

**Effect of Soil Solarization and Bio-fertilization on
Strawberry Production and pathogenic Fungi under Siwa
Oases Conditions**

BY

Gomaa, S.S. and Kobisi A. M.

Doi:10.21608/asajs.2021.143516

قبول النشر: ٢٠٢٠ / ١٢ / ٢

استلام البحث: ٢٠٢٠ / ١١ / ١٦

Abstract:

The present study was carried out during the two successive seasons of 2017-2018 and 2018-2019 in the experimental station, desert research center at Siwa oasis, Marsa Matrouh Governorate to evaluate the effect of soil solarization and bio-fertilization on soil borne microorganisms, weed characters, growth, yield and quality of strawberry. The experiment included two solarization treatments (solarize and non-solarize) and five bio-fertilizers treatments (Bio-fertilizers alone, bio-fertilizers + 0.25 mineral fertilizers, bio-fertilizers + 0.5 mineral fertilizers, bio-fertilizers + 0.75 mineral fertilizers as well as the traditional treatment as a control. The results indicated that, soil solarization increased average soil temperature and eradicated most annual broad and narrow-leaved weeds, increased microorganisms population and reduced rotted fruit caused by pythium or phytophthora as well as increasing of strawberry growth early yield and yield. Traditional treatment produced the highest yield followed by bio-fertilizers treatments with 0.75, 0.50 and 0.25 percent of chemical fertilizers compared with bio-fertilizers alone. On the other hand, bio- fertilizers produced the highest fruit quality and gave the lowest values of rotted fruits.

Kay words: Solarization, bio-fertilization, pathogen fungi, total bacteria, pythium and phytophthora.

INTRODUCTION

Strawberry (*Fragaria x ananassa* Duch.) has been widely cultivated in Egypt. It is one of the most important vegetable crops for local consumption and exportation (planted area is about 11072 hectar and total production about 407240 Ton, **FAO, 2017**). Cultivated area in Egypt has been increasing in recent years especially due to the mediterranean climate, fertile soils, and geographic location which support high production, early and profitability of such a specialty crop (**Abd-Elgawad, 2019**).

Soil-borne diseases cause heavy losses to strawberry production *i.e.* *Macrophomina phaseolina* and *Fusarium spp.* (**Benlioglu et al. 2014**), *Phytophthora cactorum*, *P. citricola* and *Verticillium dahlia* (**Hartz et al., 1993**), *Pythium* and *Rhizoctonia*, (**Camprubi et al.,2007**). Also, **Embaby, 2007; Khafagi, 1982; Tadrous, 1991** and **Tarek, 2004** under Egyptian conditions found that, *Alternaria spp.*, *Aspergillus spp.*, *Botrytis cinerea*, *Rhizopus stolonifer*, *Rhizoctonia solani*, *Phytophthora cactorum*, *Fusarium spp.*, *Penicillium spp.* And *Sclerotinia sclerotiorum* are the most fungal isolates causing strawberry fruit rots.

Soil solarization is a nonchemical soil disinfection method which harnesses solar energy for heating the soil. It involves hydro-chemical processes leading to physical, chemical and biological changes in the soil, which take place during and even after the termination of solarization, **Katan (1998)**. Solarization is a potential alternative practice for soil fumigation which has been phased out due to its environmental risks, where, solarization controlled a wide range of fungal pathogens and weed pests (**Himelrick and Dozier, 1991; Katan, 1981; Katan and DeVay, 1991; Pullman et al., 1981; Stapleton and DeVay, 1986 and Gomaa, 2008**). In this respect, **De vay (1991)** reported that, solarization commonly targets mesophyllic organisms, which include most plant pathogens and pests, without destroyed

the beneficial mycorrhizal fungi and the growth promoting (*Bacillus spp.*). So, the lethal effects are most pronounced on microorganisms which have not good soil competitors and many plant pathogens fall in to this group, since they tend to have specialized physiological requirements which are more adapted to co-existence with the host plant (Stapleton, 1991). Soil solarization is an effective soil disinfestation technique for most vegetable crops (Candido *et al.*, 2008), especially strawberry production (Hartz *et al.*, 1993; Camprubi *et al.*, 2007 and Domínguez *et al.*, 2014).

In the last twenty years, ecological farms have been used environmentally friendly agricultural practices to improve plants yield and fruit quality. A real challenge in ecological fruit production and agricultural sustainability is the reduction of chemical fertilization and chemical treatments for pests and disease control. In this direction, scientists actively search for good agricultural practices and compounds of natural origin that are natural adaptors and do not disturb plants ecological balance (Vasil'eva *et al.*, 2005, Caulet *et al.*, 2013 and Gomaa, *et al.*, 2016).

Bio-fertilization has been widely used, especially with vegetable crops vs lettuce (El Massiry, 2009), globe artichoke (Ibrahim, 2009), Jerusalem artichoke (Hafez, 2013) and thyme (Attia *et al.*, 2006) and strawberry (El-Miniawy *et al.*, 2014 and Gomaa *et al.*, 2016). On potato, Gomaa, (2008) found that organic and bio-fertilization improve yield and enhance the efficacy of solarization and unfortunately beneficial microorganisms (bio-fertilizers) which needed to add after solrization.

It could be a particularly attractive practice for strawberry production in Siwa oasis area which located in the northern part of the western desert of Egypt, where's strawberry crop is grown

as an annual, with a very warm summer fallow period (ideal conditions for solarization) followed by a fall planting through October and November. So, this study was undertaken to document the ability of soil solarization to control annual weeds and soil borne pathogens and its effect on productivity of strawberry plants treated with bio-fertilizers in the warmer planting area of Egypt.

MATERIALS AND METHODS

The present study was carried out during the two successive seasons of 2017-2018 and 2018-2019 in the experimental station, desert research center at Siwa oasis, Marsa Matrouh Governorate to evaluate the effect of soil solarization and bio-fertilization on soil borne microorganisms, weed characters, growth, yield and quality of strawberry.

Experimental design:

During July, soil experiment was ploughed and divided into rows, each one have 1.m width and 10.5m length. Organic fertilizers, rock phosphate and rock potassium (Felsibar) were applied for all plots except the traditional treatment plots (control) where contain organic fertilizers, calcium super phosphate, ammonium sulphate and potassium sulphate as recommended, then fertilizers were incorporated in rows and levelled before trickle irrigation lines were installed.

Soil experiment irrigated abundantly then the trickle lines were removed and soil covered with clear poly ethylene traps of 60 micron thickness for about 6 weeks during August and September, while an untreated soil was used as a control. Soil temperatures were measured weekly during 6 weeks of solarization, then the polyethylene traps were removed and directly soil samples has taken from 0-15 cm depth to determine the densities of microorganisms (total counts of bacteria, fungi, and pathogen fungi). Fresh strawberry transplants cv. Festival

were hand transplanted in 4 rows and 30 cm apart on the med. of October in two seasons, then bio-fertilizers which purchased from the general authority of agricultural funds and equalization, namely Biogen (a symbiotic nitrogen fixing bacteria), Phosphorin (phosphate solubilizing bacteria) and Potassumage (potassium solubilizing bacteria) were applied directly after transplanting and monthly during growth stages for all experiment except the traditional treatment plots.

