

## Studies on Some Septicemic Bacterial Diseases Affecting *Oreochromis Niloticus* in Earthen Fish Farm

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### Abstract

This study was carried out on 150 *Oreochromis niloticus* at Dakhalia governorate within winter and summer seasons of the year 2015. The clinicohistopathological picture and biochemical investigations of the diseased fish were recorded. Sixty fish were infected with various types of bacterial species, of which, (49) fish were infected with Gram-negative bacteria while (11) fish were infected with Gram-positive one. The most common bacterial infection were *Aeromonas hydrophila* (33%), *Pseudomonas fluorescens* (20.5%), *Streptococcus agalactiae* (17.8 %), *Aeromonas sobria* (12.3 %), *Vibrio anguillarum* (9.5%) and *Vibrio alginolyticus* (6.8%). The highest total prevalence of bacterial infections was recorded in summer season (39%) compared to 21 % in winter. Isolates varied in their antibiotic sensitivity pattern. Inflammatory changes associated with these bacterial diseases were obvious in histopathological sections. Disease conditions were affected by various environmental stressors especially temperature, pH, salinity, ammonia and some heavy metals.

**Keywords:** *Oreochromis niloticus*, Bacterial diseases, Diagnosis, Pathology, water quality.

### Introduction

The most healthiest and cheapest source of protein is obtained from aquaculture (Nijdam et al, 2012). So, we now trend to increase production of aquatic animal food products more than others (FAO, 2012).

Tilapia is the most well known cultured species in all over the world, as it occupies the second prevailing species after carp (FAO,

2012). Egypt is ranked 2nd one in production of Tilapia and the 7 th of top ten countries in the total production of fish (El Zokm et al, 2012).

Diseases are considered one of the hazard factors that increase the incidence the Epidemiological aquaculture (Umesh et al, 2008). Any disease usually originates from synergism of the pathogen, host and environment as temperature, pH,

nitrogenous waste products, salinity and some heavy metals (*Madhun et al, 2015*).

Bacterial diseases are considered one of the important diseases affecting fish mainly the septicemic bacterial diseases which caused great loss and mortality (*Noor El-Deen et al, 2010*). The bacterial fish pathogens comprise natural inhabitants of the aquatic environment (*El-Refaeey, 2013*). The current study was investigated the most predominant bacterial diseases in tilapia fish in earthen pond at Dakahlia province.

#### **Materials and Methods:**

##### **Fish:**

The study was carried out on 150 freshly captured *Oreochromis niloticus* of different body weight ranged from (175 to 200 g) collected from earthen ponds in private fish farms in Dakahlia governorate in Egypt showing signs of septicemia during the period of December 2014 to July 2015 (winter and summer seasons). Seventy five fish were collected per season.

##### **Bacteriological examination**

###### **Sampling and processing**

Samples were retrieved from liver, kidney, spleen, gills under fully sterile conditions. Loopfuls were cultured into tryptic soy agar, Aeromoas medium base supplemented with ampicilin, thiosulphate citrate bile salt sucrose agar (TCBS) and Blood agar medium. The inoculated plates were

incubated at 25 °C for 18-48 hours. Representative numbers of the different colonial types detected on the media were collected from plates and streaked on TSA for purity and identification.

##### **Identification of isolates**

Identification of the obtained bacterial isolates was carried out by performing biochemical and morphological characters using traditional, API 20 E and API strept kits according to (*Buller, 2004*).

##### **Water quality examination**

In summer, water samples were obtained from various locations from Dakahlia fish farm (2times) in sterile plastic bottles and stored according to methods adopted from (*APHA, 2000*). Then Physico-chemically (Temperature, pH and salinity), unionized ammonia (NH<sub>3</sub>) and heavy metals (iron and copper) analysis were measured using Thermometer, Symphony VMR PH meter, Martin conductivity meter, Cintza 101 double beam spectrophotometer and Atomic absorption spectrometer respectively.

##### **Histopathological studies**

Tissue specimens used for histopathological techniques were obtained from liver, spleen, kidney, gills, intestines and gonads of infected *O. niloticus*. Samples were conserved in 10 % buffered formalin, dehydrated by ethanol solution, then fixed with paraffin, and cut at 5 µm thick. Tissue sections were routinely processed

and stained with Hematoxylin and Eosin (H & E) and finally examined under light microscopy following the criteria reported in *Bancroft (1996)*.

### Results

Naturally infected *O. niloticus* showed: hemorrhagic patches extensively distributed on the external body surface, scales detachment, erosions, fin and tail rot. The gills were congested with presence of excessive amount of mucus. Some cases showed abdominal distention, ascites exophthalmia (fig1A). On the other hand, postmortem findings revealed paleness of liver, enlargement of gall bladder and congestion of kidney (fig1B).

### Isolation and identification of retrieved bacterial isolates

The phenotypic characteristics of obtained loopfuls from different fish organs showed a total number of seventy three bacterial isolates retrieved from sixty infected fish. Most of obtained isolates were Gram-negative 82.2 % represented as; *A. hydrophila* 33% followed by *Ps. fluorescens* 20.5 %, *A. sobria* 12.3 %, *V. anguillarum* 9.5%, *V. alginolyticus* 6.8%. On the other hand *Streptococcus agalactiae* was the incriminated Gram-positive bacterial pathogens involved in this investigation 17.8% . Full phenotypic and biochemical characteristics of retrieved isolates are reported in (tables 1, 2) and the

prevalence of the different bacterial species is summarized in (table 3).

### Antibiogram sensitivity testing

Isolates differed in their sensitivity pattern according to different antimicrobial agents tested (table4). The most effective antimicrobial agent tested has been established to be Sulphamethoxazole + Trimethoprim complex 66 %. On the other hand, the highest resistances displayed by these bacterial isolates were noticed against novobiocin 59 % and tetracycline 46.5%.

### Histopathological lesions

Inflammatory and necrotic alterations were obvious, also some bacterial colonies were observed in histological sections (Fig. 2).

Degeneration, hemorrhagic necrotic changes were commonly observed in haemopoietic tissues, liver and kidney. On other hand, gills showed congestion, necrosis and desquamation of epithelial lining of primary gill lamellae and also fusion of secondary gill lamellae.

