EFFECT OF CERTAIN PESTICIDES ON SOME ENZYMES OF CHICK EMBRYOS

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ABSTRACT

Effects of Chlorpyrifos 48 % EC (Dursban & Pestban) and Profenofos 72 % EC (Selecron & Ictacron) at two sublethal concentrations (1/100th and 1/10th of the field recommended rate) on some serum , brain and liver enzymes as well as total protein from 18 and 21 days old - chick embryos (hatching) were determined. In general, Dursban and Pestban at the two concentrations significantly decreased brain acetylcholinesterase (AChE) either at 18 or 21 days old chick embryos, except Dursban in 18 days at the low concentration. On the other hand, Selecron and Ictacron at the two concentrations caused insignificant inhibition in brain AChE activity after 18 and 21 days compared with the control groups.

Serum alanine amino transferase (ALT) specific activity was significantly increased with Chlorpyrifos and Profenofos at the used concentrations. On the contrary, Chlorpyrifos from the two formulations at the high concentration significantly inhibited the serum aspartate amino transferase (AST). There were no differences between the two formulations of Profenofos on the serum amino transferases (ALT & AST).

The results also revealed that the activity of liver acid phosphatase (AcP) in 18 days old chick embryos was insignificantly inhibited at the two concentrations of Chlorpyrifos (Dursban & Pestban). There were no significant differences between Selectron and Ictacron on the liver enzyme activity at 18 days. Chlorpyrifos and Profenofos were caused significant inhibition of 21 days old chick embryo liver (AcP) activities from the two formulations, except Selectron at the high concentration.

The activity of liver alkaline phosphatase (AIP) in the 18 days old - chick embryos was decreased insignificantly at the two concentrations of Dursban, in contrast, the activity was increased by increasing the concentration of Pestban, Selecton and Ictacron. On 21 days old - chick embryos, Chlorpyrifos (Dursban & Pestban) caused significant decrease of liver AIP activity at the two concentrations. Selecton at the two concentrations had insignificant effect on serum AIP activity, but Ictacron caused significant activation of serum AIP activity at the high concentration.

There were insignificant changes in the serum total protein between the treated groups with Dursban, Pestban, Selecton, Ictacron and control groups.

INTRODUCTION

The intensive use of organic pesticides on a wide variety of crops, fruit orchards and vegetable crops for controlling pests leads to environmental contamination and killing wild life (Flint and Bosch, 1981).

The earliest observations on the effects of external applications of pesticides to bird eggs by spraying DDT revealed few apparent toxic effects (Mitchell, 1946 and Somers et al., 1974). Paraquat proved to be highly toxic following its application to eggs of chickens and Japanese quail (Lutz-Ostertag and Henou 1975) or mallards (Hoffman and Eastin 1982). Some organophosphorus pesticides are known to cause delayed neurotoxicity in chickens (Abo-Donia and Graham 1978; EL-Sebae et al., 1980). Mallard

(Anas platyrhynchos) eggs are intensive to external exposure of organophosphorus pesticides including parathion, which results in mortality, stunted growth and teratogenicity. These effects are accompanied by inhibition of brain acetylcholinesterase (Hoffman and Eastin 1981; Hoffman and Albers, 1984). Chickens (Gallus gallus) and Japanese quail (Coturnix japonica) are also sensitive to external application of parathion (Meiniel, 1973).

Activity of brain acetylcholinesterase (AChE), plasma cholinesterase, alkaline phosphatase were inhibited, while, plasma aspartate amino transferase was increased at one or more stages of mallard development treated with the organophosphorus insecticide; phenyl phosphonothioc acid – O - ethyl – O (- 4 – nitrophenyl) ester (EPN) (Hoffman and Sileo, 1984). The *in vivo* results of brain ChE activities revealed that Profenofos was not a potent ChE inhibitor, while sulprofos was a relatively stronger ChE inhibitor insecticide at the 1/4 LD⁵⁰ levels (Enan, 1979). Dimethoate and Chlorpyrifos exposed mallard (*Anas platyrhynchos*) ducklings exhibited activity depression of brain acetylcholinesterase (AChE) activity (Martin and Forsyth 1998). Also, Chlorpyrifos, parathion, acephate and trichlorfon inhibited brain AChE of chick embryos (Lesser *et al.*, 2000).

The purpose of the present study is to investigate the potential toxic effects of Chlorpyrifos and Profenofos on some serum, brain and liver enzymes as well as the serum total protein of chick embryos by applying different formulations (exported or locally) at different concentrations to the incubating eggs.

