SYMBIOTIC NITROGEN FIXATION PERFORMANCE SCREENING OFR. MELILOTI ISOLATES FROM SALINE SOIL AGAINST DIFFERENT VARIETIES OF ALFALFA PLANTS IN TWO TYPES OF SOIL

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ABSTRACT

Two laboratory and greenhouse experiments were conducted to study the efficiency of six Rhizobium meliloti isolates (2, 4, 11, 12 and 15) and strain ARC-1 isolated from wild alfalfa plants of salt affected soil to symbiotic performance with 5 varieties alfalfa (Medicago sativa) plants (Ismailia 1, Ismailia 2, Nitrogen fixers, Diablo and El-Wady El-gedid), in two soil types.

The results showed that, all isolates withstand high concentration of NaCl more than strain ARC-1. Isolate No.15 was the most resistant one to 7.5% NaCl and

different antibiotics concentration.

The data of symbiotic parameters showed that, the highest values of nodules number, nodules dry weight and N2-ase activity on plant roots were found with plants inoculated with strain ARC-1 in two soil types. Also isolate No. 12 recorded the highest shoots dry weight and N-content in two soils. The superior interactions were found between ARC-1 and Ismailia 1 for nodulation status and isolate No. 12 and Nitrogen fixers variety for shoot dry weight and N-content.

Keywords: Salinity, Alfalfa (Medicago sativa), Rhizobium meliloti. Symbiotic nitrogen fixation.

INTRODUCTION

About one-third of land area of the earth is subjected to arid and semi-arid climates and about 15% of the arid and semi-arid lands are saltaffected (Zahran, 1999).

Alfalfa (Medicago sativa) is among the commonly cultivated fodder legumes, which grown in large areas of arid and semi-arid environments, and its production magnificence is one of the major targets concerned by agronomists (Passarakli and Huber, 1991).

Rhizobia isolated from arid lands are capable to nodulate the legume host under saline condition (Douka et al., 1978).

Although high salt concentrations inhibit nodulation and No-fixation in many legumes, some varieties of legumes, e.g., Medicago sativa can successfully fix N2 under these conditions (Zahran, 1999). Level of salinity that affects the symbiosis between R. meliloti and Lucerne (Medicago sativa) are lower than those that affect the growth and survival of individual Lucerne genotypes or Rhizobium spp. (Mohammed et al., 1989).

Tolerance of the legume host to salt is the most important factor in determining the success of compatible Rhizobia strains to form successful symbiosis under high soil salinity conditions, therefore, we need to select Five selected bacteria isolates and a strain (ARC-1) were used in the experiment and uninoculated plants served as a control. All inoculated bacterial cultures were grown to maximum growth (ca.10⁹ cfu ml⁻¹). Inoculation was singly surface applied to soil as 10 ml culture pot⁻¹ once at planting and another 10 ml inoculated after seedling stage. After complete germination, plants were thinned to ten plants pot⁻¹. All pots were arranged in a complete randomized design with three replicates.

After 60 days of planting, plants were gently uprooted, nodules were counted and weighed. Shoot dry weight 10 plants was determined and analyzed for total nitrogen according to Page et al., (1982). Nitrogenase activity (N₂-ase) of plant roots was determined using the Acetylene Reduction Assay as described by Hardy et al., (1973), using Dani 1000 Gas Chromatography. Data were subjected to analysis of variance (ANOVA) according to the procedure of Snedecor and Cochran (1980).

RESULTS AND DISCUSSION

NaCi tolerance:

Nine isolates and one reference strain were tested for their tolerance to different NaCl concentrations. Table (3) showed that all isolates did tolerate high levels of salinity compared to reference strain (ARC-1) which tolerated 3.0% NaCl. This result agree with those obtained by Hua et al., (1982) and Jian et al., (1993) who reported that Rhizobia isolated from saline soils survive in inhibitory levels of salinity better than Rhizobia isolated from non saline soils. Isolate No. 15 was the most tolerant one, which tolerated 7.5% NaCl, whereas the salt tolerance for the other isolates was in the range of 4.5 to 6.0% NaCl. The majority of tested Rhizobia isolates (fast-growers) did withstand a concentration as high as 4.5% NaCl confirming the results of El-Sheikh and Wood (1990) that fast growing Rhizobia had higher salt tolerance rate than slow-growing (Bradyrhizobium).