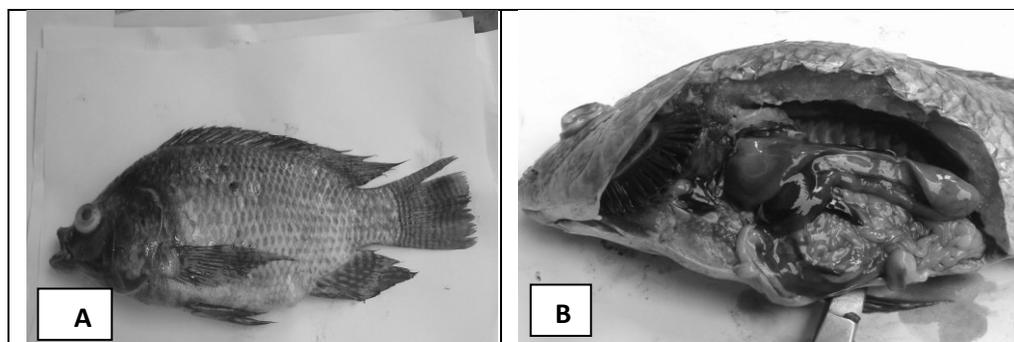
### Water quality parameters

Disease conditions reported in this study were appeared to be related to stressful environmental condition expressed by unseemly water quality parameters in examined fish farms (Table 5). Investigated fish farms at summer season showed slightly alkaline waters as the pH values measured about 8.9 and salinity recorded 2 ‰ .

Results also demonstrated presence of unfavorable values of some heavy metals such as iron which

ranged about 2.95 ppm with no presence of copper in summer season. On other hand, levels of

some nitrogenous waste products as NH<sub>3</sub> was 1mg/l which above the permissible level.



**Fig1:** (A) Naturally infected tilapia showing haemorrhages on the external body surface and exophthalmia. (B) Naturally infected tilapia showing paleness of liver, enlargement of gall bladder and congestion of kidney.

**Table1:** biochemical characteristics of Gram- negative bacterial isolates retrieved from naturally infected *O. niloticus* obtained from API20E.

| Biochemical test                               | <i>A. hydrophila</i> | <i>A. sobria</i> | <i>Ps. fluorescens</i> | <i>V. alginolyticus</i> | <i>V. anguillarum</i> |
|--|----------------------|------------------|------------------------|-------------------------|-----------------------|
| B-Galactosidase production (OPNG)              | +                    | +                | -                      | -                       | +                     |
| Arginine dihydrolase production (ADH)          | +                    | +                | +                      | -                       | +                     |
| Lysine decarboxylase production (LDC)          | +                    | +                | -                      | +                       | -                     |
| Ornithine decarboxylase production (ODC)       | -                    | -                | -                      | -                       | -                     |
| Citrate utilization (CIT)                      | V                    | V                | V                      | +                       | +                     |
| H <sub>2</sub> S production (H <sub>2</sub> S) | +                    | +                | -                      | -                       | -                     |
| Urease production (URE)                        | -                    | -                | -                      | V                       | -                     |
| Tryptophane deaminase production (TDA)         | -                    | -                | -                      | V                       | -                     |
| Indole production (IND)                        | +                    | +                | -                      | +                       | -                     |
| Acetoin production (VP)                        | +                    | +                | +                      | -                       | -                     |
| Gelatinase production (GEL)                    | +                    | +                | -                      | V                       | +                     |
| Acid from glucose (GLU)                        | +                    | +                | -                      | +                       | +                     |
| Acid from manitol (MAN)                        | +                    | +                | -                      | +                       | +                     |
| Acid from inositol (INO)                       | -                    | -                | -                      | -                       | +                     |
| Acid from Sorbitol (SOR)                       | V                    | -                | -                      | V                       | +                     |
| Acid from rhamnose (RHA)                       | -                    | -                | -                      | -                       | -                     |
| Acid from sucrose (SAC)                        | +                    | +                | -                      | +                       | +                     |
| Acid from melibiose (MEL)                      | -                    | V                | V                      | -                       | -                     |
| Acid from amygdalin (AMY)                      | +                    | V                | -                      | -                       | -                     |
| Acid from arabinose (ARA)                      | V                    | V                | -                      | -                       | +                     |

(+) Positive, (-) Negative, (V) Variable

**Table2:** Phenotypic characteristics of *S. agalactiae* obtained from naturally infected *O. niloticus* fish using API strept.

| Bacterial isolates      | Winter | %    | summer | %    | Total isolate | %    |
|-------------------------|--------|------|--------|------|---------------|------|
| <i>A. hydrophila</i>    | 6      | 30   | 18     | 34   | 24            | 33   |
| <i>A. sobria</i>        | 1      | 5    | 8      | 15   | 9             | 12.3 |
| <i>Ps. fluorescens</i>  | 11     | 55   | 4      | 7.5  | 15            | 20.5 |
| <i>V. anguillarum</i>   | 2      | 10   | 5      | 9.4  | 7             | 9.5  |
| <i>V. alginolyticus</i> | 0      | 0    | 5      | 9.4  | 5             | 6.8  |
| <i>S. agalactia</i>     | 0      | 0    | 13     | 24.5 | 13            | 17.8 |
| Total                   | 20     | 100% | 53     | 100% | 73            | 100% |