MATERIALS AND METHODS

Chemicals

The organophosphorus, insecticides used were;

I. Two formulations of Chlorpyrifos 48 % EC { (O, O diethyl - O - (3,5,6 - trichloro - 2- pyridyl) phosphorothioate } from two chemical companies:

Dursban was obtained from Dow Agro Sciences (England).

- Pestban was obtained from Agrochem Company (Egypt).
- II. Two formulations of Profenofos 72 % EC { O (4 bromo 2 chlorophenyl) O ethyl S propyl phosphorothicate) } from two chemical companies:

Selection was obtained from Sengenta

 Ictacron was obtained from International Company of Chemicals and Trade Agencies (ICCTA), (Egypt).

All other chemicals were of highest purity grade commercially available.

Tested chicken eggs:

Fertile eggs of Alexandria Strain chicken (*Gallus gallus*) obtained from the Experimental Station of Faculty of Agriculture , Alexandria University, Alexandria, Egypt. Eggs, weighting 52.2 ± 1.6 gm were placed in the incubator maintained at 37.5 ° C and 65 - 75 % relative humidity and 3-5 times rotation every day.

Eggs treatment:

On the fourth day of incubation all eggs were candled before treatment, and infertile ones and those with dead embryos were discarded. Eggs were randomly divided into ten groups (50 eggs per each group) and treated as follows:

1-Untreated control

1) Treated control with water

2- Two groups treated with Dursban 48 % EC (1/100th and 1/10th of the field recommended rate)

3- Two groups treated with Pestban 48 % EC (1/100th and 1/10th of the field recommended rate)

4- Two groups treated with Selection 72 % EC (1/100th and 1/10th of the field recommended rate)

5- Two groups treated with Ictacron 72 % EC (1/100th and 1/10th of the field recommended rate)

Eggs were treated by immersing the eggs in aqueous emulsion of pesticides for 15 sec according to the method of Meiniel, 1973. Eggs were allowed to dry for 5 min on racks and were then returned to the incubator.

Biochemical studies:

Blood samples were obtained from hatching embryos of each group by decapitation. Brain and liver of 18 and 21 days old embryos (hatching) were removed for biochemical parameter measurements. Brain acetylcholinesterase (AChE) activities was determined spectrophotometrically using the method of Ellman *et al.*, (1961), serum alanine amino transferase (ALT) and aspartate amino transferase (AST) were determined according to the method of Reitman and Frankel (1957). Also, acid phosphatase activities (AcP) in liver were determined according to Bessey *et al.*, (1946). Serum and liver alkaline phosphatase activities (AIP) were determined by the method of Hausman *et al.*, (1967).

Total protein:

Total serum protein was determined by the method of Weichsebaum, (1946).

Statistical analysis

The data were expressed as mean \pm SD. Data were statistically analyzed using one – way analysis of variance (p < 0.05) according to Dixon and Massay (1957).

RESULTS AND DISCUSSION

Acetylcholinesterase activity (AChE):

Specific activities of brain (AChE) of 18 and 21 days old embryos treated with Chlorpyrifos; Dursban and Pestban and Profenofos; Selectron and lotacron with the two concentrations were summarized in table (1). Chlorpyrifos from the two formulations (Dursban or Pestban) was caused significantly inhibition of brain AChE activities in 18 and 21 days embryo, except Dursban at 18 days with the low concentration. There were

insignificant differences between Dursban and Pestban effects on brain AChE activities of 18 and 21 days old chick embryos. Profenofos (Selecron and Ictacron) at the two used concentrations; 1/100th and 1/10th of the field recommended rate were caused insignificant inhibition of brain AChE activity either in 18 days or 21 days old embryos.

This data was supported by several reports for the effect of organophosphorus pesticides on the AChE activity in birds. Chlorpyrifos caused significant inhibition in the brain ChE from 15 days old chick embryos (Lesser *et al.*, 2000). Helal, 2000 reported that Chlorpyrifos significantly decreased AChE of Japanese quail at 1 / 50 or 1 / 100 of LD₅₀ for 30 days, when compared with the control. The *in vivo* results of brain ChE activities revealed that Profenofos was not a potent ChE inhibitor, while sulprofos was a relatively stronger ChE inhibitor insecticide at the 1/4 LD⁵⁰ levels (Enan, 1979). Esterases in some avian species were also, found to be affected by sublethal doses of organophosphate compounds (EI – Hamady *et al.*, 1996). Also, brain and serum AChE activities were strongly inhibited after the treatment with azamethiphos and methomoyl (Fossi *et al.*, 1992).

