

Efficacy of Some Ecofriendly Inducers in Controlling Barley Net Blotch

El-Nawawy, M. A.¹; Nabila A. Mosustafa² and Sherin Ph. Ibrahim¹

1- Food Sci. Dept., Faculty of Agriculture, Ain Shams University, Egypt.

2- Barley Disease Res. Department, Plant Pathology Research Institute, Agriculture Research Centre, Giza, Egypt.



ABSTRACT

The antifungal activity of resistance inducers; Mono potassium phosphate (KH_2PO_4), Chitosan clay Nano composite (ccnc), Humic acid (HA), Sodium meta silicate (Si) were tested in controlling *Pyrenophorateres* anamorph: *Drechslerateres* the causal organism of net blotch disease of barley. The evaluation was carried out at barley adult plant stages using two rowed hulled Giza 127, Giza 128 and six rowed hulls Giza 129 and Giza 130 in greenhouse and open field conditions at two locations; Itay El-Baroud and Sakha agricultural Research stations in Behaira and Kafr Elsheik Governorates, Egypt in 2013/14 growing season. Under artificial inoculation in greenhouse all concentrations of the four tested inducers were significantly reduced on disease severity (DS). Chitosan clay Nano composite (ccnc), Mono potassium phosphate (KH_2PO_4), Humic acid (HA) and Sodium meta silicate (Si) respectively compared with the control. The highest correlation was observed between DS and peroxidase, 1,3-glucanase after 15 days. Under field condition there was similar result of greenhouse. Spraying barley plants of the studied varieties with the tested inducers showed significantly correlation between disease severity and each of thousand kernel weight (TKW), plot weight, total protein, total lipids, total fiber, crude ash and total carbohydrates compared with the control. It is worth mentioning that six rowed hulls barley Giza 129 and Giza 130 were more responsive to the positive effects of inducers compared with two rowed hulled barley Giza 127 and Giza 128.

keywords: barley, *Pyrenophorateres*, antifungal activity, inducers, net blotch.

INTRODUCTION

Barley is the fourth most important cereal crop in the world (Poehlman, 1985). It has a long history of use as human food and animal feed, of health benefits and for malting and brewing industry in many countries around the world (Malcolmson, et al., 2005).

There are various forms of barley such as, hulls barley (a six-rowed), which is a good alternative to enrich food in developed countries, particularly due to its significant contents of beneficial substances, such as digestible fiber, micronutrients and other components that prevent chronic diseases (Steffenson, et al., 1991). Products with hulls barley, in comparison with products with pure wheat flour, have increased content of beta-glucans, i.e. by more than the fold. Hulled barley (two-rowed) is used mostly for malt production, Noworolnik, (2004) studied various forms of barley and indicated a definitely higher content of protein and a lower content of ash and crude fiber in hulls forms than in the hulled ones.

Net Blotch (BNB) caused by *pyrenophora teres* anamorph: *Drechslerateres*, is one of the major diseases of barley in Egypt and most of the cereal growing regions of the world. Under warm and humid conditions expression of barley net blotch (BNB) Tekauz, (2003) disease symptoms can increase rapidly, causing substantial grain yield loss 27% on an average and up to 34% when it is severe (Yitbarek and Wudneh, 1985). Typical yield losses due to net blotch nearing 100% in some highly susceptible barley cultivars. The disease is wide-spread in Northern Egypt, common in Middle Egypt while not detected in Assiout (Upper Egypt). The disease was also found to be of wide-spread along the northern coast. (Faten El-Nashar, 1983) Disease resistance has been a major strategy in controlling net blotch of barley. Several control methods against *P.teres* had been recommended, such as crop rotation, the application of fungicides and the use of resistant cultivars. The use of genetic resistance is the favoured for controlling this disease;

however; it is complicated by the existence of several pathotypes of the pathogen (Boungab, et al., 2012). An alternative control approach against net blotch would be based on economically and technically feasible and environmentally safe strategy. Many investigations have been made to understand the physiological and biochemical basis of induced systemic resistance (ISR). Much of this knowledge is due to the identification of a number of chemical and biological elicitors, some of them are commercially available for use in conventional Agriculture (Gary and Robert, 2004). Several natural and synthetic chemical agents have been described as activators of defense-related processes when applied to plants. Some of these activators may have potential application in Agriculture (Yamaguchi, 1998).

Monopotassium phosphate is the formulation with the lowest salt index and thus the foliar fertilizer of choice for many crops. It is an excellent and fast source of Potassium (K) when applied as a foliar fertilizer. Additionally, it controls powdery mildew in fruit trees and grapevines (Reuveni and Reuveni, 2001).

Application of humic acids (HA) has several benefits and agriculturists all over the world are accepting humic acids as an integral part of their fertilizer program. Humic acid is one of the major components of humus. Humates are natural organic substances, high in humic acid and containing most of the known trace minerals necessary to the development of plant life. Studies of the positive effect of humic substance on plant growth have demonstrated the importance of optimum mineral supply independent of nutrition. (Yildirim, 2007).

Chitin and chitosan are naturally-occurring compounds that have potential in agriculture with regard to controlling plant diseases. These molecules were shown to display toxicity and inhibit fungal growth and development. Fragments from chitin and chitosan are known to have eliciting activities leading to a variety of defense responses in host plants in response to microbial infections, including the accumulation of phytoalexins, pathogen-related (PR) proteins and proteinase inhibitors, lignin synthesis and callose formation

.Based on these and other proprieties that help strengthen host plant defenses, interest has been growing in using them in agricultural system to reduce the negative impact of diseases on yield and quality of crops . (El Hadrami *et al.*, 2010).Si can activate the expression of defense related genes and mayplay important role in the transduction of plant stress signal such as salicylic acid ,Jasmonic acid and ethylene (Kunzheng *et al.*, 2009).

The objectives of this study were to evaluated the response of some two – rowed (hulled barley) and six – rowed (hulls barley) genotypes to infection caused by *pyrenphora teres* ,and to estimate the improvement in of

yield component and chemical composition when some safe resistance inducers were used for controlling barley net Blotch diseases .

