Polymorphisms in the exon-3 of leptin gene using tetra-primer ARMS-PCR in Egyptian buffalo

F. A. Sharara 1,*, M. A. Aboul-Hassan 1, W. M. Shakweer 2

- ¹ Animal Production Department, Faculty of Agriculture, Al-Azhar University, Cairo, Egypt
- ² Animal Production Department, Agricultural and Biological Research Institute, National Research Centre, 33 El-Buhouth Street, P.O: 12622, Dokki, Cairo, Egypt
- * Corresponding author E-mail: fahmysharara@azhar.edu.eg (F. Sharara)

ABSTRACT

The Egyptian buffalo is one of the Egyptian genetic resources which is irreplaceable and must be preserved, because of its great adaptability to local harsh environmental conditions and resistance to infectious diseases. A total number of 74 blood samples of female Egyptian buffalo were used to assess the allelic and genotypic frequency of G>A SNP at codon 159 of leptin gene. The tetra-primer amplification refractory mutation system-polymerase chain reaction (Tetra-primer ARMS-PCR), a single nucleotide polymorphism (SNP) technique allows for the simultaneous detection of alleles using a single PCR reaction and gel electrophoresis. Results revealed three genotypes GG, AG, and AA with frequencies of 0.49, 0.39, and 0.12, respectively. Allele frequencies of G and A were found to be 0.69 and 0.31, respectively. For the first time, allelic and genotypic frequency of G>A SNP at codon 159 in Egyptian buffalo was detected with the highest allelic frequency of allele A (0.31). Further studies are needed to provide more information about the association between this SNP and production and reproduction traits.

Key words: Egyptian buffalo, Tetra-primer ARMS-PCR, Leptin hormone.

INTRODUCTION

In Egypt, the Egyptian buffalo (Bubalus bubalis) is very significant because it is exceptional at adapting to challenging environmental conditions, resistant infectious disease, and excellent at providing nutritional benefits. As a result of its flavor and composition, Egyptian consumers prefer buffalo milk (Abu El-Magd et al., 2015). According to the Egyptian census of livestock, Egypt owns 3.8 million heads of buffalo, which represents about 37% of the total livestock, and buffalo milk production represents 70% of milk production in Egypt (Statistics of Ministry of Agriculture, 2018).

The recent advances in molecular genetic techniques during the past decades allow the scientists to achieve genetic studies at the molecular level. One of the applications of molecular genetic techniques is documenting and describing the genetic variation of DNA sequences between and within populations. A nucleotide polymorphism single represents a single base variation in a DNA sequence at any given position, with an appearance rate of more than 1% in a given population. Compared with other genetic variations, SNPs spread across genomes with a very high frequency (Borodina et al., 2004). Tetra-primer ARMS-PCR (Tetra-primer ARMS), a simple and inexpensive SNP genotyping technique, allows for the

simultaneous detection of alleles using a single PCR reaction and gel electrophoresis. The technique only requires a set of four primers and does not require any expensive infrastructure or reagents (Ye *et al.*, 2001).

Leptin's primary transcript is 167 amino acids long, subsequently, the first 21 amino acid signal sequence is cleaved (Zhang et al., 1994). Leptin is primarily produced in adipose tissue and secreted into the bloodstream (Deshpande et al., 2014). Its expression also occurs in other cells and tissues, including the placenta (Hoggard et al.,1997), mammary glands (Smith-Kirwin et al., 1998), skeletal muscles (Wang et al.,1998), and pituitary glands (Morash et al., 1999), in which LEP acts as autocrine or paracrine. There are very different functions of the LEP hormone, including modulation of reproduction (Yu et al., 1997), metabolism (Agarwal et al., 2009), mammary gland development, as well as cell differentiation and proliferation (Onneta et al., 2002). The leptin gene has been mapped to chromosome 8 (BBU 8q32) in buffaloes (Vallinoto et al., 2004), and consists of three exons, of which the first exon is not translated into protein.