In general, Chlorpyrifos is neurotoxic in nature by acting as inhibitor of neuronal cholinesterase activity (Altuntas *et al.*, 2002). In contrast, Profenofos is known as non-ChE inhibitor insecticide. Classes of compounds known to have the common side chain of S-n-propyl OP's are characteristically very low in their potency as ChE inhibitors particularly *in vivo*. The producer of Profenofos state that acute toxicity of the compound to human or mammals cannot be treated or pretreated by known antidotes atropine sulfate or PAM (Enan, 1979).

Table (1): Effect of Chlorpyrifos and Profenofos on brain acetylcholinesterase (AChE) activity in 18 and 21 days chick embryos.

OD / mg protein /	min. Mean ± SD
18 day	21 day
2.56 ± 0.14°	$4.05 \pm 0.65^{\circ}$
2.59 ± 0.16^{c}	$4.03 \pm 0.11^{\circ}$
2.165 ± 0.14 ^{abc}	$1.66 \pm 0.03^{\circ}$
1.94 ± 0.13 ^{ab}	1.12 ±0.12 ^a
1.98 ± 0.20 ^{ab}	1.72 ± 0.15^{a}
1.74 ± 0.14^{a}	1.51 ±0.25 ^a
2.29 ± 0.36 ^{DC}	3.82 ± 0.36 bc
2.18±0.37 ^{oc}	3.77 ± 0.47^{5c}
2.33 ±0.14 ^{bc}	3.76 ± 0.101 bc
2.10 ± 0.38 ^{bc}	3.58 ± 0.24^{5c}
	2.56 ± 0.14^{c} 2.59 ± 0.16^{c} 2.165 ± 0.14^{abc} 1.94 ± 0.13^{ab} 1.98 ± 0.20^{ab} 1.74 ± 0.14^{a} 2.29 ± 0.36^{bc} 2.18 ± 0.37^{bc} 2.33 ± 0.14^{bc}

Significantly different from the control groups by one- way analysis of variance of (p

Each group includes nine replicates.

Transaminases activities:

Table (2) showed the effect of Chlorpyrifos and Profenofos from the two formulations at the two concentrations on the transaminases (ALT & AST) in serum of 21 days old chick embryos. ALT activities were significantly increased with Chlorpyrifos in a concentration dependent manner. Dursban increased ALT activity about 75.6 % and 153.7 % of control at low and high concentration respectively, while Pestban increased the activity about 118.5 % and 226.63 % of control, respectively. Profenofos (Selecron & Ictacron) caused significantly activation in the (ALT) activity, but there were no significant differences between them. In contrast, Chlorpyrifos from the two formulations was significantly decreased the activities of AST at the high concentration. The activity of AST decreased to 48.6 % and 45.75 % of the control at high concentration of Selecton and Ictacron respectively. The present data are also in agreement with Dieter and Wiemeyer (1978) who reported elevation of AST and ALT values in adult birds following an acute dosage of dieldrin, a known hepatotoxin. Also, Chlorpyrifos significantly elevated AST and ALT of Japanese quail when daily treated with sublethal doses (1/50 and 1/100 of LD₅₀) for 30 days (Helal, 2000).

The observed changes in serum AST and ALT activities are consistent with the possibility of cellular damage in liver. Elevated serum activities on intracellular enzymes characteristic of certain organs have indicated cellular leakage associated with probable cellular injury following exposure to pesticides during avian embryogenesis (Baker et al., 1972).

Table (2): Effect of Chlorpyrifos and Profenofos on serum transaminases

(ALT & AST) of 21 day Treatments		Jnits / L ean ± SD
	ALT	AST
Untreated control	127.15 ± 3.04^{a}	134.95 ± .3 ^e
Control treated with water	140.3 ± 6.08^{a}	133.6 ±13.2 ^e
Chlorpyrifos		
1. Dursban at low conc.	215.5 ± 3.54 ^b	126.85 ± 0.21 ^{de}
2. Dursban at high conc.	312.05 ± 15.1°	104.25 ± 2.2°
3. Pestban at low conc.	268.7 ± 12.3 ^{bc}	119.99 ± 0.7 ^{de}
4. Pestban at high conc.	401.75 ± 50.4 ^d	116.3 ± 5.6 ^d
Profenofos		
Selection at low conc.	372.8 ± 6.3 ^d	75.67 ± 7.1 ^{ab}
2. Selection at high conc.	232.25 ± 28.4 ^b	68.96 ± 1.4 ^a
3. Ictacron at low conc.	376.1 ± 11.0 ^d	76.65 ± 3.2 ^{ab}
4. Ictacron at high conc.	244.3 ± 7.64 ^b	72.85 ± 1.91ab

Significantly different from the control groups by one- way analysis of variance of (p < 0.05) .

