

HPLC DETERMINATION OF AMOXICILLIN, AMPICILLIN, PHENOXYMETHYL PENICILLIN, DICLOXA CILLIN, CLOXA CILLIN AND DICLOXA CILLIN IN PHARMACEUTICAL PREPARATIONS

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ABSTRACT

A simple high performance liquid chromatographic procedure was described for the determination of amoxicillin, ampicillin, cloxacillin, dicloxacillin, flucloxacillin and phenoxyethylpenicillin (penicillin V) in single or combined pharmaceutical formulations. The drugs were chromatographed on an Ultrasphere ODS column and (40% methanol containing 9 mM sodium heptane sulphonate, 30 mM potassium dihydrogen phosphate and 0.04% triethylamine) adjusted to pH 5 with 1 M phosphoric acid as mobile phase using penicillin G as internal standard. The flow rate was 1 ml min⁻¹ at 230 nm detection. The results show good accuracy as shown by the % recovery of 99.61, 99.53, 99.9, 100.08, 100.18 and 99.90% and high precision as shown by the relative standard deviation RSD 0.744, 0.613, 0.557, 0.731, 0.46 and 0.30%

INTRODUCTION

Amoxicillin and ampicillin are broad spectrum antibiotics effective against many gram positive and gram negative organisms. Cloxacillin, dicloxacillin and flucloxacillin are isoxazolylpenicillin resistant to penicillinase produced by staphylococci.

A co-formulation of amoxicillin and flucloxacillin combines the complementary activities of both to produce a broad spectrum bactericidal activity against a wide range of gram positive and gram negative pathogens. Also a co-formulation of ampicillin with cloxacillin or dicloxacillin extends the antibacterial spectrum to include a wide range of gram positive and gram negative organisms. Phenoxyethylpenicillin is an acid stable penicillin and used mainly in the treatment of streptococcal infections and in rheumatic fever prophylaxis.

Concerning the analysis, iodometric (1-4), potentiometric (5-7) spectro-photometric (8-10), HPTLC⁽¹¹⁾ and HPLC⁽¹²⁻¹⁶⁾ methods were reported for determination of the above penicillins in pharmaceutical preparations. The spectrophotometric and potentiometric assays are often very difficult to carry out and of low sensitivity owing to interference of other ingredients which are generally present in the pharmaceutical preparations. In the present research we introduce a new HPLC method for the determination of amoxicillin, ampicillin, cloxacillin, as well as dicloxacillin, flucloxacillin, and phenoxyethylpenicillin in different pharmaceutical preparations containing these mixtures.

EXPERIMENTAL

Apparatus:

LDC analytical, 4100 solvent delivery system, Milton Roy, Variable Wave length Detector (VWD).

Reagents

Methanol, HPLC grad, BDH, England. Water, double distilled, Lepco, Egypt. Potassium dihydrogen phosphate, Potassium Hydroxide, Triethylamine and Heptane sulphonate, Fluka, Switzerland. O-Phosphoric acid, Riedel de Haen, Germany.

Stock solutions:

Amoxicillin, ampicillin, cloxacillin and flucloxacillin, 1 mg ml⁻¹, Ribbon, Italy. Penicillin G, 2 mg ml⁻¹ and dicloxacillin, 1 mg ml⁻¹, CID, Egypt. Phenoxyethylpenicillin, 0.615 mg ml⁻¹, The Nile Co., Egypt. All of the standard solutions were dissolved in water except phenoxyethylpenicillin dissolved in methanol.

Authentic mixtures of:

Amoxicillin 1 mg ml⁻¹ and flucloxacillin 1 mg ml⁻¹. Ampicillin 1 mg ml⁻¹ and cloxacillin 1 mg ml⁻¹. Ampicillin 1 mg ml⁻¹ and dicloxacillin 1 mg ml⁻¹.

Pharmaceutical preparations:

Amplelox 500 mg capsules, from MUP, Egypt, labelled to contain 250 mg ampicillin as ampicillin trihydrate and 250 mg cloxacillin as cloxacillin sodium, B.N. 96198, average weight of capsule content 620 mg. Cloxapen 500 mg capsules, from The Nile Co., Egypt, labelled to contain 250 mg ampicillin and dicloxacillin sodium equivalent to 250 mg dicloxacillin, B.N. A.22, average weight of capsule content 566 mg.

