Nephroprotective Effect of *Malva sylvestris* Extract against CCl₄ Induced Nephrotoxicity in Albino Rats

Hanaa S. S. Gazwi and Magda E. Mahmoud

Department of Agricultural Chemistry, Faculty of Agriculture, Minia University, El-Minia, Egypt

ABSTRACT

The aim of the study was to evaluate the potential effect of *Malva sylvestris*(*M. sylvestris*) extract on nephrotoxicity caused by CCl_4 in rats. Forty rats were divided into four equal groups. Group 1: negative control group (normal).Group2: positive group, treated with CCl_4 (1ml/kg body weight (b.w)), twice a week for 8 weeks. Groups 3 and 4 were treated with the same dose of CCl_4 co-administered with *M. sylvestris* at dose 150 or 300 mg/kg b.w, respectively. Results appeared that rats treated with CCl_4 showed a significant increment in WBCs, kidney function (urea and creatinine), total lipid, cholesterol, triglycerides, glucose, malondialdehyde (MDA), and nitric oxide (NO) levels, but a significant decline in the mean values of weight gain, RBCs, PCV, Hb, uric acid, and catalase (CAT) as comparing with the control group. The treatment with *M. sylvestris*(150 and 300 mg/kg b.w) improved the hematological and biochemical parameters. These protective effects were dose dependent. The histological results confirmed these parameters that enhanced by CCl_4 Thus, the extract of *M. sylvestris* represents an encouraging chance for the curative renal injury. **Keywords:** *Malva Sylvestris*, Nephrotoxicity, Kidney function, Malondialdehyde, Nitricoxide.

INTRODUCTION

Kidneys are one of the key organs of the body which carry out several important roles in the body. Removal of waste from the bloodstream (urine formation) is considered the main function of the kidney. Also, the kidney perform many homeostatic functions example maintain volume, pH and ionic balance. Also, toxic metabolic by-products such as urea, ammonia, and uric acid are excrete (Alam et al., 2016). There are numerous therapeutic agents such as aminoglycoside antibiotics, nonsteroidal anti-inflammatory drugs (NSAIDs), antitubercular and chemotherapeutic drugs can badly influence the kidney causes severe renal failure, nephrotic syndrome and chronic interstitial nephritis (Hoitsma et al.,1991).Renal failing is the disorder where the with holding of metabolic products in reaction to the weakening of function (Pydi, 2011). Prevalence of chronic kidney disease in worldwide is estimated 8-16% in 2013 (Jha et al.,2013). In the 2015 worldwide Burden of ailment observe, kidney sickness changed into the 12th most commonplace cause of demise, accounting for 1.1 million deaths worldwide(Wang et al., 2016). Until date, available options for a remedy against renal disorders revolve around dialysis and kidney transplant. Both of them are often outside the reach of many people, especially in developing countries. Therefore, there is need to increase research toward finding a safe, facilely available and effective remedy against kidney disorder. Since this solution is the usage of plants in medicine being an age-long practice in several parts of the world for both preventive and curative. Today, it is estimated that about 80% of the planet's population rely on botanical preparations as medication to meet their health needs (Ogbera et al., 2010).

Malva sylvestris L. (Malvaceae) has been used medicinally throughout the world since 3000 BC (Henry and Piperno, 2008). It is commonly used as medicinal plant and in nurturing both animals and humans. Edible uses are attentive with folk gastronomy and with those uses covered by so-called simple nourishment (Barros *et al.*, 2010 and Guarrera, 2003). Raw young leaves are eaten in salads, shoots and leaves are consumed in soups and as boiled vegetables. Medical applications of the plant are numerous such as treating specific disorders of the respiratory system, gastrointestinal tract, musculoskeletal system, as well as skin injuries. It also has diuretic effects, laxative, lenitive, spasmolytic and choleretic impacts. Also, is used as bronchodilator, anti-diarrheal, anti-cough and highly recommended for the treatment of acne and skin care (Carvalho, 2005 and DellaGreca *et al.*, 2009).

CHECKED

Turnitl

Additionally, effective against mouth and throat diseases and can reduce swelling and soothe tooth pain, and gingivitis (Passalacqua et al., 2007). Many compounds in this plant carry the properties of analgesic and antiinflammatory (Gasparetto et al., 2012), including kaempferol (De Melo et al., 2009), apigenin (Funakoshi-Tago et al., 2011), quercetin (Kleemann et al., 2011), scopoletin (Moon et al., 2007) and ferulic acid (Kim et al., 2012). The biological activity of this plant might be ascribed to antioxidants, for example, polyphenols, vitamin E, β -carotene, vitamin C, unsaturated fatty acids (e.g. α linolenic acid), minerals and other vital phytochemicals (Barros et al., 2010). Leaves of Malva sylvestris have exceptionally antioxidant properties including the activity of radical-scavenging, diminishing force and lipid peroxidation hindrance in liposomes and brain cells homogenates.

To date, there are no data concerning the *in vivo* effect of *M. Sylvestris* extract on nephrotoxicity damage and oxidative stress induced by CCl₄. Thus, the present study evaluates the protective effect of *M. Sylvestris* against CCl₄-induced oxidative stress and nephrotoxicity in rats.