Each group includes nine replicates.

Acid phosphatase activity (AcP):

Effects of Chlorpyrifos and Profenofos in the two formulations on liver AcP in 18 and 21-day chick embryos treated at 4 day of development are summarized in table (3). The results revealed that the activity of 18 days old chick embryos liver AcP was insignificantly inhibited with the two concentrations of Chlorpyrifos from the two formulations. While with 21-days old chick embryos the activity was significantly decreased by Chlorpyrifos (Dursban, Pestban). There were no differences between selecton and lctacron on the enzyme activity in the liver of 18 and 21- days chick embryos, except for Selecton at the high concentration after 21 days.

The present results are in agreement with the results of Sati, (1996) who reported some changes in the liver AcP activity of hen after single or multiple treatments with some organophosphorus esters. The increment of AcP activity seems to result from enhanced enzyme turnover under pesticide stress, whereas the reduction of its activity may be related to leakage of the enzyme into the extracellular compartment. Barzu et al., (1973) demonstrated that Op's may cause a release of some hydrolytic enzymes from lysosomes

Table (3): Effect of Chlorpyrifos and Profenofos on liver acid phosphatase in 18 and 21 days chick embryos.

	μ mole P- nitropheno Mean	
Treatments	18 day	21 day
Untreated control	0.64 ± 0.04 ^{abc}	1.35 ± 0.098 ^e
Control treated with water	0.72 ± 0.08 ^{abcd}	1.22 ± 0.075 ^{de}
Chlorpyrifos		
1. Dursban at low conc.	0.61 ± 0.086 ^{abc}	0.923 ± 0.02 ^{ab}
2. Dursban at high conc.	0.51 ± 0.03 ^{ab}	0.915 ± 0.035 ^{ab}
3. Pestban at low conc.	0.597 ± 0.08 ^{abc}	0.796 ± 0.03^{a}
4. Pestban at high conc.	0.43 ± 0.07^{a}	0.98 ± 0.09 ^{abc}
Profenofos		
1. Selecton at low conc.	0.75 ± 0.14 bcd	1.16 ± 0.13 ^{cd}
2. Selecron at high conc.	0.93 ± 0.13 ^d	1.4 ± 0.05°
3. Ictacron at low conc.	0.67 ± 0.08 ^{abcd}	1.061 ± 0.04 ^{bcd}
lctacron at high conc.	0.842 ± 0.23 ^{cd}	1.1 ± 0.18 ^{bcd}

Significantly different from the control groups by one- way analysis of variance of < 0.05).

Each group includes nine replicates.

Alkaline phosphatase activity (AIP):

The activity of liver alkaline phosphatase (AIP) of 18 - days old chick embryos was decreased insignificantly with the two concentrations of Dursban, while, the activity was increased by increasing the concentration of Pestban, Selecton and Ictacron (table 4). On 21- days, Chlorpyrifos caused significant decrease of liver AIP activity, but Profenofos inhibited the activity at

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low concentration and changed to activation with the high concentration. The data indicated that the activities of serum AIP treated with Dursban and Pestban were decreased in a concentration dependent manner. Selecton had insignificant inhibition on serum AIP activity, but Ictacron caused significant activation of serum AIP activity. The present results are in agreement with some authors (Tag EI-Din et al., 1996 and Helal, 2000). Liver is often the primary target for the toxicity of various toxicants. The assessment of liver enzymes in blood is generally a more sensitive measure of hepatotoxicity and can be assessed within a shorter time (Korsrud et al., 1972). The reduction in AIP activity might be due to tissue damage, while the enhanced activity could be related to the influence of glucocorticoides (Murphy, 1966), or could be attributed to its release from ruptured cells due to the effect of pesticide (Shaffi, 1980).

Table (4): Effect of Chlorpyrifos and Profenofos on serum and liver alkaline phosphatase in 18 and 21 days chick embryos.