Dipenacid 500 mg capsules, from CID, Egypt, labelled to contain 250 mg ampicillin as ampicillin trihydrate and 250 mg dicloxacillin as dicloxacillin sodium, B.N. 985101, average weight of capsule content 586.2 mg. Dipenacid 500 mg vials, from CID, Egypt,

labelled to contain 250 mg ampicillin as ampicillin sodium and 250 mg dicloxacillin as dicloxacillin sodium, B.N. 1294/93A, average weight of vial content 420 mg.

Plumox 500 mg capsules, from Lepien, Egypt, labelled to contain 250 mg amoxicillin as amoxicillin trihydrate and 250 mg flucloxacillin as flucloxacillin sodium monohydrate, B.N. 9814/9, average weight of capsule content 402 mg. Plumox 1 g vials, from Lepien, Egypt, labelled to contain 500 mg amoxicillin as amoxicillin sodium starch and 500 mg flucloxacillin as flucloxacillin sodium starch, B.N. 9807/8, average weight of vial content 1240 mg.

Osopen 1000 tablets, from the Nile Co., Egypt, labelled to contain 1,000,000 IU phenoxymethyl penicillin, B.N. 9/47, average weight of tablet 760 mg.

Chromatographic conditions :

Mobile phase 40% methanol containing (9 mM-sodium heptane sulphonate, 30 mM-potassium dihydrogen phosphate and 0.04% triethylamine) adjusted to pH 3 with 1 M-phosphoric acid, degassed and filtered using 0.45 μ m membrane filter.

Column 5 μ Ultrasphere ODS (2.5) x 4.6 mm)

Detector 230 nm, 0.2 a.u.f.s

Flow rate 1 ml min⁻¹, Pressure 1500 psig

Temperature ambient, Injected volume 10 μ l

Procedures:

A-Preparation of calibrations curves :

From the standard solution of each drug volumes from 0.2-1 ml were pipeted into 10 ml volumetric flasks, then 0.5 ml of the internal standard was added to each flask and completed to volume with water. 10 μ l of each sample was injected into the column and all measurements were repeated three times at each concentration. The calibration curve was based on the peak area ratio of each compound to that of the internal standard against concentration.

B-Authentic mixtures :

Different aliquots from the authentic mixture stock solutions prepared in ratios claimed to be present in the pharmaceutical preparations within the range recorded in the calibration were transferred to 10 ml volumetric flasks and completed as described under procedure A.

C-Commercial dosage forms :

1- Capsules and Tablets :

The contents of 10 Ampiclox, Cloxapen, Dipenacel, Plumox capsules and Osopen tablets were weighed and the average weight of each was determined, mixed and powdered. An accurately weighed portions of the powders equivalent to 1/10 the average weight of each product were transferred to

100ml volumetric flasks, dissolved in water (except osopen which dissolved in methanol) with the aid of an ultrasonic bath, completed to volume and filtered. The first portion of the filtrate discarded.

Different aliquots from the filtrate were transferred to different 10 ml volumetric flasks and completed as under procedure A.

2- Vials :

The contents of 10 Dipenacel or Plumox vials were weighed and the average weight of each was determined and mixed. An accurately weighed portions of the powders equivalent to 1/10 the average weight of each product were transferred to 100 ml volumetric flasks and dissolved in water, completed to volume and filtered. The first portion of the filtrate discarded. Different aliquots from the filtrate were transferred to 10 ml volumetric flasks and completed as under procedure A.

RESULTS AND DISCUSSION

A mixture containing amoxicillin, ampicillin, cloxacillin, penicillin G, penicillin V, flucloxacillin and dicloxacillin was chromatographed on an Ultrasphere ODS column. A number of mobile phases composed of methanol - phosphate buffer - heptane sulphonate were tested in the presence and absence of triethylamine. The mobile phase composed of 40% methanol containing 9mM-sodium heptane sulphonate and 30 mM-potassium dihydrogen phosphate were failed to separate cloxacillin and flucloxacillin (Fig. 1A).

The separation was improved by increasing the morality of heptane sulphonate but the peaks become more tailed (Fig. 1B). The tailing was overcome by addition of triethylamine. It was found that the mobile phase composed of 40% methanol containing (9mM-sodium heptane sulphonate, 30 mM-potassium dihydrogen phosphate and 0.04% triethylamine) adjusted to pH 5 with 1 M-phosphoric acid flowing at 1ml min⁻¹ would result in a good baseline and symmetrical peaks with good capacity and resolution factors.

