DETERMINATION OF METHOMYL RESIDUES IN SLUG TISSUES AND DIFFERENT ORGANS IN ALBINO RATS

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ABSTRACT

High performance liquid chromatographic (HPLC) analysis of methomyl was used in order to measure the residue amount of methomyl as ng/g tissue of both terrestrial slug, Lehmenia marginata and male white albino rat, Raltus norvegicus var. albus. The results indicated high performance liquid chromatographic methomyl response after 48 hrs of topical application and poisoned food technique methods at different concentrations against gastropod slug, L. marginata. The distributed amounts of methomyl in different organs of male white albino rate, R. norvegicus var. albus due to LC₅ methomyl subcutaneously injection could be summarized as follows:

a) Cerebellum tissue contained higher amounts of methornyl than cerebrum by 1.8 folds.

b) Kidney had the highest methomyl residue of 6.99 ng/g tissue.

c) Both liver and heart had the same residual amount of methomyl; the recorded values were 4.87 and 4.82 ng/g tissue, respectively.

d) Cerebrum was the organ that contained the lowest methomyl residue.

e) It was obvious that the amount of methomyl recovered remarkably attenuated by increasing the interval after treatment.

 Haematological investigation showed marked decrease in both red blood cells and white blood cells.

INTRODUCTION

Two important polyphagous animal pests, belonging to two different categories of animal kingdom, were selected in this study. Terrestrial gastropods (molluscus: stug and snails) are being abundantly distributed in north coast, new reclaimed lands in addition to delta region of Egypt (Kassab and Daoud, 1964; El-Okda and Khalil, 1981 and Abo-Bakr, 1997). Land slugs belong to phylum Mollusca, Class Gastropoda and Order Pulmonata. They attack plant seedlings, roots of growing trees, excrete undesirable slim material, decrease the quality of crops, causing death of young seedlings and mature buds and finally transfer plant-causing diseases such as fungi and/or nematodes from plant to another (El-Wakil and Radwan, 1991; Abdallah et al., 1992 and Kassem, 1993). Methomyl is used to control land slugs, therefore the dissipation and distribution of methomyl were measured using High Performance Liquid Chromatographic analysis in terrestrial slug as well as in different organs of male white albino rat (cerebrum, cerebellum, heart, liver and kidney). Moreover, the haematological effect of methomyl was also investigated.

MATERIALS AND METHODS

1. Pesticides used

Methomyl, Lannate 90% S.P., S-methyl-N-[(methyl-carbamoyl) oxy]-thio-acetimidate.

2. Tested pests

2.1. Terrestrial slugs

L. marginata slugs were collected from ornamental nurseries and vegetable fields in Damanhour district, Behera Governorate. They were acclimatized for a week on the laboratory conditions and were fed on lettuce (Lactuca sativa) ad libitum.

2.2. White albino rats

Strain of white albino rats with an average age of 2-3 months and an average weight of 125-150 g was purchased from rodent laboratory, Extension Dept., Faculty of Agriculture, Alexandria University. The animals were housed in metallic cages in the laboratory and kept under observation at least two weeks for acclimatization.

3. Laboratory experiments

3.1. Determination of methomyl residues in land slug

The following concentrations of methornyl were prepared for lettuce leaves dipping method and topical application, 10, 30, 50, 100 and 200 ppm. Determination of the residues of methornyl in land slug, *Lehmania marginata* (Müller) was performed after 48 hrs and compared with the control groups (EI-Wakil *et al.*, 1999 and Lokman and Harpy, 1999).

3.2. Toxic effect against albino rats

a) Albino rats were injected subcutaneously with LC₅ (1/10 of LC₅₀) of methomyl (1 mg/kg body weight) dissolved in corn oil in DMSO (Dimethyl sulphoxid). The animals were killed by decapitation after different time intervals (12, 24, 72, 120 and 168 hrs) of trea ment. Blood samples were obtained for biochemical studies as well as different organs; brain (cerebellum & cerebrum), liver, kidney and heart to determine the pesticide residues of methomyl (El-Nabarawy et al., 2003 and Zidan et al., 1998).

Determination of tested pesticide residues in slug tissue and different organs of rats

4.1. Extraction process

At each time interval for rat and one time for slug after treatment by methomyl, the soft tissues were pooled and homogenized as 1:10 (w/v) with acetone for five minutes. The homogenate was filtered through a Whatman filter paper No. 1, then the filter paper was washed several times with acetone (10 ml portion). The filtrate was collected and made up to 100 ml with acetone, then dried over anhydrous sodium sulphate at 40°C. The pesticide residue was dissolved in 3 ml of methanol (Rotary vacuum evaporator, Yamato model RE-46 with flask 500 ml).