The experimental design was split plot with 5 replicates. The plot area was 10.5 m² included 140 plants. Soil solarization was assessed in main plot while bio-fertilizers were assessed in sub plot. The experiment includes 10 treatments which were the combination between two solarization treatments (solarize and non-solarize) and five bio-fertilizers treatments (Bio-fertilizers alone, bio-fertilizers + 0.25 mineral fertilizers, bio-fertilizers +0.5 mineral fertilizers, bio- fertilizers + 0.75 mineral fertilizers as well as the traditional treatment as a control. The traditional treatment plots received 300 kg calcium super phosphate (15.5 % P₂O₅) applied during soil preparation then 300 kg ammonium nitrate (33.5 %), 50 kg phosphoric acid (85% P₂O₅) and 250 kg potassium sulfate (48.5 % K₂O) / feddan were divided into 10 equal parts and applied weekly through fertigation system during the growing season starting fifteen days transplanting later. Fertigation occurred four times every week and the bio treatments bio 4 (0.75%) , bio 3 (50%), bio 2 (0.25%) received the fertigation 3, 2, and 1 time every week respectively as well as no received for bio 1.

Soil temperature: during solarization period, soil temperatures at 0, 5, 10 and 15cm were recorded weekly during day hours at 8 am to 8 pm.

Soil microorganisms: soil samples were taken before and after solarization to determine total microbial counts using nutrient

agar medium, PDA-Rose Bengal medium and PDA-PCNB medium to culture bacteria, total fungi and pathogen fungi, respectively. Samples were examined for total fungi and pathogen fungi using the dilution method (Talyour, 1962) and Plate count technique (Johnson *et al.*, 1959). Martine medium Martine (1950) and Nash and Synder (1962) were used to determine fungi and pathogen fungi respectively. Total bacteria was determined by using method of Holt *et al.*, (1994).

Weed measurements: during the second season broad and narrow-leaved were taken from a randomly quadratic meter after 4 and 8 weeks from transplanting to determine average number and total fresh weight.

Growth characters: Two weeks after transplanting, strawberry plants per plot were counted then survival ration were calculated. Six weeks after transplanting, randomly samples of 5 plants from each plot were taken to determine shoot high, shoot fresh weight and leaves number per plant.

Yield components: Strawberry fruits were harvested two times weekly during the growing seasons, counted, and weighed to calculate average fruit number and weight. The early yield per plant was determined as weights of all harvested fruits during the first five harvesting times. Total yield per plant was calculated.

Fruit quality: Twenty five fruits were randomly collected from each treatment in the middle of the growing seasons and fruit firmness was measured using Shatillon penetrometer. Soluble solid content (SSC) was determined by using digital refractometer (Abbe Leica model) and L ascorbic acid content was determined according to the methods described by A.O.A.C. (2005).

Disease incidence Disease incidence was assessed as a total number of rotted fruits as compared with total fruits number from beginning to the end of harvesting time in all treatments.

The diseases of fruits were separated according to different symptoms: gray mold (*Botrytis cinerea*) and dry rot (*Rhizoctonia solani*). Number of fruits in each group were counted and total fruit rot numbers was counted, then the lost yield was calculated.

Statistical analysis: Data were subjected to statistical analysis by M-STAT C (Russell, 1991). The differences among means were performed using least significant difference (LSD) at 5% level.

RESULTS AND DISCUSSION

Soil temperatures: Temperature reading daily recorded every two hours at day time (8 am to 6 pm) ones every week during solarization period at four depths 0, 5, 10, and 15 cm (Fig 1). An increasing in the temperature of solarized soil was observed up to a maximum of 63.8 and 60.2 c° at 2 pm for soil surface compared to 60.2 and 56.6 c° for non-solarized soil surface in first and second seasons respectively. High temperatures at 5 cm depth were 57.2 and 55.8 at 4 pm in solarized plots compared with 49.0 and 49.2 for non-solarized at the same time and depth in two seasons respectively. At 10 and 15 cm depths solarized soil plots recorded (55.5, 54.2) and (52.4, 50.4) at 4 pm in the first and second season, while temperatures in non-solarize treatment recorded (44.2, 41.5) and (41.2, 40.0 c° for first and second season respectively. From the previous data, it was clearly that, covering soil with transparent plastic traps raised the average absolute soil temperatures at the four depths with an increment values (3.6, 8.2, 11.3 and 11.2 c°) and (3.6, 6.6, 12.7 and 10.4) compared with bare soil in the first and second seasons respectively. Although maximum soil temperature decreased with increasing soil depth at all, the deferent between solarize treatments increased with increasing soil depth (Fig. 1). These results are similar with results obtained by other investigators

(Bicici *et al.*,2000; Campiglia *et al.*,2000; Shukla *et al.*, 2000; Peachey *et al.*, 2001; Rieger *et al.*,2001).

Soil microorganisms: Data presented in Table (1) showed, total fungi, total bacteria and pathogen fungi before and justly after solarization treatment. Population of microbes at 15 cm of soil depth drastically reduced with solarize treatments compared with non-solarize. The reduction percentages were 72.37, 87.08 and 92.66 percent for total fungi, total bacteria and pathogen fungi respectively. **Similar results were found by Triki *et al.*, (2001); Hamada, (2002); El-Sheshtawy (2006) and Gomaa, (2008).**

Regarding bio-fertilization effect and development of total fungi, total bacteria and pathogen fungi counts throughout planting season, data presented in table (2) showed that, population of all microorganisms was relatively higher at end of season (April) compared with it at transplanting (October). Also, its counts with non-solarize treatment was higher than solarize. The most pronounced effect was pathogen fungi count, which increased on April compared with October and drastically decreased with solarization compared with non-solarization. Total count of bacteria sharply increased with solarization on April samples compared with October samples. It may be worth to mention that, most increment of bacteria population with solarization belonged to bio-fertilizers supplied which considered a beneficial organisms. From the previous data, we notice that, decreasing of soil microorganisms after solarization may be due to chemical and microbial activities, which led to generation of toxic compounds in vapor and liquid phases and consequently accumulate under plastic mulch especially near soil surface which in turn become more effective against soil flora (**Gamliel *et al.*, 2000**). The effect of solarization was most pronounced on mesophyllic group which include most plant pathogens and pests (**Abu-Gharbieh, 1998**). While most beneficial organisms

belonged to thermophyllic group which can be survive and even flourish under solarization (De Vay and Stapleton 1998).