Complete analysis of the seven component mixture was achieved within 25 minutes (Fig. 1C). The retention times were 2.61, 4.55, 5.98, 9.51, 14.29, 15.64 and 24.91 minutes for amoxicillin, ampicillin, penicillin G, phenoxymethylpenicillin, cloxacillin, flucloxacillin and dicloxacillin, respectively. The internal standard was chosen on the basis that it should be eluted between the peaks and should not interfere with any of the previously mentioned components. Penicillin G was found to be the most suitable internal standard with a retention time of 5.98 minutes.

The peak area ratio of amoxicillin, ampicillin, penicillin V, cloxacillin, flucloxacillin and dicloxacillin

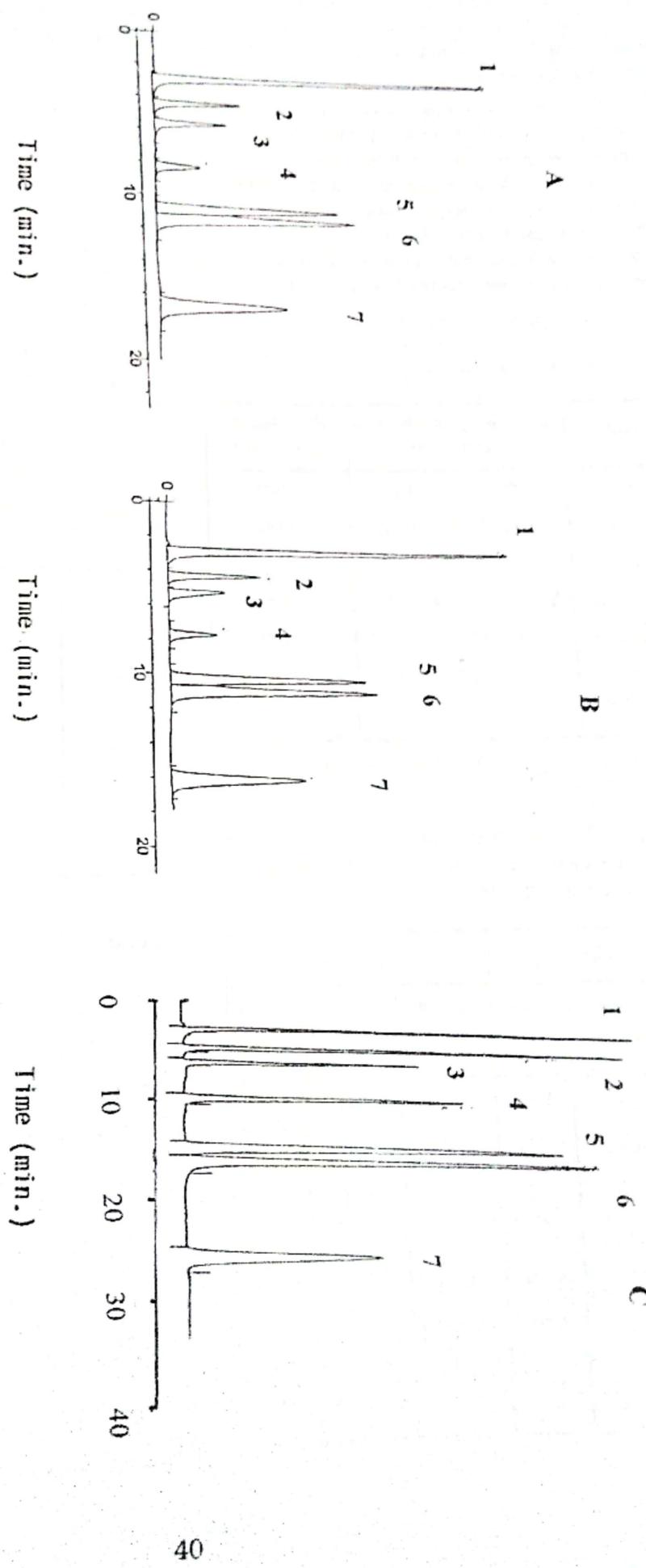


Fig. 1 Effect of molar concentration of heptane sulphonate sodium salt in the mobile phase on the separation of 1- amoxicillin 2-ampicillin 3- penicillin V 4-penicillin G (internal standard) 5- cloxacillin 6- flucloxacillin 7- dicloxacillin.