MATERIALS AND METHODS

1) Plant materials

The grains of Egyptian Barley (*HordiumVulgare*) cultivars (Giza 127 andGiza 128) as two rowed hulled, (Giza 129 and Giza 130) as six rowed hulls were obtained from Barley Research Section ,field cropInstitute, Agriculture Research Center (ARC) Giza , Egypt

Table (1) shows the pedigree of barley cultivars under investigation

No.	Cultivars	Pedigree
1	Giza 127	“w 12291’/’ Bags’// Harmal- 02 and ‘W 12291’/4’ 11012-2’/’70 -22425’/3’/’Apm’/’IB65’//’A16’
2	Giza 128	“w 12291’/’ Bags’// Harmal- 02 and ‘W 12291’/4’ 11012-2’/’70 -22425’/3’/’Apm’/’IB65’//’A16’
3	Giza 129	“DeirAlla 106/Cell//As46/Aths*2”
4	Giza 130	“Comp. cross 229//Bco Mr/DZ02391/3/DeirAlla 106”

2) source of four chemical inducers:

Mono potassium phosphate (KH₂PO₄) (50 & 100 µg/ml) , Chitosan clay nano composite (ccnc) (15 &30 mg/ml) , Humic acid (1.5 & 2.5 m/l) , and Sodium meta silicate (Si) (1&1.5g/l), were tested to evaluate their capabilities to induce resistance against net blotch (BNB) caused by *pyrenphora teres* anamorph : *Drechslera teres*.

Mono potassium phosphate (KH₂PO₄),Chitosan clay Nano composite and Humicacid,were obtained from central Laboratory of Organic Agriculture , Agriculture Res. Center ,Giza , Egypt

Sodium meta silicate (Si) was obtained from Sigma Aldrich Chemical Co. (St. Louis, MO, USA) .

The following experiments were conducted under greenhouse condition (artificial infection) and field conditions (natural infection)

3- Greenhouse experiments :

a-Isolation of the pathogen:

Samples of naturally infected barley leaves showing net blotch symptoms with necrotic lesions were collected from different locations. Small pieces (5mm) were cut from each sample and sterilized with sodium hypochlorite 1% for 1 min and dried between folds of sterilized filter papers and placed on potato dextrose ager plats (PDA) supplemented with streptomycin sulphate (100 mg/ml) . Petri dishes were incubated at 24°C for 48-72 hours. After 4 days incubation they were kept at 4 °C until use in further experiments.

Barley plants were grown in the clay pots (30 cm diameter) in a greenhouse at 20±2°C all the cultural practices applied . Adult barley plants(70 days old) with three replicates were assigned for each treatment were sprayed with one of the following chemical compounds : Mono potassium phosphate (KH₂PO₄) (50 &100µg/ml) , Chitosan clay Nano composite (ccnc)(15 &30 mg/ml) , Humic acid(1.5 & 2.5 m/l) , and Sodium meta silicate(Si) (1& 1.5 g/l) ,control plants were only sprayed with distilled water . five days after treatment , plants were inoculated with *Drechslera teres*.by spraying a suspension

of the fungus adjusted to a concentration of 2 × 10⁴ conidia / ml (Gupta et al., 2003) .

The inoculated plants were incubated in the dark for 48 h at 20±2C° and 100 % relative humidity ; they were then transferred to a growth chamber maintained at 20±2C° and 70-80 % RH with 12 hr daylight per day . Disease severity was determined as the percentage of infected leaf area 9 days after inoculation.

b-Disease assessments

Net blotch were carried out 12-14 d after inoculation (DAI). The net-like necrosis was assessed visually using the rating scale of Tekauz (1978) as illustrated in Figure

Inoculated barley plants were estimated and calculated using formula suggested by Townsend and Heuberger , (1943) as flows :

$$\text{Disease severity (\%)} = \frac{\sum (\text{NPC} \times \text{CR})}{\text{NIP} \times \text{MSC}} \times 100$$

Where : NPC = no. of plants in each class rate

CR = Class Rate

NIP = No. of tested plants.

MSC = Maximum severity class rate .

C. Determination of enzyme activities :

1. Peroxidase activity :

Leaf samples of two rowed and six rowed barley cultivars were used to determine the activity of peroxidise (PO),the extraction procedure was essentially based on the methods described by Biles and Martyn, (1993).Peroxidase activity was expressed as changes in absorbance min⁻¹g⁻¹ fresh weight at 425 nm by using spectrophotometer (Unico – 2100) .

2- Extraction and assay of β – 1,3 glucanase :

The method of Pan *et al.*(1991) was used to determine β – 1,3 glucanase activity . Laminarine was used as substrate and dinitro salicylic acid as reagent to measure reducing sugars .the method was carried out as 0.5 ml of enzyme extract was added to 0.5 ml of 0.05 M of potassium acetate buffer (pH5) containing 2 % laminarin . The mixture was incubated at 50° C for 60 minutes. The reaction was stopped by adding 1 ml of dinitrosalicylic acid reagent and heating the tubes for 5

minutes at 100° C. The tubes were cooled and 3 ml of distilled water were added before assay. The optical density was read at 500 nm. β - 1,3glucanase activity was expressed as mM glucose equivalent released /gram fresh weight tissues /60 minutes.

Field experiments

Field experiments were carried out under natural infection at experimental farms of Itay EL-Baroud (Behaira Governorate) and Sakha (KafrElsheik Governorate), in 2013/14 growing season. The experiment was conducted in a randomized complete block design (RCBD), with three replicates to each treatment. The plot size was 3 x3.5 m2 (1/400 Fadden) and each plot included 6 rows with 20 cm between rows. Each row was sown by 5g barley grains. The tested elicitors were prepared by mixing with sterile distilled water containing 0.01 % Tween 20 to obtain the desired final concentration. In control treatments, sterilized distilled water and Tween 20 were used instead of the inducers. The different treatments with four inducers were sprayed after 70 days from planting to determine the effect of different concentrations of four inducers while the control plots were sprayed with distilled water.

Yield components and chemical analyzed:

At maturity plants were harvested and Grain yield / plot (kg) and one thousand – kernel weight (TKW) (g) were determined. samples from different treatments to all varieties were chemically analyzed for protein, lipids, fiber, ash and carbohydrates according to the methods described in the A.O.A.C (2005).

Disease assessment of field :

The reaction of barley plants to net blotch was recorded at growth stage ; 10.5(Large, 1954) according to double – digit scale 00-99. The first digit gives the relative height of the disease, while the second digit shows the disease severity as a percentage of leaf area affected in terms of 0-9, where 0 = 0%, 2 = 20%, 30 = 3%, (Eyal et al., 1987).