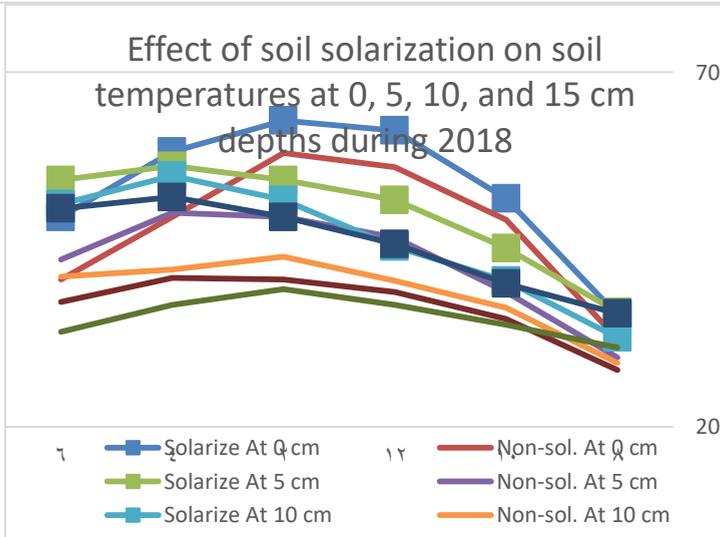
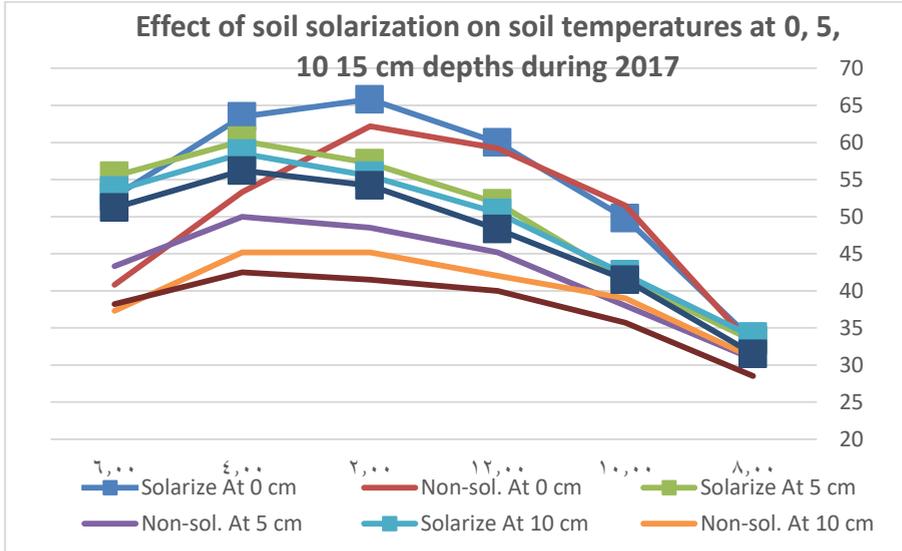


Figure (1) Effect of solarization on soil temperatures at soil surface, 5 cm, 10 cm and 15 cm depth during 2017 and 2018 seasons.

Table (1): Effect of soil solarization on total count of fungi, bacteria and pathogen fungi (CFU/ g dry) pre and post solarization treatment (on July and September) at 15 cm depth.

		Total Fungi (10^4)	Total Bacteria (10^6)	Pathogen Fungi (10^4)
Solarize	Before (July)	188.35	320,86	12.64
	After (September)	04,02	44,20	1,37
Non-Solarize	Before (July)	184,60	331,34	12,38
	After (September)	197,31	342,36	18,67

Table (2): Effect of soil solarization, bio-fertilization and interaction on total count of fungi, bacteria and pathogen fungi (CFU, g dry) on October and April.

Solarization	October (at transplanting)			April (end of season)			
	Total fungi (10^4)	Total bacteria (10^6)	Pathogen fungi (10^4)	Total fungi (10^4)	Total bacteria (10^6)	Pathogen fungi (10^4)	
Solarize	Bio 1	53.5	43.18	1.61	47.5	268.22	4.36
	Bio 2	48.3	44.42	1.64	45.5	261.15	3.38
	Bio 3	47.5	44.51	2.11	44.3	255.57	3.68
	Bio 4	46.3	46.41	1.34	46.6	214.38	3.92
	Bio 5	45.7	45.13	1.67	52.4	86.47	6.18
Non-Solarize	Bio 1	188.3	327.41	16.41	174.5	338.14	22.14
	Bio 2	185.4	335.18	14.32	177.3	347.65	24.35

Bio 3	178.3	312.08	16.42	178.6	365.15	24.22
Bio 4	185.6	314.21	17.17	178.6	344.81	22.35
Bio 5	188.4	315.42	16.54	214.5	317.45	26.15
L.S.D at 0.05 for solarization	15.1	16.2	1.62	5.11	44.46	1.23
L.S.D at 0.05 for fertilization	N.S	N.S	N.S	3.27	16.81	1.34
L.S.D at 0.05 for interaction	N.S	N.S	N.S	4.62	23.77	N.S

Weeds control: The effect of soil solarization, bio-fertilization and their interaction on broad and narrow leaved numbers and fresh weight after four and eight weeks during 2017 growing season are presented in Table (3). Solarization significantly decreased broad-leaved and narrow-leaved numbers and fresh weight compared with non-solarize treatment. On other hand, fertilization treatment and interaction did not have significant effect except broad and narrow-leaved fresh weight after eight weeks from transplanting. Conventional fertilizer treatment produced heaviest, followed by bio 4, bio 3 and bio 2 treatments for broad-leaved or bio 4 and bio 3 for narrow-leaved as compared with bio 1 or bio 1 and bio 2 for broad and narrow-leaved respectively. Concerning interaction effect, conventional fertilizer treatment with non-solarize produced the heaviest broad and narrow-leaved fresh weight after eight weeks compared with other treatments. The most pronounced decreased weeds weight obtained when soil solarized with all fertilizer treatments or with bio 1 and bio 2 for broad and narrow-leaved respectively. Our results indicated that, soil solarization with clear poly ethylene has strong effect on weed germination, as well as fresh weight of broad and narrow-leaved weed after four and eight weeks (Table 3). The reducing of weed number and weight attributed to raising the temperature of soil to lethal levels for weed seed germination (De Vay and stapleton, 1998) or

attributed to chemical, physical and biological changes which caused in the soil that provide effective management of weed control (**Abu-Gharbieh, 1998**). Moreover, we noticed that, increasing narrow- leaved weed numbers compared with broad-leaved after four and eight weeks as well as increasing number and weight of total weeds in general after eight weeks compared with four weeks. These results may be due to that, narrow-leaved seed weed is more tolerant and adapted to lethal effect of high temperature and relatively removing solarisation effect after eight weeks as compared with four weeks. Similar results were obtained by (**Hamada, 2002 and El-Sheshtawy, 20006**).

Table: (3) Effect of soil solarization and bio-fertilization on number and fresh weight of broad-leaved and narrow-leaved at 4 and 8 weeks after transplanting, 2018 season).