**Table3:** Prevalence of bacterial isolates obtained from infected fish

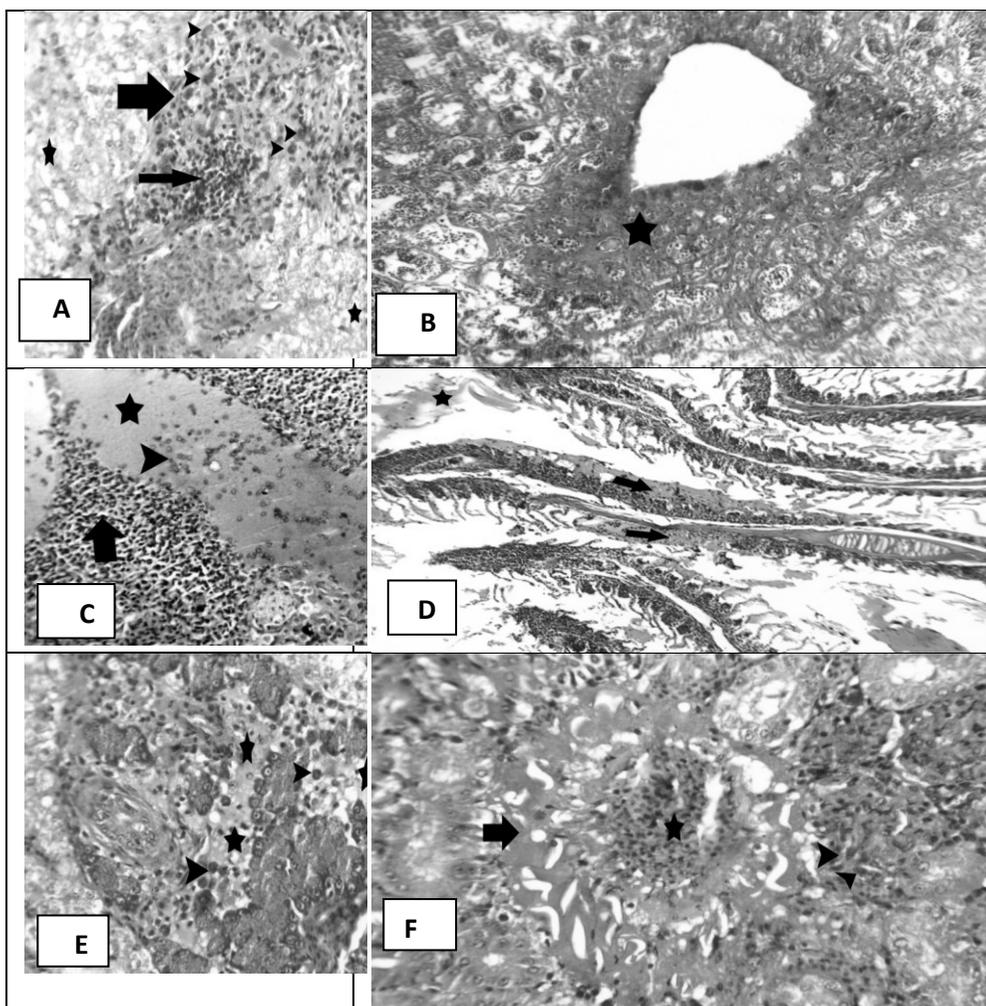
|   |                                   |   |
|---|-----------------------------------|---|
| <b>Colony characters</b>                                      | Onto marine and TSAS agar         | Small pin point, whitish, rounded and slightly raised |
|   | Onto Blood agar                   | β Haemolysis  |
| <b>Microscopical Examination</b>                              | Gram- stain and cell form         | Gram positive cocci arranged in short chains          |
| <b>Biochemical characteristics obtained from API 20 Strep</b> | Voges-Proskauer ( VP )            | +   |
|   | Hippurate ( hip )                 | +   |
|   | Aesculin ( ESC )                  | -   |
|   | pyrrolidonyl arylamidase ( PYRA ) | -   |
|   | α galactosidase ( α-GAL )         | -   |
|   | β glucuronidase ( β GUR )         | Variable  |
|   | β galactosidase ( β GAL )         | -   |
|   | Alkaline phosphatase ( PAL)       | +   |
|   | leucine arylamidase ( LAP)        | +   |
|   | Arginine dihydrolase ( ADH)       | +   |
|   | Ribose ( RIB)                     | +   |
|   | Arabinose ( ARA)                  | -   |
|   | Mannitol ( MAN)                   | -   |
|   | Sorbitol ( SOR)                   | -   |
|   | Lactose ( LAC)                    | Variable  |
|   | Trehalose ( TRE)                  | Variable  |
|   | Inulin ( INU)                     | -   |
|   | Raffinose ( RAF)                  | -   |
| Amygdalin ( AMD)  | Variable                          |   |
| Glycogen ( GLYG)  | -                                 |   |