4.2. Cleanup process

Cleanup was carried out using a chromatographic column filled with silica get (2 g), florisit (2 g) and anhydrous sodium sulphate (1 g). The sample extract was transferred to the column, preconditioned by passing 10 ml of acetone through it. The column was eluted with 50 ml of methanol and the elute was collected and concentrated to 3 ml too. The samples were subjected to HPLC analysis.

4.3. HPLC analysis

The running conditions for High Performance Liquid Chromatography of methomyl was standardized and the optimum conditions were presented in Table (1). However, the chromatographic column in the present work was Ultrasphere Si-Column (ODS) 25 cm x 4.6 mm and the solvent system was methanol. The detector involved in this study was UV 245 nm wavelength with sensitivity of 0.02 Aufs. The chart speed was 0.5 cm/min and the injection volume was 10 $\mu l.$ Identification was accomplished by retention times and compared with the standards of methomyl at the same conditions. For quantification, the peak areas corresponding to each injected sample were compared with that of standard solutions of the insecticide. Detection limits for methomyl using the employed method were determined according to the analyte concentrations that produce a chromatographic peak equal to three times of base line noise.

4.4. Determination of residues using HPLC instrument

 The HPLC instrument involved in the present work was Beckman HPLC system consisting of:

a) Beckman 110 B Solvent Delivery.

- b) Beckman analog interface module 406.
- c) Beckman programmable UV detector module 166.

Hamilton 50 micro syringe fixed needle.

3. HPLC grade methanol.

 Stock solutions for methomyl (90% purity) were prepared in HPLC grade methanol.

Table (1): Running condition of high performance liquid

ltem	Value
Column	Ultra sphere-Si-column (ODS) 25 cm x 4.6 mm
Solvent system	Methanol
Flow rate	1 ml/min
Detector	245 nm wave length
Sensitivity	0.02 Aufs
Chart speed	0.5 cm / min
Inject volume	10 µl
Att.	32

RESULTS AND DISCUSSION

Haematological effects of methomyl

The haematological effects of LC_5 methomyl subcutaneously injection on adult male white albino rat, *Rattus norvegicus* var. albus were investigated. Haemoglobin, creatinine, white blood cells (WBCs) and red blood cells (RBCs) counts were recorded. The following observations could be noticed (Table 2):

- Methomyl decreased both blood cell counts by 46.49 and 57.69% for RBCs and WBCs, respectively.
- b) Methomyl treatments induced insignificant changes in both haemoglobin and creatinine contents.

Residues analysis

High performance liquid chromatographic (HPLC) analysis of methomyl was used in order to measure the residue amount of methomyl as ng/g tissue of both terrestrial slug, Lehmania marginata and male white albino rat, Rattus norvegicus var. albus. The use of HPLC is a potentially useful technique for insecticide residue analysis (Strait et al., 1991 and El-Nabarawy et al., 2003). Beckman HPLC system was used for analysis of methomyl. The optimum conditions of methomyl analysis is recorded in Table (1).

Table (2): Effect of LC₅ methomyl treatment on blood parameters of male white albino rat (*Rattus norvegicus* var. *albus*) after different time intervals

Blood	Control ±S.D	Time intervals (hrs)					Mean	1.00
parameters		12	24	72	120	168	±S.D	L.S.D _{0.05}
RBCs x10 ⁸	3.85±0.270	2.15	2.25	2.70	1.95	1.75	2.06+0.190	0.251
WBCs x10 ³	9.10+0.620	9.90	7.20	2.90	1.80	2.95	3.85+0.340	0.469
Hb, mg%	10.50+0.940	9.20	10.30	9.50	7.30	9.20	9.10+0.770	1.110
Creatinine, mg/dl	0.61 <u>+</u> 0.046	0.66	0.72	0.76	0.84	0.78	0.75 <u>+</u> 0.067	0.091

Values are means of 4 replicates.

The results in Table (3) show the high performance liquid chromatographic methomyl response after 48 hrs of topical application and poison food technique methods at different concentrations against gastropod slug L. marginata. Untreated slugs exhibited very minute amount of methomyl 0.003 ng/g tissue. Treatments by topical application and poison food technique showed insignificant difference in methomyl residue due to the method of application. In addition, increasing concentrations exhibited a remarkable increase of methomyl amount recovered. Both methods of treatment exerted a degree of fortification of the methomy! recovered that increasing concentration 3 times increased methomyl recovered by 18,7 and 26.5 times higher due to topical application and poison food tichnique, while increasing concentration 20 times enhanced the amount of methomyl found by 81.12 and 182.4 times higher due to topical application and poison food technique, respectively. These results are in agreement with those reported by Andrews (1989), Kassem (1993), Abdallah, et al. (1998) and Hanafy et al.(1998).