MATERIALS AND METHODS

Materials

Carbon tetrachloride were obtained from El-Gomhorya Pharmaceutical Company, Cairo, Egypt and Chemical kits for uric acid, blood urea nitrogen (BUN),creatinine, protein, albumin, total lipids total cholesterol (TC), triacylglycerol (TG),glucose, malondialdehyde (MDA), catalase (CAT) and nitric oxide (NO) were purchased from Biodiagnostic Company, Cairo, Egypt. All other chemicals were considered among the best available commercial grades. Leaves of *M. Sylvestris* were collected from Faculty of Agriculture Farm.

Preparation of aqueous extract of M. Sylvestris leaves

M. Sylvestris leaves were dried in shade at room temperature for two weeks and ground into fine powder. 100 g of powder was mixed with 1L of boiling distilled

water for 10 min under continuous stirring. The mixture was filtered twice through a mesh and through whatman No .1 filter paper, and the obtained liquid was evaporated to dryness under vacuum using a rotary evaporator, then collected in glass Petri dishes, dried in a vacuum desiccator and finally stored in airtight glass containers in a refrigerator at 4°C for use in the experiments.

Animals and Experimental Design

Forty female Sprague-Dawley albino rats (150 \pm 5g) were obtained from the Animal House of Faculty of Agriculture, Minia University. They were housed under standardized environmental conditions, fed with standard diet and left to acclimatize to the environment (22 \pm 2°C under a 12/12 hr light/dark cycle) for one week before the experiment was beginning.

Animals were divided into four groups (ten rats each) and treated as follows: Control group (negative), rats were injected with the respective vehicle (0.5 ml/kg b.w paraffin oil in saline). Rats of the second group (positive) were injected with CCl₄ (1 ml/kg, 1:1 mixture with paraffin oil, i.p. (Marsillach *et al.*, 2009)), each third days for 8 weeks to induce liver fibrosis. The third and fourth groups (CCl₄+*M. Sylvestris*150 and 300 mg/kg respectively) were received *M. Sylvestris* extract (150 and 300 mg/kg respectively) daily for two weeks. Then injected with CCl₄ as described in positive group.

After eight weeks rats were fasted overnight and anesthetized by diethyl ether to take the blood samples from the retro-orbital plexus (Schermer, 1967) from all animals of each group. Each sample was divided into two portions: the first portion was immediately taken in heparinized tube for hematological study and the second portion of the sample was taken in glass tube and left for 20 min to coagulant at room temperature and then centrifuged at 3000 rpm for 15 min, to obtained serum samples which kept at -20°C until used for the assessment of kidney function and lipid profile tests. Then, rats were sacrificed and kidney tissues were dissected, washed with ice-cold saline, weighed and stored at -20°C. Thereafter, kidney tissues were homogenized in saline and the homogenate was used for assessment of oxidative stress markers catalase (CAT), NO and lipid peroxides (MDA). In addition, specimens from kidney tissues were fixed in 10% formalin for histopathological examination.

Determination of Hematological Parameters

The red blood cells (RBC) and white blood cells (WBC) counts were carried out by Neubauerhemocytometer method (Dacie and Lewis 1991). The hemoglobin (Hb) concentration was determined according to Jain (1986), using the cyanmethemoglobin method. The packed cell volume (PCV) was determined by the microhaematocrit method according to Dacie and Lewis (1991).

Biochemical estimations

Kidney function tests Serum uric acid, urea and creatinine levels were assaved in the samples by a colorimetric method (Fossati *et*

assayed in the samples by a colorimetric method (Fossati *et al.* 1980; Fawcett and Scott 1960 and Szasz*et al.*, 1979, respectively).Serum albumin level was assayed using the method described by Dumas *et al.*, (1972).Creatinine

/albumin ratio (C/A) was calculated from the results obtained.

Determination of total lipids, cholesterol, triacylglycerol and glucose.

Serum samples were used for determination of total lipids (Zollner and Kirsch, 1962) total cholesterol (TC) (Allain *et al.*, 1974), triacylglycerol (TG) (Fossati and Prencipe, 1982) and glucose Trinder (1969).

Determination of lipid peroxide level

Lipid peroxidation level in the kidney homogenate was determined as thiobarbituric acid reactive substances (TBARS) by measuring malondialdehyde (MDA) level spectrophotometrically in kidney homogenates according to Mihara *et al.*,(1978) and catalase (CAT) and nitric oxide (NO) were determined according to Yoshioka *et al.*,1979. and Green *et al.*, (1982)

Histopathological examination

Kidney specimens were fixed in 10% formalin and processed for paraffin sections of 4 μ m thickness. Sections were stained with hematoxylin and eosin (H&E) for routine histopathological examination and Masson's trichrome for demonstration of collagen fibers.

Statistical analysis

The results obtained in the present study were evaluated by One Way ANOVA test followed by Tukey Dunken SPSS. The results were expressed as mean \pm standard error and values of P<0.05 were considered statistically significant (Snedecor and Cochran, 1986).

RESULTS AND DISCUSSION

Effect of *M. sylvestris* on weight gain and average kidney weight in albino rats

Body weight changes may provide an indicator of drug effect and are used for assessment of responses to the drug therapy (Asuquo *et al.*, 2012). The effect of *M. sylvestris* on weight gain (g) and average kidney weight (g) are represented in Table 1. Treatment of rats with CCl₄ lead to a significant reduce in weight gain comparing with control group (P< 0.05). Treatment with *M. sylvestris* markedly improved the growth. The average weight of the kidney was significantly increase in CCl₄ group comparing with normal group (P<0.05). Administration of *M. sylvestris* leads to a significant reduce in the average kidney weight comparing with CCl₄ group (P<0.05).