**Table 4: Antibiogram of obtained bacterial isolates**

|  | Conc     | <i>A. hydrophil</i> |     | <i>A. sobria</i> |   | <i>Ps. fluorescen</i> |   | <i>V. anguillar</i> |     | <i>V. alginolyti</i> |    | <i>S. agalacti</i> |    | total |    |
|--|----------|---------------------|-----|------------------|---|-----------------------|---|---------------------|-----|----------------------|----|--------------------|----|-------|----|
|  |          | N:2<br>4            | %   | N:9              | % | N:15                  | % | N:7                 | %   | N:5                  | %  | N:13               | %  | N:73  | %  |
| Tetracycline<br>R<br>I<br>S                                    | 30<br>µg |                     | 4   | 4                | 4 | 9                     |   |                     |     |                      |    | 6                  |    |       |    |
|  |          | 1                   | 12. | 4                | 2 | 3                     | 4 | 57                  | 2   | 40                   | 9  | 3                  | 34 | 46.   |    |
|  |          | 3                   | 5   | 2                | 2 | 1                     | 7 | 1                   | 14  | -                    | 0  | 4                  | 1  | 11    | 5  |
|  |          | 20                  | 83  | 3                | 3 | -                     | 0 | 2                   | 28. | 3                    | 60 | -                  | 0  | 28    | 38 |
| Novobiocin<br>R<br>I<br>S                                      | 30<br>µg |                     |     | 6                | 6 | 6                     | 7 |                     |     | -                    |    | 4                  |    |       |    |
|  |          | 18                  | 75  | 6                | 7 | 10                    | 2 | 3                   | 43  | 4                    | 0  | 6                  | 3  | 43    | 59 |
|  |          | 4                   | 17  | 3                | 3 | 3                     | 0 | 3                   | 43  | 1                    | 80 | 4                  | 1  | 21    | 29 |
|  |          | 2                   | 8   | -                | 3 | 2                     | 1 | 1                   | 14  |                      | 20 | 3                  | 2  | 9     | 12 |
|  |          |                     |     | 0                | 0 | 3                     | 3 |                     |     |                      |    | 0                  |    |       |    |
| Streptomycin<br>(S)<br>R<br>I<br>S                             | 10<br>µg |                     |     | 5                | 5 | 4                     | 7 |                     |     |                      |    | 0                  |    |       |    |
|  |          | 5                   | 21  | 5                | 2 | 7                     | 1 | 2                   | 28. | 3                    | 60 | -                  | 2  | 22    | 30 |
|  |          | 4                   | 17  | 2                | 2 | 2                     | 3 | 1                   | 5   | 2                    | 40 | 3                  | 3  | 14    | 19 |
|  |          | 15                  | 62. | 2                | 2 | 6                     | 4 | 4                   | 14  | -                    | 0  | 10                 | 7  | 37    | 51 |
|  |          |                     |     | 2                | 2 | 0                     | 0 |                     |     |                      |    | 7                  | 7  |       |    |
| Sulphamethoxa<br>zole<br>+Trimethoprim<br>(SXT)<br>R<br>I<br>S | 25<br>µg |                     | 0   | 0                | 3 | 2                     | 7 |                     |     |                      |    | 0                  |    |       |    |
|  |          | -                   | 10  | -                | 6 | 4                     | 3 | 1                   | 14  | 4                    | -  | -                  | 5  |       |    |
|  |          | -                   | 0   | 3                | 6 | 5                     | 4 | 5                   | 71  | 1                    | 80 | 2                  | 8  | 9     | 12 |
|  |          | 24                  |     | 6                | 6 | 6                     | 0 | 1                   | 14  |                      | 0  | 11                 | 5  | 48    | 66 |
| Amoxicillin<br>AML<br>R<br>I<br>S                              | 10<br>µg |                     |     | 4                | 4 | 2                     | 7 |                     |     |                      |    | 2                  |    |       |    |
|  |          | 12                  | 50  | 4                | 4 | 3                     | 0 | -                   | 0   | 4                    | 80 | 3                  | 3  | 26    | 36 |
|  |          | 10                  | 42  | 1                | 1 | 7                     | 4 | 6                   | 86  | 1                    | 20 | 3                  | 2  | 28    | 38 |
|  |          | 2                   | 8   | 4                | 4 | 5                     | 7 | 1                   | 14  | -                    | 0  | 7                  | 3  | 19    | 26 |
|  |          |                     |     | 4                | 4 | 3                     | 3 |                     |     |                      |    | 5                  | 5  |       |    |
|  |          |                     |     | 4                | 4 | 3                     | 3 |                     |     |                      |    | 4                  | 4  |       |    |
| Doxycycline<br>(DO)<br>R<br>I<br>S                             | 30<br>µg |                     |     | 7                | 7 | 6                     | 7 |                     |     |                      |    | 1                  |    |       |    |
|  |          | 10                  | 42  | 7                | 8 | 9                     | 0 | 2                   | 28. | 3                    | 60 | 2                  | 5  | 33    | 45 |
|  |          | 3                   | 12  | 1                | 1 | 4                     | 2 | 2                   | 5   | -                    | 0  | 9                  | 6  | 19    | 26 |
|  |          | 11                  | 46  | 1                | 1 | 2                     | 7 | 3                   | 28. | 2                    | 40 | 2                  | 9  | 21    | 29 |
|  |          |                     |     | 1                | 1 | 1                     | 1 |                     |     |                      |    | 1                  | 1  |       |    |
|  |          |                     |     | 1                | 1 | 3                     | 3 |                     |     |                      |    | 5                  | 5  |       |    |
| Kanamycin (K)<br>R<br>I<br>S                                   | 30<br>µg |                     |     | 1                | 1 | 2                     | 7 |                     |     |                      |    | 3                  |    |       |    |
|  |          | 7                   | 29  | 1                | 5 | 4                     | 2 | 2                   | 28. | 1                    | 20 | 5                  | 3  | 20    | 27 |
|  |          | 6                   | 25  | 5                | 5 | 3                     | 0 | 5                   | 5   | 1                    | 20 | 4                  | 1  | 24    | 33 |
|  |          | 11                  | 46  | 3                | 3 | 8                     | 5 | -                   | 71  | 3                    | 60 | 4                  | 3  | 29    | 40 |
|  |          |                     |     | 3                | 3 | 3                     | 3 |                     |     |                      |    | 1                  | 1  |       |    |

**Table 5: Mean water quality measures in examined fish farm**

| Parameters             | Summer season |
|------------------------|---------------|
| Salinity (‰)           | 2 ‰           |
| Temperature (°C)       | 31.5          |
| pH                     | 8.9           |
| NH <sub>3</sub> (mg/l) | 1             |
| Iron (ppm)             | 2.95          |
| Cu (ppm)               | 0             |



**Fig3.**A: Liver infected with *A. hydrophila* shows hydropic degeneration in hepatocytes (asterisks), perivascular fibrosis with mononuclear cells (thin arrow) and eosinophilic granulocytes infiltration (arrowheads) (H&E, x200).

B: Testis infected with *A. sobria* shows perivascular necrosis and fibrosis (asterisk) (H&E, x100).

C: Spleen infected with *Ps. fluorescence* shows bacterial colonies (arrowhead) inside area of necrosis (asterisk) surrounded by leukocytes infiltration (thick arrow) (H&E, x200).

D: Gills infected with *V. anguillarum* shows necrosis (arrows) and desquamation of epithelial lining primary gill lamellae (asterisk) (H&E, x100).

E: Liver infected with *V.alginolyticus* shows focal necrosis (asterisks) and fibrosis in hepatopancreas with few eosinophilic granulocytes infiltration (arrowheads) (H&E, x200).

F: Caudal kidneys infected with *S.agalactia* shows congested renal blood vessel (asterisk) with perivascular fibrosis (arrow) and basophilic bacterial colonies trapped inside glomerulus (arrow heads) (H&E, x200).

## Discussion

Bacterial disease is the most critical sources of disease problems in all types of fish. Specific bacterial pathogens in every type of fish either freshwater or marine are responsible for severe mortalities (Moustafa et al., 2015; Elgandy et al., 2015). Freshwater aquaculture is considered the main investment in Egypt and other countries (Saad et al., 2014). Tilapias (*Oreochromis* sp.) are one of the most important cultured fish in freshwater aquaculture industry (Bostock et al., 2010).

