

Chemical, Microbiological and Enzymatic Evaluation of Mastitic Milk

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Abstract

A total of 400 quarter milk samples were collected from 100 dairy animals bred in three dairy farms in Suez Canal area examined by California Mastitis Test (CMT) and found that 102 samples were positive for subclinical mastitis (SCM). Positive samples microbiologically revealed that the most common bacterial isolates from subclinical mastitis cases were *Staphylococcus aureus* 92 (90.20%), *Staphylococcus epidermidis* 18 (17.65%), *Streptococcus* spp. 79 (77.45%) and *E. coli* 35 (34.31%). On the other hand, total yeast and mould count were 74 (72.54). Chemical and enzymatic examinations in positive CMT milk samples revealed that mean level of milk lactose % was 3.16 ± 0.79 , while the mean level of milk chloride % was 0.13 ± 0.04 significantly higher in positive CMT milk samples, mean value of LDH and ALP enzyme was 503.52 ± 14.21 IU/ML and 723.77 ± 21.30 IU/ML which significantly higher in positive CMT samples than negative CMT samples. Therefore, our study concluded that milk lactose, chloride and enzymes are considered to be suitable diagnostic methods for diagnosis of SCM in dairy animals.

Key words: Subclinical mastitis, etiology, milk LDH, ALP, lactose, chloride; *Staphylococcus aureus*, *Streptococcus* spp., *E. coli*.

Introduction

Milk considered as good supplement of nutrients for human diet. It contains all the food constituents required. Its composition is affected by the breed of animals, species, health of udder, stage of lactation and diseases affected udder (Sharif *et al.*, 2007). Mastitis is an inflammatory change of the mammary gland characterized by an increase in

somatic cells in the milk and pathological changes in mammary tissues. (Souto *et al.*, 2010). It is a dangerous disease of dairy animals which is found in clinical and subclinical forms causing great economic losses and of public health concern. (Sharif *et al.*, 2007). Mastitis is the first cause of elevated somatic cell count, so affects both quality and quantity of milk. Elevated milk SCC is associated with changes in protein

quality, change in fatty acid composition, lactose, and mineral concentration, increased enzymatic activity and PH of rawmilk (Auldistet *al.*, 1996 and Coulon *et al.*, 2002). Bacteria that causing mastitis in cattle are transmitted through raw milk from infected udder and cause disease problems in human., such type of bacteria includes Mycobacterium, Brucella and Staphylococcus and Streptococcus species. (Dagnaw, 2015). Many studies were revealed that some changes detected in enzyme activities due to mastitis (Andrei *et al.*, 2011). Detection of enzyme activity in milk considered as reliable markers for early diagnosis of subclinical mastitis (Babaeiet *al.*, 2007; Guhaet *al.*, 2012). Milk enzymes ALP and LDH were markedly increased in mastitis and they considered both as the early indicators of acute mastitis (Larsen *et al.*, 2010).

There for the aim of this study was based on: 1- Detection of subclinical mastitis in dairy animals by using screening test (CMT),2- Determination of lactose and chloride percent,3- Determination of lactate dehydrogenase enzyme and alkaline phosphates enzyme,4- Microbiological analysis of positive CMT samples.

Materials And Methods

1. Animals:

A total of 400quarter milk samples from100 dairy animals breaded in three dairy farms at Suez Canal

area, Egypt were subjected to this investigation. Animals selection based on the age and stage of lactation.

2.Sampling:

Before milking each udder was washed with water and soap then washed with potassium permanganate 1/1000 solution, dried with clean towel, The teat was disinfected with 70% ethyl alcohol. The first three streams were rejected then 150 ml of milk for each sample were sent aseptically in a sterile screw capped bottlesto be examined. Each sample was thoroughly mixed before divided into three subsamples used for screening tests, microbiological and chemical examination.

3. California mastitis test (CMT) APHA, (2004)

Equal amount (2ml) of milk and frieso-test reagent were mixed thoroughly in a cup of black plastic paddle and swirl gently of the paddle for 10seconds. Results were recorded according to the tendency of gel formation and expressed as strong positive (+++), positive (++) , weak positive (+)or negative (-).