This increase in weight of the kidney may be imputed to lesions and injuries related to xenobiotics (Wong *et al.*, 2010) like CCl₄ which peroxidizes proteins of cell that way stimulating pathway of the inflammatory. Also, these results came in agreement with Abdel Moneim and El-Deib, (2012) and Sahar and Dalia, (2014) who found that CCl₄caused a significant decrease in body weight while increases kidney weight and relative weight of kidney. The enlargement of the kidney was significantly reduced in *M. sylvestris* groups, suggesting that the *M. sylvestris* includes some protecting phytomedicinals. This observation of the effect on body weights of *M. sylvestris* groups can be explicated by its effect on appetite center in the hypothalamus.

Parameter	Control	CCl ₄	M. sylvestris (150 mg/kg b.w)	M. sylvestris (300 mg/kg b.w)
Weight gain (g)	52.45 ±4.81	22.24 ^a ±1.32	$38.17^{ab} \pm 3.82$	40.28 ^b ±4.69
Average kidney weight (g)	1.37±0.04	$1.59^{a} \pm 0.08$	$1.35^{b}\pm0.06$	1.32 ^b ±0.07

Table 1. Effect of <i>M. sylvestris</i> on weight gain and average kidney weight in albino ra	ats
---	-----

Data represent the mean \pm S.E. of observation from 10 rats.^a significantly different from control group at P \leq 0.05.^b significantly different from CCL₄ group at $P \le 0.05$.

Effect of *M. sylvestris* on the hematological parameter

Hematological parameters are used to provide useful information for diagnosis in the routine clinical evaluation of the state of patient health. This parameters (RBC, PCV, Hb, and WBC) are used to evaluate the effects of *M. sylvestris* on the blood of rats (Table 2). The group was injected with CCl₄ significantly decreased the levels of RBC, PCV, and Hb, while increased the level of WBC compared to the control group (Table 2). These could be attributed to CCl4 toxicity and its direct effect on the hematopoietic system (Zeynab and Shereen, 2012 and Al-Mashhadani, 2017). And decreasing hemoglobin in

injected rats with CCl₄ is an indication of hemolysis and the decreasing in hemoglobin has a corresponding elevation in methemoglobin content which affects the oxygen-carrying the blood, caused by the toxicant(Tilak et al, 2007). The increasing in WBC can be due to the stimulation of the immune defense system(Kashinath 1990) or increasing antigen concentration in the body (Hoeney, 1985), the low values PCV may be attributed to anemic conditions. While groups pretreated with 150 and 300 M. sylvestris significantly increased ($p \le 0.05$) RBC, PCV, and Hb levels while decreased in WBC levels when compared with untreated rats (Table2).

Table 2. Effect of *M. sylvestris* on a hematological parameter in albino rats

Parameter	Control	CCl ₄	M. sylvestris (150 mg/kg b.w)	M. sylvestris (300 mg/kg b.w)
RBCs ($x10^{6}$ /cmm)	6.92±0.17	6.75±0.16	6.9±0.00	6.92±0.14
PCV%	49.1±1.38	41.0 ^a ±2.69	45.5±0.00	45.8±2.26
Hgb g/dl	15.85±0.53	$13.50^{a} \pm 0.00$	$13.3^{a} \pm 1.06$	$14.78^{b} \pm 0.72$
WBCs (x10 ³ /cmm)	10.15±0.85	13.6±2.78	10.0±1.06	10.9±2.98

Data represent the mean \pm S.E. of observation from 10 rats.^a significantly different from Control group at P \leq 0.05.^b significantly different from CCl_4 group at $P \le 0.05$.

Effect of *M. sylvestris* on renal function:

Table (3) show that urea, uric acid, creatinine, albumin levels and creatinine/ albumin ratio were used as biochemical markers for evaluation of kidney injury and these parameters were significantly increased in CCl4treated animals (P < 0.05). This result agrees with that of Al-Seeni et al., (2016). These increases could be attributed to impairment in renal functions. Uric acid, the metabolic end outcome of purine metabolism, has tried to be a selective antioxidant, capable particularly of reacting with free radicals and hypochlorous acid (Hasugawa and

Table 3. Effect of M. sylvestris on renal function

Kuroda, 1989). The increasing levels of creatinine and urea may be due to a diminish in glomerular filtration rate caused by acute renal dysfunction (Rahmat et al., 2014).

In addition, reduced albumin concentration in CCl₄treated rats resulted in significant leakage due to hyperplasia in glomeruli and tubules (Adewole et al., 2007). Whereas, treated rats with M. sylvestris concomitantly with CCl₄ afforded significant protection against CCl₄-intoxication (Table 3). The ameliorative effect against renal toxicity may be ascribed to high levels of polyphenols and other antioxidants like flavonoids.

Parameter	Control	CCl ₄	M. sylvestris (150 mg/kg b.w)	M. sylvestris (300 mg/kg b.w)
Uric acid (mg/dl)	2.78 ± 0.17	$1.64^{a} \pm 0.07$	$2.24^{ab} \pm 0.05$	2.67 ^b ±0.01
urea(mg/dl)	22.13 ± 2.67	42.26 ^a ±3.59	28.87 ^b ±3.59	25.13 ^b ±2.21
Creatinine (mg/dl)	0.72 ± 0.02	$1.51^{a} \pm 0.08$	$1.04^{ab} \pm 0.05$	$0.98^{ab} \pm 0.03$
albumin(mg/dl)	5.02 ± 0.42	$3.77^{a} \pm 0.23$	3.93 ^a ±0.36	4.41 ^a ±0.21
Creatinine/Albumin ratio	0.14 ± 0.01	$0.4^{a}\pm0.01$	$0.27^{ab} \pm 0.03$	$0.22^{ab} \pm 0.02$

Data represent the mean \pm S.E. of observation from 10 rats.^a significantly different from Control group at P \leq 0.05.^b significantly different from CCl₄ group at $P \le 0.05$.

