1977; Domsch et al, 1981; Nelson et al, 1983 ; Pitt, 1985, 2000).

Production of 2ry metabolites (crude toxins)

The selected fungus (*Fusarium graminearum*) was grown on plates of potato dextrose agar for one week, inoculum of the tested fungus was transferred to test tubes containing 10 ml of sterilized phosphate buffered saline mixed thoroughly , then adjusted to

approximately 10^5 C.F.U/ml. One ml of fungul spore suspension was inoculated into autoclaved conical flask containing 100 ml of Sabouraud's dextrose broth liquid media , incubated for 10 days at 28 °C , filtered under aseptic conditions using bacterial filter (0.2 µl pore size Millipore – USA)

The supernatant was processed for crude toxin determination according to the method of Biuret method (Henry, 1964) using ready made kit (Biocon-Germany). Protein concentrations was expressed as µg/dl. Then the secondary metabolites were kept in clean sterilized vials in the fridge for experimental procedures.

Treatment of experimental animal :

Eighteen wister male albino rats (10-12 weeks of age) weighing 120- 150 g were obtained from animal house research unit at the Egypt National Research Center, Dokki, Giza. The animals were housed in plastic cages with corn cob bedding which was changed weekly. Food and water were provided. Animals were divided into two groups, (9 rats in each group), control and treated. The control animals received normal food and water for 8 weeks. Treated animals received normal Food and water plus oral doses of 1.5 ml fungal secondary metabolites / 100 g body weight/day for 8 weeks.

Hormonal determination

Serum sample

At the end of the experiment, blood samples from 9 control animals and 9 treated animals were allowed to clot for 30 minutes at 37°C before centrifugation for 15 minutes at 1000 x g. to obtain serum then stored at -20°C.

Quantitative measurement of rat follicle stimulating hormone by ELISA system (Closset, and Hennen 1988 and Teerds, et al, 1989).

Quantitative determination of testosterone in serum by enzyme immunoassay system (Zirkin, and Chen, 2000 and Sakuma, 2009).

Results

Fungi isolated from various samples:

As shown in Table 1, a total of 12 species of fungi belonging to 8 genera were isolated and identified from wheat, barley, rice, sorghum and maize. two species of *Aspergillus* (*A.flavus* and *A.niger*); Two species of *Penicillium* (*P. digitatum* and *P. notatum*); Three *Fusarium* species (*F. graminearum*, *F. moniliforme* and *F. solani*); one *Cladosporium* species (*C. herbarum*);; one species of *Alternaria* (*A. altrnata*); one species of *Rhizopus* (*R. stolonifer*); one species of *Tricoderma* (*T. viridi*); and one species of *Cunninghamella* (*C. elegans*).

Aspergillus (*A.flavus* and *A.niger*) and *Fusarium moniliforme* were

the most commonly isolated fungi from all the tested samples followed by *Penicillium notatum* which was isolated from 4 samples (wheat, barley, sorghum and maize); *Tricoderma viridi* was isolated from 4 samples (wheat, barley, rice and sorghum); *Penicillium digitatum* was commonly isolated from 3 samples (wheat, sorghum and maize); *Alternaria altrnata* was isolated from 3 samples (wheat, barley and sorghum); *Fusarium graminearum* was isolated from 2 samples (barley and rice); *Cladosporium herbarum* commonly isolated from 2 samples (wheat and rice); *Rhizopus stolonifer* was isolated from 2 samples (wheat and barley); *Fusarium solani* was commonly isolated from only 1 sample (barley); *Cunninghamella elegans* was isolated from only 1 sample (sorghum).

Production of 2ry metabolites (crude toxins)

The concentration of fungal (crud toxin) in terms of protein content in the fungal 2ry metabolites determined by spectrophotometric method megarments was equal to 637 µg/dl

Testosterone measurements

Table 2 and Figure 1 showed that the mean concentration of serum testosterone in treated experimental animal model was greatly reduced (P<0.01) compared to the control group (0.57 , 4.46 ng/ml, respectively).