The present study confirmed that tilapia are susceptible to many bacterial pathogens which able to cause diseases. In agreement with other studies relating to freshwater bacterial diseases (Akinbowale et al., 2006; Al-Harbi and Uddin, 2010) most of retrieved bacterial isolates were Gram-negative 82.2%. On the other hand gram-positive bacteria were about 17.8%. Results of bacteriological examination established that *A.hydrophila* 33%, *Ps. fluorescens* 20.5%, *S.agalactia* 17.8%, *A.sobria* 12.3%, *V.anguillarum* 9.5% and *V.alginolyticus* 6.8%.

*Vibrio* spp, *Ps. fluorescens*, *S.agalactia* and *Aeromonas* spp

were the most common isolated bacteria from this study. These results are similar to those reported by (Tatsuro et al, 2004; Abdel hamid et al, 2013; Elgandy et al, 2013).

*A.hydrophila* were the most prevalent bacterial pathogens representing 33% in this study which in accordance to those reported by Al-Laham, et al (2014); Jovanović et al (2015). *A. hydrophila* recorded 39% in summer, but 21% in winter season. Also, *A.sobria* was reported in summer (15%) while winter season (5%). This result can be explained as the maximized activity of cytotoxins, hemolysin and enterotoxins above 22 °C increase the incidence of disease in summer (Krovacek et al, 1991).

Pseudomonads septicemic infections are widely spread. The high frequency of *Ps. fluorescens* infection was recorded in winter 55%, but in summer 7.5%. This can be explained as proteinases and lipases of *P. fluorescens* have ability to produce at 7 °C which trigger incidence of disease in winter season (Wang and Jayarao, 2001).

Vibriosis has great importance in marine water fish but also affect

freshwater fish (*Dyer and Oliver, 2008; Elgendy et al, 2015*). Seasonally, the highest prevalence of *V. anguillarum* infection was recorded during the winter and summer 10% and 9.4% respectively. On the other hand the highest prevalence of *V. alginolyticus* infection was recorded in summer 9.4% with no presence in winter. This explained as *V. alginolyticus* may lose its ability to cause disease in cold season (*Yan et al, 2007*).

Streptococosis has great economic importance due to high fish mortality (*Haghighi et al, 2010*). *S. agalactia* has high incidence in summer about 24% with no prevalence in winter season. This result can be explained that Raising of water temperature (above 26 °C) is optimal for increase the prevalence of streptococosis (*Mian et al, 2009*).

Prevalence of bacterial isolates was higher in summer 53 % than in winter season 20 %. This may be explained as higher levels of un ionized ammonia was detected in fish farm water during this period

In regards to antibiogram sensitivity testing, freshwater aquaculture is a promising aquaculture sector needing pioneering scientific and technical developments. Excessive using of antibiotics for prophylactic and therapeutic led to increase bacterial resistance (*Sarter et al, 2007*). Reservoirs of antibiotic resistance can interact between various environmental conditions

and transfer of resistant bacteria from animals to humans (*Witte, 2000; Vaseeharan et al., 2005*).

*A. hydrophia* isolates showed higher resistance to novobiocin 75% and Amoxicillin 50% in compared to other antibiotic tested , On other hand , isolates show intermediate susceptibility to isolates superior susceptibility to Doxycycline 42% .Also, it shows maximum susceptibility in Sulphamethoxazole + Trimethoprim complex 100 % ,tetracyclin82%, Streptomycin 62.5% and Kanamycin42%. Results are nearly in accordance with *Laith and Najiah (2013) and Al Laham and Al Fadel (2014)*.

*A. sobria* isolates demonstrated extreme resistance to Doxycycline 75%, novobiocin 67%, Streptomycin 55%, tetracycline 44% were observed in this isolates. Moreover, higher intermediate sensitivity was showed in case of Kanamycin 55.5%. On other hand, high susceptibility in Sulphamethoxazole + Trimethoprim complex 66% was characteristic. Results are nearly in accordance with *Guz and Kozinska (2004) and Çiftci et al (2015)*. In respect to *Ps. fluorescens* , revealed resistance were reported to tetracyclin 93%,novobiocin 67% , Doxycycline60% ,Streptomycin 47%.On contrary, prominent sensitivity was detected in kanamycin 53%and Sulphamethoxazole +Trimethoprim combination 40%,but intermediate resistant to Amoxicillin

47%. Results are nearly in accordance with *Eissa et al (2010)*; *Foysal et al (2011)*.

Regarding *V.anguillarum*, higher sensitivity was recorded in Streptomycin 57% and Doxycycline 43%. Furthermore, superior intermediate sensitivity was maximum in Amoxicillin 86% and both Sulphamethoxazole +Trimethoprim combination and Kanamycin 71%, but high resistance to tetracycline 57%. Results are nearly in accordance with *Vaseeharan et al (2004)*; *Jayasree et al (2006)*.

In concern to *V. alginolyticus* strains showed reputed sensitivity to both Kanamycin and tetracycline 60%, but maximum intermediate sensitivity to novobiocin 80%. In addition, isolates exploited superior resistance to both Amoxicillin and Sulphamethoxazole +Trimethoprim reach up to 80%, Doxycycline and Streptomycin about 60%. Results are nearly in accordance with *Li et al (1999)*; *Jayasree et al (2006)*.

Concerning the antibiogram of *S.agalactia* isolates displayed higher sensitivity to Sulphamethoxazole+Trimethoprim 85%, Streptomycin 77%, Amoxicillin 54%. On other hand, isolates displayed prominent resistance was detected in tetracycline 69%, novobiocin 46%, Kanamycin 37%, on contrary, intermediate sensitivity to Doxycycline 69%. Results are nearly in accordance with *Al-*

*Marzouk et al (2005)* ; *Abuseliana et al (2010)*.

In concern to water quality parameters reported in this study, in agreement with *Eissa et al (2013)*; *Moustafa et al (2015)* unfavorable aquatic environmental condition as low value of water quality parameters increase the prevalence of disease noticed in this study.