Characters	Broad-leaved weeds No		Narrow-leaved weeds No		Broad-leaved weeds fresh weight		Narrow-leaved weeds fresh weight		
	Seasons	4 weeks	8 weeks	4 weeks	8 weeks	4 weeks	8 weeks	4 weeks	8 weeks
Solarization									
Solarize	1.80	٤,٧٢	٤,٢٦	٧,٤٣	١١٧,٠٩	٢٢١,٨٤	٤٦,٨٤	١١٢,٠٩	
Non-solarize	13.02	٣٩,١٠	٤١,٣٠	٦٩,٩١	٥٧٢,٦٧	١٦٩٨,٧٢	٣٧١,٦٩	١٠٤٦,١٠	
LSD at 0.05	1.20	١٠,١	٣,٢٦	٨,٣٧	٤٥,٢٣	٣٩٢,٢٠	٨٣,٤٣	٩٩,٢١	
Bio-fertilization									
Bio-1	٧,٠٢	20.46	21.85	35.30	327.36	724.80	200.11	317.66	
Bio-2	٧,٦٢	22.62	22.74	37.60	356.01	880.94	209.36	338.37	
Bio-3	٧,٥٤	22.69	24.18	42.00	351.01	986.49	221.97	629.98	
Bio-4	٧,٤٢	22.33	23.10	39.61	345.23	989.38	212.42	665.91	
Bio-5	٧,٤٥	21.45	22.03	38.84	344.80	1219.80	202.46	943.57	
LSD at 0.05	N.S	N.S	N.S	N.S	N.S	١٧٣,٤٧	N.S	٥٧,٤١	
Interaction									
Solarize	١,٧٧	4.50	3.49	7.04	114.83	211.66	38.39	63.39	
	١,٩٦	4.75	4.71	7.64	127.40	223.25	51.81	68.79	
	١,٨٣	5.34	4.38	7.38	119.17	250.98	48.14	110.65	
	١,٨١	4.62	4.49	7.49	117.43	217.30	49.39	112.35	
	١,٦٤	4.38	4.22	7.60	106.60	206.02	46.46	205.29	
Non-solarize	١٢,٢٧	36.41	40.20	63.55	539.88	1237.94	361.83	571.92	
	١٣,٢٩	40.49	40.77	67.55	584.61	1538.62	366.90	607.95	
	١٣,٢٥	40.05	43.98	76.62	582.85	1722.01	395.79	1149.30	
	١٣,٠٢	40.03	41.72	71.73	573.03	1761.47	375.45	1219.47	
	١٣,٢٥	38.51	39.83	70.08	583.00	2233.58	358.47	1681.84	
LSD at 0.05	N.S	N.S	N.S	N.S	N.S	٢٤٥,٣٣	N.S	٨١,١٨	
Characters									
Characters	Broad-leaved weeds No		Narrow-leaved weeds No		Broad-leaved weeds fresh weight		Narrow-leaved weeds fresh weight		
	Seasons	4 weeks	8 weeks	4 weeks	8 weeks	4 weeks	8 weeks	4 weeks	8 weeks
Solarization									
Solarize	1.80	٤,٧٢	٤,٢٦	٧,٤٣	١١٧,٠٩	٢٢١,٨٤	٤٦,٨٤	١١٢,٠٩	
Non-solarize	13.02	٣٩,١٠	٤١,٣٠	٦٩,٩١	٥٧٢,٦٧	١٦٩٨,٧٢	٣٧١,٦٩	١٠٤٦,١٠	
LSD at 0.05	1.20	١٠,١	٣,٢٦	٨,٣٧	٤٥,٢٣	٣٩٢,٢٠	٨٣,٤٣	٩٩,٢١	

Bio-fertilization								
Bio-1	٧,٠٢	20.46	21.85	35.30	327.36	724.80	200.11	317.66
Bio-2	٧,٦٢	22.62	22.74	37.60	356.01	880.94	209.36	338.37
Bio-3	٧,٥٤	22.69	24.18	42.00	351.01	986.49	221.97	629.98
Bio-4	٧,٤٢	22.33	23.10	39.61	345.23	989.38	212.42	665.91
Bio-5	٧,٤٥	21.45	22.03	38.84	344.80	1219.80	202.46	943.57
LSD at 0.05	N.S	N.S	N.S	N.S	N.S	١٧٣,٤٧	N.S	٥٧,٤١
Interaction								
Solarize	١,٧٧	4.50	3.49	7.04	114.83	211.66	38.39	63.39
	١,٩٦	4.75	4.71	7.64	127.40	223.25	51.81	68.79
	١,٨٣	5.34	4.38	7.38	119.17	250.98	48.14	110.65
	١,٨١	4.62	4.49	7.49	117.43	217.30	49.39	112.35
	١,٦٤	4.38	4.22	7.60	106.60	206.02	46.46	205.29
Non-solarize	١٢,٢٧	36.41	40.20	63.55	539.88	1237.94	361.83	571.92
	١٣,٢٩	40.49	40.77	67.55	584.61	1538.62	366.90	607.95
	١٣,٢٥	40.05	43.98	76.62	582.85	1722.01	395.79	1149.30
	١٣,٠٢	40.03	41.72	71.73	573.03	1761.47	375.45	1219.47
	١٣,٢٥	38.51	39.83	70.08	583.00	2233.58	358.47	1681.84
LSD at 0.05	N.S	N.S	N.S	N.S	N.S	٢٤٥,٣٣	N.S	٨١,١٨

Vegetative growth: Effect of soil solarization, fertilization treatments and their interaction on transplant survival ratio, leaves number, plant height and plant fresh weight are presented in Table (4). Transplant survival ratio significantly affected by solarization treatment compared with non-solarize treatment, while fertilization treatments or interactions effects were not significant in both seasons.

Concerning vegetative growth characters, data in table 4 showed that, soil solarization affected positively on strawberry plant height and plant fresh weight compared with non-solarize treatment. While its effects on leaves number was not significant. All vegetative growth characters significantly affected by fertilization treatments. Conventional fertilizer produced the highest leaves number, highest plant height and weightiest plants followed by bio 4 treatment compared with bio 1 which produced the lowest values in this respect, followed by bio 2 and bio 3 in both seasons. Regarding to interaction, data

showed non-significant effects on vegetative growth characters except plant fresh weight character, since conventional fertilizer treatment with solarization gave the heaviest plant fresh weight compared with bio 1 fertilizer with non-solarize treatment in both seasons.

Table: (4) Effect of soil solarization and bio-fertilization on plants survival ratio, average leaves number, plant height and fresh weight of strawberry plants, 8 weeks after transplanting.

Characters	Survival ratio		Leaves No.		Plant height (cm)		Plant fresh weight (g)	
Seasons	1 St Season	2 nd Season						
Solarization								
Solarize	93,03	92,67	8,70	8,04	10,22	10,07	30,10	36,40
Non-solarize	78,46	76,24	8,22	8,10	13,06	14,10	28,70	32.73
LSD at 0.05	1,48	1,28	N.S	N.S	1,26	0,18	1,72	1,97
Bio-fertilization								
Bio-1	85.12	84.05	7.41	7.04	13.16	13.16	27.20	30.71
Bio-2	86.38	84.40	7.87	7.77	13.52	13.64	28.63	33.56
Bio-3	85.36	84.40	8.37	8.47	14.72	15.04	33.66	34.72
Bio-4	85.95	83.33	9.28	9.03	15.32	15.37	34.61	36.49
Bio-5	85.92	86.07	9.36	9.42	15.24	15.73	35.53	37.47
LSD at 0.05	N.S	N.S	0.48	0,23	0.62	0.76	1.05	0,88
Interaction								
Solarized	91.67	93.81	7.73	7.11	14.10	13.90	31.44	34.06
	93.00	92.38	8.07	8.00	14.39	14.35	33.32	35.44
	93.33	93.33	8.55	8.62	15.41	15.40	35.47	36.16
	94.05	90.00	9.55	9.31	16.21	15.75	36.95	37.94
	93.10	93.81	9.59	9.66	16.00	15.97	38.33	38.66
Non-solarized	78.57	74.29	7.09	6.98	12.23	12.41	22.96	27.36
	79.76	76.43	7.67	7.54	12.64	12.93	23.93	31.69
	77.38	75.48	8.18	8.32	14.03	14.67	31.85	33.29
	77.86	76.67	9.01	8.74	14.42	14.98	32.27	35.03
	78.74	78.33	9.13	9.17	14.48	15.48	32.73	36.28
LSD at 0.05	N.S	N.S	N.S	N.S	N.S	N.S	1,49	1,24