4. Microbiological examination:

2.1-Preparation of samples for microbiological examination

according to APHA (2004): Milk samples were microbiologically examined forEnumeration, Isolation and identification of Staphylococcus spp. according to Deibel and Herrttman (1984) on Baird-Parker agar plates andincubated at 37°C for 48

hours and isolation of *E. coli* was carried out according to APHA (2004) on eosin methylene blue agar (EMB) plates and incubated at 37 °C for 24 hours. Isolation and count of streptococcus spp. according to APHA (2004) on aulinazid agar medium plates and incubated at an inverted position at 36± 1° C for 48 hours and identification of streptococcus species done according to Koneman *et al.* (1988) and Quinn *et al.* (1994). Total Yeast and mould count was carried out using Sabouraud dextrose agar medium according to APHA (2004).

5. Chemical examination:

Estimation of milk lactose and chloride levels according to *Analysis of milk and its product* (2005).

Measurement of alkaline phosphatase and lactate dehydrogenase enzyme according to Bergmeyer (1974) and Goldberg and Spooner (1983) respectively: Milk samples were skimmed by centrifugation at 10,000 gm for 20 min at 4°C. Defatted milk samples were used for enzyme activity estimations of lactate dehydrogenase enzyme (LDH) and alkaline phosphatase (ALP) activity were assayed by spectrophotometer.

Results

Table 1: Incidence of subclinical mastitis in examined quarter milk samples according to California mastitis test.

NO. of animals	No. of quarters milk Samples	Positive samples NO.	Positive samples %
100	400	102	25.5

Table 2: Correlation between positive CMT score, % of mastitic milk samples and microbiological results.

Positive CMT scores	No. of samples	% of mastitic milk samples	NO. of Microbiological positive samples	Accuracy %
+	38	37.25	38	100%
++	39	38.24	39	100%
+++	25	24.51	25	100%
Total	102	100	102	100%

Table 3: Incidence and count of some microorganisms in examined positive CMT quarter milk samples (n=102).

Isolated organism	Incidence		Total count(cfu/ml)		
	NO	%	Min	Max	Mean± SE.
<i>S. aureus</i>	92	90.20	6×10 ²	5×10 ⁶	2.8×10 ⁵ ± 8.7×10 ²
<i>S. epidermidis</i>	18	17.65	2×10 ²	1×10 ⁵	1.2×10 ⁴ ± 1.7×10 ²
Strept.spp.	79	77.45	1×10 ²	2×10 ⁹	2.6×10 ⁷ ± 1.5× 10 ²
<i>E.coli</i>	35	34.31	1×10 ²	1.3×10 ⁶	8.4×10 ⁴ ± 0.53×10 ²
Total yeast and mold count	74	72.54	1×10 ²	1×10 ⁷	1.7×10 ⁵ ± 1×10 ²

Table4: Statistical analytical results of Milk Chemical and Biochemical parameters values based on CMT in examined quarters' milk samples (n=400).

Biochemical Parameters	CMT positive milk samples(n=102)			CMT negative milk samples(n=298)		
	Min	Max	Mean ± SE	Min	Max	Mean ± SE
Lactose%	2.23	5.7	*3.16 ± 0.79	3.15	6.17	4.12±0.20
Chloride%	0.07	0.24	*0.13 ± 0.04	0.03	0.10	0.06±0.001
ALK.Ph. Enzyme	276.18 IU/ML	2561 IU/ML	***723.77 ± 21.30	81.70 IU/ML	256.40 IU/ML	186.4±15.6
LDH. Enzyme	165 IU/ML	844 IU/ML	***503.52±14.21	68.40 IU/ML	190.17 IU/ML	158.7±7.04

*Significance at P< 0.05. *** Significance at P < 0.001.

Discussion

1- Incidence of subclinical mastitis by California Mastitis Test (CMT).