In respect to the pH values, the farm water were slightly alkaline 7.5 in summer season which increase the prevalence of vibriosis reported in this study as the low pH value increase the mucus secretion which increase adhesion of vibrios (*Balebona et al, 1995*). On the other hand, high pH increase the toxicity by non ionised ammonia (*Ludwig et al, 2007*). The high pH values was (7.5) in summer due to increasing photosynthetic activity, which decrease levels of CO<sub>2</sub> and also low levels of dissolved oxygen. These findings are supported by (*Goher, 2002*).

In regards to salinity, the high salinity noticed in this study (2 ‰) which also explained the presence of vibriosis in tilapia. This was in agreement to those reported by *Al-Sunaiher et al (2010)*; *Abou El-Geit et al (2013)*. High level of salinity can be explained by increasing evaporation rate in summer (*Herbs, 2002*).

Regarding ammonia, the ammonia level reported in this investigation ranged 1.1ppm which was higher than permissible limit (0.02mg/l). This was in agreement to those

reported by *Sachidan and Yajurvedi (2006)*; *Moustafa et al (2015)*. Increasing level of ammonia triggers the invasion of bacteria especially immuno-suppression fish (*Moustafa et al, 2015*). While, prolonged low levels of ammonia irritates skin and gills facilitating parasitic infestations and opportunistic bacterial infections (*Goher, 2002*).

In regard to heavy metals reported in this study, iron ranged 2.95 mg/l which above the permissible limits (0.3 mg/l) according to *Anzec (2000)* with no presence of copper in summer season. This was in agreement in case of levels of iron but not in case of copper levels to those reported by *Karadede and Ünlü (2007)*; *Moustafa et al (2015)*. The high level of iron increases the incidence of some bacterial disease as vibrios since the virulence of vibro spp. enhance with higher iron (*Muiño et al, 2001*).

In summary the most important bacterial pathogens causing diseases in *Oreochromis niloticus* in the studied farm were *A. hydrophila*, *A. sobria*, *Ps. fluorescens*, *V. anguillarum*, *V. alginolyticus* and *S. agalactia*. Septicemic bacterial infections were related to the noticed unfavorable aquatic environmental stressors. The role of heavy metals including iron in exacerbating bacterial fish diseases is evident. Polluted water weakens the fish host defenses allowing

increased opportunities for bacterial infections to affect fish populations.

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### References

- Abdel hamid A.M., El-Barbary M.I., and El-Deweny, M. E.,(2013)**: Bacteriological status of ashtoum El-Gamil protected area. Egypt. J. Aquat. Biol. & Fish., Vol. 17, No. 3:11-23.
- Abou El-Geit, E.N, Saad, T.T., Abdo, M. H.,and Zaki ,M. S.,(2013)**: Microbial infections among some fishes and crustacean species during blooming phenomenon in Qaroun Lake-Egypt. Life Science Journal 10(2): 1217-1224.
- Abuseliana,A., Daud,H., Abdul Aziz,S., Bejo, S. K., and Alsaïd ,M., (2010)** *Streptococcus agalactiae* the Etiological Agent of Mass Mortality in Farmed Red Tilapia (*Oreochromis sp.*).Journal of Animal and Veterinary Advances;9(20):2640-2646.
- Akinbowale ,O.L. , Peng, H., and Barton, M.D. ,(2006)**: Antimicrobial resistance in bacteria isolated from aquaculture sources in Australia. Journal of Applied Microbiology 100( 5): 1103–1113.
- Al Laham,S.A., and Al Fadel,F.M.,(2014)**: Antibacterial Activity of Various Plants Extracts

- Against Antibiotic-resistant *Aeromonas hydrophila*. Jundishapur J Microbiol. Jul; 7(7): e11370.
- Al-Harbi, A.H., and Uddin, M.N., (2010):** Bacterial population of African catfish, *Clarias gariepinus* (Burchell 1822) cultured in earthen ponds. Journal of Applied Aquaculture 22:187–193.
- Al-Marzouk, A., Duremdez, R., Yuasa, K., Sameer, A.Z., Al-Gharabally, H., and Munday B., (2005):** Fish kill of mullet *Lizaklunzingeri* in Kuwait Bay: The role of *Streptococcus agalactiae* and the influence of temperature. In P. Walker, R. Lester and M.G. Bondad-Reantaso (eds). Diseases in Asian Aquaculture V, pp. 143-153. Fish Health Section, Asian Fisheries Society, Manila.
- Al-Sunaiher, A.E., Ibrahim, A.S.S., and Al-Salamah, A. A., (2010):** Association of *Vibrio* Species with Disease Incidence in Some Cultured Fishes in the Kingdom of Saudi Arabia World Applied Sciences Journal 8 (5): 653-660.
- Anzec (2000):** Australian and New Zealand Guidelines for Fresh and Marine Waters, National Water Quality Management Strategy. Australian and New Zealand Environmental and Conservation Council.
- APHA (2000):** Standard Methods for the Examination of Water and Wastewater, D.C.
- Balebona, M.C., Morinoigios, M.A. and Borrego, J.J., (1995):** Role of extracellular products in the pathogenicity of *Vibrio* strains on cultured gilt-head sea bream, *Sparus aurata*. Microbiol. SEM., 11: 439–446.
- Bancroft, G.D. and Stevens, A., (1996):** Theory and practice of Histological techniques, 4th edition. Edinburgh, Churchill living stone.
- Bostock, J., McAndrew, B., Richards, R., Jauncey, K., Telfer, T., Lorenzen, K., Little, D., Ross, L., Handisyde, N., Gatward, I., and Corner, R., (2010):** Aquaculture: global status and trends. *Philosophical Transactions of the Royal Society B* 365: 2897–2912.
- Buller, N.B., (2004):** Bacteria from fish and other aquatic animals: a practical identification manual, Cabi.
- Çiftci, A., Onuk, E.E., Çiftci, G., Findik, A., Sogut, M. Ü., and Gulhan, T., (2015):** The Comparative Analysis of Phenotypic and Genotypic Properties of *Aeromonas sobria* Strains Isolated from Rainbow Trout (*Oncorhynchus mykiss*, Walbaum, 1972): Kafkas Univ Vet Fak Derg 21(4): 585-592.
- Dyer, B.K., and Oliver, J. D., (2008):** The ecology of *Vibrio vulnificus*, *Vibrio cholera*, and *V. parahaemolyticus* in North Carolina estuaries. *The Journal of Microbiology*, 46 (2), 146-153.
- Eissa, A.E., Tharwat, N.A. and Zaki, M.M., (2013):** Field assessment of the mid winter mass kills of trophic fishes at Mariotteya stream, Egypt: Chemical and

biological pollution synergistic model. *Chemosphere*, 90: 1061–1068.