Effect of *M. sylvestris* and CCl₄ on total lipid, cholesterol, triglycerides and glucose.

The data in Table 4 shows that treatment with CCl₄ to rats significantly raised the levels of total lipid, triglycerides, cholesterol, and glucose comparing with control.

These results came in agreement with Nwidu et al. (2017) who found that oxidative stress caused by CCl_4 increased the lipid profile levels. On the other hand, it may be presumed that hypercholesterolemia in rats treated with CCl₄ resulted from the damage of hepatic parenchyma cells, leading to an imbalance of lipid metabolism (Havel et al., 1986). However, M. sylvestris significantly improved the lipid profile of rats treated with CCl₄.

Hanaa S. S. Gazwi and Magda E. Mahmoud

Parameter	Control	CCl ₄	M. sylvestris (150 mg/kg b.w)	M. sylvestris (300 mg/kg b.w)
Total Lipid (mg/dl)	430.15±17.67	$670.16^{a} \pm 28.91$	$500.61^{ab} \pm 11.72$	$450.21^{b} \pm 11.89$
Cholesterol (mg/dl)	6.27 ± 95.25	$136.62^{a} \pm 7.41$	112.87 ± 8.52	$101.69^{b} \pm 7.26$
Triglycerides (mg/dl)	66.63 ± 4.65	$87.26^{a} \pm 3.65$	979. $1^{a} \pm 2.4$	$73.84^{b} \pm 2.39$
Glucose (mg/dl)	70.76 ± 4.99	$98.78^{a} \pm 5.01$	$80.65^{b} \pm 5.62$	$72.43^{b} \pm 6.16$

Table 4. Effect of <i>M</i> .svlvestris and CCL	on total lipid cholesterol, triglycerides, and glucose.

Data represent the mean \pm S.E. of observation from 10 rats .^a significantly different from Control group at P \leq 0.05.^b significantly different from CCL group at P \leq 0.05.

Effect of *M. sylvestris* on malondialdehyde, catalase and nitric oxide in albino rats

but not significant compared to CCl_4 group. It seems that *M. sylvestris* protect the kidney rats.

Table 5, indicate that the level of kidney malondialdehyde in CCl_4 group was significantly higher than the control group. Our result came in agreement with other reporters who demonstrated that CCl_4 significantly increased the renal MDA levels comparing with control group (Abdel Moneim and El-Deib, 2012; Sahar and Dalia, 2014 and Hamid *et al.*,2018). The increase in the level of MDA indicates that peroxide enhancement leads to tissue injury and the failure of antioxidant mechanisms to stop the production of excessive free radicals (Furfaro *et al.*, 2012; Satoh *et al.*,2013 and Garcia-Nino and Pedraza-Chaverri , 2014). Noorah and Mousa, (2014) reported that the increase in MDA concentration due to increased oxidative stress .Concomitant *M. sylvestris* with CCl_4 markedly improved levels of MDA compared with CCl_4 group

As shown in Table 5, the activity of CAT was decreased in rats treated with CCl_4 (0.367±0.03U/g) compared to control group (0.979 ±0.02 U/g). However, treatment with *M. sylvestris* markedly increased its activity

Oxidative stress is an imbalance between the antioxidant mechanisms and the production of Reactive oxygen species (ROS) (Frei, 1994). The results of the present study have appeared that oxidative stress caused by CCl_4 is also evident from the significant depletion of the antioxidant catalase enzymes. Decreased CAT activity in the kidney may be caused by accumulation of ROS in kidney tissue in this study. Similar results were reported (Ottu *et al.*, 2013 and Sylvia *et al.*, 2017). The protective effects of *M. sylvestris* against the CCl₄ could be ascribed to its high concentration of phenolic compounds and other antioxidants

Treated with CCl_4 increased NO activity (14.29±1.23) compared to control group (4.84±0.59), treatment with *M. sylvestris* significantly decreased the activity of NO compared to the CCl_4 group. Our findings came in agreement with Khan *et al.*,(2009) and Abdel-Moneim and El-Deib (2012) who showed that NO renal levels significantly increased in the CCl_4 group comparing with control group.

Table 5. Effect *of M. sylvestris* on hepatic lipid peroxide as (MDA) and catalase activities (CAT) in CCl₄-induced hepatoxicity in rats.

	(150 mg/	(kg b.w) (300 mg/kg	b.w)
/±0.49 10.54	$4^{a} \pm 0.45$ 6.84^{ab}	± 0.44 8.21 ^{ab} ± 0.2	30
9±0.02 0.36	$7^{a} \pm 0.03$ 0.613 ^{ab}	± 0.02 0.776 ^{ab} ± 0	.02
±0.59 14.2	9 ^a ±1.23 12.91 ^a	± 1.82 9.56 ^{ab} ± 1 .	24
	9±0.02 0.36 ±0.59 14.29	$\begin{array}{cccccccccccccccccccccccccccccccccccc$	$y \pm 0.49$ 10.54 $^{a} \pm 0.45$ $6.84^{ab} \pm 0.44$ $8.21^{ab} \pm 0.3$ $y \pm 0.02$ $0.367^{a} \pm 0.03$ $0.613^{ab} \pm 0.02$ $0.776^{ab} \pm 0.3$

Data represent the mean \pm S.E. of observation from 10 rats .^a significantly different from Control group at P \leq 0.05.^b significantly different from CCL4 group at P \leq 0.05.