Mobile phase : aqueous methanol 40% containing 30mM KH_2PO_4 and 6 nM heptane sulphonate

adjusted to pH 5 with potassium hydroxide

aqueous methanol 40% containing 30mM KH_2PO_4 and 7 mM heptane sulphonate

adjusted to pH 5 with potassium hydroxide

aqueous methanol 40% containing 30mM KH_2PO_4 , 9 mM heptane sulphonate and

0.04 % triethylamine adjusted to pH 5 with o- phosphoric acid

(A).
(B).

0.04 % triethylamine adjusted to pH 5 with o- phosphoric acid

(C).

Column

5 μ Ultrasphere C18, 250 \times 4.6 mm

Detector

UV at $\lambda = 230$

to that of penicillin G (the internal standard) exhibit rectilinear relationship within concentration range 2-10 mg% for amoxicillin, ampicillin, cloxacillin, flucloxacillin and dicloxacillin and 1.23 - 6.15 mg % for phenoxymethylpenicillin.

The regression equations and correlation coefficients (*r*) are given in table 1. The validity of the listed regression equations was tested by the assay of authentic penicillin V or a laboratory made authentic mixtures containing known quantities of amoxicillin and flucloxacillin, ampicillin and cloxacillin, ampicillin and dicloxacillin in ratios equivalent to those found in commercial dosage forms, tables 2, 3, 4 and 11.

Table (1): Regression analysis

Compound	Regression equations	Correlation coefficient (<i>r</i>)
Amoxicillin	$Y = 0.418 X + 0.1192$	0.999
Ampicillin	$Y = 0.219 X - 0.006$	0.999
Cloxacillin	$Y = 0.481 X + 0.022$	0.9999
Dicloxacillin	$Y = 0.453 X + 0.052$	0.999
Flucloxacillin	$Y = 0.392 X + 0.0029$	0.999
Phenoxy-methylpenicillin	$Y = 0.285 X + 0.003$	0.999

Y=Peak area ratio

X= Concentration (mg %)

Table (2) Determination of amoxicillin and flucloxacillin in authentic mixture using the proposed HPLC method.

Amoxicillin			Flucloxacillin		
Amount added, mg%	Amount found, mg%	Recovery %	Amount added, mg%	Amount found, mg%	Recovery %
2	2.01	100.5	2	1.98	99
5	4.962	99.24	5	5.01	100.2
10	9.91	99.1	10	9.94	99.4
Mean		99.613	Mean		99.533
SD		0.771	SD		0.611
RSD		0.774	RSD		0.613

The results show good accuracy as shown by the % recovery 99.61, 99.53, 99.9, 100.08, 100.18 and 99.908 for amoxicillin, flucloxacillin, ampicillin, cloxacillin, dicloxacillin and phenoxy-methylpenicillin, respectively and high precision as shown by the relative standard deviation, RSD 0.744, 0.613, 0.557, 0.731, 0.46 and 0.308 for amoxicillin, flucloxacillin, ampicillin, cloxacillin, dicloxacillin and phenoxy-methylpenicillin, respectively.

The proposed method was applied to assay commercial tablets, vials and capsules as in Figs. (2-6). Also results from tables 5, 6, 7, 8, 9, 10 and 12 show good accuracy and precision.

Table (3): Determination of ampicillin and cloxacillin in authentic mixture using the proposed HPLC method.

Ampicillin			Cloxacillin		
Amount added, mg%	Amount found, mg%	Recovery %	Amount added, mg%	Amount found, mg%	Recovery %
2	2.01	100.5	2	2.015	100.75
5	4.97	99.4	5	5.01	100.2
10	9.98	99.8	10	9.93	99.3
Mean		99.8	Mean		100.08
SD		0.556	SD		0.73
RSD		0.557	RSD		0.73

Table (4) Determination of ampicillin and dicloxacillin in authentic mixture using the proposed HPLC method.

Ampicillin			Dicloxacillin		
Amount added, mg%	Amount found, mg%	Recovery %	Amount added, mg%	Amount found, mg%	Recovery %
2	1.98	99	2	2.0044	110.22
5	5.01	100.2	5	5.031	100.62
10	9.97	99.7	10	9.97	99.7
Mean		99.63	Mean		100.18
SD		0.602	SD		0.461
RSD		0.604	RSD		0.46

Table (5) Determination of amoxicillin and flucloxacillin in Flumox capsules using the proposed HPLC method.