		Mean ± S_	
Treatments	*Liver (S	S.A X 10 3)	**Serum of
	18 day	21 day	Hatched
Untreated control	216.6 ± 9.5 ^a	205.9 ± 2.3 ^{bc}	416.7 ± 29.3 ^{cd}
Control treated with water	215.5 ± 3.6 ^a	213.7 ± 12.8 ^{bc}	410.0 ± 10.0 ^{cd}
Chlorpyrifos			
1. Dursban at low conc.	213.6 ± 7.7 ^a	142.1 ± 34.4°	328.6 ± 14.8 ^{bc}
2. Dursban at high conc.	194.5 ± 6.4°	116.9 ± 14.1 ^a	261.2 ± 19.4 ^b
3. Pestban at low conc.	321.7 ± 49.1 ab	155.6± 1.4°	287.5 ± 17.5 ^b
4. Pestban at high conc.	588.4 ± 162.7 ^d	110.9 ± 4.1 ^a	171.0 ± 22.0°
Profenofos			
1. Selecron at low conc.	314.1 ± 27.3 ^{ab}	198.9 ± 38.2 ^b	387.7 ± 16.0 ^{cd}
2. Selecron at high conc.	429.3 ± 61.7 ^{bc}	274.2 ± 29.9 ^d	398.8 ± 45.2 ^{cd}
3. Ictacron at low conc.	465.4 ± 58.1°	151.2 ± 7.3 ^a	442.2 ± 49.9 ^d
4. Ictacron at high conc.	746.7 ± 20.2°	247.9 ± 13.9 ^{cd}	524.0 ± 84.5°

^{*} Specific activity expressed as (Units / g wt.)

Total protein:

Table (5) illustrates the effects of the tested pesticides on serum total protein of chick embryos and the control groups. The data showed that there were insignificant differences in the serum total protein of the treated groups with Dursban; Pestban; Selectron and Ictacron at both concentrations and the control groups. This data is similar to those of Mandal *et al.*, (1992) who reported that fenvalerate at 5 mg / kg did not change the level of sheep serum total protein.

^{**} Specific activity expressed as (Units / L)

Significantly different from the control groups by one-way analysis of variance of (p < 0.05).

Each group includes nine replicates

Table (5): Effect of Chlorpyrifos and Profenofos on the serum total protein of 21- days chick embryos treated on the 4th dav.

						Treatment				
Total	Untreated	Untreated Control		Chlo	Chlorpyrifos			Pro	Profenofos	
Protein	Control	with								
		water	Dursban	pan		Pestban	Selecton	ron	Cto	Ictacron
(g/g)			Low conc.	High cone	- Tong	Low conc. High conc. I am sens.	1			
					LOW COILC.	righ conc.		High conc.	Low conc. High conc. Low conc. High conc.	High conc.
Mean	2.32									
+1	±0.31ª	2.36±0.29	2.36±0.29° 2.26±0.65° 2.18±0.69° 2.34±0.65° 2.23±0.25°	2.18±0.69ª	2.34±0.65ª	2.23±0.25ª	2.27±0.19ª 2.16+0.1ª 2.54+0.10ª 2.40.0.23ª	2.16+0.1ª	2 54+0 103	2 40.0 278
SD										2.4010.3/
Clanifica	41.									
Each oro	antily differen	nt from the	Each group includes also control groups by one-way analysis of variance of (p< 0.05).	s by one-wa	y analysis of	f variance of (0< 0.05).			
B	and along morning seplicates.	s mine repli	cates.							

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Also, Fayez and Kilgore (1992) reported that the serum total protein of male rat was not affected by the single acute oral dose of 1, 2.1 and 3.5 mg /kg of oxamyl. The insignificant effect of pesticides on the lipids and protein implies the possibility of the absence of any tendency of these chemicals to exert cytotoxic effects, which are highly dependent on interference with lipoprotein levels and rate of biosynthesis (Radwan *et al.*, 1993).