Results summarized in table (1) revealed that 102 out of 400 examined quarter milk samples (25.5%) were positive for CMT. The obtained result was nearly similar to those obtained by *Islam (2011) and Saidiet al. (2013)*, while relatively higher incidence were obtained by *Ayanoet al. (2013), Murugaiyahet al. (2014) and Rahmanet al.(2014)*. Whereas comparatively lower incidence were recorded by *Hashemiet al. (2011) and Hussein (2012)*. California

mastitis test generally used as rapid test for detection of sub-clinical mastitis which detects somatic cell nuclear material, depending on a threshold of 300,000 SCC per milliliter (*Radostitset al., 2000*). Results given in table (2) proved that 100% positive CMT samples in all scores were microbiologically positive. The obtained results clarified a good correlation between CMT in all scores and microbiological results. Lower correlation was reported by *Hussein (2012)*. Nearly similar results were recorded by *Saidiet al. (2013) and Sanotheranet al. (2016)*, they found that 97%, 95% and 93.9% of CMT yielded

bacterial growth, respectively. Inspection of table (2) showed that 38(37.25%), 39(38.24%) and 25(24.51%) of examined samples were positive for CMT scores (+), (++) and (+++) respectively. These results were being disagreed with those obtained by *Sabuncuet al. (2013)* who reported that 82.58%, 14.83% and 2.58% were positive of CMT score (+), (++) and (+++), respectively. Nearly similar result was reported by *Nabih and Abd-El. Rahman (2015)*, they reported that 16%, 32% and 52% of examined samples were positive for CMT score (+), (++) and (+++) respectively.

2- Microbiological evaluation of positive samples.

It was evident that *S. aureus* could be isolated from 92 out of 102 (90.2%) of microbiologically positive samples in single and/or mixed infection. Results in table (3) revealed that the total *S. aureus* count in microbiologically positive mastitis milk samples ranged from 6×10^2 to 5×10^6 with a mean count value of $2.8 \times 10^5 \pm 8.7 \times 10^2$ cfu / g. The results indicated that *S. aureus* was the first major pathogenic organism incriminated in subclinical mastitis and this was potentiated by what had been reported by several authors; (*Nagwaet al., 2015; Abdel Tawabet al., 2016 and Sanotharan et al., 2016*). Data tabulated in table (3) revealed that *S. epidermidis* count in microbiologically positive mastitis milk samples was ranged

from 2×10^2 to 1×10^5 with a mean count of $1.2 \times 10^4 \pm 1.7 \times 10^2$ cfu / g. *S. epidermidis*, is often regarded as a culture contaminant but its importance as a pathogen has been recognized in recent years. *S. epidermidis* is a common cause of infection indwelling foreign devices, surgical wound and bacteremia in immunocomprised patients (*Blum and Rodvold, 1987*). Results given in table (3) revealed that the total Streptococcus species count in microbiologically positive mastitis milk samples ranged from 1×10^2 to 2×10^9 with a mean count of $2.6 \times 10^7 \pm 1.5 \times 10^2$ cfu / g. streptococcus spp. classified to *Strept. pyogenes* was isolated from 44(43.14) where organism was isolated from subclinical mastitis milk samples by many authors with different incidence (*Saidiet al., 2013, Murugaiyahet al. 2014 and Mureithi and Njuguna 2016*). While *Strept. agalactiae* was isolated from 30 (29.41%) Nearly similar result was recorded by *Ahmed et al. (2008)*. Higher results were recorded by *Plozzaet al. (2011) and Ramirez et al. (2014)*. Lower records were obtained by *Ayanoet al. (2013), El Sayedet al. (2015) and Sztachanskaet al. (2016)*. *E. coli* represent the third important causative bacterial agents isolated from examined mastitis milk samples in this work, Inspection of table (3) revealed that the total *E. coli* count in microbiologically positive mastitis milk samples

ranged between 1×10^2 and 1.3×10^6 with a mean count of $8.4 \times 10^4 \pm 0.53 \times 10^2$ cfu/g., Nearly similar results were reported by *Plozzaet al. (2011)*, *Hussien (2012)* and *Abd-ELrahman (2013)*. Lower findings were recorded by *Ali et al. (2015)* and *Sanotharan et al., (2016)*. Higher results were recorded by *Ahmed et al. (2008)* and *Nagwaet al. (2015)*. Result recorded in table (3) revealed that the total yeast and mould counts of examined samples ranged from 1×10^2 to 1×10^7 with a mean count value of $1.7 \times 10^5 \pm 1 \times 10^2$ cfu / g., Lower results of yeast and mould counts were recorded by *Rajeev et al. (2011)* and *Murugaiyahet al. (2014)*. On contrary, sporadic incidence of subclinical mastitis due to yeast had been reported by *Dudkoet al. (2003)* and *EbrahimandNikookhah. (2005)*.