Amoxicillin			Flucloxacillin		
Amount added, mg%	Amount found, mg%	Recovery %	Amount added, mg%	Amount found, mg%	Recovery %
2	2.02	101	2	1.964	98.2
5	4.953	99.06	5	4.91	98.2
10	9.96	99.6	10	9.78	97.8
Mean		99.88	Mean		98.066
SD		1	SD		0.23
RSD		1	RSD		0.235

Table (6) Determination of amoxicillin and flucloxacillin in Flumox vials using the proposed HPLC method.

Amoxicillin			Flucloxacillin		
Amount added, mg%	Amount found, mg%	Recovery %	Amount added, mg%	Amount found, mg%	Recovery %
2	2.04	102	2	2.05	102.5
5	5.15	103	5	5.2	104
10	10.04	100.4	10	10.25	102.5
Mean		101.8	Mean		103
SD		1.31	SD		0.866
RSD		1.29	RSD		0.84

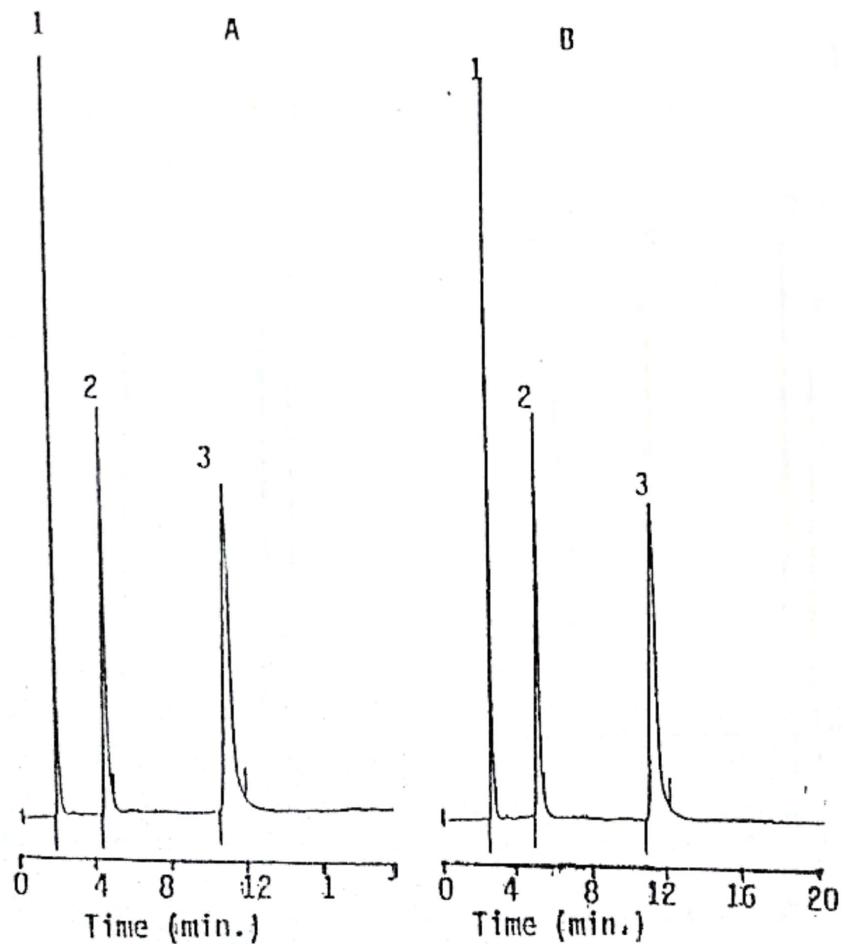


Fig. (2) HPLC Chromatogram of Flumox Vials (A) and Flumox Capsules (B).

1- Amoxicillin

2- Penicillia G

3- Flucloxacillin.

Table (7) Determination of ampicillin and cloxacillin in Ampiclox capsules using the proposed HPLC method.

Ampicillin			Cloxacillin		
Amount added, mg%	Amount found, mg%	Recovery %	Amount added, mg%	Amount found, mg%	Recovery %
2	1.99	99.5	2	2.03	101.5
5	2.028	100.56	5	5.16	103.2
10	9.97	99.7	10	10.12	101.2
Mean		99.2	Mean		101.96
SD		0.563	SD		1.078
RSD		0.563	RSD		1.057