In conclusion, the external exposure of chicken eggs to the organophosphorus insecticide; Chlorpyrifos at the sublethal concentrations was found to be ChE inhibitor, while Profenofos is low in their potency as ChE inhibitor. The low potency in Profenofos can be attributed to less persistence in vivo and / or less affinity to the biochemical targets. We can be considering that the limiting factors for anticholinesterase effects are expected to be lipid solubility, stability and affinity to the ChE (Enan et al., 1981). Chlorpyrifos and Profenofos were found to be exerting hepatic action as showed by affecting phosphatases and transaminases. There were no differences between the two formulations of Profenofos in their effects on the studied biochemical targets. Certain Egyptian carriers are available and suitable to formulating pesticides with high good physics and chemical properties and without any apparent phytotoxicity that can serve the national pesticide industry (El-Sebae et al., 1980). While the differences between the two formulations of Chlorpyrifos on the AChE activity and the other enzymes may be du to the differences of the additives. Also, toxic impurities in the pesticide products might be formed during the manufacturing process, during storage, or after opening the sealed pesticide container.

REFERENCES

- Abou Donia, M.B.and D.G. Graham (1978). Delayed neurotoxicity of O ehtyl O 4 nitrophenyl phosphorothioate; subchronic (90 day) oral administration in hens. Toxicol. Appl. Pharmacol., 45: 685-700
- Altuntas, I.; N. Delibas, M. Demirci; I. Kilinc and N. Tamer (2002). The effect of methidathion on lipid peroxidation and some liver enzymes: role of vitamin E and C.Arch.Toxicol.,76: 470-473.
- Baker, F.D.; C. F. Tumasonis and J. Barron (1972). Mixed function oxidase activity in the chick embryo and the adult mouse. Bull. Environ. Contam. Toxicol., 9: 329-336.
- Barzu, T.; B. Cuuparencu and A. Hantz (1973). Activity of organophosphorus compounds on cell organelles-I. Effect of tetraethyl dithiopyrophophate on lysosomal hydrolyses. Biochem. Pharmacol., 22: 185-194.
- Bessey, D.A.; O.H. Lowry and M. I. Broch (1946): Determinations in serum with p-nitophenyl phosphate. J. Biol. Chem., 164: 321-329.
- Dieter, M. P. and S. N. Wiemeyer (1978). Six different plasma enzymes in bald eagles (*Haliaeetus Leucocephalus*) and their usefulness in pathological diagnosis. Comp. Biochem. Physiol., 61C: 153-155.
- Dixon, W. T. and J. H. Massay (1957) Introduction to statistical analysis 2 nd Ed., Mc Graw-Hill Book Co. Inc., New York.

Aly, Nagat M. and S. M. Abd - El Rahman

EL - Hamady, S. E.; F. A. Ahmed and R.B. Abo – Arab (1996). Side effects of some pesticides used to control water hyacinth. Proc. Of 6th Int. Conf. On Environ. Part is A must. N. I. O. F. and M. S. P. D. Alex.

Ellman, G.L.; D.K. Courtney; M.Jr. Andero and R. M. Feathestone (1961): A new and rapid calorimetric determination of acetylcholinesterase

activity. Biochem. Pharmacol., 7:88-95.

EL-Sebae A. E.; M. A. S. Othman; M. Hammam Soheir; G. Tantawy and S. A Soliman (1980). Delayed neurotoxicity of cyanofenphos in chickens. J.

Environ, Sci. Health, B15 (3): 267-285.

Enan, E. E. (1979). The effect of pre-exposure to some pesticides, synergists and induces on some biological functions in albino rats. Doctor thesis of Public Health Sc. High Institute of Public Health, Alexandria University.

Enan, E. E.; O. H. Enan and A. H. EL-Sebae (1981). Biochemical targets affected by sublethal doses of organophosphorus insecticides. Pest

Control (3): 121-123.

Fayez, V. and W. W Kilgore (1992). Acute toxicity effects of oxamyl in the rat. Fund. Appl. Toxicol., 18 (1): 155-159.

Flint, M.L. and R. V. D. Bosch (1981). Intruduction to integrated pest mangment. Plenunn Press New York and London pp.240.

Fossi, M. C.; C. Leonzio; A. Massi; L. Lari and S. Casini (1992). Serum esterase inhibition in birds: A non-destructive biomarker to assess organophosphate and carbamate contamination. Arch. Environ. Contam. Toxicol., 23(1): 99-104.

Hausman, T. U; W.P. Helger and W. Gross (1967): Optima condition for the determination of serum alkaline phosphatase by a new kinetic method.

Clinica. Chimica. Acta., 15: 241-245.

Helal, N. A. (2000). Toxicological studies of some chemical compounds against some environmental biotics. M.Sc. Thesis, Fac. of Agric. Tanta Univ.