Table (2) The observation and response of barley plants to net blotch infection at adultplant stage :

No.	Reaction	Observation	Response value
1	No disease	HR	00
2	Resistant	R	0.1
3	Resistant – Moderately resistant	R- MR	0.2
4	Moderately resistant	MR	0.3
5	Mod. Res. – Mod.Sucep.	MR – MS	0.4
6	Moderately susceptible	MS	0.6
7	Mod.Susc – Suscep	MS – S	0.8
8	Susceptible	S	0.9
9	Highly susceptible	HS	1.0

Statistical analysis

All experiments were carried out as a randomized complete block design with three replicates. Data were subjected to analysis of variance (ANOVA) by using MSTAT-C statistical package and the least significant differences (LSD) was used to compare means. Correlation analysis was performed with SPSS 6.0 statistical package.

RESULTS

1.Green house experiments

Anova of Table (3) showed highly significant effects for inducers, concentrations and varieties while all the interactions between concentration, variety and inducers were non Significant. Data in Table 4 show that the effect of spraying barley varieties with different inducers on net blotch disease severity. A significant reduction of the disease was recorded to all inducers and at higher concentrations. Chitosan clay was the best treatment in reducing net blotch severity followed by KH₂PO₄, Humic acid and Si respectively. Moreover, reduction of net blotch severity was higher on cultivars Giza 130 followed by Giza 129 (hulls barley) than varieties Giza 128 followed by Giza 127 and control.

Table (3) Analysis of variance of disease severity (DS) of net blotch disease when mono potassium phosphate (KH₂PO₄), Chitosan nanoparticles, Humic acid and Silicon were applied at three concentrations on four barley cultivars

Source of variation	D. F.	M.S.	F.VALU	P > F
Concentration (c)	2	4.709	20.0345	0.0000
Inducer (I)	3	1.480	6.2943	0.0006
C X I	6	0.488	2.0745	0.0632
Variety (v)	3	1.335	5.6784	0.0013
C X V	6	0.210	0.8933	
I X V	9	0.147	0.7408	
C X V X I	18	0.069	0.2919	
Error	96	0.235		

Table (4) Effect of spraying barley plants on severity of net blotch disease severity under greenhouse condition .

substance	Conc.	Disease severity %				General mean
		Giza 127	Giza 128	Giza 129	Giza 130	
KH ₂ PO ₄	50µg/ml	43.3	36.6	20.0	13.3	29.98
	100 µg/ml	30.0	16.6	13.3	10.0	17.48
	Mean	36.65	26.6	16.65	11.65	23.73
CCNC	15mg/ml	36.6	33.3	23.3	10.0	25.8
	30mg/ml	20.0	13.3	10.0	0.0	10.83
	Mean	28.3	23.3	16.65	5.0	18.32
Humic acid	1.5m/l	50.0	40.0	26.6	16.6	33.3
	2.5m/l	30.0	26.6	16.6	13.3	21.63
	Mean	40.0	33.3	21.6	14.95	27.47
Si	1.0 g/l	53.3	43.3	30.0	23.3	37.48
	1.5 g/l	56.6	30.0	23.3	16.6	31.63
	Mean	54.95	36.65	26.65	19.95	34.56
control		85.0	80.0	60.0	50.0	68.75
	Genral Mean	40.81	29.96	17.48	12.89	29.30

Activity of peroxidase and β 1-3 gluconseenzymes :

Four treatments were used to study their effect on peroxidase and β 1-3 gluconse activities which play a main role as plant defence mechanisms. Results in table(5 and 6) indicate that all treatment stimulated enzymes activity at second concentration after 15 day.

The highest activity was obtained with chitosan followed by KH₂PO₄ compared to other treatment. Meanwhile, in β 1-3 gluconse the highest activity was obtained with KH₂PO₄ followed by chitosan, Humic acid and silicon as compared with control.

Varieties Giza 130 and Giza 129 (hulls barley) were highly affected with enzymes activities than Giza 128 and Giza 127 (hulled barley).

Table (5) Effect of spraying different inducers on peroxidase activity of barley varieties (Ciza 127 ,Ciza 128 , Ciza 129 andCiza 130) under greenhouse conditions (artificial infection)

substance	Con.	*Peroxidase Activity %								Mean
		Giza 127		Ciza 128		Ciza 129		Ciza 130		
		3day	15 day	3day	15 day	3day	15 day	3day	15 day	
KH ₂ PO ₄	50µg/ml	0.65	1.160	1.28	2.45	1.79	2.020	1.85	1.99	1.64
	100 µg/ml	0.72	1.22	1.92	3.68	1.99	2.34	2.42	3.83	2.27
Mean		0.69	1.19	1.6	3.06	1.89	2.18	2.14	2.91	1.96
ccnc	15mg/ml	0.77	1.43	1.20	2.21	1.43	2.68	1.22	2.40	1.66
	30mg/ml	0.96	1.93	1.90	3.18	1.95	3.30	2.40	3.60	2.40
Mean		0.87	1.68	1.55	2.69	1.69	2.99	1.81	3.0	2.03
Humic acid	1.5m/l	0.55	0.97	0.69	1.09	0.98	1.05	0.52	1.80	0.95
	2.5m/l	0.81	1.57	0.89	1.44	1.12	2.04	0.98	2.02	1.35
Mean		0.68	1.27	0.79	1.26	1.05	1.55	0.75	1.91	1.15
Si	1.0 g/l	0.62	0.92	0.72	1.07	0.97	1.03	1.10	1.72	1.01
	1.5 g/l	0.89	1.24	0.85	1.40	1.12	2.02	1.90	1.40	1.35
Mean		0.76	1.08	0.79	1.24	1.05	1.53	1.5	1.56	1.18
control		0.45	0.80	0.58	0.9	0.65	0.81	0.72	0.98	0.73
LSD p ≤ 0.05		0.18	0.71	0.78	0.92	0.79	0.89	0.78	1.2	

*peroxidase activity expressed as absorbance/3.0 min 1.0 g fresh weight .