Bio-5	85.92	86.07	9.36	9.42	15.24	15.73	35.53	37.47
LSD at 0.05	N.S	N.S	0.48	٠,٢٣	0.62	0.76	1.05	٠,٨٨
Interaction								
Solarized	91.67	93.81	7.73	7.11	14.10	13.90	31.44	34.06
	93.00	92.38	8.07	8.00	14.39	14.35	33.32	35.44
	93.33	93.33	8.55	8.62	15.41	15.40	35.47	36.16
	94.05	90.00	9.55	9.31	16.21	15.75	36.95	37.94
	93.10	93.81	9.59	9.66	16.00	15.97	38.33	38.66
Non-solarized	78.57	74.29	7.09	6.98	12.23	12.41	22.96	27.36
	79.76	76.43	7.67	7.54	12.64	12.93	23.93	31.69
	77.38	75.48	8.18	8.32	14.03	14.67	31.85	33.29
	77.86	76.67	9.01	8.74	14.42	14.98	32.27	35.03
	78.74	78.33	9.13	9.17	14.48	15.48	32.73	36.28
LSD at 0.05	N.S	N.S	N.S	N.S	N.S	N.S	١,٤٩	١,٢٤
Characters	Survival ratio		Leaves No.		Plant height (cm)		Plant fresh weight (g)	
Seasons	1 st	2 nd	1 st	2 nd	1 st	2 nd	1 st	2 nd
	Season	Season	Season	Season	Season	Season	Season	Season
Solarization								
Solarize	٩٣,٠٣	٩٢,٦٧	٨,٧٠	٨,٥٤	١٥,٢٢	١٥,٠٧	٣٥,١٠	٣٦,٤٥
Non-solarize	٧٨,٤٦	٧٦,٢٤	٨,٢٢	٨,١٥	١٣,٥٦	١٤,١٠	٢٨,٧٥	32.73
LSD at 0.05	١,٤٨	١,٢٨	N.S	N.S	١,٢٦	٠,١٨	١,٧٢	١,٩٧
Bio-fertilization								
Bio-1	85.12	84.05	7.41	7.04	13.16	13.16	27.20	30.71
Bio-2	86.38	84.40	7.87	7.77	13.52	13.64	28.63	33.56
Bio-3	85.36	84.40	8.37	8.47	14.72	15.04	33.66	34.72
Bio-4	85.95	83.33	9.28	9.03	15.32	15.37	34.61	36.49
Bio-5	85.92	86.07	9.36	9.42	15.24	15.73	35.53	37.47
LSD at 0.05	N.S	N.S	0.48	٠,٢٣	0.62	0.76	1.05	٠,٨٨
Interaction								
Solarized	91.67	93.81	7.73	7.11	14.10	13.90	31.44	34.06
	93.00	92.38	8.07	8.00	14.39	14.35	33.32	35.44
	93.33	93.33	8.55	8.62	15.41	15.40	35.47	36.16
	94.05	90.00	9.55	9.31	16.21	15.75	36.95	37.94
	93.10	93.81	9.59	9.66	16.00	15.97	38.33	38.66
Non-solarized	78.57	74.29	7.09	6.98	12.23	12.41	22.96	27.36
	79.76	76.43	7.67	7.54	12.64	12.93	23.93	31.69
	77.38	75.48	8.18	8.32	14.03	14.67	31.85	33.29
	77.86	76.67	9.01	8.74	14.42	14.98	32.27	35.03

	78.74	78.33	9.13	9.17	14.48	15.48	32.73	36.28
LSD at 0.05	N.S	N.S	N.S	N.S	N.S	N.S	1,49	1,24
Characters	Survival ratio		Leaves No.		Plant height (cm)		Plant fresh weight (g)	
Seasons	1St	2nd	1St	2nd	1St	2nd	1St	2nd
	Season	Season	Season	Season	Season	Season	Season	Season
	Solarization							
Solarize	93,03	92,67	8,70	8,04	10,22	10,07	30,10	36,40
Non-solarize	78,46	76,24	8,22	8,10	13,06	14,10	28,70	32.73
LSD at 0.05	1,48	1,28	N.S	N.S	1,26	0,18	1,72	1,97
	Bio-fertilization							
Bio-1	85.12	84.05	7.41	7.04	13.16	13.16	27.20	30.71
Bio-2	86.38	84.40	7.87	7.77	13.52	13.64	28.63	33.56
Bio-3	85.36	84.40	8.37	8.47	14.72	15.04	33.66	34.72
Bio-4	85.95	83.33	9.28	9.03	15.32	15.37	34.61	36.49
Bio-5	85.92	86.07	9.36	9.42	15.24	15.73	35.53	37.47
LSD at 0.05	N.S	N.S	0.48	0,22	0.62	0.76	1.05	0,88
	Interaction							
Solarized	91.67	93.81	7.73	7.11	14.10	13.90	31.44	34.06
	93.00	92.38	8.07	8.00	14.39	14.35	33.32	35.44
	93.33	93.33	8.55	8.62	15.41	15.40	35.47	36.16
	94.05	90.00	9.55	9.31	16.21	15.75	36.95	37.94
Non-solarized	93.10	93.81	9.59	9.66	16.00	15.97	38.33	38.66
	78.57	74.29	7.09	6.98	12.23	12.41	22.96	27.36
	79.76	76.43	7.67	7.54	12.64	12.93	23.93	31.69
	77.38	75.48	8.18	8.32	14.03	14.67	31.85	33.29
	77.86	76.67	9.01	8.74	14.42	14.98	32.27	35.03
	78.74	78.33	9.13	9.17	14.48	15.48	32.73	36.28
LSD at 0.05	N.S	N.S	N.S	N.S	N.S	N.S	1,49	1,24

These results indicated that, the changes in soil properties due to solarization may have positive effect vegetative growth characters of strawberry plants, in addition that, soil temperature effects, soil microorganisms and weed control revealed, solarization caused beneficial conditions such as enhancement of soil physiology properties, availability of nutrients, weed eradication, inhibition of pathogens and stimulation of beneficial microorganisms which were add as a bio-fertilizers (Stapleton,

1991; Hartz *et al.*, 1993; Camprubi *et al.*, 2007). However, conventional treatment produced the vigorous plants compared with other treatments especially bio-1 and bio-2 which received least amount of chemical fertilizers and consequently gave lowest vegetative growth. Similar results were found by (El-Miniawy *et al.*, 2014 and Gomaa *et al.*, 2016).

Yield and its component:

Data presented in Table (5) showed that, soil solarization and bio-fertilization significantly affected on early yield per plant, total yield per plant, total yield per plot and average fruit weight in both season, while interaction effects were not significant except early yield character in both seasons. Soil solarization increased early yield, total yield per plant, total yield per plot and average fruit weight as compared with non-solarize treatment in both seasons. Conventional fertilization and bio-4 treatments gave the highest early and total yield per plant followed by bio-3 compared with bio-1 treatment, which gave the lowest values in this respect. Also, conventional treatment significantly increased total yield per plot and average fruit weight followed by bio-4 then bio-3 treatment compared with bio-1 which gave the lowest values followed by bio-2 in both seasons. Regarding interaction effect on early yield, conventional and bio-4 with solarization produced the highest significant early yield compared with bio-1 with solarize or non-solarize in both seasons. Generally, solarization enhanced strawberry yield characters and the increment was most pronounced with increasing chemical fertilizer rate by one hundred percent and three quarter. Also, solarization superiority was evident on total yield per plot compared with other characters may be due to it's a positive effects on plants survival ratio, fruits rot percent and weeds growth suppression and consequently absent the competition especially at the early

growing stage. In order to that, strawberry plants had advantage to increasing growth compared with non-solarize treatment. These results indicated that, the changing in soil properties resulted from solarization may have a positive effect on transplants standing and improve its survival which resulted more plants per unit area (**Candido et al.,2008**), improvement vegetative growth (**Porras et al., 2007**). Our previous data on soil temperatures, soil microorganisms and weed control (Tables 1,2,3 and 4) revealed that, solarization caused a good conditions such as improving chemical and physical properties and availability of nutrients, eradication of annual weeds as well as inhibition of pathogens and stimulation of beneficial microorganisms. This conditions led stimulate strawberry growth especially at early stages and consequently increased growth, early yield and total yield of strawberry (**Domínguez et al., 2014 and Ozyilmaz et al., 2016**). Moreover, adding bio fertilizers relatively enhanced soil population of microorganism at end of season compared with transplanting time, where, the most counts of microorganisms in solarized plots belonged to beneficial groups (**Stapleton, 1991**). So, adding bio-fertilizers enhance the efficacy of solarization and gave unfortunately for beneficial microorganisms (bio-fertilizers) to living and flourish (**Gomaa, (2008)**). Increasing strawberry growth and yield and enhancement of its fruit quality were reported by many researchers (**El-Miniawy et al., 2014 and Gomaa et al., 2016**).