Effect of *M. sylvestris* on histopathological changes in the kidney

Several studies showed that plants possessing the properties of free radical scavenging can play an important role in protecting oxidative damage in various organs such as kidney, liver, and brain; caused by environmental and chemical toxic substances through metabolic activation to highly reactive substances such as free radicals (Kengar *et al.*, 2017).

Fig 1 and 2 show the histopathological changes of kidney section. A photomicrograph of kidney section of group (A) showed renal corpuscles, glomeruli (G) formed of capillary tufts is surrounded by Bowman's capsules (arrows). Note the proximal (P) and distal (D) convoluted tubules cells intact with acidophilic cytoplasm and vesicular nuclei.CCl₄ intoxicated rat kidney showed renal structural disruption with empty areas of completely degenerated renal corpuscles (*) (B).Renal structural disruption with extensive collagen fibers deposition around renal tubules and blood vessels(arrows). These results

agree with Majno and Joris, (2004) and Kengar et al.,(2017) who showed CCl₄ caused significant glomerular and tubular degenerations. Glomerular congestion has been observed with disintegrated Bowman's capsules. Vacuolar and degenerative changes were also evident in the endothelial lining of the glomerular tuft and in the epithelial lining of renal tubules. Swollen proximal tubules and dilated Bowman's capsule were detected. Swollen collecting dust, distinct intertubular connective tissue and thickening of collecting tubules are an indicator of CCl₄ toxicity .While administration of M. sylvestris regained the histological alternations produced by CCl₄ into normal histological kidney structure with a small dilatation in some tubules (H&E, scale bar=50µm). Nephroprotective effects of *M. sylvestris* related to the existence of active components such as phenolics and flavonoids which can control the oxidation and peroxidation process of lipids, resulting in reduction of tissue damage and help to continue the blood flow in the kidney and improvement of kidney functionality (Rafieian-Kopaei et al., 2013).

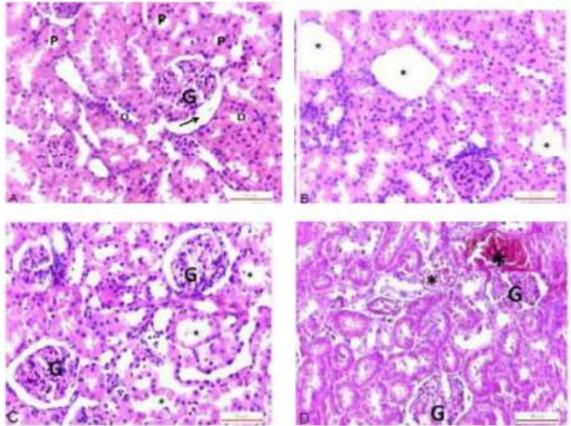


Fig. 1. Representative photographs of kidney sections stained by H&E(scale bar=50μm).(A) Control, (B) rats treated with CCl₄, (C) rats treated with *M. sylvestris* (150mg/kg b.w) + CCl₄ and (D) rats treated with *M. sylvestris* (M. sylvestris 300mg/kg b.w) + CCl₄

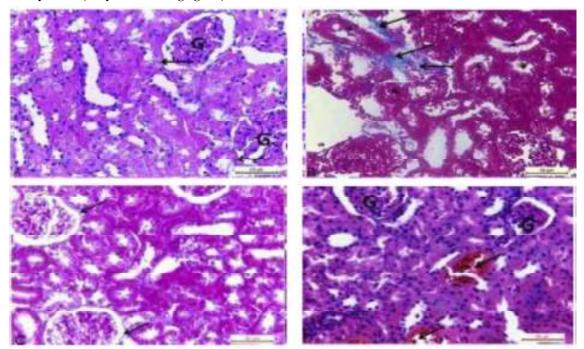


Fig. 2. Representative photographs of kidney sections stained by Masson trichrome (scale bar=50μm).(A) Control, (B) rats treated with CCl₄, (C) rats treated with *M. sylvestris* (150mg/kg b.w) + CCl₄ and (D) rats treated with *M. sylvestris* (*M. sylvestris* 300mg/kg b.w) + CCl₄

CONCLUSION

The current study indicates that M sylvestris supplements prevent biochemical change and pathological dissection caused by CCl₄. This renal protective effect of M. sylvestris can be ascribed to the presence of antioxidant contents, for example, phenol compounds and flavonoid that cause a significant reduction of the oxidative threat leading to a normal physiological function. The results support the use of M. sylvestris to treat kidney disease.