Antibiotic resistance:

Results of resistance and sensitivity to different antibiotics among the examined Rhizobial isolates and (ARC-1) strain are illustrated in Table (4). Isolate 15 was highly resistant to all antibiotics examined, while the strain ARC-1 was sensitive to the most antibiotics, *i.e.* ampicillin, kanamycin and streptomycin and intermediate to bactriocin and erythromycin. Strain ARC-1 and isolate 15 exhibited similar behaviors against NaCl concentration test. These results are in accordance with those obtained by Zahran (1991) who reported that the response to salt and antibiotics stress in the *Rhizobium meliloti* might be genetically controlled.

Isolates 11, 12 and 14 were resistant to ampicillin, bactriocin and erythromycin, while were intermediate to kanamycin and streptomycin. In this concern, Somasegaran and Hoben (1994) and Anne et al., (2004) investigated the sensitivity to different antibiotics at different ranges of concentration varied between *Rhizobium* spp. and it was suggested that such variation may be a useful taxonomic character. So, antibiotic resistance

was used as one of the traditional techniques for identification at the species level and the genus level for bacteria (Nakayama, 1999).

Table (3): Minimum inhibitory concentration (MIC) of NaCl on R. meliloti isolates

150	lates								_	
NaCl conc.				l:	solates	5				Strain
(%)	2	3	4	7	8	11	12	14	15	ARC-1
Control	+	+	+	+	+	+	+	+	+	+
0.5	+	+	+	+	+	+	+	+	. +	, ±.
1.0	+	+	+	+	+	+	+	+	+) +
1.5	+	+	+	+	+	+	+	+	+	+
2.0	+	+	+	+	+	+	+	+	+	+
2.5	+	i + 1	+	+	+	+	+	+	+	+
3.0	+	+ ;	+	+	+	+) +	+	+	+
3.5	+	+	+	+	+	+) +	+	+	
4.0	+	+	+	+	+	+	+	+	+	-
4.5	+	+	+	+	+	+	+	+	+	-
5.0	+	-	+	+	+	+	+	+	+	-
5.5	+		-	- 1	+	+	+	+	+	-
6.0] -	- 1	-	-	-) +	+	+	+	
6.5	-	-	-	i -	_	-	} -	i -	+	} -
7.0		- 1	-	- 1	-] -	} -	(-	(+	} -
7.5	-			۱.	-	-] -	ł -	+	} -
8.0	-	-	-		-				<u> </u>	<u>-</u>

Table (4): Resistance and sensitivity of R. meliloti isolates to different antibiotics

	11(10101100				
	Ampicitiin	Bactriocin	Erythromycin	Калатусіп	Streptomycin (50 µg)
isolates	(10 µg)	(10 µg)	(15 µg)	(30 µg)	
		Mean diameter	er of the clear inh	nibition z <u>one (n</u>	nm)
ARC-1	35	10	18	21	41
Isolate 2	30	0.8	0.6	24	42
Isolate 3	15	0.6	0.6	22	25
Isolate 11	10	0.6	0.8	21	14
Isolate 12	10	0.6	0.5	16	13
Isolate 14	0.8	0.4	0.6	12	14
Isolate 15	0.4	0.0	0.0	1.0	<u>1</u> .1

Nodulation status and N2-ase activity:

Data presented in Table (5) show the effect of inoculation with five different *Rhizobium meliloti* isolates and one reference strain (ARC-1) on nodulation status of five varieties of alfalfa plants. Results showed that, in clayey soil, the greatest number of nodules (171.6) and dry weight of nodules (162.9 mg pot⁻¹) were obtained with plants inoculated with strain ARC-1 and isolate 12 which recorded (169.5 and 160.3 mg pot⁻¹) for number and dry weight of nodules, respectively. The fewer number and dry weight of nodules were found with plants inoculated with isolate 14 (127.4 nodules and 112.9 mg pot⁻¹). On the other hand, the superior interactions were found between strain ARC-1 and variety Ismailia 1 (189.7 nodules and 151.7 mg pot⁻¹) and isolate 12 and variety Ismailia 2 (187.7 nodules and 186.0 mg pot⁻¹) for number and dry weight, respectively.

The same trend was shown in sandy soil as in clayey soil especially, the behavior of strain ARC-1 (142.6 nodules and 110.6 mg pot⁻¹) and isolate 12(112.2 nodules and 107.5 mg pot⁻¹) for number and dry weight of nodules, respectively.