Table: (5) Effect of soil solarization and bio-fertilization on average fruit weight, early yield/plant, total yield/plant and total yield/plot of strawberry plants.

Characters	Average fruit weight (g)		Early yield/plant (g)		Total yield/plant (g)		Total yield/plot (kg)	
	1 St Season	2 nd Season	1 St Season	2 nd Season	1 St Season	2 nd Season	1 St Season	2 nd Season
Solarization								
Solarize	١٢,٢٩	١٢,٤٢	٢١٠,٩	٢٢٥,٦	٤١٦,٦	٤٢٢,٢	٥٤,٠٤	٥٤,٣٣
Non-solarize	١٠,٨٣	١١,٢٠	١٧٤,١	١٨٣,٨	٣٧٠,٤	٣٨٤,٥	٣٩,٥٧	٣٩,٥٠
LSD at 0.05	٠,١٥٤	٠,٤٢	١٢,٢	١٤,٦	٢٦,٨	٢٥,٩	٣,٨٧	٢,٢٥
Bio-fertilization								
Bio-1	9.67	9.79	152.8	172.5	324.7	336.7	38.58	38.51
Bio-2	10.86	11.16	187.8	203.2	367.5	377.9	44.20	44.32
Bio-3	11.81	12.06	196.7	208.3	391.7	405.0	46.48	47.41
Bio-4	12.28	12.61	208.4	215.0	427.5	439.8	50.80	50.43
Bio-5	13.19	13.42	216.6	224.7	456.1	457.1	53.87	53.91
LSD at 0.05	0.48	٠,٥٧	8.6	١٢,٥	١٤,٤	24.9	2.27	٢,٩٧
Interaction								
Solarize	10.50	10.16	155.3	175.3	341.8	341.1	43.67	44.63
	11.63	11.96	206.9	224.4	386.6	393.7	50.12	50.63
	12.51	12.68	218.6	235.3	417.6	430.9	54.30	56.00
	13.20	13.53	233.1	243.1	449.2	460.5	58.84	57.56
	13.63	13.76	240.4	250.3	488.0	484.4	63.09	62.84
Non-solarize	8.84	9.41	150.3	169.9	307.6	332.3	33.49	32.40
	10.09	10.35	168.8	182.2	348.4	362.1	38.29	38.01
	11.11	11.44	174.7	181.4	365.8	379.1	38.66	38.83
	11.36	11.69	183.6	186.9	405.8	419.0	42.75	43.29
	12.75	13.08	192.9	199.6	424.1	429.8	44.65	44.99
LSD at 0.05	N.S	N.S	12.1	١٧,٦	N.S	N.S	N.S	N.S

Fruits quality:

The effect of soil solarization, bio-fertilization and interaction on strawberry fruit firmness, T.S.S and L. Ascorbic acid content are presented in Table, (6). Solarization significantly enhanced fruit firmness compared with non-solarize treatment, while the same treatment had not have significant effect on strawberry fruit T.S.S or L. Ascorbic acid content in both seasons.

Concerning of bio-fertilization effect, data in Table (6) showed that, fruit firmness, T.S.S and L. Ascorbic acid content significantly affected by bio-fertilization treatments. Bio-3 in first season, bio-3 with bio-2 in the second gave the highest fruit firmness followed by other bio-fertilizer treatments as compared with conventional treatment which produced the lowest fruit firmness in both seasons. Bio-3 alone or bio-3 and bio-2 gave the highest T.S.S values in first and second seasons respectively compared with conventional treatment which gave the lowest value. Moreover, Highest L. Ascorbic acid content obtained with bio-3 or bio-3 and bio-4 in the first and second seasons respectively compared with bio-1 and conventional treatments which produced the lowest fruit L. Ascorbic acid content in both seasons. All interactions effect on the tree characters were not significant in both seasons.

Table: (6) Effect of soil solarization and bio-fertilization on Fruit firmness, T.S.S content and L. Ascorbic acid (mg/100g F.W.) in 2017 and 2018 seasons.

Charact ers	Fruit firmness (g/cm ²)		T.S.S content		L. Ascorbic acid (mg/100g F.W.)	
	1 st Season	2 nd Season	1 st Season	2 nd Season	1 st Season	2 nd Season
Solarization						
Solarize	٤٤٣,٢	٤٥٢,٢	٨,٣٨	٨,٤٧	٦٤,٨٨	٦٣,٨٩
Non-solarize	٤٢٦,٨	٤٢٤,٦	٨,٢٣	٨,١٣	٦٣,٥١	٦٥,٤٣
LSD at 0.05	١٤,٩	١١,٦	N.S	N.S	N.S	N.S
Bio-fertilization						
Bio-1	445.7	447.9	8.37	8.37	61.50	62.60
Bio-2	451.3	468.5	8.46	8.52	63.18	63.37
Bio-3	466.4	462.6	8.61	8.55	69.12	68.23
Bio-4	424.1	406.6	8.18	8.40	64.83	66.95
Bio-5	379.9	406.2	7.93	7.94	62.37	62.15
LSD at 0.05	١٣,٣	٩,١	٠,١٣	٠,١١	١,٩٨	٢,٦٤
Interaction						
Solarize	450.9	469.2	8.41	8.47	61.86	60.67
	455.7	473.7	8.55	8.56	63.32	63.18
	472.4	476.8	8.77	8.71	68.75	68.36
	442.3	420.9	8.23	8.52	66.08	66.66
	379.5	420.2	7.97	8.12	64.41	60.58
Non-solarize	440.4	426.7	8.33	8.27	61.15	64.53
	446.9	463.3	8.36	8.49	63.04	63.56
	460.4	448.3	8.45	8.39	69.48	68.10
	405.8	392.2	8.12	8.29	63.58	67.24
	380.3	392.3	7.89	7.76	60.32	63.72
LSD at 0.05	N.S	N.S	N.S	N.S	N.S	N.S

Fruit rot and absent yield: Data in Table, 7 showed that, soil solarization significantly decreased rotted fruits even gray mold rot caused by *Botrytis cinerea* or dry rot caused by *Phytophthora cactorum* compared with non-solarize treatments in both seasons. This resulted in more lost fruit yield especially in the first season. Also, rotted fruits significantly affected by bio fertilizer treatments, where the rotted fruits increased with increasing chemical fertilizers ratio and bio 1 treatment (without chemical fertilizer) produced the lowest value compared with other treatments. On the other hand, control treatment (traditional) gave the highest number of rotted fruits in both seasons. Regarding lost yield per plot, data showed the same trend, where, control treatment produced the highest value compared with bio-fertilizers treatments and the lost yield was increased with increasing chemical fertilizer ratio in both seasons.