Hoffman, D. J. and P. H. Albers (1984): Evaluation of potential embryo toxicity and teratogenicity of 42 herbicides, insecticides, and petroleum contaminants to mallard eggs. Arch. Environ. Contam. Toxicol., 13: 15-27.

Hoffman, D. J. and WC. Jr. Eastin (1981): Effects of malathion, diazinon, and parathion on mallard embryo development and cholinesterase activity.

Environ. Res., 26: 472-485.

Hoffman, D. J. and WC. Jr. Eastin (1982): Effect of lindane, paraquat, toxaphene, and 2,4,5- trichloro phenoxy acetic acid on mallard embryo development. Arch. Environ. Contam. Toxicol., 11: 79-86.

Hoffman, D. J. and L. Sileo (1984): Neurotoxic and teratogenic effects of organophosphorus insecticide (phenyl phosphonothioic acid – O – (4nitrophenyl) estr) on mallard development. Toxicol. Appl. Pharmacol., 73: 284–294.

Korsrud , G. O. ; H. C. Grice and J. M. Mc Laughan (1972). Sensitivity of several serum enzymes in detecting carbon tetrachloride liver damage

in rats. Toxicol. Appl. Pharmacol., 22:474-483.

J. Agric. Sci. Mansoura Univ., 29 (11), November, 2004

Lesser, J.; D. Blodgett and M. Enrich (2000). Comparison of oxime inhibited reactivation of organophosphorus – inhibited acetylcholinesterase i9n brain of avian embryos. J. of Toxicol and Environ. Health. Part- A. 59(1): 57-66.

Lutz – Ostertag Y. and C. Henou (1975) : Paraquat : mortalitè embryonnaire et effects sur l'appareil pulmonaire de l'embryon de Poulet et de Caille.

CR Hebd Acad Sc Ser 281 (D): 439 - 442.

Mandal, T.K.; A. Bhattacharya; A. K. Chabruborty and D.K.Basak (1992): Disposition kinetics cytotoxicity and residues of fenvalerate in tissues following oral administration to goats. Pestic. Sci., 35(3): 201-207.

Martin, P.A. and D.J. Forsyth (1998). Effect of exposure to vegetation sprayed with dimethoate or Chlorpyrifos on ducklings (Anas platyrhgnchos).

Ecotoxicol., 7(2): 81-87.

Meiniel, R. (1973): L'action tèratogène du parathion chez l'embryon d'oiseau. Arch. Anat.Histol. Embryol. Norm. Exp., 56: 97-109.

Mitchell, R.C. (1946). Effects of DDT spray on egg and nestling of birds. J.

Wildl Mgt, 10: 192-195.

Murphy, S.D. (1966). Liver metabolism and toxicity of triphosphate insecticides in mammals, avian, and piscine species. Proc. Soc. Exp. Biol. Med., 123:393.

Radwan, M. A.; K. A. Osman and A. K. Salama (1993). Biochemical response of the brown garden snail, *Helix aspersa* to chlorfluazuron and

flufenxuron. J. Environ. Sci. Health, B28 (63): 291-303.

Reitman , A. and S. Frankel (1957): A colorimetric method for determination serum glutamic oxaloacetic and glutamic pyrovic transaminase. Am. J. Clin. Path., 28: 53-55.

Sati, J. M. D. (1996). Liver enzymes as biomarker of exposure to organophosphorus pesticides. Alex. Sci. Exch., 17 (4): 351-360.

Shaffi, S. (1980). Thiodon toxicity. Non-specific phosphonosterases in nine fresh water toleosts. Toxicol. Lett., 6: 399.

Somers, J.; E. T. Moran; B. C. Reinhart and G.R. Stephenson (1974). Effect of external application of pesticides to the fertile egg on hatching success and early chick performance. 1. Preincubation spraying with DDT and commercial mixtures of 2,4-D; Picloran and 2,4,5 T. Bull. Environ. Contam. Toxicol., 11:33-38.

Tag El-Din, M. H.; Bayoumi and S. E. El-Hamady (1996). Efficiency of chlorpyrifos, carbosulfan and cypermethrin against the adults of rice weevil Sitophilus oryzae with respect to their detrimental side effects on

white rats. J. Agric. Sci. Mansoura Univ., 21(6): 2335-2342.

Weichsebaum, T. E. (1946): Determination of protein in small amounts of blood serum and plasma. Am. J. Clin. Pathol., 16: 40-44.