In 2007, Orrù *et al.* sequenced the leptin coding region (exon2 and exon3) on a panel of 32 Italian River Buffalo and two Egyptian River Buffalo. Twelve new SNPs were detected. One of the new SNPs, G>A in exon 3, causes a change in the second position of

codon 159, resulting in arginine (R) to glutamine (Q) substitution of the primary transcript (138 in mature peptide). However, this polymorphism was found in the heterozygous state in only one sample of Italian buffalo, whereas the other animals showed a GG homozygote genotype (Orrù et al., 2007). The existence of an A allele was confirmed by DNA sequencing of 390 Murrah and Philippine Carabao buffaloes, with an allele frequency of 0.18 (Seong and Kong, 2012). Also, Kale et al., 2013 detected the same variation by SSCP in Murrah, Surti and Bhadawari, with a high allele frequency of 0.3. There is no study about the genotypic frequency of the A > G SNP at codon 159 in Egyptian buffalo. Therefore, the aim of the present studies was to assess the allelic and genotypic frequency of SNP A > G at codon 159 in Egyptian buffalo by Tetra-primer ARMS-PCR.

MATERIALS AND METHODS

DNA samples

A total of 74 blood samples were collected from Egyptian buffalo. The samples were randomly taken from the Agricultural Experiments Station (AES) which belongs to the Faculty of Agriculture, Cairo University, Giza, Egypt. Approximately 5 ml of blood per animal was obtained by jugular venipuncture in a K3-EDTA tube containing anticoagulant. The genomic DNA was extracted from whole blood using a commercial kit according to the manufacturer's protocol for ISOLATE II Genomic DNA Kit, Bioline, Cat No. BIO-52066.

Genotyping

The Tetra-primer ARMS-PCR and primers for G>A SNPs at codon 159 were designed by available online the freely program http://primer1.soton.ac.uk/primer1.html The list primer sequences of annealing temperatures, and the sizes expected of PCR products are given in Table 1. In a final volume of 20µl, a PCR reaction containing 1µl genomic DNA, 2µl each primer, 0.5µl MgCl₂, 10 µl GoTaq® Green Master Mix (Promega, Madison, USA), and nuclease-free water up to 20µl were performed. The PCR program included a 5-minute denaturing step at 95°C, 35 cycles of [Denaturation 95°C for 30 Sec, Annealing 64°C for 30 Sec, Extension 72°C for 1-minute] and a final 7-minute extension step at 72 °C.

Statistical analysis

Allelic and genotypic frequencies were calculated by simple counting. Hardy-Weinberg equilibrium was tested by gene-calc web http://gene-calc.pl/hardy-weinberg-page.

RESULTS AND DISCUSSION

The tetra-primers were designed for the detection of the G>A point mutation at codon 159 of the leptin gene in Egyptian buffalo. Three genotypes are shown in Fig. 1 for GG, AG, and AA with frequencies of 0.49, 0.39, and 0.12, respectively. Allele frequencies of G and A were found to be 0.69 and 0.31, respectively. The accordance of the mentioned SNPs with HWE was authorized by the gene-Calc web at 0.05 of the level of significance.

The allelic and genotypic frequency of the amino acid substitution arginine (R) to glutamine (Q) at position 159 was detected in buffaloes for the first time by Orrù et al., (2007) with an allelic frequency of 0.01, whereas the heterozygote GA genotype was not detected. The higher frequency (0.18) was detected by Seong and Kong (2012) in Murrah and Philippine Carabao buffaloes, with a low frequency of homozygote genotype AA (0.03). The highest allelic frequency of allele A (0.3) was detected by Kale et al., (2013), and the same results were reported in the present Single-Strand Conformation study. The Polymorphism (SSCP) for leptin exon 3 of Egyptian buffalo failed to detect the G > A point mutation at codon 159 (Ghoneim et al., 2016), whereas the tetra-primer (ARMS-PCR) in our study was declared for fast and efficient detection of the G > A SNP at codon 159 in the leptin gene.

The R159Q substitution is located at position 138 in the C-terminal region of the mature peptide, within helix D (Reicher *et al.*, 2011). Arginine is a polar, positively charged (basic amino acid) and has an uncharged R group, while glutamine is polar, uncharged and has an amide R group. The expected effect of the R159Q SNPs on protein functions was studied by Mahrous *et al.*, (2020). The R159Q lowered the stability of the mature leptin peptide tertiary structure by 0.25 kcal/mol and classified this mutation as a neutral mutation with a total PredictSNP expected accuracy of 83%.

CONCLUSION

In this study, for the first time, allelic genotypic frequencies of G>A SNP at codon 159 in Egyptian buffalo were detected, with

the highest allelic frequency of allele A (0.31). Further studies are needed to provide more information about the association between this SNP, production and reproduction traits.