Follicle stimulating hormone measurements

Mean value of follicle stimulating hormone (Tables 2 and Figure 1)

showed increased value ($P < 0.01$) in treated experimental animal model than in case of control (9.143, 4.020 ng/ml, respectively)

Table 1. Frequency distribution of fungi isolated from samples

Fungi \ Samples	wheat	barley	rice	sorghum	maize	Total No.
<i>Aspergillus flavus</i>	+	+	+	+	+	5
<i>Aspergillus niger</i>	+	+	+	+	+	5
<i>Penicillium digitatum</i>	+	-	-	+	+	3
<i>Penicillium notatum</i>	+	+	-	+	+	4
<i>Fusarium graminearum</i>	-	+	+	-	-	2
<i>Fusarium moniliforme</i>	+	+	+	+	+	5
<i>Fusarium solani</i>	-	+	-	-	-	1
<i>Cladosporium herbarum.</i>	+	-	+	-	-	2
<i>Alternaria altrnata</i>	+	+	-	+	-	3
<i>Rhizopus stolonifer</i>	+	+	-	-	-	2
<i>Trichoderma viridi</i>	+	+	+	+	-	4
<i>Cunninghamella elegans</i>	-	-	-	+	-	1

Table (2) Statistical analysis for the effect of crude toxin on hormonal level in control and treated samples

	Mean testosterone level (ng/ml)	Mean follicle stimulating hormone level (ng/ml)
Control	4.467	4.020
Treated	0.577 **	9.143 **

** Significant at 1% level ($P < 0.01$)

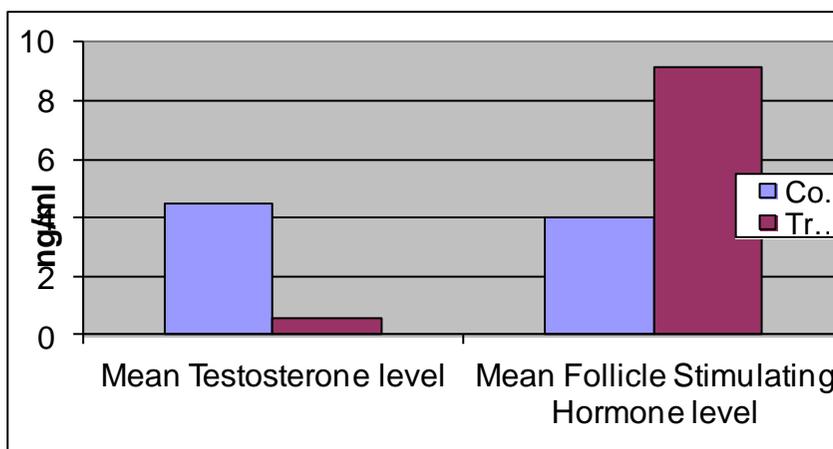


Figure (1) The effect of crude toxin on hormonal level in control and treated samples .

Discussion

Many species of fungi produce mycotoxins in food. Molds can grow and mycotoxins can be produced during storage and transport. Mold growth and mycotoxin production are related. To inadequate storage conditions . environmental conditions, e.g. heat, water and insect damage consider a predisposing factor in mycotoxin production and concentrations (*Dowd, 2004*).

Molds can grow over a temperature range of 10 - 40°C, a pH range of 4 - 8 and 12 - 15% moisture. The conditions most suitable for mold growth and for mycotoxin formation are not necessarily the same. The *Fusarium* molds associated with Alimentary Toxic Aleukia have been reported to grow prolifically at 25 - 30° C without producing much mycotoxin, but at near freezing

temperatures, they produce large quantities of mycotoxins with minimal mold growth (*Joffe, 1986*). In the present study a total of 12 fungal species belonging to 8 genera were isolated and identified from the examined samples; wheat, barley, rice, sorghum and maize. The isolated fungi included two species of *Aspergillus* (*A.flavus* and *A.niger*); two species of *Penicillium* (*P. digitatum* and *P. notatum*); three *Fusarium* species (*F. graminearum*, *F. moniliforme* and *F. solani*); one *Cladosporium* species (*C. herbarum*); one species of *Alternaria* (*A. altrnata*); one species of *Rhizopus* (*R. stolonifer*); one species of *Trichoderma* (*T. viridi*); one species of *Cunninghamella* (*C. elegans*). *Aspergillus* and *Fusarium* were the most commonly isolated fungi from the collected and examined grains followed by *Penicillium notatum* .