**Eissa, N.M.E., AbouEl-Ghiet, E.N., Shaheen, A.A., and Abbass, A. (2010):** Characterization of *Pseudomonas* Species Isolated from Tilapia "*Oreochromis niloticus*" in Qaroun and Wadi-El-Rayan Lakes, Egypt . *Global Veterinaria* 5 (2): 116-121

**El Zokm, G.M., ELGohary, S.E., and Abd-ElKhalik, D.E., (2012):** Studies of some heavy metals in water and sediment in El-Max fish farm, Egypt. *World Appl.Sci. J.*, 18(2): 171-180.

**Elgendy, M.Y., (2013):** Epizootiological studies on some bacterial infections in marine fishes. Ph.D thesis In: *Fish Diseases and Management*. Cairo, Faculty of Veterinary Medicine.

**Elgendy, M.Y., Moustafa, M. , Gaafar, A.Y. , and Borhan, T. ,(2015):** Impacts of extreme cold water conditions and some bacterial infections on earthen-pond cultured Nile tilapia, *Oreochromis niloticus*. *RJPBCS.*, 6: 136-145.

**El-Refaey, A.M.E., (2013):** Studies on major bacterial diseases affecting fish ; Tilapia *Oreochromis niloticus* , Catfish, *Clarias gariepinus* and mullets in Port Said, Egypt with special references to its pathological alterations. *Researcher*, 5(2):5-14.

**FAO, F., (2012):** *Yearbook 2010: Fishery and aquaculture Statistics*. Food and Agriculture

Organisation of the United Nations, Rome, 78.

**Foysal, M.J., Rahman, M.M., and Alam, M., (2011):** Antibiotic sensitivity and *in vitro* antimicrobial activity of plant extracts to *pseudomonas fluorescens* isolates collected from diseased fish. *International Journal of Natural Sciences*, 1(4):82-88.

**Goher, M. E., (2002):** Chemical studies on the preceptation and dissolution of some chemical elements in lake Qarun. Thesis Ph.D Fac. Sien.Al-azhar Univ.

**Guz, L., and Kozinska, A., (2004):** Antibiotic susceptibility of *Aeromonas hydrophila* and *A. sobria* isolated from farmed carp (*Cyprinus Carpiol.*). *Bull Vet Inst Pulawy* 48, 391-395.

**Haghighi, S., Soltani, M., Gh, N.B., Gha, M., and Skall, D., (2010):** Molecular epidemiology of zoonotic streptococcosis/lactococcosis in rainbow trout (*Oncorhynchus mykiss*) aquaculture in Iran. *Iranian J Microbiol* 2:198–209.

**Hagi, T., Tanaka, D., Iwamura, I., and Hoshino, T., (2004):** Diversity and seasonal changes in lactic acid bacteria in the intestinal tract of cultured freshwater fish. *Aquaculture* 234 (1–4): 335–346.

**Herbs, D. B., (2002):** Gradients of salinity stress, environmental stability and water chemistry as a template for defining habitat types and physiological strategies in inland salt water. *Hydrobiol.*, 466: 209-219.

- Jayasree, L., Janakiram, P., and Madhavi, R., (2006):** Characterization of *Vibrio* spp. Associated with Diseased Shrimp from Culture Ponds of Andhra Pradesh (India). Journal of the World Aquaculture Society Volume 37, Issue 4, pages 523–532.
- Jovanović, B., Whitley, E. M., Kimura, K., Crumpton, A., and Palić, D., (2015):** Titanium dioxide nanoparticles enhance mortality of fish exposed to bacterial pathogens. Environmental Pollution Volume 203, Pages 153–164.
- Karadede, H., and Ünlü, A. E., (2007):** Heavy Metal Concentrations in Water, Sediment, Fish and Some Benthic Organisms from Tigris River, Turkey Environmental Monitoring and Assessment, Volume 131, Issue 1, pp 3.
- Krovacek, K., Faris, A., and Mansson, I., (1991):** Growth of and toxin production by *Aeromonas hydrophila* and *Aeromonas sobria* at low temperatures. Int. J. Food Microbiol. 13: 165-176.
- Laith, A. R., and Najjiah, M., (2013):** *Aeromonas hydrophila*: Antimicrobial Susceptibility and Histopathology of Isolates from Diseased Catfish, *Clarias gariepinus* (Burchell) .J Aquac Res Development, 5:215.
- Li, J., Yie, J., Foo, R. W. T., Ling, J. M. L., Xu, H., and Woo, N. Y. S., (1999):** Antibiotic Resistance and Plasmid Profiles of *Vibrio* Isolates from Cultured Silver Sea Bream, *Sparus sarba*. Marine Pollution Bulletin 39( 1–12): 245–249.
- Ludwig, G. M., M. Hobbs and P. Perschbacher. (2007):** Ammonia, pH, and plankton in sunshine bass nursery ponds: the effect of inorganic fertilizer or sodium bicarbonate. *North American Journal of Aquaculture* 69:80-89.
- Madhun, A. S., Biering, E., Isachsen, C. H., Omdal, L. M., Einen, A. C. B., Wennevik, and V., Svasand, T., (2015):** Annual report on health monitoring of wild anadromous salmonids in Norway. Report from the Norwegian Veterinary Institute and the Institute of Marine Research, Bergen, Norway. 10:29:58Z.
- Mian, G. F., Godoy, D. T., Lea, C. A. G., Yuhara, Y. T., Costa, G. M., and Figueiredo, H. C. P., (2009):** Aspects of the natural history and virulence of *S. agalactiae* infection in Nile tilapia. Vet. Microbiol., 136: 180-183.
- Moustafa, M. M., Eissa, A. E., Laila, A. M., Gaafar, A. and Elgendy, M. Y. (2015):** Mass Mortalities in Mari-Cultured European Sea Bass (*Dicentrarchus labrax*) at Northern Egypt. Research Journal of Pharmaceutical, Biological and Chemical Sciences, vol. 5, issue (4), pp. 95-109.
- Muiño, L., Lemos, L. M., and Santos, Y., (2001):** Presence of high-affinity iron uptake systems in fish-isolated and environmental strains of *Vibrio anguillarum* serotype O3 Federation of European

Microbiological Societies Volume 202, Issue 1, pages 79-83.