Decreasing of disease incidence due to solarization may be attributed to the production of NH₃ and an increase in soil microbial activity, which can help control soilborne pathogens through competition, antibiosis, parasitism/predation, etc. (Nunez-Zofio *et al.* 2011; Martinez *et al.* 2011). Most pathogens affected was *Verticillium dahliae* (Daugovish *et al.* 2011), *P. cactorum* (Porras, *et al.*, 2007). Microbiological changes in soil environment have also been documented as a mechanism of pathogen suppression and resulting improved crop productivity (Mazzola, 2011).

Table: (7) Effect of soil solarization and bio-fertilization on Total rotted fruit/ plot, Botrytis gray mold rot, Dry rot and Absent yield in 2017 and 2018 seasons.

Characters	Total rotted fruit/ plot		Botrytis gray mold rot		Dry rot		Absent yield	
	1 St Season	2 nd Season	1 St Season	2 nd Season	1 St Season	2 nd Season	1 St Season	2 nd Season
Solarization								
Solarize	٢٤,٣٣	٢٩,٢٦	١٩,٥٨	٢٣,٤٩	٤,٧٤	٥,٧٧	٣٠٢,٦	419.7
Non-solarize	٩٣,٢٦	١٠٣,٣٢	٨٦,٠٢	٩٥,٦٩	٧,٢٤	٧,٦٣	٢٠٩٨,١	١٢٥٨,٢
LSD at 0.05	٧,٥٠	٢,٩٤	٧,٢٩	٣,٥٧	٠,٩٤	١,١١	١٣٢,١	٤٢,٠
Bio-fertilization								
Bio-1	29.22	36.11	29.68	27.05	8.35	9.06	273.9	348.6
Bio-2	37.55	42.95	48.61	36.15	6.25	6.81	438.4	506.6
Bio-3	57.22	61.04	86.54	55.73	4.18	5.32	618.2	802.6
Bio-4	76.56	88.18	123.19	81.89	5.23	6.29	892.9	1072.4
Bio-5	93.44	103.17	142.09	97.14	5.96	6.03	1309.4	1464.5
LSD at 0.05	٣,٩٢	٥,١٩	٣,٥٢	٥,٠٣	١,٠٩	١,٢٢	72.6	٧٣,٢
Interaction								
Solarize	18.76	22.42	12.05	15.38	6.71	7.04	196.7	228.21
	19.45	23.57	13.98	18.36	5.47	5.21	226.1	281.97
	22.43	25.76	19.53	22.24	2.90	3.53	259.7	370.35
	23.44	31.36	19.48	25.64	3.96	5.72	309.6	459.02
	37.55	43.18	32.87	35.84	4.68	7.34	520.7	759.03
Non-solarize	39.67	49.79	29.68	38.71	9.99	11.08	351.2	469.04
	55.65	62.33	48.61	53.94	7.04	8.40	650.7	731.32
	92.00	96.32	86.54	89.22	5.46	7.10	976.8	1234.89
	129.68	145.01	123.19	138.14	6.49	6.87	1476.2	1685.77
	149.32	163.16	142.09	158.44	7.23	4.72	2098.1	2169.96
LSD at 0.05	٥,٥٤	٧,٣٤	٤,٩٨	٧,١١	N.S	١,٧٢	102.6	١٠٣,٥

Conclusion: the present results indicate that soil solarization has the potential for nonchemical management of soilborne diseases of strawberry and it may be possible to grow strawberries at Siwa oasis with soil solarization and without chemical fertilizers or with limited

amounts of fertilizers. Although the highest yield was obtained by soil solarization with adding recommended chemical fertilizers, we can achieve a proper strawberry yield with application of bio-fertilizers combine with half or one-fourth of recommended doses. Moreover, soil solarization with bio-fertilizers produced the highest fruits quality as well as lowest rotted fruits ratio. Finally, solarization has potential as a component in an integrated pest management program of fruit rot diseases in strawberry production, particularly at areas have hot summer like Siwa oasis.

REFERENCES

AOAC (Association of Official Analytical Chemists-International), (2005). Official Methods of Analysis. 18th edn., eds.: W. Hortwitz, G. W. Latimer, AOAC-Int. Suite 500, 481 North Frederick Avenue, Gaithersburg, Maryland, USA.

Abd-Elgawad, M. M. M. (2019). Plant-parasitic nematodes of strawberry in Egypt: a review. Bulletin of the National Research Centre 43:7 <https://doi.org/10.1186/s42269-019-0049-2>.

Abu-Garbieh, W. (1998). Pre and post-plant soil solarization. FAO Plant Production and Protection Paper, 147: 15-34.

Benlioglu, S. ; A. Yildiz ; O. Boz and K. Benlioglu (2014) Soil disinfestation options in Aydin province, Turkey, strawberry cultivation. *hytoparasitica*, 42:397–403.

Becici, M.;S. Toker; Y.Canihos; A. Erkilic; R. Albajes and E. Sekeroglu (2000). Integrated disease management in tomato crops grown in high tunnels. Bulletin –OILB-SROP. 23 (1): 15-19. (c.f CAB Abstracts 2000-2001).

Campiglia, E.; O. Temperini; R. Mancinelli; F. Saccardo; P.J.Stoffella; D.J.Cantliffe and G. Damato (2000). Effect of soil solarization on the weed control of vegetable crops and on the cauliflower and fennel production in the open field. *Acta Horticulturae*.533 : 249-255.(c.f. CAB Abstracts 2000-2001).

Camprub, A; V. Esta; M.A. El Bakali; F.Garcia-Figueresand , C. Calvet(2007). Alternative strawberry production using solarization, metham sodium and beneficial soil microbes as plant protection methods *Agron. Sustain. Dev.* 27 : 179–184

Candido,V.; T. D. Addabbo; M. Basile; D. Castronuovo and V. Miccolis (2008) Greenhouse soil solarization: effect on weeds, nematodes and yield of tomato and melon *Agron. Sustain. Dev.* 28 221–230.

Caulet, R. P., A. Morariu; D. Iurea and G. Gradinariu (2013). Growth and photosynthetic characteristics of two

strawberry cultivars in response to furostanol glycosides treatments. *Not. Bot. Horti. Agrobo.* 41(1):231-237.

Daugovich, O, Shennan C, Muramoto J, Koike S (2011). Anaerobic soil disinfestation for southern California strawberry. 2011, Proceedings of Annual International Research Conference on Methyl Bromide Alternatives and Emissions Reductions. San Diego(CA),pp2-1/2-4

De vey, J. E. (1991). Use of soil solarization for control of fungi and bacterial plant including biocontrol. *FAO plant production and protection paper 109:* 79-128.

De vey, J. E. and J. J. Stapleton (1998). Soil solarisation: past, present and future. *FAO Plant Production and Protection Paper 147* 1-5.

Domínguez, P.; L. Miranda and C. Soria, (2014). Soil biosolarization for sustainable strawberry production. *Agron. Sustain. Dev.* 34:821–829.

El-Sheshtawy, A. A. (2006). Weed control in faba bean crop using soil solarization. M. Sc. Thesis, Faculty of Agriculture, Al-Azhar Univ. Cairo, Egypt. Pp 65-72.

Embaby, E.M. (2007) Pestalotia fruit rot on strawberry plants in Egypt. *Egypt. J. Phytopathol.*, 35 (2): 99-110.

FAO, FAOSTAT/Statistical database (2017).<http://faostat.fao.org/site/535/ desk top Default. aspx? Page ID=535#ancor>.