ACKNOWLEDGMENTS

The authors would like to thank the Faculty of Agriculture, Cairo University, Giza, Egypt for kindly providing blood samples and animal records.

REFERENCES

- El-Magd, M.A., Zaki, A.G., Kahilo, K.A., Barakat, M.E.S., Hassan, I.F. 2014: Effect of SNPs in prolactin promoter on milk traits in egyptian buffalo. Advances in Dairy Research, 3:1-4.
- Agarwal, R., Rout P.K., Singh, S.K. 2009: Leptin: A biomolecule for enhancing livestock productivity. 8:169–176.
- Borodina, T.A., Lehrach, H., Soldatov, A.V. 2004: Ligation detection reaction-TaqMan procedure for single nucleotide polymorphism detection on genomic DNA. Analytical biochemistry, 333(2): 309-319.
- Deshpande, M, Rank, D.N., Vataliya, P.H., Joshi, C.G. 2014: Study of leptin gene polymorphism in mehsana buffaloes (*Bubalus bubalis*). Buffalo Bull 33:115–119.
- Ghoneim, M.A., Ogaly, H.A., Gouda, E.M., El-Behairy, A.M. 2016: Prediction of desirable genotype patterns in Baladi beef cattle and water buffalo by identification of new leptin gene SNPs. Livestock Science, 194:51-56.
- Hoggard, N., Hunter, L., Duncan, J.S., Williams, L.M., Trayhurn, P., Mercer, J.G. 1997: Leptin and leptin receptor mRNA and protein expression in the murine fetus and placenta. Proceedings of the National Academy of Sciences, 94(20):11073-11078.
- Kale, D.S., Yadav, B.R., Mukherjee, A., Prasad, J. 2013: Exploring DNA polymorphisms of leptin gene within Indian water buffaloes. Journal of Advanced Veterinary Research, 3 (1):20-26.
- Mahrous, K.F., Aboelenin, M.M., Rashed, M.A., Sallam, M.A., Rushdi, H.E. 2020: Detection of polymorphism within leptin gene in Egyptian river buffalo and predict its effects on different molecular levels. Journal of Genetic Engineering and Biotechnology, 18(1):1-11.

- Morash, B., Li, A., Murphy, P.R., Wilkinson, M., Ur, E. 1999: Leptin gene expression in the brain and pituitary gland. Endocrinology, 140(12):5995-5998.
- Bonnet, M., Delavaud, C., Laud, K., Gourdou, I., Leroux, C., Djiane, J., Chilliard, Y. 2002: Mammary leptin synthesis, milk leptin and their putative physiological roles. Reproduction Nutrition Development, 42(5):399-413.
- Orrù, L., Terzano, G.M., Napolitano, F., Savarese, M.C., De Matteis, G., Scatä, M.C., Moioli, B. 2007: DNA polymorphisms in river buffalo leptin gene. Italian Journal of Animal Science, 6 (sup2), 342-344.
- Reicher, S., Gertler, A., Seroussi, E., Shpilman, M., Gootwine, E. 2011: Biochemical, in vitro biological significance of natural sequence variation in the ovine leptin gene. Gen Comp Endocrinol 173:63–71.
- Seong, J., Kong, H.S. 2012: Polymorphisms of LEP, LGB and PRLR in water buffalo. CNU J Agric Sci 39:577–581.
- Smith-Kirwin, S.M., O'Connor, D.M., Johnston, J., Lancey, E.D., Hassink, S.G., Funanage, V.L. 1998: Leptin expression in human mammary epithelial cells and breast milk. J Clin Endocrinol Metab 83:1810–1813.
- Vallinoto, M., Schneider, M.P.C., Silva, A., Iannuzzi, L., Brenig, B. 2004: Molecular cloning and analysis of the swamp and river buffalo leptin gene. Animal genetics, 35(6):462-463.
- Wang, J., Liu, R., Hawkins, M., Barzilai, N., Rossetti, L. 1998: A nutrient-sensing pathway regulates leptin gene expression in muscle and fat. Nature, 393(6686): 684-688
- Ye, S., Dhillon, S., Ke, X., Collins, A.R., Day, I.N. 2001: An efficient procedure for genotyping single nucleotide polymorphisms. Nucleic acids research, 29 (17), e88-e88.
- Yu, W.H., Kimura, M., Walczewska, A., Karanth, S., McCann, S.M. 1997: Role of leptin in hypothalamic–pituitary function. Proceedings of the National Academy of Sciences, 94(3): 1023-1028.
- Zhang, Y., Proenca, R., Maffei, M., Barone, M., Leopold, L., Friedman, J.M. 1994: Positional cloning of the mouse obese gene and its human homologue. Nature, 372(6505): 425-432.