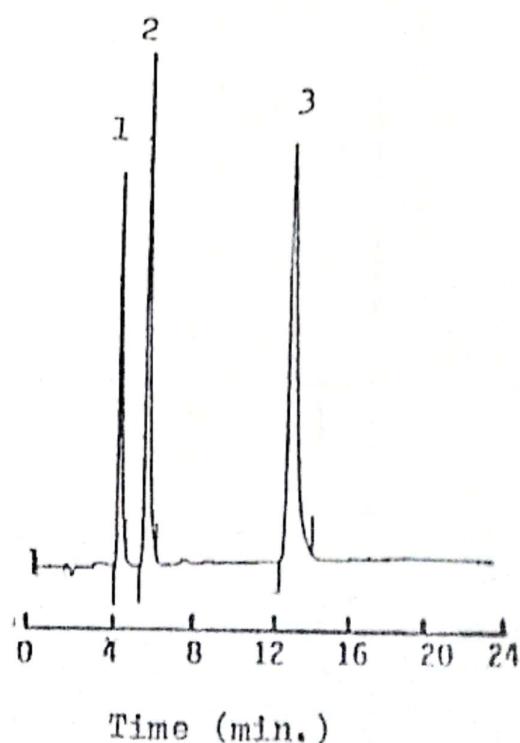
Table (8) Determination of ampicillin and dicloxacillin in Cloxapen capsules using the proposed HPLC method.

Ampicillin			Dicloxacillin		
Amount added, mg%	Amount found, mg%	Recovery %	Amount added, mg%	Amount found, mg%	Recovery %
2	1.95	97.5	2	1.94	97
5	4.9	98	5	4.81	96.2
10	9.74	97.4	10	9.61	96.1
Mean		97.63	Mean		96.43
SD		0.32	SD		0.49
RSD		0.32	RSD		0.51

(A)



(B)

**Fig. (3)** HPLC Chromatogram of authentic mixture of ampicillin and cloxacillin (A) and Ampiclox capsules (B).

1-Ampicillin

2-Penicillin G

3-Cloxacillin

Table (9) Determination of ampicillin and dicloxacillin in Dipenacid capsules using the proposed HPLC method

Ampicillin			Dicloxacillin		
Amount added mg%	Amount found mg%	Recovery %	Amount added mg%	Amount found mg%	Recovery %
2	1.96	99	2	1.96	99
5	4.98	99.2	5	4.92	98.4
10	9.88	98.1	10	9.78	97.8
Mean		98.76	Mean		98.066
SD		0.986	SD		0.305
RSD		0.99	RSD		0.311

Table(10) Determination of ampicillin and dicloxacillin in Dipenacid vials using the proposed HPLC method

Ampicillin			Dicloxacillin		
Amount added mg%	Amount found mg%	Recovery %	Amount added mg%	Amount found mg%	Recovery %
2	1.97	98.5	2	1.95	97.50
5	4.9	98	5	4.86	97.2
10	9.71	97.1	10	9.71	97.10
Mean		97.9	Mean		97.266
SD		0.655	SD		0.208
RSD		0.67	RSD		0.214