These findings were also observed by Zahran (1999) and Clen and David (2004) who reported that some legumes such as *Medicago sativa* produce indeterminate (meristematic) nodules that were more salt-tolerant than determinate (non-meristematic) ones formed on roots of other legumes. Furthermore, Rhizobia isolated from saline soil survive in inhibitory levels of salinity better than the other collected from soils with no salinity stress. Thus, salt tolerant Rhizobia efficiently colonize roots of leguminous plants grown in saline environments (Wall and Favelukes, 1991).

Regarding N_2 -ase activity, data in Table (6) declared that isolate 12 recorded the highest values of N_2 -ase activity to be 68.85 and 14.90 µmole C_2H_4 /mg of nodules hr⁻¹, in clayey and sandy soil, respectively. However, the lowest values of N_2 -ase activity was found in plants inoculated with isolate 2 which recorded 21.10 and 7.24 µmole C_2H_4 /mg of nodules hr⁻¹, in clayey and sandy soil, respectively.

Shoot dry weights and N-content:

Shoot dry weights and N-contents were significantly different depending upon both the R. meliloti isolates and alfalfa cultivars (Table, 7). In general, alfalfa plants grown in clayey soil produced significantly greater total dry weight and N-content in all treatments than those in sandy soil.

Among alfalfa varieties, Nitrogen fixers was found the most responsive one recording 5.55 g pot 1 for shoot dry weight and 17.62 mg pot 1 for shoot N-content in clayey soil and 3.18 g pot 1 and 9.12 mg pot 1 for shoot dry weight and shoot N-content in sandy soil, respectively. The lowest dry weight and N-content were obtained with uninoculated plants (control) were 3.74 g pot 1 and 8.99 mg pot 1 in clayey soil and 1.90 g pot 1 and 4.82 mg pot 1 in sandy soil, respectively.

On the other hand, isolate 12 supported the highest dry weight and N-content in alfalfa shoot cultivars to be 5.91 g pot⁻¹ for dry weight and 20.41 mg pot⁻¹ for N-content in clayey soil, respectively, and 3.13 g pot⁻¹ and 10.73 mg pot⁻¹ in sandy soil followed by strain ARC-1 which recorded 4.78 g pot⁻¹ and 21.14 mg pot⁻¹ in clayey soil and 3.08 g pot⁻¹ and 9.35 mg pot⁻¹ in sandy soil. These results agree with those obtained by Zahran (1999) and Pieter *et al.*, (2002) who reported that Rhizobia isolated from wild legumes of arid or saline lands might be superior to homologous strains of Rhizobia in effectively nodulating their legume host.

The superior interaction was found between isolate 12 and variety Nitrogen fixers which recorded 8.84 g pot⁻¹ for dry weight and 25.41 mg pot⁻¹ for N-content in clay soil and 3.77 g pot⁻¹ and 16.45 mg pot⁻¹ in sandy soil.

Levels of salinity that inhibit legumes-Rhizobia symbiosis differ from these harmful for growth of the individual symbionts. Legumes are generally more susceptible to osmotic stress than their specific microsymbionts (Kristin and Walker, 2001).

Table (5): Nodules number and dry weights of alfalfa cultivars inoculated with R. meliloti isolutes in clayey and sandy soils