3- Chemical and enzymatic evaluation of positive CMT quarter milk samples:

Table (4) revealed that lactose content in examined positive CMT quarter milk samples ranged from 2.23 to 5.7% with a mean value of 3.16 ± 0.79 , while in negative CMT quarter milk samples was ranged from 3.15 to 6.17 % with mean value of 4.12 ± 0.20 . lactose content showed significant ($p < 0.05$) decreased in positive CMT milk samples ($P < 0.05$), The obtained result, similar to those obtained by *Sharif et al. (2007)*, *Hamid et al. (2012)* and *Nagwaet al. (2015)*.

chloride content in examined positive CMT quarter milk samples was ranged from 0.07 to 0.24 with a mean value of 0.13 ± 0.04 , while in negative CMT milk samples ranged from 0.03 to 0.10 with a mean value of 0.06 ± 0.001 ., Results obtained in this work showed that chloride content increased significantly ($P \leq 0.05$) in positive CMT quarter milk sample. Inspection of table (4) revealed that Alkaline phosphatase enzyme in examined positive CMT quarter milk samples ranged from 276.18 IU/ML to 2561 IU/ML with a mean value of 723.76 ± 21.30 . While in negative CMT samples ranged from 81.70 IU/ML to 256.40 IU/ML with a mean value of 186.40 ± 15.60 . Also lactate dehydrogenase enzyme content in examined positive CMT quarter milk samples ranged from 165 IU/ML to 844 IU/ML with a mean value of 503.52 ± 14.21 , while in negative CMT quarter milk samples ranged from 68.4 to 190.17 with a mean value of 158.7 ± 7.04 . The obtained results indicate that Alkaline phosphatase enzyme and lactate dehydrogenase enzyme increased significantly ($P \leq 0.001$) in positive CMT quarter milk samples. The obtained results were nearly similar to those obtained by *Aliaaet al. (2013)* and *Nagwaet al. (2015)*. Lower results were reported by *Zeinhomet al. (2013)* and *Nabih&abd. El Rahman (2015)*.

Conclusion and Recommendations

The obtained results revealed that CMT and lactose and chloride as well as enzyme evaluation of mastitis milk can be considered as efficient tests for detection of subclinical mastitis in cows and buffaloes. Therefore, in order to minimize the risk of infection of milk and to safeguard consumers, the following suggestions should be applied:

1- The herd should be periodically examined for subclinical mastitis using screening tests and confirmed by detection of enzymes level in milk.

2- Separation of the infected animals from healthy one and milked last or by special precautions and the milk from infected quarter should not be mixed with the bulk milk and discarded.

3- Efficient treatment of infected animals using effective drugs and retest them after suitable time to prove their complete cure.

4- Application of good herd management and strict hygienic measures including:

a- Functionally adequate milking machine should be used in a correct manner.

b- Proper hand –milking procedures with efficient washing and drying of milker's hand and udder.

c- Good cleaned and sanitized milking equipments should be used.

d- Application of teat dip after milking using a suitable and effective antiseptic solution.

5- Using a suitable scheme for prophylactic treatment of udder during drying period.

References:

Abd- Elrahman, A. H. (2013): Mastitis in housed dairy buffaloes: incidence, etiology, clinical finding, antimicrobial sensitivity and different medical treatment against *E. coli* mastitis. Life Science Journal;10 (1). Abstract(6-9).

Abd El-Tawab, A.A.; Ammar, A.M.; Sleim, M. Abd El-Hakeem; El Hofy, Fatma, I. and Salem, Heba. S. S. (2016) Prevalence and antimicrobial susceptibility pattern of *Staphylococcus aureus* isolated from dairy cattle's subclinical mastitis in EL-Sharkia Governorate, Benha veterinary medical journal, vol. 30, no. 1:11-19, march, 2016.

Ahmed, W.M.; Abd El-Moez, Sherein I. and Nabil, Ghada M. (2008): observations on sub-clinical mastitis in Buffalo –cows with Emphasis on measuring of milk Electrical Resistance for its early detection ., Global Veterinaria 2(1) ;41-45.

Ali, Z.; Dimri, U. and Jhambh, R. (2015): prevalence and antibiogram of bacterial pathogens from subclinical mastitis in buffaloes, Buffalo Bulletin (March 2015) Vol.34 No.1, abstract (15-21).