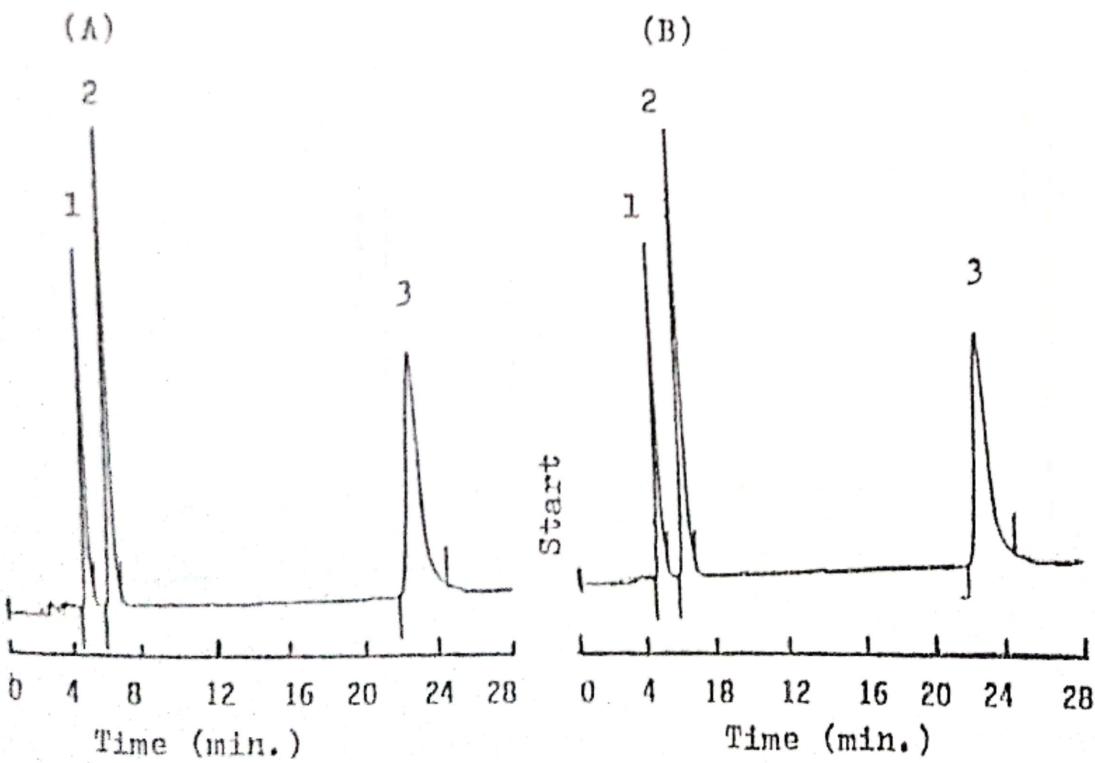


Fig. (4) HPLC Chromatogram of authentic mixture of ampicillin and dicloxacillin (A) and Cloxapen capsules (B).

1- Ampicillin

2- Penicillin G

3- Dicloxacillin

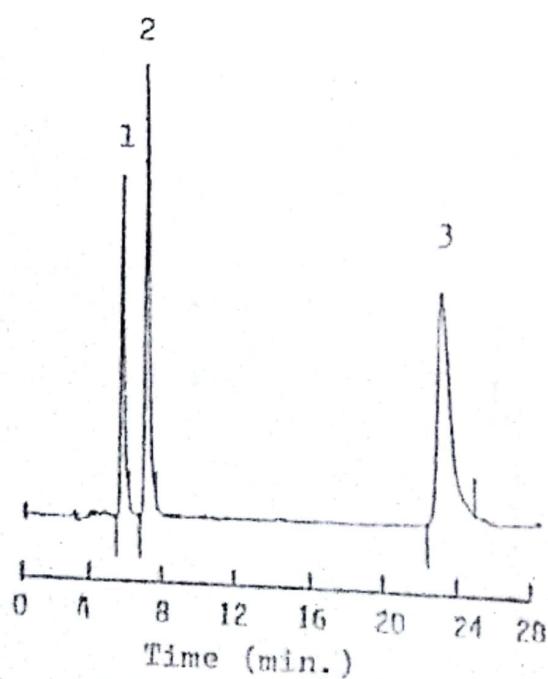
Table (11) Determination of authentic phenoxyethylpenicillin using the proposed HPLC method.

Amount added mg%	Amount found mg%	Recovery %
1.23	1.233	100.244
3.075	3.07	99.84
6.15	6.128	99.64
Mean		99.908
SD		0.3076
RSD		0.308

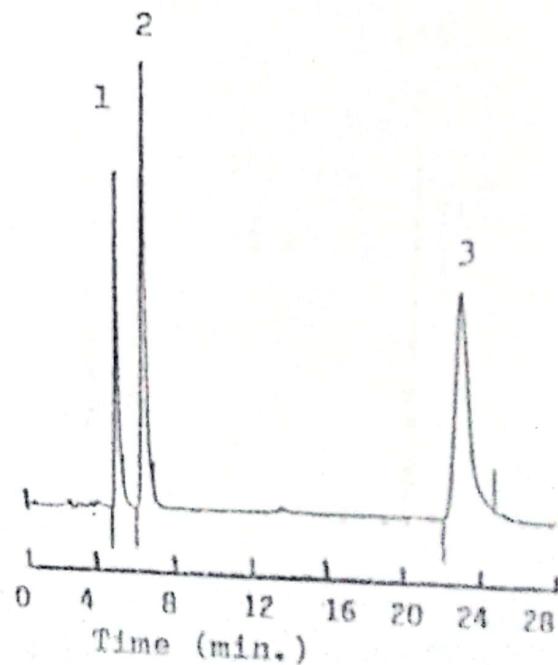
Table (12) Determination of phenoxyethylpenicillin in Ospen tablets using the proposed HPLC method.