			Num	ber of nod	Number of nodules (No. pot.1)	pot_)			Dry we	Dry weight of nodules (mg pot"	dules (mg	pot.)	
isolates	tes	٧,	V2	V3	٧4	۸۶	×	٧,	V2	٧3	۸	Vs	×
							Clayey soil	y soil					
Isolate 2		169.0	147.7	127.7	157.7	1727	154.9	170.0	140.0	117.0	108.7	174.7	142.1
Isolate 11		164.0	1650	136.0	1470	130.0	148.4	168 0	144.7	124 0	74.0	75.0	117.1
Isolate 12		173.0	187 7	166 7	1550	165.0	169.5	166.0	186.0	138.0	166.0	145.7	160.3
Isolate 14		100.7	132.7	98.7	142.7	162.0	127.4	100.0	122.7	106.3	104.7	131.0	112.9
Isolate 15		184 0	136.7	165.0	187 7	137.7	162 2	139.7	104.7	1490	146.0	154.0	138.7
ARC-1		189.7	184.0	155 0	162.7	166.7	171.6	151.7	173.3	159.0	161.7	168.7	162.9
Control		0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00
×		140.0	136.3	121.3	136 1	133.4		127.9	124.5	113.3	108.7	121.3	
	>			11.10						13.57			
L.S.D 0.05	lso			13 03				2002		16.06	,		
	V x Iso.			29.13		000		No. of the last of	Company of	35.91			
							Sand	Sandy soil					
Isolate 2		114.7	117.7	88.0	108.0	1167	109.0	83.7	149.7	69.7	94.0	95.7	98.6
Isolate 11		112.0	84.0	104.0	133.7	94.0	105.5	110.0	108.7	95.7	92 0	100.7	101,4
Isolate 12		107.0	141.0	72.0	97.0	144.0	112.2	103.0	132.0	65.0	113.7	124.0	107 5
tsolate 14		93.7	0.96	131.0	105.0	124 0	109.9	93.7	93.0	102.7	98.0	109.7	99 4
Isolate 15		103.0	0.69	134.0	64 7	76.0	89.3	92.0	7.97	106.7	59.0	73.7	81.6
ARC-1		153.6	160.0	147.0	147.7	104.7	142.6	129.7	85.7	120.7	124.0	93.0	110.6
Control	100	0.00	00.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00
×		97.9	95.5	96.8	93.7	94.3		87.4	92.3	80.04	82.95	85.24	
	>			N.S						8.36			
L.S.D 0.05	lso.			12.27						9.89			
	V x Iso.			27.44						22.12			
, · · · ·	V ₁ : Ismailia 1	V ₂ : Isı	V ₂ : Ismailfa 2	V ₃ : Nj	V ₃ : Nitrogen fixers	STS		V4: Diablo	0		Vs: El-Wad	Vs: El-Wadi El-gedid	

	N ₂ -ase a	ctivity µmo	N2-ase activity µmole C2H2 Img of nodules hr1	g of nodul	es hr.1	STATE OF THE PARTY OF	TO THE		25000		200	2000
Isolates	Clayey soil	oil	10.00		10 M	488	Sandy soil	oil	1000	03.0		8
	٧,	٧2	٧3	٧,	Vs	×	٧,	V2	٧,	۸,	Vs	×
Isolate 2	13.33	12.13	23.89	16.15	39.87	21.10	7.80	7.00	12.40	2.10	6.90	7.24
solate 11	46.83	10.94	70.61	54.58	31.58	42.91	6.20	3.75	2.38	11.89	13.90	7.60
Isolate 12	11.79	67.70	88.10	56.71	63.99	68.85	8.72	11.47	11.66	24.70	17.84	14.90
Isolate 14	22.97	49.13	38.10	24.10	70.00	40.86	17.00	99.6	6.27	6.31	4.32	8.71
Isolate 15	64.72	46.10	54.18	23.27	79.75	53.60	11.11	6.58	2.07	7.61	11.40	8.35
ARC-1	25.53	33.46	25.38	107.2	52.65	48.84	11.10	10.76	15.93	14.25	26.70	15.75
Control	0.00	0.00	00.00	00.00	00.00	0.00	0.00	0.00	0.00	0.00	00.00	0.00

Table (7): Shoot biomass and N-content of alfalfa plants inoculated with various R. mellioti Isolates in different two types of soil