Table (6) Effect of spraying different inducers on β1-3 gluconase activity of barley varieties (Ciza 127 ,Ciza 128 , Ciza 129 ,Ciza 130) under greenhouse conditions (artificial infection)

substance	Con.	* β1-3 gluconaseActivity %								Mean
		Giza 127		Ciza 128		Ciza 129		Ciza 130		
		3day	15 day	3day	15 day	3day	15 day	3day	15 day	
KH ₂ PO ₄	50µg/ml	0.51	0.69	0.29	0.47	0.35	0.51	0.42	0.55	0.47
	100 µg/ml	0.713	0.93	0.55	0.65	0.48	0.68	0.52	0.79	0.66
Mean		0.61	0.81	0.42	0.56	0.41	0.59	0.47	0.67	0.56
ccnc	15mg/ml	0.20	0.41	0.18	0.43	0.22	0.44	0.33	0.49	0.33
	30mg/ml	0.32	0.55	0.45	0.60	0.35	0.66	0.45	0.68	0.50
Mean		0.26	0.48	0.31	0.51	0.28	0.55	0.39	0.58	0.42
Humic acid	1.5m/l	0.197	0.57	0.12	0.26	0.14	0.41	0.29	0.45	0.30
	2.5m/l	0.28	0.71	0.34	0.48	0.46	0.64	0.39	0.60	0.48
Mean		0.23	0.64	0.23	0.37	0.30	0.52	0.34	0.52	0.39
Si	1.0 g/l	0.12	0.32	0.08	0.25	0.22	0.34	0.25	0.35	0.18
	1.5 g/l	0.18	0.41	0.32	0.45	0.35	0.49	0.36	0.59	0.39
Mean		0.15	0.36	0.20	0.35	0.28	0.83	0.30	0.47	0.36
contro		0.10	0.09	0.21	0.18	0.18	0.23	0.11	0.14	0.15
LSD p ≤ 0.05		0.15	0.17	0.15	0.17	0.16	0.17	0.15	0.17	

*β – 1,3gluconase activity expressed as mM glucose equivalent released / gram fresh weight/60 min .

Data in Table 7 show that the disease severity observed between DS and β1,3-gluconase,peroxidase ,after (DS) was significantly and negatively correlated with each 15 day rating -0.531 and-0.351 respectively. of the biochemical traits . the highest correlation was

Table (7) Correlation between disease severity (DS) of net blotch disease and activities of peroxidase andβ1,3-gluconase under greenhouse condition (artificial infection)

Variable.	Variable			
	Peroxidase 3 d.	Peroxidase 15d	β1,3-gluconase3d	β1,3-gluconase15
Peroxidase 3 d.				
Peroxidase 15d	0.827** ^a			
β1,3-gluconase3d	0.509**	0.595**		
β1,3-gluconase15	0.509**	0.628**	0.841**	
D.S	-0.321**	-0.351**	-0.343**	-0.531**

^a(**) linear correlation coefficientis significant at p≤ 0.01

Field experiments :

Data in Table 8 showed highly significant correlation between disease severity and each of 1000 kernel weight , plot weight , total protein , total lipids , total fiber , crude ash and total carbohydrates . The highest negative correlation between disease severity and 1000 kernel weight , plot weight and total fiber which rating -0.648 , -0.493 and -0.362 respectively implies that the substance , which significantly decreased disease severity will induce significant increase in these trails .

Data Table 9 showed highly significant correlation between disease severity and each of 1000 kernel weight , plot weight , total protein , total lipids , total fiber , crude ash and total carbohydrates . The highest negative correlation between disease severity and 1000 kernel weight plot weight and total fiber which rating -0.716, -0.688 and -0.501 respectively implies that the substance , which significantly decreased disease severity will induce significant increase in 1000 kernel weight , plot weight and total fiber .

Table (8) Correlation between disease severity of net blotch , agronomic traits and technological traits under field conditions in Etay –el baroud location .

No.	variable	Variable						
		1000 Kernel weight	plot weight	total protein	total lipids	total fiber	crude ash	totalcarbo.
1	1000 kernel weight							
2	plot weight	0.539**a						
3	total protein	- 0.067	-0.435**					
4	total lipids	0.058	-0.011	0.558**				
5	total fiber	0.256**	-0.039	0.797**	0.836**			
6	crude ash	- 0.012	-0.253**	0.790**	0.721**	0.844**		
7	total carbohydrates	-0.138	-0.093	0.221**	0.819**	0.475**	0.498**	
8	disease severity	-0.648**	-0.493**	-0.061	-0.127	-0.362**	-0.035	0.112

a) linear correlation coefficient (r) is significant at $p \leq 0.01$ (**) or $p \leq 0.05$ (*) and $n=48$

The obtained results revealed that all the tested treatments reduced significantly disease severity compared to the control .(table 10 and 11) The second concentration showed the best effected against the disease . In this respect KH_2PO_4 was the most effective treatment , where it recorded the lowest degree of disease severity followed by chitosan , humic acid and Si .All tested genotypes showed also the highest degree

of disease reduction over the highest degree of disease reduction over the control , 64.2 – 81.5 % in Giza 127, 51.1 – 74.2 % Giza 128 45.0-86.0% in Giza 129 and 62.8 – 81.1% in Giza 130 at Etay El boardod location . Also , at Sakha, net blotch control over non- inducers applied treatment ranged from 63.6 – 85.4 % in Giza 127, 54.4 – 81.1% in Giza 128 , 37.5 – 85.0 % in Giza 129 and 53.3 – 80.0% in Giza 130.

Table (9) correlation between disease severity of net blotch , agronomic traits technological traits under field condition in Sakha location .

No.	variable	Variable						
		1000 Kernel weight	plot weight	total protein	total lipids	total fiber	crude ash	totalcarbo.
1	1000 kernel weight							
2	plot weight	0.739**a						
3	total protein	0.089	-0.005					
4	total lipids	0.183*	0.144	0.580**				
5	total fiber	0.355**	0.282**	0.789**	0.862**			
6	crude ash	0.053	0.004	0.784**	0.724**	0.835**		
7	total carbohydrates	-0.106	-0.171*	0.224**	0.807**	0.508**	0.502**	
8	disease severity	-0.716**	-0.688**	-0.400**	-0.263**	-0.501**	-0.243**	0.124

a) linear correlation coefficient (r) is significant at $p \leq 0.01$ (**) or $p \leq 0.05$ (*) and $n=48$

Table (10) Effect of spraying barley genotype with some inducers on disease severity plants cvs. (Giza 127, Giza 128) as two rowed hulled and (Giza 129, Giza130) as six rowed Hulled under field conditions during 2013/2014 growing season in Etay El boardod location