**Nijdam, D., Rood, T., and Westhoek, H., (2012):** The price of protein: review of land use and carbon footprints from life cycle assessments of animal food products and their substitutes. *Food Policy* 37:760–770.

**Noor El- Deen, A.E., Atta, N.S., and Abd El Aziz, M.A., (2010):** Oral Vaccination of Nile Tilapia, *Oreochromis niloticus*, Against Motile Aeromonas Septicemia. *Nature and Science*, 8(2): 21-26.

**Roberts, R.J. (2012):** Fish pathology, Wiley Blackwell, W.B. Saunders, Philadelphia, PA.

**Saad, T.T, Ketkat, S.A., and Mohammed, F. A., (2014):** Changes associated with *Pseudomonas* infection in cultured *Oreochromis species* and its relations to economic losses of fish production farms. *Adv. Res. Agri. Vet. Sci.* Vol. 1, No. 3: 127-137.

**Sachidanandamurthy, K.L., and Yajurvedi, H.N., (2006):** A study on physicochemical parameters of an aquaculture body in Mysore city, Karnataka, India *Journal of Environmental Biology* 27(4) 615-618.

**Sarter, S., Nguyen, H.N., Hung, L.T., Lazard, J. and Montet, D., (2007):** Antibiotic resistance in Gram negative bacteria isolated from farmed catfish. *Food, Cont.*, 18: 1391-1396.

**Tatsuro, H., Daichi T., Yasutada I., and Takayuki H. (2004):** Diversity and seasonal changes in

lactic acid bacteria in the intestinal tract of cultured freshwater fish. *Aquaculture* 234 (1–4): 335–346.

**Umesh, N.R., Mohan, C.V., Phillips, M.J., Bhat, B.V., Ravi Babu, G., Chandra Mohan, A.B. and Padiyar, P.A. (2008):** Risk analysis in aquaculture experiences from small-scale shrimp farmers of India. In M.G. Bondad-Reantaso, J.R. Arthur and R.P. Subasinghe (eds). *Understanding and applying risk analysis in aquaculture. FAO Fisheries and Aquaculture Technical Paper*. No. 519. Rome, FAO. pp. 247–264.

**Vaseeharan, B., Lin, J., and Ramasamy, P., (2004):** Effect of probiotics, antibiotic sensitivity, pathogenicity, and plasmid profiles of *Listonella anguillarum*-like bacteria isolated from *Penaeus monodon* culture systems. *Aquaculture* Volume 241, Issues 1–4, Pages 77–91.

**Vaseeharan, B., Ramasamy, P., Muruganc, T. and Chen, J.C., (2005):** In vitro susceptibility of antibiotics against *Vibrio sp.* and *Aeromonas sp.* isolated from *Penaeus monodon* hatcheries and ponds *Int. J. of Antimicrob. Agents.*, 26: 285–291.

**Wang, L., and Jayarao, B.M., (2001):** *J Dairy Sci*; 84: 1421-1429.

**Witte, W., (2000):** Ecological impact of antibiotics use in animals on different complex microflora environment. *Intern. J. Anti. Agent*, 14: 321-325.

Yan, Q., Chen ,Q., Ma ,S., *alginolyticus* to the intestinal mucus of large yellow croaker (Zhuang, Z., and Wang, X., *Pseudosciaena crocea*). (2007): Characteristics of adherence of pathogenic *Vibrio* Aquaculture, 269: 21-30.

### الملخص العربي

دراسات عن بعض امراض التسمم الدموي التي تصيب اسماك البلطي النيلي في مزارع الاسماك الترايبية  
فيولا حسن زكي، ايمان زهران، رحمة عوض

أجريت هذه الدراسة على عدد ١٥٠ سمكة من أسماك البلطي النيلي تم تجميعها مباشرة من مزارع الاسماك الترايبية بمحافظة الدقهلية حيث تم تجميع ٧٥ سمكة من اسماك البلطي خلال فصلي الشتاء والصيف لسنة ٢٠١٥. قد تم تسجيل كل من الأعراض الاكلينيكية والباثولوجية للأسماك المريضة. كشفت نتائج الفحص البكتريولوجي عن عدد ٦٠ سمكة مصابة اصابه طبيعياً بمختلف العترات البكتيرية (٤٩ سمكة) وكانت معظم البكتيريا المعزولة سالبة الجرام حوالي ٨٢,٢ % في حين البكتيريا ايجابية الجرام تمثل نحو ١٨,٩١%. كان ميكروب الايرومونس هيدروفيليا الاكثر انتشارا ٣٣% تلاها ميكروب السيدوموناس فلوريسينس ٢٠% يليها بالاستريبتوكوكس اجالكتيا ١٧%، الايروموناس سويريا ١٢%، الفيبرو انجيولارم ٩,٥% بينما كان معدل اصابه بفيبرو الجينوليتيكس ٦,٨% فقط. كشفت الدراسة ان أعلى معدل انتشار العدوى البكتيرية في موسم الصيف من فصل الشتاء ، ٣٩% من اسماك البلطي مصاب في الصيف ولكن ٢١% فقط مصاب في فصل الشتاء. وتختلف العترات المعزولة في حساسيتها لمختلف المضادات الحيوية. التغيرات الهيستوباثولوجي كانت واضحة جدا مع الامراض البكتيرية. الامراض تتأثر بالتغيرات البيئية مثل الحرارة، درجة الحموضة، الملوحة، الامونيا، وبعض العناصر الثقيلة.