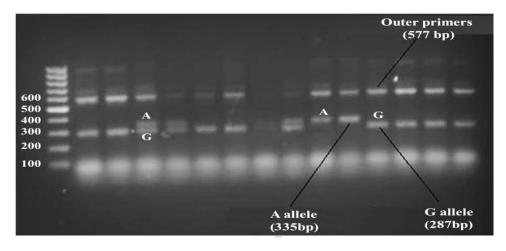


Figure 1: The amplified products by Tetra-Primer ARMS-PCR on agarose gel

Table 1: Nucleotide sequences of primers, Annealing temperature, and the sizes expected of PCR products.

Primer	Primer sequence	Annealing temperature
LEP inner-F (G allele)	GGGTCACTACAGGACATGTT <u>T</u> C G	64 C
LEP inner-R (A allele)	CCAGGACTGAGGTCCAGCT <u>T</u> CT	
LEP outer-F	CAGAGGGTCACTGGTTTGGACTT	
LEP outer-R	CTCCGGTCTACTGTTTGCTGGAT	
Product size for G allele: 287		
Product size for A allele: 335		
Product size of two outer primers: 577		

Allele specificity is conferred by A second mismatches T instead of G and T instead of C (demonstrated by underlined letter) are introduced at the third position from the 3' end of each of the Forward and Reverse inner primers.

تعدد الأشكال في إكسون ٣ لجين اللبتين باستخدام Tetra-primer ARMS-PCR في الجاموس المصري

فهمي أحمد شرارة ^{۱۰}، محمد أبو الحسن أحمد ١، وليد محمد شقوير ٢

ا قسم الانِتاج الحيواني، كلية الزراعة بالقاهرة، جامعة الأزهر، مصر. القسم الانِتاج الحيواني، المركز القومي للبحوث، الدقي ، الجيزة

البريد الاليكتروني للباحث الرئيسي:fahmysharara@azhar.edu.eg

الملخص العربي

يحظى الجاموس المصري بأهمية كبيرة في مصر، لما له من قدرة كبيرة على التكيف مع الظروف البيئية القاسية، ومقاومة الأمراض المعدية، كما يفضل المستهلكون في مصر لبن الجاموس لما له من مذاق مميز ومكونات جيدة. لهرمون اللبتين وظائف كثيرة ومتعددة، بما في ذلك التأثير على التكاثر، والخثيل الغذائي، وتطور الغدة اللبنية، وكذلك تمايز الحلايا وتكاثرها ويتم إنتاج هرمون اللبتين بصورة أساسية في الأنسجة الدهنية ويتم التعبير عنه أيضًا في بعض الأنسجة الأخرى، بما في ذلك المشيمة والغدد اللبنية والعضلات الهيكلية والغدد النخامية. (Tetra-primer ARMS-PCR (Tetra-primer ARMS) PCR واحد هي تقنية بسيطة وغير مكلفة لدراسة الاختلافات الوراثية، حيث تساعد هذه الطريقة في الكشف المتزامن للأليلات باستخدام تفاعل PCR واحد وتفريد كهربي للجيل. لذلك، كان الهدف من هذه الدراسة هو تقدير تكرار الأليل وتكرار التركيب الوراثي لـ G>A SNP في الكودون رقم 109 و AG و وكرار التركيب الوراثية طرز وراثية هي GG و A و وكرار التركيب الوراثية على الترتيب. لأول مرة، تم تقدير تكرار الأليل وتكرار التركيب الوراثية للاختلاف الوراثي هي G>A للكودون 10 في الجاموس المصري ، مع أعلى تكرار أليلي للأليل (0.31) A هناك حاجة إلى مزيد وتراد التركيب الوراثية للاختلاف الوراثي هذا الاختلاف الوراثي وصفات الإنتاج والتكاثر.

الكلهات الاسترشادية: الاختلافات الوراثية، هرمون اللبتين، الجاموس المصرى.