Table (3): Methomyl residues in land slug, Lehmania marginata (Müller) afte, 48 hrs at different concentration treatments, shown as ng/g tissue.

<u></u>						
Total	(Mean + S.D.				
Treatment	10	30	50	100	200	Mean _ S.D.
Control	0.003	0.003	0.003			0 003 ± 0 0
Topical application technique	0.062	1.160	1.870	7,930	5 030	3.210 <u>+</u> 1.60
Dipping poison food technique	0.054	1.430	1,140	3 120	9.850	3.118 <u>+</u> 1.46

- Data represent means of 3 replicates + S.D.

- Retention time (RT) = 2.66 min.

The distributed amounts of methomyl in different brain regions and organs of male white albino rat, *R. norvegicus* var. *albus* due to LC₅ methomyl subcutaneously injection are summarized in Table (4) as well as Figure (1).

The results of methomyl residue analysis in different organs of rat revealed that:

- a) Cerebellum tissue contained higher amounts of methomyl than cortex by 1.8 folds
- b) Kidney had the highest methomyl residue of 6.99 ng/g tissue.
- c) Both liver and heart had nearly the same amount of methomyl recovered, the values were 4.870 and 4.826 ng/g tissue for liver and heart, respectively.
- d) The organ that contained the lowest methomyl residue was the cortex (cerebrum) of the rat brain.
- e) It was obvious that increasing time of measurements after treatments attenuated the amount of methomyl recovered remarkably, the amount of methomyl was decreased descendingly by increasing time interval. This might be due to detoxification and/or rapid excretion of the methomyl of the body corresponding to increasing time.

These results are in agreement with those reported by Strait et al.(1991) and El-Dashlouty et al.(2000).

Table (4): Methomyl residues in different organs of male white albino rat, Rattus norvegicus var. albus treated with LC₅ at different time intervals, shown as ng/α tissue.

Time		Brain			Kidney	Total amount
intervals (hrs)	Cortex	Cerebellum	Heart	Liver		
Control	0.18	0.09	0.006	0.08	0.19	0.346
12	1.32	1.79	1.65	1.62	1.21	7.59
24	0.87	1.44	1.36	1.04	1.22	5.93
72	0.71	2.59	1.13	1 15	1.24	6.82
120	0.34	0.62	0.40	0.78	2.56	4.70
168	0.30	0.18	0.28	0.20	0.57	1.53
Total amount	3.72	6.71	4.826	4.87	6.99	27.116

* Data represent means of 3 replicates.

* Retention time (RT) = 2.66 min.

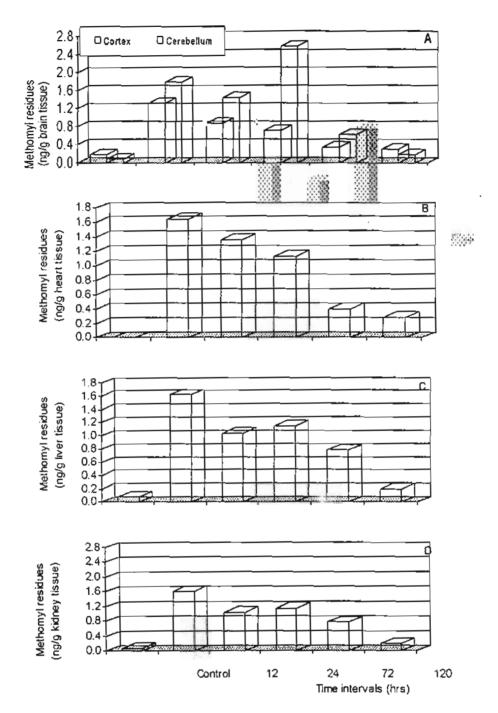


Fig. (1): Methomyl residues in different organs: A) Brain, B) Heart, C)
Liver, and D) Kidney of male white albino rat, Rattus
norvegicus var. albus treated with LC5 at different time
intervals, shown as ng/g tissue.