Gamliel, A.; M. Austeweil; G. Krritzman; J. Katan; N.Aharonson; E. Cohen; B. Rubien and G. A. Matthews (2000). Non-chemical approach to soilborne pest management organic amendments. XIVth international plant protection congress, Jerusalem, Israel, 1999. *Crop Protection* 19 (8-10): 847-853.

Gomaa, S. S., (2008). Effect of organic and bio-fertilization and soil solarization on potato production under north Sinai

conditions. Ph.D. Thesis, Faculty of Agriculture, Ain Shams Univ., Cairo, Egypt. PP. 40-45.

Gomaa, S. S.; R. H. A. Ghodia and M. K. M. Agha (2016). Evaluation and improvement of some strawberry cultivars (*fragaria ananassa* duch.) at siwa oasis. J. Biol. Chem. Environ. Sci., 7(3): 287-302.

Hamada, M. M. (2002). Use of solarization and organic fertilizers for reducing pollution in Table beet under Ras Sudr conditions. M. Sc. Thesis, Faculty of Agriculture, Ain Shams Univ. Cairo, Egypt. pp 75-78.

Hartz, T.K.; J.E. DeVay, and C.L. Elmore (1993). Solarization is an Effective Soil Disinfestation Technique for Strawberry Production. Hortscience 28(2):104-106.

Himelrick, D.G. and W.A. Dozier. (1991). Soil fumigation and soil solarization in strawberry production. Adv. Strawberry Production 10: 12-28.

Holt, J. G.; N. R. Crrieg; P.H. A. Sneath; G. T. S. Taley and S.T. Williams (1994). Bergey is Manual of Determinative Bacteriology. Williams and Wilkins, Baltimore, USA.

Johnson, L. E.; A. A. Crul; E. A. Bond and H. A. Fribourg (1995). Methods of studing soil microflora plant disease relationship. Minneapolis publishing co. USA: pp 1-178.

Katan, J. (1981). Solar heating (solarization) of soil for control of soil-borne pests. Annu. Rev. Phytopathol. 19:211-236.

Katan, J. and J.E. DeVay (1991). Soil solarization. CRC Press, Boca Raton, Fla.

Katan, J (1998). Adaptation and implementation of soil solarization: achievements and difficulties. International workshop on management of soilborne pathogens March 1-5, 1998.

Khafagi, Y.S. 1982. Studies on fruit rot diseases of strawberry in ARE. M.Sc.Thesis, Fac. of Agric., Moshtohor, Zagazig Univ., Benha Sector. 134 pp.

Page, A.L., R.H. Miller and D.R. Keeney, 1982. Methods of soil analysis-chemical and microbiology properties, SSSA Inc., Mad., WI., USA.

Martine, J. P. (1950). Use of acid, Rose Bengal and Streptomycin in the plate method for estimating soil fungi. Soil Sci.: pp 69-215.

Martínez M. A, Martínez, M. C, Bielza P, Tello J, Lacasa A. (2011). Effect of biofumigation with manure amendments and repeated biosolarization on *Fusarium* densities in pepper crops. Microbiol Biotechnol 38:3–11.

Mazzola, M (2011). Potential of biofumigation of soilborne pest control in strawberry. 2011 Proceedings of Annual International Research Conference on Methyl Bromide Alternatives and Emissions Reductions. San Diego (CA), pp 47-1/47-2

Nash, S. M. and W.C. Synder (1962). Quantitative estimation by plate counts of propagules of bean root rot *Fusarium* in field soils. Phytopathology. 52 : 567-570.

Nunez-Zofio M, Larregla S, Garbisu C (2011) Application of organic amendments followed by soil plastic mulching reduces the incidence of *Phytophthora capsici* in pepper crops under temperate climate. Crop Prot 30:1563–1572. doi:10.1016/j.cropro.2011.08.020.

Ozyilmaz, U. ; K. Benlioglu ; A. Yildiz and H. S. Benlioglu (2016). Effects of soil amendments combined with solarization on the soil microbial community in strawberry cultivation using quantitative real-time PCR. Phytoparasitica, 44:661–680

Porrás, M., Barrau, C., Arroyo, F. T., Santos, B., Blanco, C., and Romero, F. 2007. Reduction of *Phytophthora cactorum* in strawberry fields by *Trichoderma* spp. and soil solarization. Plant Dis. 91:142-146.

Peachey, R.E.; J.N. Pinkerton; K.L. Ivors; M.L. Miller and L.W. Moore (2001). Effect of soil solarization, cover crops and metham on field emergence and survival of buried annual bluegrass (*Poa annua*) seeds. *Weed Technology*, 15 (1): 81-88. (c.f. CAB Abstracts 2000-2001).

Piper, C.S., 1950. Soil and plant analysis.1st Ed. Interscience Publishers Inc., New York, USA, pp 30-59.

Pullman, G.S., J.E. DeVay, R.H. Garber, and A.R. Weinhold (1981). Soil solarization: Effects on verticillium wilt of cotton and soil borne populations of *Verticillium dahliae*, *Pythium* spp., *Rhizoctonia solani* and *Thielaviopsis basicola*. *Phytopathology* 71:954-959.

Russell, D. F., (1991). In “MSTATC, Directory crop soil science Department” Michigan University. USA.

Reger, M.; G. Krewer and P. Lewis (2001). Solarization and chemical alternatives to methyl bromide for pre plant soil treatment of strawberries. *HortTechnology*. 11(2):258-264 (c.f. CAB Abstracts 2000-2001).

Shukla, L.; D. K. Singh; N.T. Yaduraju; T.K. Das and S.P. Magu (2000).Effect of soil solarization on microflora and soil enzymatic activity. *Annals of plant protection sciences*. 8 (2): 2018-222 (c.f. CAB Abstracts 2000-2001).

Stapleton, J.J. (1991). Soil solarization in tropical agriculture for pre- and post-plant applications. *FAO Plant Production and Protection Paper 109*: 220-228.

Stapleton, J.J. and J.E. DeVay (1998). Soil solarization: A non-chemical approach for management of plant pathogens and pests. *Crop Protection* 5:190-198.

Tadrous, A.Z. 1991. Pathological studies on strawberry fruit rots. M.Sc. Thesis, Fac. Agric., Suez Canal Univ. 116pp.

Tarek, S.S.S. 2004. Integrated control for minimizing postharvest diseases of strawberry. MSc. Thesis, Fac. of Agric., Ain Shams Univ. 165pp.

Talyour, J. (1962). The estimation of numbers of bacteria by ten fold dilution series. J. Appl. Bact. 25: 54-56.

Thomas, R.L., R.W. Sheard and J.R. Moyer, 1967. Comparison of conventional and automated procedures for nitrogen, phosphorus and potassium analysis of plant materials using a single digestion. Agron. J., 59: 240-243.

Triki, M.A.; S.Priou and M. El-Mahjoub (2001). Effect of soil solarization on soil borne population of *Pythium aphanidermatum* and *Fusarium solani* and on the potato crop in Tunisia. Potato-Research. 44(3): 271-279.(c.f. CAP Abest. 2002).

Vasil'eva I. S., S. A. Vanyushkin, S. V. Zinov'eva, Z. V. Udalova, Y. V. Bolychevtseva and V. A. Paseshnichenko, (2003). Photosynthetic pigments of tomato plants under conditions of biotic stress and effects of furostanol glycosides. App. Biochem Microboil. 39(6):606-612.