Fusarium species are ubiquitous in soils. They are commonly considered as field fungi invading more than 50% of the crop grains before harvest (**Robledo-Robledo, 1991**) and a main cause of diseased maize and wheat (**Doko et al, 1996, Munkvold and Desjardins, 1997, and Orsi et al, 2000**). Grains contamination by fungi not only renders grains unfit for human consumption by discoloration and reduction of nutritional value, but can also lead to mycotoxin production. Mycotoxins are poisonous secondary metabolites produced by some fungi in staple foods and foodstuffs. (**Pitt, 2000**).

Testosterone and follicle-stimulating hormone (FSH) are required to obtain full reproductive potential. In the tests, hormonal harmony are required by germ cells as they mature into spermatozoa (**Walker, and Cheng, 2005**).

On average, in adult males, levels of testosterone are about 7–8 times as great as in adult females, but, as the metabolic consumption of testosterone in males is greater, the daily production is about 20 times greater in men. Females are also more sensitive to the hormone. Testosterone is observed in most vertebrates. (**Mechoulam et al, 1984**).

The in vivo experiment in this study shows obvious and significant reduction in mean concentration of serum testosterone level in treated experimental animal model in case

of control than treated (0.57 , 4.46 ng/ml) respectively . Also the obtained results indicate that the mean value of follicle stimulating hormone was significantly increased in treated experimental animal model than in case of control (9.143, 4.020 ng/ml) respectively, which disturb the normal spermatogenesis process.

In Conclusion, The current in vivo study confirm that mycotoxicosis significantly affect the male fertility via affecting the values of hormonal levels including testosterone and follicle stimulating hormone throughout disturbing spermatogenesis which is the main factor contributing in successful fertilization process.

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تأثير السموم الفطرية على خصوبة هرمونات الذكورة

الملخص العربي

الفطريات المعزولة من العينات ؛ تم تحديدها في ١٢ نوعا تنتمي إلى ٨ أجناس و. كان الاسبرجلس والفيوزيريم الجنسان الاكثر شيوعا في العينات التي تم العزل منها تم زراعة الفيوزيريم جرامنيريم على وسط غذائي مناسب لإنتاج السموم الفطرية الخام وفي نهاية مدة الزراعة تم فصل نواتج الايض الثانوية بفلتره الوسط الغذائي ومن ثم استخدمت نواتج الايض في تعريض مجموعة الحيوانات الاختبارية المعالجة فقط لمدة ٨ اسابيع على التوالي وفي نهاية فترة التعريض تم سحب عينات دم لاستخلاص السيرم من كلتا المجموعتين (المجموعة المعالجة والمجموعة المعيارية الغير معالجة) وتم استخدام اختبارات الفحص الهرموني باستخدام المقاييس المناعية الإنزيمية لتحديد نسبة هرمون التستوستيرون و الهرمون المحفز للجريبات في مصل الدم للعينات القياسات الهرمونية لكل من التستوستيرون و الهرمون المحفز للجريبات اوضحت تناقص حاد في معدل التستوستيرون في العينات المعالجة بالمقارنة بالقياسات في العينات المعيارية الغير معالجة وعلى النقيض فقد ازدادت معدلات الهرمون المحفز للجريبات في العينات المعالجة عن ما هو عليه في العينات المعيارية الغير معالجة وبما ان الاتزان الهرموني عامل حاسم في عملية تخليق الحيوانات المنوية في الخصية وبدورة تخليق الحيوانات المنوية يعتبر اهم معيار للخصوبة عند الرجال.