Amount added mg%	Amount found mg%	Recovery %
1.23	1.236	100.487
3.075	3.014	98.016
6.15	6.095	99.105
Mean		99.202
SD		1.238
RSD		1.248

(A)



(B)

**Fig. (5)** HPLC Chromatogram of Dipenacid vials (A) and Dipenacid capsules (B).
1- Ampicillin 2- Penicillin G 3- Dicloxacillin

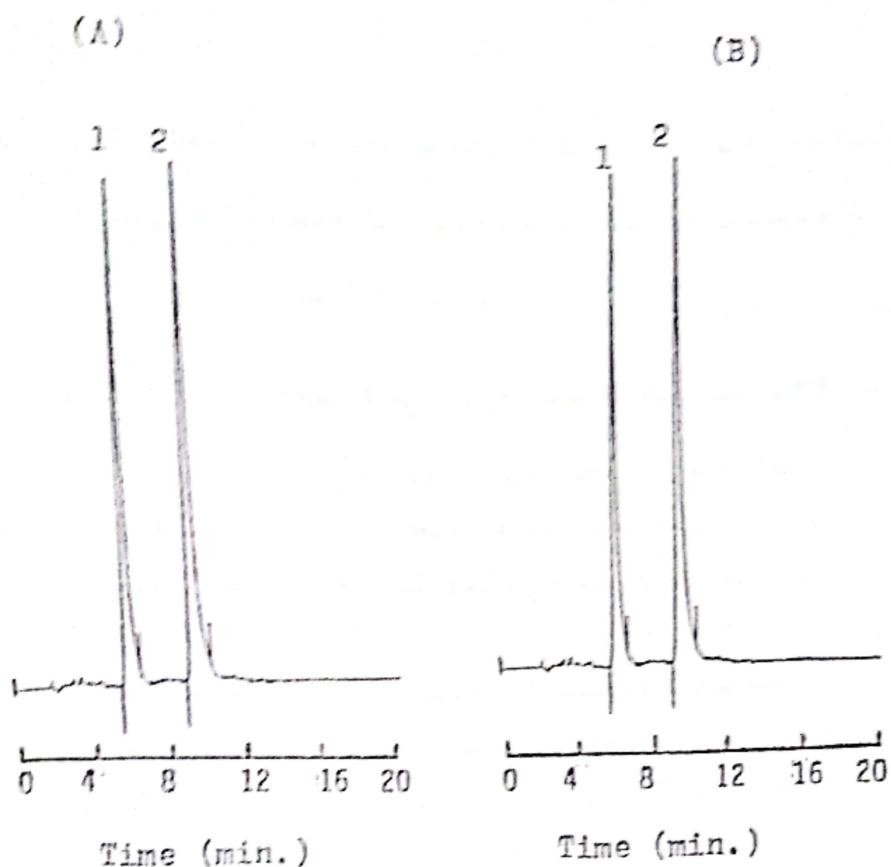


Fig. (6) HPLC Chromatogram of authentic sample of Penicillin V (A) and Ospen tablets (B).

1- Penicillin V

2- Penicillin G

CONCLUSION

In conclusion, the proposed method is selective, precise, accurate, time saving and can be used in quality control laboratories for analysis of antibiotic preparations containing these compounds in single or combined.

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تقدير مركبات الأموكسيسيلين والأمبيسيللين والبنيسييللين - ف و الفلوكساسيللين والكلوكساسيللين والداى كلوكساسيللين فى المستحضرات الصيدلية باستخدام كروماتوجرافيا السائل ذو الضغط العالى

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يصف هذا البحث طريقة لتحليل خليط من الأموكسيسيلين والأمبيسيللين والكلوكساسيللين
والفلوكساسيللين والبنيسييللين - ف باستخدام كروماتوجرافيا السائل ذو الضغط العالى وتم التطبيق على حقن وكبسولات
الفلوموكس والداى بيناسيد وحقن الكلوكسابين وكبسولات الأبيكلوكس وأقراص الأوسين وقد جئت النتائج على درجة
عالية من حيث الدقة والحساسية .

وقد أظهرت النتائج نسبة عالية من استرجاع الدواء (٩٩٥٣ - ٩٩٩ - ٩٩٦١ - ٩٩٠٨ - ٩٩١٠ - ١٨)
وكذلك أظهرت النتائج أقل معامل إنحراف معياري نسبي (٧٤٤ - ٦١٣ - ٥٥٧ - ٠٩٩٩ - ٠٩١٠)
وعلى فالطريقة الجديدة تساعد على سرعة تحويل مثل هذه المركبات بمفردها أو مخلوطه
مع الدقة فى النتائج .