The results in Table (5) and Figures (2 and 3) show the high-performance liquid chromatographic response of methomyl when 50, 100, 150, 200 and 250 μg of such insecticide were injected under the previous conditions. These results indicate the linearity of response within the range of 50 μg to 250 μg of such compound. The values of peak area were calculated to be 1.9×10^4 , 3.6×10^4 , 6.8×10^4 , 11.3×10^4 and 15.6×10^4 (BC) when 50, 100, 150, 200 and 250 μg , respectively were injected into the high-performance liquid chromatography as shown in Table (5).

Table (5): Concentration-response relationship for methomyl using High

Performance Liquid Chromatography.

No.	Concentration of methomy! (µg)	Peak area (BC)
1	50	1.9 x 10⁴
2	100	3.6×10^4
3	150	6.8 x 10 ⁴
4	200	11.3 x 10 ⁴
5	250	15.6 x 10 ⁴

⁻ Retention time (RT) = 2.66 min.

⁻ Each value is an average of 3 replicates.

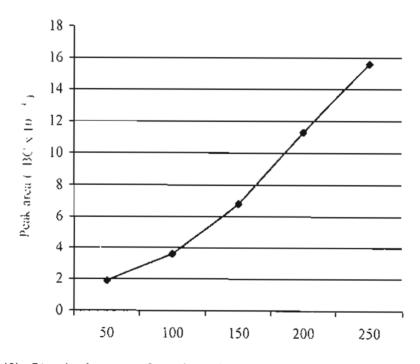


Fig. (2): Standard curve of methomyl using high performance liquid chromatography peak area (BC).

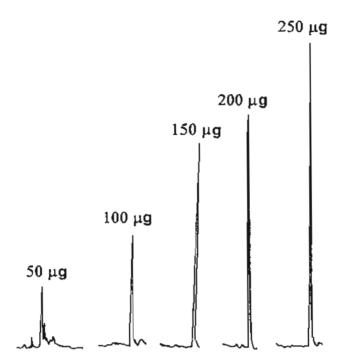


Fig. (3): High performance liquid chromatographic response for different concentrations of methomyl.

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تقدير متبقيات الميتوميل في أنسجة البزاقات والأعضاء المختلفة في الفئران البيضاء

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اشتملت الدراسة على دراسة التأثير على قياسات الدم في الفئران البيضاء وأوضحت النتائج أنه عند المعاملة بــ LCs لذكور الفئران البيضاء حقنا تحت الجد وكذلك دراسمة تركيسز الهيموجلوبين والكرياتينين وعدد خلايا الدم الحمراء والبيضاء وقد لوحظ الأتي:

 ا) نقص فى كلا من كرات الدم الحمراء والبيضاء بمعدل ٤٦,٤٩%، ٢٩,٧٩% على التوالى عند المعاملة بمبيد الميثوميل.

 لمعاملة بالميثوميل أحدثت تغيرات غير معنوية في كمل من محتوى الهيموجلوبين والكرياتينين.

كذلك إشتملت الدراسة على دراسة اختفاء وتوزيع مبيد الميثوميسل بإسستخدام جهاز الكروماتوجرافي السائل ذو الكفاءة العالمية ونلك في البزاقات الأرضية والانسجة المختلفة من نكور الفئران البيضاء إشتملت على الغص الأمامي والغص الخلفي للمخ وكذلك القلب والكبد والكلى. ومن نتائج الدراسة وجد أنه عند تقدير كميات متبقيات مبيد الميثوميل معبرا عنها بالنانو جرام لكل جرام من النسيج (ng/g tissue) في البزاقات الأرضية لم تظهر نتائج معنوية بعد ٤٨ ساعة على تركيزات مختلفة باستخدام الطريقتين وهما التطبيق السطحي والطعم السام لمبيد الميثوميل، أما الكميات المنتشرة من المبيد في مناطق المخ والأعضاء لذكور الفنران البيضاء المعاملة بـ كلام المبيد حقنا تحت الجلد تتلخص في الأتي:

انسجة القص الخلفي Cerebellum تحتوى على كمية أعلى من الميثوميل أكثر من القص الأمامي Cerebrum بـ ١,٨ أضعاف.

٢- الكلى بها متبقيات عالية حوالي ٦,٩٩ نانوجرام من المبيد/جرام من النسيج.

٣- تساوت كميات الميثوميل تقريبا في كلا من الكبد والقلب حيث القيم همي ١٨٧٦، ٢٦٨،٤ نساوت الميثوميل تقريبا في كلا من الكبد والقلب على التوالي.

٤- بزيادة زمن القياس بعد المعاملات انخفضت كمية الميثوميل المقدرة تتازليا بمرور الزمن.