REFERENCES

- Abdel-Moneim A.E and El-Deib K.M.(2012). The Possible protective effects of Physalisperuviana on carbon tetrachloride-induced nephrotoxicity in male albino rats. *Life SciJ.*,9: 1038-1052.
- Adewole S.O.; Abdulkadir A.S.; Oladepo W.D and Thajasvarie N.(2007). Effect of Melatonin on Carbon Tetrachloride-Induced Kidney Injury in Wistar Rats. *African J. Biom. Res.*,10: 153–164.
- Alam A.M.; Quamri M.A; Siddiqui M.A; Hai U and Sofi G .(2016).Nephroprotective Effect and Unani Medicine: A Review. J. Nephrol Ther 6: 236.
- Allain C; Poon L.S; Chan C.S; Richmond W and Fu P.C. (1974).Enzymatic determination of Total Serum Cholesterol. *Clin. Chem.*,20: 470-475.
- Al-Mashhadani F.A.(2017).Effect of Fenugreek Seed and Leaves on Some Hematological and Biochemical Parameters in CCl4-induced Liver Injury. Int.J. Curr. Microbiol .App.Sci. 6(4): 2328-2337.
- Al-Seeni M.N; Haddad A.E;Mazin A.Z and Abeer M.A. (2016). The hepatoprotective activity of olive oil and Nigella sativa oil against CCl4 induced hepatotoxicity in male rats. BMC Complementary and Alternative Medicine., 16:438.
- Asuquo O.R; Ekanem T.B; Eluwa M.A; Oko O.O and Ikpi D.E.(2012). Evaluation of toxicological effects of Spondiasmombin in adult male Wistar rats J. Nat. Sci. Res., 2:144-51.
- Barros L; Carvalho A.M and Ferreira I.C. (2010).Leaves, flowers, immature fruits and leafy flowered ste M. sylvestris of *Malva sylvestris*: a comparative study of the nutraceutical potential and composition. *Food ChemToxicol* 48: 1466–1472.
- Carvalho A.M.(2005).EtnobotánicadelParque Natural de Montesinho.Plantas, tradición y saber popular en unterritorio del nordeste de Portugal. Universidad Autónoma, Madrid.
- Dacie J.V and Lewis SM (1991). Practical Haematology. 7th Edition, Churchil Livingstone, Edinburgh: 54-79
- De Melo G.O; Malva; D.C; Vanderlinde F.A; Rocha F.F; Pires P.A; Costa E.A; de Matos L.G; Kaiser C.R and Costa S.S. (2009). Antinociceptive and antiinflammatory kaempferol glycosides from Sedum dedroideum. J. Ethnopharmacology., 124: 228–232.
- DellaGreca M; Cutillo F; D'Abrosca B; Fiorentino A; Pacifico S and Zarrelli A.(2009).Antioxidant and radical scavenging properties of *Malva sylvestris*. Nat. Prod. Com. 4:893-896.
- Dumas B.T and Biggs H.G .(1972). In Slandered Methods of Clinical Chemistry 7: PP. Academic Press New York.
- Fawcett J and Scott J.A. (1960).rapid and precise method for the determination of urea. J ClinPathol. 13:156–9.
- Fossati P and Prencipe L. (1982). Serum triglycerides determined colorimetrically with an enzyme that produces hydrogen. *Clin Chem.*,28(10):2077-80.

- Fossati P; Principe L and Berti G .(1980). Use of 3,5dichlorohydroxybenzenesulfonic acid 4 aminophenazone chromogenic system in direct enzymatic assay of uric acid in serum and urine. *Clin Chem.*, 26:227–31.
- Frei B (1994). Am. J. Med. 97: 13-22.
- Funakoshi-Tago M; Nakamura K; Tago K; Mashino T and Kasahara T.(2011).Antiinflammatory activity of structurally related flavonoids, apigenin, luteolin, and fisetin. *International Immunopharmacology* 11:1150– 1159.
- Furfaro A.L ;Macay J.R; Marengo B; Nitti M ; Parodi A; Fenoglio D; Marinari U.M; Pronzato M.A; Domenicotti C; Traverso N .(2012).Resistance of neuroblastoma GI-ME-N cell line to glutathione depletion involves Nrf2 and heme oxygenase-1 Free Radic.*BiolMed.*,52:488– 496.
- Garcia-Nino W.R and Pedraza-Chaverri J. (2014).Protective effect of curcumin against heavy metals-induced liver damage .*Food Chem Toxicol* 69: 182–201.
- Gasparetto J.C; Martins C.A; Hayashi S.S; Otuky M.F and Pontarolo R.(2012). Ethnobotanical and scientific aspects of MS L.: a millennial herbal medicine. *Journal* of Pharmacy and Pharmacology.,64:172–189.
- Green L.C; Wagner D.A; Glogowski J; Skipper P.L; Wishnok J.S and Tannenbaum S.R.(1982). Analysis of nitrate, nitrite, and [15N] nitrate in biological fluids, *Analytical Biochemisty* 126: 131–138.
- Guarrera P.M.(2003).Food medicine and minor nourishment in the folk traditions of Central Italy (Marche, Abruzzo and Latium).*Fitoterapia*.,74: 515–544.
- Hamid M Abdulrahim Y; Liu D; Qian G; Khan A and Huang K.(2018). The hepatoprotective effect of seleniumenriched yeast and gum arabic combination on carbon tetrachloride-induced chronic liver injury in rats. J Food Sci.83(2):525-534
- Hasugawa T.M and Kuroda S .(1989). A new role of uric acid as an antioxidant in human plasma. Jap. J. Clin. Pathol. 37: 1020–27.
- Havel R.J.(1986).Functional activities of hepatic lipoproteins receptors. Ann Rev Physiol.,48:119–134
- Henry A.G and Piperno D.R. (2008).Using plant microfossils from dental calculus to recover human diet: a case study from Tell al-Raqa'i, Syria. *Journal of Archaeological Science.*, 35: 1943–1950.
- Hoeney M .(1985). Introduction to Clinical Immunology. Butterworth, London. 3.
- Hoitsma A.J; Wetzels J.F and Koene R.A. (1991). Druginduced nephrotoxicity.Aetiology, clinical features and management. *Drug Saf* 6: 131-147.
- Jain N.C. (1986) .Schalm's veterinary haematology 4th edition (ed N.C. Jain) Lea and Febiger, Philadelphia, 1221.
- Jha V; Garcia-Garcia G; Iseki K ;Li Z; Naicker S ; Rajiv S.M.D; Yee-Moon A.W.M.D and Chih-Wei Y.M.D. (2013).Chronic kidney disease: global dimension and perspectives. *Lancet.*, 382: 260-272.
- Kashinath R.T. (1990). Hypolipidemic effect of disulphide in rats fed with high lipids diets and/or ethanol. Ph.D. Thesis University of Bangalore, 221-225.
- Kengar S; Dattatray T and Jaywant J. (2017). Nephroprotective effect of amaranthusspinosus root extract in carbon tetrachloride-induced histological toxicity in male albino rat. *Int J. Drug Devand Res.*, 9:(2).5-7.
- Khan M.R; Rizvi W; Khan G.N; Khan R.A and Shaheen S. (2009). Carbon tetrachloride-induced nephrotoxicity in rats: Protective role of Digeramuricata. J. Ethnopharmacology., 122(1) 91-99.