Aliaa A. E. Mohamed, Ahlam K.A Wahba, Ragaa A.S.R.faisal and Yousreya H. M (2013): Some

- Bacteriological and Biochemical Studies on Subclinical Mastitis in Buffaloes. New York Science Journal 2013; 6 (7).Discussion(table3).
- American public Health association “APHA” (2004):** Compendium of methods for microbiological examination of food. 17thEd., APHA, Washington D.C.USA.
- Analysis of Milk and Its Products (2005):** A lab Manual Milk Industry Foundation (U.S.), Milk industry foundation U.S. Biotech Books 2005., ISBN 8176221279, 9788176221276, No. page 629.
- Andrei, S.; Matei, S.; Fit, N.; Cernea, C.; Ciupe, S.; Bogdan, S. and Groza, I.S. (2011):** Glutathione peroxidase activity and its relationship with somatic cell count, number of colony forming units and protein content in subclinical mastitis cows milk. Romanian Biotechnological Letters. 16(3): 6209-6217.
- Auldist, M. J.; Coats, S.; Sutherland, B.J.; Mayes, J.J. and McDowell, G.H. (1996):** Effect of somatic cell count and stage of lactation on raw milk composition and the yield and quality of cheddar cheese. Journal of Dairy Research 63, 269-280.
- Ayano, A. A.; FikiruHiriko; MollaSimyalew A., and Yohannes, A. (2013):** Prevalence of subclinical mastitis in lactating cows in selected commercial dairy farms of Holetadistrict., Journal of Veterinary Medicine and Animal Health Vol. 5(3), pp. 67-72.
- Babaei, H.; Mansouri-Najand, L.; Molaei, M.M.; Kheradmand, A. and Sharifian, M. (2007):** Assessment of lactate dehydrogenase, alkaline phosphatase and aspartate aminotransferase activities in cow’s milk as an indicator of subclinical mastitis. Vet. Res. Commun., 31, 419-425.
- Bergmer, H.U. (1974):** Methods of enzymatic analysis, 2ndEd. A Cadmic press, London, pp.574-582-869.
- Blum, R.A. and Rodvold, K.A.(1987):** Reconition and importance of staphylococcus epidermidis infections. Clin.pharm.1987 Jun, 6 (6):464-475.
- Coulon, J.B., Gasqui, P., Barnouin, J., Ollier, A., Pradel, P. and Pomiès, D. (2002):** Effect of mastitis and related-germ on milk yield and composition during naturally occurring udder infections in dairy cows. Animal Research 51, 383-393.
- Dagnaw, G. G. (2015)** Public Health Significance of Bovine Mastitis, World J. Biol. Med. Science, Volume 2 (4), 20-31, 2015.
- Deibel, I. and Harrtman, K. (1984):** .Compendium of methods for microbiological examination of foods. American public Health Assoc .Washington D.C. USA.
- Dudko, P (2003):** The microbiological examination of milk samples results conducted in

the Northern Great Poland region during the bovine mastitis control. *Annales Univ. Med.Vet.*58,103-116.

Ebrahimi, A. and Nikookhah, F (2005): Identification of fungal agents in milk sample on mastitic cow. *Indian Vet. J.*, 82:52-54.

Elsayed, M. S.; El-Bagoury, A. M. and Dawoud, Mai A. (2015): Phenotypic and genotypic detection of virulence factors of *Staphylococcus aureus* isolated from clinical and subclinical mastitis in cattle and water buffaloes from different farms of Sadat City in Egypt, *Veterinary World* 8(9):1051-1058.abstract(4-8)page.

Goldberg, D.M. and Spooner, R. J. (1983): Methods of enzymatic analysis. Bergmer H.V. 3 rd ed. no3, 258-265.

Guha, A., Gera, S. and Sharma, A. (2012): Evaluation of milk trace elements, lactate dehydrogenase, alkaline phosphatase and aspartate aminotransferase activity of subclinical mastitis as an indicator of subclinical mastitis in riverine buffalo (*Bubalus bubalis*). *Asian-Aust. J. Anim. Sci.*, 25(3):353-360.