			Dry w	Dry weight of shoot (g pot"	shoot (g	(, jod (Ż	N-content (mg pot	(mg po	()	
Isolates		, '×	V2	V3	٧4	Ve	×	٧,	V2	٧,	٧,	Vs	×
			n.	0.0	37		Clayey	y soil	2	rien.	6		
Isolate 2	2	.67	3.90	5.55	4.40	2.51	3.81	9.70	15.79	18.00	13.10	13.10	13.94
Isolate 11	<u>ښ</u>	39	4.29	6.40	2.83	3.39	4.10	17.43	15.58	12.63	13.56	96.6	13.84
Isolate 12	4	36	6.39	8.84	6.51	3.44	5.91	18.28	21.02	25.41	25.10	12.23	20.41
Isolate 14	m	69	2.81	5.36	6.35	3.45	4.33	4.36	18.19	19.64	20 85	10.74	14.76
Isolate 15	4	9.	4.60	3.63	5.95	3.37	4.23	11.58	14.51	12.10	18.13	11.22	13.50
ARC-1	رې ک	31	4.52	5.39	4.56	4.10	4.78	17.43	23.37	25.53	18.70	20.66	21.14
Control	ю	66.	3.33	3.71	4.40	3.27	3.74	5.89	6.98	10.32	13.26	8.52	8.99
×	ෆ	3.93	4.26	5.55	4.93	3.36	87.	12.10	16.49	17.62	.17.52	12.35	
			95.	0.147			28	55		0.586			
L.S.D _{0.05} Iso.	7			0.174			- 17	371		0.693			
	30.			0.389				610	6	1.550			
	_				1.3		Sand	Sandy soil	19	G-			
Isolate 2	2	54	3.40	2.25	3.53	2.10	2.76	6.94	10.40	8.16	7.89	6.12	7.90
Isolate 11	.2	36	3.47	4.30	2.93	1.73	2.96	9.13	10.86	3.31	7.21	6.85	7.47
Isolate 12	<u>ښ</u>	12	3.36	3.77	3.59	1.79	3.13	9.05	12.87	16.45	9.72	5.57	10.73
Isolate 14	2	-71	2.72	3.25	3.14	2.56	2.88	11.30	7.91	8.03	8.49	7.85	8.71
Isolate 15	<i>∠</i> i	14	2.46	2.62	1,39	2.49	2.29	5.60	6.51	8.34	9.36	4.72	6.91
ARC-1	w.	.20	3.21	3.43	3.54	2.01	3.08	8.71	7.99	12.20	10.83	7.04	9.35
Control	1	.71	2.30	2.62	1.74	1.13	1.90	4.14	5.71	7.35	3.74	3.15	4.82
×	2	2.58	2.98	3.18	2.84	1.97		7.83	8.89	9.12	8.18	5.90	
>			- 1	0.126	S 00 E		34			0.562			
L.S.D _{0.05} Iso.				0.149						0.665			
V x Iso.	Ö,			0.334						1.488			
V ₁ : Ismallía		V ₂ : Ismailia 2	ailia 2	V ₃ : Nit	V ₃ : Nitrogen fixers	ers		V4: Diablo	0		Vs: El-Wad	V ₅ : El-Wadi El-gedid	

REFERENCES

- Anne, M.A.; C. Teresa; H. Fasahath; H. Matthew and M. Rajeev (2004) Antibiotic-sensitive to 1C Mutants and their suppressers. J. Bacteriol., 186: 1851-1860.
- Atlas, R.M.; A.G. Brown; K.W. Dobra and L. Miller (1984). Experimental Microbiology: Fundamentals and Applications. Collier Macmillan, Canada, Inc.
- Clen, E. C. and H. K. David (2004). Sinorhizobium meliloti sulfotronsferase that modifies lipopolysaccharide. J. Bacteriol., 186: 4168-4176.
- Cordovilla, M.P.; A. Ocana; F. Ligero and C. Lluch (1995). Salinity effects on growth analysis and nutrient composition in four grain legumes-Rhizobium symbiosis. J. Plant Nut., 18: 1595-1609.
- Douka, C.E.; C.G. Apostalakis and V.D. Sharloy (1978). Studies of Rhizobium meliloti isolated from salt-affected soils. Ann. Appl. Biol., 88: 457-460.
- El-Sheikh, E.A.E. and M. Wood (1990). Effect of salinity on growth, nodulation and nitrogen yield of chickpea (*Cicer arientinum* L.). J. Exp. Bot., 41: 1263-1270.
- Hardy, R.W.F.; R.C. Burns and R.D. Holsten (1973). Application of the acetylene-ethylene assay for measurement of nitrogen fixation. Soil Biol. Biochem., 5: 47-81.
- Hua, S.S.T.; V.Y. Tsai; G.M. Lichens and A.T. Noma (1982). Accumulation of amino acids in *Rhizobium* sp. strain WR1001 in response to sodium chloride salinity. Appl. Environ. Microbiol., 44: 135-140.
- Jian, W.U.; Y. Susheng and Li. Jilun (1993). Studies on the salt tolerance of Rhizobium meliloti. Acta. Microbiol. Sin., 33: 260-267.
- Kristin, L. and C.W. Walker (2001). Genetic Analysis of the Sinorhizobium meliloti BacA Protein: Differential Effects of Mutations on Phenotypes. J. Bacteriol., 183: 6444-6453.
- Mohammad, M.; W.F. Campbell and M.D. Rumbaugh (1989). Acetylene reduction in salt-tolerant alfalfa and *Rhizobium*. Arid Soil Res. Rehabil, 3: 469-476.
- Nakayama, Y. (1999). Kanamycin resistant reporter gene. Bulletin of the National Institute of Agrobiological Resources, Kannondal, Tsukuba Ibaraki, 305-8602, Japan.
- Page, A.L.; R.H. Miller and D.R. Keeney (1982). "Methods of Soil Analysis". II- Chemical and Microbiological Properties. Soil Sci. Amer., Madison Wisconsin, USA.
- Passarakli, M. and J.T. Huber (1991). Biomass production and protein synthesis by alfalfa under salt stress. Plant. Nut., 14: 283-293.
- Pieter, V.D.; J.V. Pablo and T. Nicolas (2002). Effect of a *Sinorhizobium meliloti* strain with a Modified Put A gene on the rhizosphere microbial community of alfalfa. Appl. Environ. Microbiol., 68: 4201-4208.
- Piper, C.S. (1950). "Soil and Plant Analysis". 1st Ed. Interscience Publishers Inc., New York, 30-229.