Sub.	Con.	Giza 127		Giza 128		Giza 129		Giza 130	
		DS*	Decrease%	DS	Decrease%	DS	Decrease%	DS	Decrease%
KH_2PO_4	50µg/ml	15.6	76.0	13.0	71.0	12.7	74.6	10.0	71.4
	100 µg/ml	12.0	81.5	11.6	74.2	7.0	86.0	6.6	81.1
CCNC	15mg/ml	16.6	74.5	15.0	66.6	13.3	73.4	11.0	68.5
	30mg/ml	13.3	79.5	13.0	71.0	7.8	84.4	7.2	79.4
humic acid	1.5m/l	20.0	69.0	17.2	61.7	27.0	45.0	12.0	65.7
	2.5m/l	15.0	76.9	15.6	65.3	16.7	66.6	8.4	76.0
Si	1.0 g/l	23.3	64.2	22.0	51.1	23.3	53.4	13.0	62.8
	1.5 g/l	17.6	72.9	20.0	55.5	17.0	66.0	9.3	73.4
control		65.0	-----	45.0	-----	50.0	-----	35.0	-----
LSD $p \leq 0.05$		2.3		2.3		2.3		2.3	

*DS : disease severity

Table (11) : Effect of spraying barley varieties with some inducers on disease severity plants cvs. under field conditions during 2013/2014 growing season in Sakha location

substance	Con.	Giza 127		Giza 128		Giza 129		Giza 130	
		*DS	Decrease%	DS	Decrease%	DS	Decrease%	DS	Decrease%
KH_2PO_4	50µg/ml	13.0	76.4	10.2	77.3	11.0	72.5	10	66.6
	100 µg/ml	8.0	85.4	8.5	81.1	6.0	85.0	6.0	80.0
CCNC	15mg/ml	15.0	72.7	13.5	70.0	13.5	58.8	11	63.3
	30mg/ml	9.0	83.6	9.5	78.8	8.0	80.0	7.9	73.6
humic acid	1.5m/l	20.0	63.6	19.4	56.8	23.0	42.5	13	56.6
	2.5m/l	12.0	78.2	11.2	75.1	16.0	60.0	9.9	67.0
Si	1.0 g/l	18.0	67.2	20.5	54.4	25.0	37.5	14	53.3
	1.5 g/l	14.0	74.5	13.5	70.0	18.0	55.0	11	63.3
control		55.0	-----	45.0	-----	40.0	-----	30	-----
LSD $p \leq 0.05$		1.8		1.8		1.8		1.8	

*DS : disease severity

2- yield component :

Data in Table 12 at Etay El board location show that all tested treatments significantly improved barley 1000 grain weight and plot weight over the control. KH₂PO₄ treatment was more effective in increasing crop parameters followed by chitosan ,humic

acid and Si , respectively . The second concentration showed the best effected on the crop parameters compared to the control . On the other hand genotypes Giza 129 and Giza 130 (Hull- less) barley were the most improving in 1000 grain weight and plot weight than genotypes Giza 128 and Giza 127 (Hulled barley) .

Table (12) : Effect of inducers on yield components on Giza 127 and Giza 128 , Giza 129 and Giza 130 under field conditions during 2013/2014 growing season in Etay El board location .

substance	Con.	Giza 127				Giza 128			
		1000 Grain Weight (g)	Increase %	Plot Weight (kg)	Increase %	1000 Grain Weight (g)	Increase %	Plot Weight (kg)	Increase %
KH ₂ PO ₄	50µg/ml	51.9	21.1	4.5	45.2	52.8	20.5	6.8	78.9
	100 µg/ml	57.3	34.5	4.9	58.1	58.3	33	7.5	97.4
CCNC	15mg/ml	51.3	20.4	4.3	38.7	51.6	17.8	5.9	55.3
	30mg/ml	56.3	32.2	4.8	32.3	57	30.1	6.3	65.7
humic acid	1.5m/l	50.7	19	4.1	32.3	50.8	15.9	5.8	52.6
	2.5m/l	55.9	31.2	4.4	41.9	56.9	29.9	6.1	60.5
Si	1.0 g/l	50.2	17.8	3.9	25.8	50.5	15.3	5.5	44.7
	1.5 g/l	55.5	30.3	4.3	38.7	55.2	26	5.9	55.2
control		42.6	-----	3.1	-----	43.8	-----	3.8	-----
LSD p ≤ 0.05		2.4		0.059		2.4		0.06	

Table (12) con.

substance	Con.	Giza 129				Giza 130			
		1000 Grain Weight (g)	Increase %	Plot Weight (kg)	Increase %	1000 Grain Weight (g)	Increase %	Plot Weight (kg)	Increase %
KH ₂ PO ₄	50µg/ml	50.1	42.3	6.7	86	51.4	32.1	4.8	71.4
	100 µg/ml	55.2	56.7	7.6	111	57.35	47.4	5.7	103.6
CCNC	15mg/ml	48.6	38	6.2	72.2	49.6	27.5	4.2	50.0
	30mg/ml	54.17	53.8	6.7	86	55.8	43.4	4.7	67.9
humic acid	1.5m/l	46.9	33.2	6.1	69.4	45.9	17.9	3.8	35.7
	2.5m/l	50.9	44.6	6.4	77.7	53.2	36.7	4.5	60.7
Si	1.0 g/l	45.9	30.4	5.1	41.7	45.2	16.2	3.1	10.7
	1.5 g/l	48.6	38	5.7	58.3	52.6	35.2	4.0	42.9
control		35.21	-----	3.6	-----	38.9	-----	2.8	-----
LSD p ≤ 0.05		2.4		0.06		2.4		0.059	

Results in Table 13 at Sakha location with minor differences , where genotypes Giza 129 and Giza 130 were the most improving in 1000 grain weight than cvs.Giza

127 and Giza 128 . In contrast , improving plot weight was highly ranking in cvs.Giza 127 and Giza 128 than cvs.Giza 130 and Giza 129 compared to the control .