Hamid, I.M.B.; Shuiep, E.S.; El Zubeir, E.M.I.; Saad, A.Z. and El Own, O.A.O. (2012): Influence of *Staphylococcus aureus* Mastitis on Milk Composition of Different Dairy Breeds of Cattle in Khartoum State, Sudan *World's Vet. J.* 2(2): 13-16.

Hashemi. M.; Kafi, M and Safdarian, M. (2011): The

prevalence of clinical and subclinical mastitis in dairy cows in the central region of Fars province, south of Iran., *Iranian Journal of Veterinary Research*, Shiraz University., Vol. 12, No. 3, Ser. No. 36, introduction(1-10).

Hussein, Suha A. (2012): Prevalence and Bacterial Etiology of Subclinical Mastitis in Dairy Cows in Al Sulaimaniyah District. *Kufa Journal for vet.Medical Sciences* Vol. 3 no.1.

Islam, M. A; Islam, M. Z.; Islam, M. A.; Rahman, M. S. and Islam, M. T. (2011): prevalence of subclinical mastitis in dairy cows in selected areas of Bangladesh, *Bangl. J. Vet. Med.* 9 (1) : 73-78, abstract (3:7).

Koneman, E. W.; Stephin, D.; William, M. J; Poul, L. P; Washington, C.and Winn, V. R. (1988): Diagnostic microbiology .3rd ed., Lippincott, J.B Company Phphiladephia.

Larsen, T.; Rontved, C.M.; Ingvarsten, K.L.; Vels, L. and Bjerring, M. (2010): Enzyme activity and acute phase proteins in milk utilized as indicators of acute clinical *E. coli* LPS-induced mastitis. *Animal*, 4(10): 1672-1679.

Murugaiyah, M.; Abdullah, FaezFirdaus J.; Mohammed, K.; SarvananthanPoshpum, S. D/O; Adamu, L.; Osman, A. Y.; Abba, Y. and Tijjani, A. (2014): prevalence and antimicrobial resistance assessment of subclinical mastitis in milk samples from selected dairy farms. *American*

Journal of Animal and Veterinary Sciences 9 (1): 65-70, 2014 ISSN: 1557-4555.

Mureithi, D. K. and Njuguna, M.N. (2016): Prevalence of subclinical mastitis and associated risk factors in dairy farms in urban and peri-urban areas of Thika Sub County, Kenya, *Livestock Research for Rural Development* 28 (2) 2016.

Nabih, A.M. and Abdel EL Rahman, Ghada H.A. (2015): Role of milk tri-iodothyronine (T₃) and some biochemical parameters on udder status in dairy buffaloes, *Assiut Vet. Med. J.* Vol. 61 No. 144 January 2015.

Nagwa A. B. El Hakem and salama, E. M., and Saad A.H. (2015): Some studies on diagnosis of subclinical mastitis., *Animal Health Research J.*,3(2):160-167.

Plozza, K.; Lievaart, J.J.; Potts, G. and Barkema, H.W. (2011): Subclinical mastitis and associated risk factors on dairy farms in New South Wales., *Australian Veterinary Journal* Volume 89, No 1–2, January.

Quinn, P. J.; Carter, M. E.; Markrkey, B. K. and Carter, G. R. (1994): Clinical veterinary microbiology. Mosby year book Europ limited, Lynton House, London., 109-126.

Radostits, O. M., Gay C. C.; Blood D .C. and Hinchcliff, K. W. (2000): Mastitis in: *Veterinary Medicine* 9 ed., Saunders, Edinburg, p603–622.

Rahman M. M.; Munsu M. N.; Ekram M. F.; Kabir M. H.;

Rahman M. T. and Saha S.(2014): Prevalence of Subclinical Mastitis in Cows at Anwara, a Coastal Upazila of Chittagong District in Bangladesh, *J Vet Adv*,4(6):594-598.

Ramírez, N.F.; Keefe, G.; Dohoo, I.; Sánchez, J.; Arroyave, O.; , Cerón, J.; Jaramillo, M. and Palacio, L.G.(2014): Herd- and cow-level risk factors associated with subclinical mastitis in dairy farms from the High Plains of the northern Antioquia, Colombia., 2014 *American Dairy Science Association*. Volume 97, Issue 7, Pages 4141–4150.