- Kim E.O; Min K.J; Kwon T.K; Um B.H; Moreau R.A and Choi S.W.(2012). Anti-inflammatory activity of hydroxycinnamic acid derivatives isolated from corn bran in lipopolysaccharide-stimulated Raw 264.7 macrophages. *Food & Chemical Toxicology*.,50: 1309– 1316.
- Kleemann R; Verschuren L; Morrison M; Zadelaar S; van Erk M.J; Wielinga P.Y and Kooistra T.(2011).Antiinflammatory, anti-proliferative and antiatherosclerotic effects of quercetin in human in vitro and in vivo models. *Atherosclerosis.*, 218: 44–52.
- Majno G and Joris I. (2004).Cells, tissues and disease. principals of general pathology (2nd ed). Oxford University Press, New York, USA.
- Marsillach J; Camps J; Ferré N; Beltran R; Rul A; Mackness B; Mackness M and Joven J. (2009). Paraoxonase-1 is related to inflammation, fibrosis and PPAR delta in experimental liver disease. *BMC Gastroenterol.*, 9 (3):1-13.
- Mihara M and Uchiyama M (1978). Determination of malonaldehyde precursor in tissues by thiobarbituric acid test, *Analytical Biochemistry* 86: 271–278.
- Moon P.D; Lee B.H; Jeong H.J; An H.J; Park S.J; Kim H.R; Ko S.G; Um J.Y; Hong S.H and Kim H.M. (2007). Use of scopoletin to inhibit the production of inflammatory cytokines through inhibition of the IkB/NF-kB signal cascade in the human mast cell line HMC-1.*European Journal of Pharmacology* 555: 218–225.
- Noorah S .A and Mousa H.M.(2014). Ameliorative effect of olive leaf extract on carbon tetrachloride-induced nephrotoxicity in rats. *Life Science J*. 11(5):238-242.
- Nwidu L.L;Ekramy E ; Oboma I .Y and Wayne G. C .(2017).Hepatoprotective effects of hydromethanolic leaf and stem extracts of spondiasmombin in carbon tetrachloride-induced- hepatotoxicity and oxidative stress. J. Basic & Clin. Pharm. 8: S2 S0 11- S019.
- Ogbera A.O ;Dada O; Adeleye F and Jewo P. I. (2010). Complementary and alternative medicine use in diabetes mellitus. *West African Journal of Medicine.*, 29(3): 158-162.
- Ottu; O.J; Atawodi; S.E and Onyike E.(2013). Antioxidant, hepatoprotective and hypolipidemic effects of methanolic root extract of *Cassia singueana* in rats following acute and chronic carbon tetrachloride intoxication *Asian Pac. J. Trop. Med.* 6: 609–615.
- Passalacqua N.G; Guarrera P.M and De Fine G. (2007).Contribution to the knowledge of folk plant medicine in Calabria region (Southern Italy). *Fitoterapia.*,78: 52–68.
- Pydi R. (2011).Nephroprotective medicinal plants- A review. Int J Univ Pharm Life Sci., 1: 266-281.
- Rafieian-Kopaei M; Baradaran A; Rafieian M. (2013).Oxidative stress and the paradoxical effects of antioxidants. J Res Med Sci., 18(7):629.