Table (13) : Effect of inducers on yield components on Giza 127 and Giza 128, Giza 129 and Giza 130 under field conditions during 2013/2014 growing season in Sakha .

substance	Con.	Giza 127				Giza 128			
		1000 Grain Weight (g)	Increase %	Plot Weight (Kg)	Increase %	1000 Grain Weight (g)	Increase %	Plot Weight (kg)	Increase %
KH ₂ PO ₄	50µg/ml	51.7	24	4.9	105	56	30.2	4.9	81.5
	100µg/ml	65	56	5.8	152	59.3	37.9	5.5	103.7
CCNC	15mg/ml	50	19.9	3.9	52.6	55	27.9	4.1	51.8
	30mg/ml	62	48.7	4.4	78.9	58.7	36.5	5.1	88.8
humic acid	1.5m/l	49	17.5	3.7	42.1	46.7	8.6	3.9	44.4
	2.5m/l	60	43.9	4.1	63.2	56.7	31.9	4.9	81.5
Si	1.0 g/l	46.7	12	3.3	21.1	45.7	6.3	3.1	14.8
	1.5 g/l	53.3	27.8	4	57.8	55.7	29.5	4.1	51.8
control		41.7	-----	2.9	-----	43	-----	2.7	-----
LSD p ≤ 0.05		1.7		0.03		1.7		0.03	

Table (13) : con.

substance	Con.	Giza 129				Giza 130			
		1000 Grain Weight (g)	Increase %	Plot Weight (Kg)	Increase %	1000 Grain Weight (g)	Increase %	Plot Weight (kg)	Increase %
KH ₂ PO ₄	50µg/ml	55	38	6.9	81.6	63.6	47.9	7.3	69.8
	100 µg/ml	66.7	67	7.9	107.8	66.7	55.1	7.9	83.7
CCNC	15mg/ml	51.7	29	6.3	65.8	63.3	47.2	6.9	60.5
	30mg/ml	65	63	6.8	78.9	65.6	52.6	7.1	65.1
Humic acid	1.5m/l	50	25	6	57.9	53.3	23.9	6.1	41.9
	2.5m/l	56.7	42	6.6	73.7	60	39.5	6.7	55.8
Si	1.0 g/l	43.3	8	4.9	28.9	51.3	19.3	5.9	37.2
	1.5 g/l	55	15	5.6	47.4	58.3	35.6	6.3	46.5
control		40	-----	3.8	-----	43	-----	4.3	-----
LSD p ≤ 0.05		1.7		0.03		1.7		0.03	

DISCUSSION

Application of Mono potassium phosphate KH_2PO_4 (50 & 100 $\mu\text{g}/\text{ml}$), Chitosan clay nanocomposite (ccnc) (15 & 30 mg/ml), Humic acid (1.5 & 2.5 m/l) and Sodium meta silicate (Si) (1 & 1.5 g/l) reduced the injurious effect of net blotch (BNB) on barley where Mono potassium phosphate and Chitosan clay Nano composite were the most effective while Humic acid was moderately effective.

Data of the present study showed that, under greenhouse and field conditions all treatments and their concentrations were significantly reduced disease severity and consequently improving the chemical composition of grains and yield component in barley varieties as compared to the untreated control. In this research Chitosan clay nano composite and KH_2PO_4 treatment showed the higher efficacy in reducing net blotch disease may be due to increasing the activities of peroxidase and β -1,3glucanase moreover it is associated with increase in activities of many class of PR proteins (Abou-Taleb, 2001 and Mosa, 2002).

Glucanase are known to play a direct role in plant defence mechanism by hydrolyzing structural components from fungal cell wall and act as elicitors which lead the accumulation of a phytoalexin and glyceollin (Sharp *et al.*, 1984). Also, peroxidase play important role in oxidation process of phenol compounds to oxidized products (quinones) which may limit the fungal growth. peroxidase produce free radicals and hydrogen peroxide which are toxic to many microorganisms (Pena and Kue, 1992).

The 1000 Kernel weight is a highly important parameter for yield and technological quality in barley. Przulj and Momcilovic (2006) Data in this study showed that all treatment improving crop parameters, In this respect KH_2PO_4 following chitosan were the most effective treatment to increase 1000 kernel weight and plot weight where efficacy of KH_2PO_4 against plant infection to trigger plant defense through a process involving the consequent of calcium from host issue, elicits the release of signal triggers of plant response and makes the plant more resistance and more responsive after subsequent infection (Gottstein and Kuc, 1989, Mucharromah and Kuc, 1991). Moreover KH_2PO_4 led to increase synthesis of host metabolites such as phytoalexins and their production could be induced by many chemicals and increase of enzymes activities and total phenols (Nighat *et al.*, 2011).

chitosan also used in plant disease control as a powerful elicitor. Its direct toxicity remains dependent on properties. chitosan is known to act as potent inducers, enhancing a battery of plant responses both locally around the infection sites and systemically to alert healthy parts of the plant. These include early signaling events as well as the accumulation of defense-related metabolites and proteins such as phytoalexins and PR-proteins (El Hadramiet *et al.*, 2009 and Hammerschmidt, 1999). It can be formation of physical barrier preventing the pathogen from spreading and

invading other healthy tissues (El Hadramiet *et al.*, 2009). Chitosan is applied to plants to prevent diseases because it can chelate nutrients and minerals (i.e., Fe, Cu) preventing pathogens from accessing them (Bornet, 2007). Its activity of peroxidase and β -1,3 glucanase which realized in this study by increasing the activity of both enzymes due to chitosan treatment, it is agreement with El Hadrami *et al.*, (2010) who found that, chitosan elicited peroxidase and polyphenoloxidase activities and increased the level of phenolic compounds.

Data obtained showed that there were negative correlation between disease severity and 1000 kernel weight, plot weight and total fiber where varieties hullless barley Giza 129 and Giza 130 were moderate resistance to net blotch than hulled barley Giza 127 and Giza 128 thus the highest value of these traits were in hullless barley than hulled barley, so the yield components depend on disease severity and varieties, this finding is in harmony with Gaunt (1980) who described that, the yield components affected by *Pyrenophorateres* depend on severity and duration of disease on barley in addition to complex physiologic process of the host. Similar results have been demonstrated early by Jordan (1981) who reported that, satisfactory yield can still be obtained with genotypes possessing in complete resistance.

A previous study showed that six-rowed genotype may possess higher level of resistance than two-rowed. This similarly with Tekauz, (2002) who reported that six-rowed genotypes are more resistant than two-rowed varieties.

The opinion of many authors that the chemical composition of barley grain is modified not only by weather conditions but also it depend on the plant genotypes Noworolink (2004).