Quinn, P.J.; M.E. Carter; B.K. Markey and J.R. Carter (1994). Clinical Vet. Microbiology, Wolf, Publishing, Tavistock, London, 220-242.

Snedecor, G.W. and W.G. Cochran (1980), "Statistical Methods", 7th Ed., Iowa

State Univ. Press, Amr., USA, pp. 255-269.

Somasegaran, P. and H.J. Hoben (1994). To Collect Nodules and Isolate Rhizobium. pp. 4-21. In: Methods in legume. Rhizobium Technology, United States Agency for International Development.

Vincent, J.M. (1970). "A Manual for the Practical Study of the Root-Nodule Bacteria. In: International Biological Programme, Handbook No. 15. Blackwell Scientific Publications, Ltd., Oxford and Edinburgh, U.K.

Wall, L.G. and G. Favelukes (1991). Early recognition in the Rhizobium meliloti-alfalfa symbiosis: Root exudate factor stimulates root adsorption of homologues Rhizobia. Bacteriol. J., 3492-3499.

Zahran, H.H. (1991). Conditions for successful Rhizobium-legume symbiosis

in saline environments. Biol. Fertil. Soils, 12: 73-80.

Zahran, H.H. (1999). Rhizobium-legume symbiosis and nitrogen fixation under severe conditions and in an arid climate. Microbiol. Mol. Biol. Rev., 968-989.

اختبار كفاءة تثبيت الأزوت تكافليا بين بعض عزلات ريزؤبيا البرسيم الحجازي المعزولة من أراضي ملحية في وجود بعض أصناف البرسيم المنزرعة في نوعين من التربة

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٢ - قسم بحوث العلف - معهد بحوث المحاصيل الحقلية - مركز البحوث الزراعية - الجيزة -

أجريت تجربتان معمليتان وكذلك تحربة أصص لاختبار كفاءة تثبيت النيتروجين لمد ٦ عزلات من ريزوبيا البرسيم الحجازي (٢ ، ٤ ، ١١ ، ١١ ، ١٠) بالاضافة الي ARC-1 والتي تم عزلها صدر نباتسات البرسيم الحجازى الذامية في أراضي ملحية و كذاك على أداء العلاقة التكافلية مع أصناف مختفة من البرسيد الحجازي (اسماعيلية ١ ، أسماعيلية ٢ ، مثبت للأزوت ، ديابلو ، الوادي الحديد) و ذلك باستخدام نو عين مسن النربة. وقد أوضحت النتائج ما يلى:

تمكنت جميع العزلات من تحمل العلوحة بدرجة أكبر من السلانة ARC-1 وكانت العزلة رقم ١٥ أكثر العزلات تحملا للملوحة (٧٠٥ كلوريد صوديوم) وكذلك تحملها للتركيزات المختلفة مسن المسضادات

كما أوضحت النقائج أن أعلى زيادة في عدد ووزن العقد الجذرية الجافة لنباتات النرسيم الحجازي وجدت عند تلقيح النباتات بالسلالة ARC-1. كما أن العزنة رقع ١٢ أعطت أعلى فيم للوزن الجاف للمجموع الخضري والمحتوى النيتروجيني للنباتات وكذلك نشاط أنزيم النيتروجينيز وذلك بالنسبة لمكلا أنسوعين مسن التربة المستخدمة في الدراسة.

كما أشارت النتائج الى أن أفضل التداخلات بين العزلات والأصناف كانت بين السلالة ARC-1 والصنف أسماعيلية ١ وذلك بالنسبة لعند العقد الجذرية ووزنها الجاف وانعزلة رقسم ١٢ والسصنف مثبت للأزوت بالنسبة للوزن الجاف للمجموع الخضري والمحتوي النيتروجيني.