- Rahmat A.A; Dar F.A; Choudhary I.M .(2014). Protection of CCl4-Induced Liver and Kidney Damage by Phenolic Compounds in Leaf Extracts of *Cnestis ferruginea* (de Candolle).*Pharmacognosy Res.*, 6: 19-28.
- Sahar; A.A. and Dalia H.A.A.(2014). The Protective Effect of Date Seeds on Nephrotoxicity Induced by Carbon Tetrachloride in Rats. Int. J. Pharm. Sci. Rev. Res., 26(12): 62-68.
- Satoh T; McKercher S.R; Lipton S.A. (2013).Nrf2/are-mediated antioxidant actions of pro-electrophilic drugs Free Radic. *BiolMed.*, 65: 645–657.
- Schermer S.(1967). In: Davis, F.A. (Ed.), The Blood Morphology of Laboratory Animals, third ed. Davis Co. Pub., Philadelphia, USA, pp. 10–42.
- Snedecor G.W and Cochran W.G.(1986). Statistical Methods, 7th Edition, Iowa
- State University Press, Ames, USA, Page 90.
- Sylvia O .I; Noghayin; E.J.O and Kingsley O.(2017). Pre-Exposure to Dennettiatripetala Ethanolic Fruit Extract Prevents Biochemical Alterations in Rats Subsequently Exposed to a Single Dose of Carbon Tetrachloride. International Journal of Pharmacology, *Phytochemistry* and Ethnomedicine., 6:8-16
- Szasz G; Borner U; Busch E.W and Bablok W.(1979). Enzymatic assay of creatinine in serum: comparison with Jaffe methods (author's transl), J. Clin Chem Clin Biochem 17: 683-687.
- Tilak K.S; Veeraiah K and Reju M.P. (2007). Effects of ammonia, nitrite, and nitrate on hemoglobin content oxygen consumption of freshwater fish, Cyprinuscarplo (Linnaeus) J. Envron. Biol. 28 (1): 45 – 47.
- Trinder P .(1969).Determination of blood glucose using 4amino phenazone as oxygen acceptor. J. ClinPathol. 22(2): 246.
- Wang H ;Naghavi M ; Allen C , et al . (2016). Mortality and Causes of Death Collaborators. Global, regional, and national life expectancy, all-cause mortality, and causespecific mortality for 249 causes of death, 1980-2015: a systematic analysis for the Global Burden of Disease Study 2015. Lancet .,388:1459–544.
- Wong F. and Salerno F.(2010). Beta-blockers in cirrhosis: friend and foe. *Hepatology* 52:811–813.
- Yoshioka T; Kawada K; Shimada T and Mori M. (1979). Lipid peroxidation in maternal and cord blood and protective mechanism against activated oxygen toxicity in the blood. Am. J. Obstet. Gynecol.,135:372-376.
- Zeynab K and Shereen B. G.(2012). Hepato-renal protection of silymarin in comparison with vitamin E in rats. *Global J. Pharmacology.*, 6 (3): 236-244
- Zöllner N and Kirsch K. (1962).Colorimetric method for determination of total lipids J. Experimental Medicine., 135: 545-550.

تأثير مستخلص الخبيزة على التسمم الكلوي الناجم عن CCl4 في الجرذان البيضاء هناء سالم صالح جازوي وماجدة عويس محمود قسم الكيمياء الزراعة –كليه الزراعة –جامعه المنيا –المنيا- مصر

يسبب التعرض لـ CCl التخليق المغرط لأنواع الأكسجين التفاعلية (ROS) في العديد من الأنسجة ، مثل الكلية. تؤدى أحيانا الشقوق الحرة إلى العجز الكلوي وربما الفشل الكلوي. الهدف من هذه الدراسة تقييم التأثير المحتمل لمستخلص الخبيزة (ROS) في العديد من الأنسجة ، مثل الكلية. تؤدى أحيانا الشقوق الحرة إلى أربع مجموعات متساوية. المجموعة ١: المجموعة الضابطة (الكنترول). المجموعة ٢: المجموعة ٢: المجموعة ١: المجموعة ٢: المجموعة ٢: المجموعة ٢: المجموعة ٢: المجموعة ٢: المجموعة ١: المجموعة ١: المجموعة ١: المجموعة ٢: المجموعة ١: المجموعة ٢: المجموعة ٢: المجموعة ١: المجموعة ١: المجموعة ١: المجموعة ١: المجموعة ٢: المجموعة ٢: المجموعة ٢: المجموعة ٢: المجموعة ١: المجموعة ١: المجموعة ١: المجموعة ١: المجموعة ٢: المجموعة ١: المحموعة ٢ المرضة وهى المعاملة بـ CCl (مل / كجم من وزن الجسم)، مرتين في الأسبوع لمدة ٨ أسابيع. المجموعتين ٣ و ٤: تم إعطانهم نفس الجرعة من ٢ إلى مستخلص الخبيزة بتركيز 100 أو ٢٠٠ ملجم / كجم من وزن الجسم يوميا ، على التوالي. وفي نهاية التجربة ، تم إجراء بعض التقديرات في الم والسيرم والكلى المرضاة إلى مستخلص الخبيزة بتركيز (100 / كجم من وزن الجسم)، مرتين في الأسبوع لمدة بـ الحال حدث لها زيادة عمل إلى مستخلص الخبيزة بتركيز (100 / عدم من وزن الجسم عنويا ، على التوالي. وفي نهاية التجربة ، تم إجراء بعض التقديرات في المو والسيرم والكلى والكن مستخلص الخبيزة بتركيز (100) معاملة بـ الحال الموالي والكلي والوريا والكريانينين)، والدهون الكلية، الكولسترول، الجلسيريدات الثلاثية ، والجلوكوزو (MDA) Malodialdehyde، واكسير النيتريك (NO) ، بينما انخفض معدل الزيادة في الوزن، كرات الدم الحمراء، PCV، حصن اليوريك ، والكتاليز (CAT) ، بالمعاولة بالحمولية ألك الموران، واكر نكريز والحرم إلى أورن راحس المعاملة بالخبيزة بتركيز ون معام المعاملة والدن المعاملة بالحمولي المالية الخبين المو من وزن الجسم) إلى أن معظم المعاملات البيوكيميائية والدموية المقاسة كانت قريبة من قيم المجموعة الصابطة (الكنترول). وأكد ذلك نتائج الفحص المعاملة والحرم المعاملة بالحموم المعاملة بالحموى الكليز ورحما لمعام ورن الخبين ورال معاملة بالخبيزة بتركيز (٢٠٥ م