Chitosan following by humic acids (HA) were the most effective for improving chemical composition as total fiber. where efficacy of humic acid (HA) in overcoming the harmful effects of barley net blotch (BNB) plant may be due to the increase in chitinase activity and stimulation plant growth through increased cell division, as well as optimized uptake of nutrients and water (Atiyeh *et al.*, 2002 and Chen *et al.*, 2004) also, regulate hormone level, improve plant growth and enhance stress tolerance (Piccolo *et al.*, 1992). HA is a suspension, based on potassium humates, which can be applied successfully in many areas of plant production as a plant growth stimulant or soil conditioner for enhancing natural resistance against plant diseases and pests (Scheuerell and Mahaffee, 2006) which consequently increase yield of plant. Foliar application of (HA) consistently enhanced antioxidants such as α -tocopherol, β -carotene, superoxide dismutases, and ascorbic acid concentrations in turf grass species (Zhang, 1997). These antioxidants may play a role in the regulation of plant development, flowering and chilling of disease resistance (Ziadi *et al.*, 2001).

finally ,In short , use of chemical inducers ,as safe substance , may improve yield component and reduce disease severity of net blotch in barley plants .

REFERENCES

- Abou- Taleb, M.A.(2001). Biochemical changes associated with the application of some resistance – inducing compounds for controlling powdery mildew of cucumber. Egypt J. Appl.Sci., 16 :387-405 .
- A .O . A .C (2005) . Official methods analysis of Association of Official Agriculture Chemists . 18th End Washing D. C.
- Atiyeh , R.M.; Lee, S.; Edwards, C.A.; Arancon , N.Q. and Metzger, J.D. (2002). The Influence of humic acids derived from earthworm proceed organic wastes on Plant growth . BioresourceTechnology , 84: 7-14 .
- Biles , C.L. and Martyn , R.D. (1993) . Peroxidase ,polyphenoloxidase and shikimate Dehydrogenozymes in relation to tissue type, maturity and pathogen induction of Watermelon seeding . Plant physiol .Biochem., 31 : 499-506.
- Bornet , A. T .(2007). Chitosan , chitin – glucan and chitin effect on minerals (iron , lead , cadmium) and organic (ochratoxin A) contaminants in wines . Eur. Food Res. Technol.226(4), 681-689.
- Boungab,K; Belabid, L. ; Fortas, Z.; Bayaa ,B. (2012).Pathotype diversity among Algerian isolates of *Pyrenophora teres f . teres* .Phytopathol .Medit ., 5 (3), 577-586
- Chen, Y., Nobili ,M. De and Aviad, T. (2004) . Stimulatory effect of humic substances on plant growth. In: Magdoff F, Weil RR, editors. Soil organic matter in sustainable agriculture . Boca Raton:CRC Press : 103-130
- El Hadrami , A.; Lorne R. A ; El Hadrami , I. and Fouad ,D. (2009) . Suppression of induced plant defense responses by fungal pathogens. In Molecular – Plant Microbe Interactions ;Brisson , N., Daayf, F., Eds; CABI: Wallingford , UK, Chapter 10 , pp. 231-268.
- El Hadrami , A.; Lorne R. A ; El Hadrami , I. and Fouad, D. (2010).Chitosan in plant protection .Marine Drugs ,8 : 968-987 ..
- Eyal , Z; Scharen ,A.L.; Prescott, J . M. and Van Cinkel, M .(1987). The septoria disease of wheat; concepts and methods of disease management . Mexico D.F.,CIMMYT, PP.46 .
- Faten El-Nashar , K. (1983) . Net blotch disease of barley caused by *Drechslera teres*(Sacc.) Shoem . M.Sc. Thesis , Fac. Of Agric. Cairo Univ.
- Gary , E . V. and Robert , M .G.(2004) Systemic Acquired Resistance and Induced Systemic Resistance in Conventional Agriculture. Crop Sci.,44: 1920- 1934
- Gaunt , R . E.(1980) . Physiological basis of yield loss. Pages 98-111 in : proceeding of E.C Stakman Commemorative Symposium on Assessment of Losses which constrain production and crop improvement in Agriculture and Forestry . Univ. Minn. Agric. Exp . Stn. Misc. Publ.7.
- Gottstein , H.R. and Kuc, J. (1989) . Induction of systemic resistance to anthracnose in cucumber Byphosphales. Phytopathology, 79: 176 – 179 .
- Gupta, S .and loughman, G.J .(2003) . RCM Lance . Aust. J. Agr. Res., 54,1379-1386.
- Hammerschmidt, R.(1999) .Phytoalexins : what have we learned after 60 years ? Ann Rev. Phytopathol. 37, 285-306 .
- Jordan , V. W. L (1981) .A etiology of barley net blotch caused by *pyrenophora teres* and some effects on yield . Plant Pathology 30: 77-87.
- KunzhengCai, D. G. and Shiming,L (2009) . Probing the mechanisms of silicon – mediated pathogen resistance .Plant Signal Behav.4 (1) :1-3 .
- Large, E.C.(1954). Growth stages in cereals .IIIustration of the Feek'sScale . Plant Pathol.,3:123-129 .
- Mosa , A.A. (2002). Induced resistance in rice against blast disease using abiotic and biotic agents. Ann. Agric. Sci.Ain Shams Univ.,47:993-1008.
- Malcolmson , L., Nowkirk , R. and Carson , G. (2005) . Expanding opporatunities for barley food and feed through product innovation .feed and quality ; 18th National American Barley Research Workshop 4th Canadian Barley symposium . pp : 2-4 .
- Mucharromah , E. and Kuc, J. (1991) . Oxalate and phosphates induced systemic resistance against Disease caused by fungi , bacteria and viruses in cucumber. Crop Protection , 10:265-270.
- Nighat , S ; Zahid , M.H; Ashfaq , S. and Jamil , F.F.(2011). Induced systemic resistance in Chickpea against Ascochyta blight by safe chemicals . Pak. J.Bot., 43(2):1381-1387.
- Noworolnik, K. (2004).nitrogen fertilization efficiency of buckwheat grown at various soil conditions . the 6th international symposium on buckwheat.Current Advances in Buckwheat Research , 601-604.
- Pan, S.Q., Ye, X.S., and Kuc, J. (1991) .Association of $\beta - 1,3$ glucanase activity and isoform Pattern with systemic resistance to blue mold in tobacco induced by stem injection with *Peronosporatabacina*or leaf inoculation with tobacco mosaic virus. Physiol . Mol. PlantPathol., 39:25-39.
- Pena , M. and Kuc, J . A.(1992) . Peroxidase – generated hydrogen peroxidase as a sources of antifungal activity in vitro and on tobacco leaf disk .Phytopathology , 82: 696 – 699 .
- Piccolo, A.; Nardi, S. and Concheri , G. (1992) .Structural characteristics of humic substances As regulated to nitrate uptake and growth regulation in plant system. Soil Biochem ., 24: 373-380 .