Rajeev, R. A.; Gupta, M. K. and Singh, K. K.(2011) Study of bovine mastitis in different climatic conditions in Jharkhand, India, *Veterinary world*, vol.4(5):205-208.abstract (1-5).

Sabuncu, A.; Enginler, S.O. and EmekDumen (2013): The effect of parity, age and season on somatic cell count of dairy cows with subclinical mastitis. *Journal of animal and veterinary advances* 12(4):475-477.

Saidi, R.; Khelef, D. and RachidKaid (2013): Subclinical mastitis in cattle in Algeria: Frequency of occurrence and bacteriological isolates., *Journal of the South Africa Veterinary Association*.,84(1), Art. #929, 5 pages.

Sanotharan, N., Pagthinathan, M. and Nafees, M.S.M (2016): Prevalence of Bovine Subclinical Mastitis and its Association with

Bacteria and Risk Factors in Milking Cows of Batticaloa District in Sri Lanka, International Journal of Scientific Research and Innovative Technology ,ISSN: 2313-3759 ,Vol. 3 No. 6; June 2016.

Sharif, T. A.; Bilal, M Q.; Yousaf, A. and Muhammad, G. (2007): review of effect of severity of sub-clinical mastitis on somatic cell count and lactose contents of buffalo milk, Pakistan vet. j., 27(3): 142-144.

Souto, LI; Minagawa, CY; Telles, EO; Garbuglio, MA; Amaku, M; Melville, PA; Dias, RA; Sakata, ST and Benites, NR (2010): Correlation between mastitis occurrence and the count of

microorganisms in bulk raw milk of bovine dairy herds in four selective culture media. J. Dairy Res., 77: 6370.

Sztachanska, M; Baranski, W; Janowski, T; Pogorzelska, J; Zdunczyk, S (2016): Prevalence and etiological agents of subclinical mastitis at the end of lactation in nine dairy herds in North-East Poland. Pol J Vet Sci; 19(1): 119-24, 20.

Zeinhom, M.M.A; Abed, A.H. and Hashem, K.S (2013) A contribution towards milk enzymes, somatic cell count and bacterial pathogens associated with subclinical mastitis cow's milk, Assiut Vet. Med. J. Vol. 59 No. 138 July 2013.abstract (1-12).

التقييم الكيميائي والميكروبي والانزيمي للبن الناتج عن التهاب الضرع

سارة احمد السيد عبد الرحمن* و احمد حسن سعد** و جيهان اسماعيل ابراهيم

قسم الرقابة الصحية علي الأغذية, كلية الطب البيطري, جامعه قناة السويس, الاسماعيليه, مصر

اجريت الدراسه علي 400 عينه لبن من ارباع الضرع للحيوانات التي تم تجميعها من 100 حيوان من ثلاث مزارع مختلفه في منطقه قناه السويس وفحصها باختبار كاليفورنيا ووجد ان 102 عينه موجب لالتهاب الضرع تحت السريري وكشف الفحص الميكروبيولوجي للعينات الموجه ان المكورات العنقوديهالذهبية كانت 92(90.20%) , المكورات العنقوديه كانت 18 (17.65%) ,المكورات السبقيه 79(77.45%) ,ميكروب الايشرشيا كولاي(القولونيه)35(34.31%) و للفطريات والخمائر كانت 74 (72.54%) .وقد كشف الفحص الكيميائي والانزيمي للعينات الموجه ان متوسط نسبه اللاكتوز للبن كانت 0.79 ± 3.16 اقل من النسبه الطبيعيه بينما كان متوسط نسبه الكلوريد للبن 0.04 ± 0.13 اكثر من النسبه الطبيعيه, وكان متوسط نسبه انزيم اللاكتات ديهيرو جيناز 14.21 ± 503.52 وحده دوليه /مللي و كان نسبه انزيم الفوسفاتيز القلوي 21.35 ± 723.76 وحده دوليه /مللي, حيث كانت الانزيمات نسبتها مرتفعه في العينات الموجه لاختبار الكاليفورنيا عن نسبتها في العينات السالبه لاختبار كاليفورنيا, من النتائج السابقه نستنتج ان قياس نسبه اللاكتوز والكلوريد والانزيمات للبن يعتبر طرق تشخيص مناسبه للكشف عن التهاب الضرع في قطيع الحيوانات الحلابه