تأثير بعض المبيدات على بعض النظم الإنزيمية في أجنة الدواجن نجاة محمد على و صفاء مصطفى عبد الرحمن المعمل المركزى للمبيدات – مركز البحوث الزراعية – الصبحية – الإسكندرية

يهدف البحث دراسة تأثير تجهيزات مختلفة لبعض المبيدات الفسفورية ؛ الكلوربيريفوس ٨٤ % مركز قابل للاستحلاب (دورسبان - بستبان) والبروفينوفوس ٧٧ % مركز قابل للاستحلاب (سيليكرون - اكتاكرون) بتركيزين (١٠٠/١، ١/١، من المعدل الموصى به حقلياً) على بعض إنزيمات السيرم والمخ والكبد وكذلك البروتين الكلى في السيرم لأجنة الدجاج عند عمر ١٨، ٢١ يسوم (الفقس) مقارنة بالمجموعات الضابطة (الغير معامل والمعامل بالماء) . ويمكن توضيح النتائج في الآتي :

لقد احدث التجهيزات المختبرة لمبيد الكلوربيريفوس (الدورسبان - البستبان) انخفاضا معنويا لنشاط ابزيم الأسيتايل كولين استيريز (AChE) في المخ بكلا التركيزين المستخدمين في الأجنة بعد ١١، ٢١، ٢١ مناور، فيما عدا المعاملة بالتركيز المنخفض من الدورسبان بعد ١٨ يوم حيث كان الإنخفاض غير معنوى مقارنة بالمجموعات الضابطة. وعلى العكس من ذلك فإن مبيد البروفينوفوس يتجهيزتيه (سيليكرون اكتاكرون) لم يحدثا تأثيرا معنويا على نشاط ابزيم الأسيتايل كولين استيريز في المخ سواء عند ١٨ أو ٢١

حدثت زيادة لنشاط إنزيم الألأنين أمينو ترانسفيريز (ALT) في سيرم الأجنة المعاملة بكل من الكوربيريفوس (دورسبان بستبان) والبروفينوفوس (سيليكرون – اكتاكرون) وبكلا التركيزين المستخدمين زيادة معنوية . وعلى العكس من ذلك فإنها أحدثت تتبيطا معنويا لنشاط إنزيم الإسبارتيت أمينو ترانسفيريز (AST) في السيرم للأجنة المعاملة فيما عدا التركيز المنخفض من كل من الدورسبان و البستبان فكان الانخفاض غير معنوى مقارنة بالمجموعات الضابطة .

كما أوضحت النتائج أيضا حدوث تتبيط غير معنوى لنشاط إنزيم الفوسفاتيز الحامضى (ACP) فى الكبد لأجنة الدجاج المعامل بكل من الدورسبان و البستبان بالتركيزات المختبرة و بعد ١٨ يوم ، بينما السيليكرون والإكتاكرون فقد أحدثا تتشيطا غير معنويا لنشاط الإنزيم عند التركيز العالى (١/ ١من المعدل الحقلي). بينما أحدث كل من الدورسبان و البستبان و الإكتاكرون بالتركيزين المختبرين انخفاضا معنويا لنشاط الإنزيم . وعلى العكس فإن مبيد السيليكرون بالتركيز العالى أحدث زيادة غير معنوية في نشاط الإنزيم في الكبد للأجنة بعد ٢١ يوم من المعاملة.

ولقد حدثت زيادة في نشاط ابزيم الفوسفاتيز القاعدى (AIP) عند ١٨ يوم في الكبد المعامل بكل من البستبان ، السيليكرون و الإكتاكرون بعلاقة طردية مع التركيز . بينما انخفض نشاط الإنزيم انخفاضا معنويا في كبد الأجنة عمر ٢١ يوما من المعاملة بمبيد الكلوربيريفوس سواء عند ١٠٠١ أو ١٠٠١ من المعدل الحقلي . سبب كل من الدورسبان و البستبان تثبيطا لنشاط الإنزيم في السيرم يزيد بزيادة التركيز . وقد أحدث مبيد السيليكرون انخفاضا غير معنوى لنشاط إنزيم الفوسفاتيز القاعدى في السيرم بينما أحدث مبيد الإكتاكرون زيادة لنشاط هذا الإنزيم في السيرم وبعلاقة طردية مع التركيز .

ولم تظهر أية تأثيرات على مستوى البروتين الكلى فى السيرم للأجنة المعاملة بكل المبيدات المختبرة وبكلا التركيزين وذلك بالمقارنة بالمجموعات الضابطة.