- Poehlman , J. M .(1985) . Adaptation and distribution. Pp.1-17. In: Barley .Rasmusson , D. C.(ed.). Agronomy Monograph No. 26, ASA . Madison. WI.
- Przulj,N.andMomcilovic,V.(2006).Barly breeding for yieldand quality.GlasnikZastiteBilja., 29: 1, 49-57.
- Reuveni, M. andReuveni, R., (2001). Monopotasssium Phosphate fertilizer (peak)—a component in integrated Control of powdery mildews in fruit trees and grapevines . Acta. Hort., 594: 619- 625.
- Scheuerell , S.J. and Mahaffee, W.H. (2006).Variability associated with suppression of graymould (*Botrytis cinerea*) on geranium by foliar applications of no aerated and aerated compost Tekas. Plant Dis. , 90: 1201 – 1208
- Sharp , J. K., Valent , B. and Albersheim, P. (1984). Purification and partial characterization of a β – glucan fragment that elicits phytoalexin accumulation in soybean . J. Biol .chem. 259, 11312 - 11320.
- Steffenson, B. J., Webster, R. K., and Jackson , L.F. (1991) . Reduction in yield loss using incomplete resistance to *Pyrenophorateresf.teres*in barley plant Dis. 75:96-100
- Tekauz A (1978): Incidence and severity of net blotch of barley and distribution of *Pyrenophora teres* biotypes in the Canadian prairies in 1976.*Canadian Plant Disease Survey* 58, 9-11.
- Tekauz, A (2002).Spot blotch(*Cochlibolussativus*) infection response in selected North American barley cultivars at the seedling and adult- plant stages . Pp. 428-435.In: Proceedings of the Second International Workshop on Barley Leaf Blight . ICARDA , Aleppo, Syria .
- Tekauz, A (2003). Spot blotch caused by *Bipolaris sorskiniana* : an emerging problem in barley In western Canada. In : proceedings of 8thInternational Congress of plant Pathology Christchurch, New Zealand .Pp. 103.
- Towsend , G. K. and Heuberger , T.W. (1943). Methods for estimating losses caused by disease experiments . Plant Dis. Rept., 27: 340-343.
- Yildirim,E., (2007). Foliar and soil fertilization of humic acid affect productivity and quality of Tomato.Acta Agric. Scandinavia . Section B. Soil and Plant Sci., 57 (2): 182-186.
- Yitbarek , S.and Wudneh ,E. . (1985). Preliminary studies on the yield losses due to net blotch in Barley ,pp 47-52 In Proceedings of the 10th EPC annual meetings . Addis Ababa. Ethiopia .
- Yamaguchi, I. (1998) . Activators of systemic acquired resistance . In: Fungicidal activity: Chemical and biological approaches to plant protection .Hutson D.H. Miyamoto J., eds. New York :Wileyev& s611s Inc., pp. 193-219 .
- Zhang , X. (1997) . Influence of plant growth regulator on turfgrass growth antioxidant status ,And drought tolerance . Ph.D. Thesis, Fac. Of Virginia polytechnic, 131p.
- Ziadi , S.; Barbedette , S., Godard, J.F.; Monoti, C.; Corre , L.E.D.; Silue, D. and Lecorre, D. (2001) . Production of pathogenesis related protein in the cauliflower (*BrassicaoleaceaVar. botrytis*) downy mildew (*Peronosporaparasitica*) pathosystem treated with Acidbenzolar -5- methyl. Plant Pathol.50 (5):579-586.

فاعلية بعض مستحضات المقاومة الصديقة للبيئة في مقاومة مرض التبغ الشبكي في الشعير

محمد عبد الرازق النواوي¹, نبييلة احمد مصطفى² و شرين فيليب²

1- قسم علوم الاغذية – كلية الزراعة – جامعة عين شمس - مصر

2- قسم بحوث امراض الشعير – معهد بحوث امراض النبات – مركز البحوث الزراعية – الجيزة - مصر

تناول هذا البحث دراسة تأثير اربعة مستخلصات وهم احادى فوسفات البوتاسيوم والكيوتوزان وحمض الهيومك والسليكات وذلك لتقليل الضرر الناتج عن الاصابة بمرض التبغ الشبكي المتسبب عن الفطر دريشسليرا تيرز لنوعين مختلفين من الترا كيب الوراثية وهي اصناف عارية (ذو ستة صفوف) مثل جيزة 129 وجيزة 130 واصناف مغطاة (ثنائية الصفوف) مثل جيزة 127 وجيزة 128 فى محطتى بحوث ايتاى البارود وسخا للموسم الزراعى 2013 / 2014. وجد تحت ظروف العدوى الصناعية بالصوبة أن هناك نقص معنوى فى شدة الاصابة نتيجة المعاملة رشاً بالكيوتوزان واحادى فوسفات البوتاسيوم وحمض الهيومك والسليكات على الترتيب مقارنة بالغير معاملة . كما وجد أن الرش بالمستحضات المختلفة ادى الى زيادة فى نشاط انزيمى البيروكسيداز وبيتا 1 - 3 جلوكانيز . وتحت ظروف العدوى الطبيعية بالحقل ، كانت هناك نتائج متشابهة مع تجارب الصوبة . فقد وجد أن المعاملة بهذة المستحضات أدت الى خفض فى شدة الاصابة وزيادة فى مكونات المحصول (وزن الالف حبة – وزن الحوض) وكذلك تحسين فى مكونات الحبة مثل البروتينات والدهون والالياف والرماد والكربوهيدرات مقارنة بالكنترول (الغير معاملة) كما وجد أن الاصناف العارية اكثر قابلية للتحسن مقارنة بالاصناف المغطاة . لذا ننصح باستخدام احادى فوسفات البوتاسيوم ، الشيتوزان ، هيومك اسد ، والسيلكون لمقاومة التبغ الشبكي نظرا لفاعليتها وكونها مواد صديقة للبيئة .

