**Original Paper****Detection of cadmium and copper in fresh and marine fishes**Mohamed A. Hassan¹, Engy El-bahy², Loaloe Shokr²¹Department of Food Hygiene, Faculty of Veterinary Medicine, Benha University, Egypt²Department of Food Hygiene, Animal Health Research Institute, Dokki, Giza, Egypt**ARTICLE INFO****ABSTRACT****Keywords**

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Cadmium and copper are very toxic heavy metals found in the environment and have public health hazard. Therefore, the current study was performed on 100 samples of freshwater fish (*Clarias lazera* and *Oreochromis niloticus*) and marine fish (*Sardina pilchardus* and *Pagrus pagrus*) collected at different times from various fish markets in Kafr El-sheikh governorate, Egypt. The collected samples were analyzed for heavy metal residues (cadmium and copper) by Atomic Absorption spectrophotometer (AAS). The obtained results showed that the mean values of cadmium concentrations (mg/kg) were 0.25 ± 0.01 , 0.17 ± 0.01 , 0.09 ± 0.01 and 0.08 ± 0.01 in *C. lazera*, *O. niloticus*, *S. pilchardus* and *P. pagrus*, respectively. While, the average concentrations of copper were 3.05 ± 0.27 , 2.62 ± 0.19 , 2.10 ± 0.13 and 1.73 ± 0.08 mg/kg in such examined samples. The sources of contamination of freshwater and marine fishes with heavy metals and their public health significance as well as some recommendations to control such serious pollutants were discussed

1. INTRODUCTION

Fish is very important source for high quality protein, minerals, vitamins and other nutrients as well as omega-3 that can decrease the danger of cardiac diseases and neurodevelopment in infants (Olmedo et al., 2013). From the nutritional point of view, consumption of fish regulates the blood glucose level (Patterson and Rashjitha, 2009).

Fish can accumulate heavy metals more than water because they their nutrients are organic matter from aquatic system, algae and other small organisms (Olaifa et al., 2004). The accumulation of heavy metals differs according to the age, sex, size and species of the fish. Furthermore, the highly active organs as gills, kidneys and liver are the target organs (Shivakumar et al., 2014).

The sources of heavy metals are earth crusts, weathering, fertilizers, and pesticides, leaching from harbor activities and sailing as well as from industries and water wastes (Elhussien and Adwork, 2018).

Cadmium causes elevation in AST, ALP and ALT activities of liver; also, it increases urea and cholesterol levels. On contrary, it reduces the levels of albumin and protein that will lead to severe damage in many organs as spleen, testis and kidneys (Rhaman et al., 2011). However, cadmium compounds as cadmium oxide, cadmium chloride and cadmium sulphate are carcinogenic. In addition, cadmium

chloride can cause impairment in fertility and can harm the unborn fetus (Court et al., 2011).

Copper has hepatotoxic effect in mice causing severe variation in the levels of ascorbic acid and glutathione. Further, copper can cause changes in the levels of glutathione and malonaldehyde concentrations due to the impairment of cell lining. Copper sulphate can cause liver damage as it results in granular damage, necrosis and dysfunction in hepatic cells. Also, copper has severe toxic effect to all living organisms (Oguz et al., 2010). However, copper gluconate is added to the food of infant as supplement. In addition, chromium copper arsenate (CCA) can be used as protecting for wood building substance and can cause kidney damage, and tubular dysfunction and necrosis (Rita et al., 2009).

Therefore, the aim of this study is to determine the concentrations of cadmium and copper in some freshwater fish and marine fish in Kafr El-sheikh governorate. Also, the hazardous toxic effects of these heavy metals and public health importance were studied.

2. MATERIAL AND METHODS*Collection of samples:*

A random 100 samples of fresh and marine fishes represented by *C. lazera*, *O. niloticus*, *S. pilchardus* and *P. pagrus* (25 of each) were purchased from fish markets in Kafr El-sheikh governorate, Egypt.

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All collected samples were examined for their cadmium and copper concentrations based on wet weight method using Atomic Absorption Spectrophotometer (AAS).

Washing procedures

In general, all equipment and instruments were perfectly rinsed to avoid their contamination with heavy metals according to Lars (2003).

Digestion technique:

The technique recommended by Staniskiene et al. (2006) was applied one gram of each sample was digested by 10ml of digestion mixture (60ml of 65% HNO₃ and 40ml of 70% HCL) in screw capped tube for estimation of their cadmium and copper levels.

The tubes were shaken and left overnight at room temperature. Further, the tubes were put in water bath at temperature starting from 60°C to 110°C for 4 hours to ensure complete digestion of the samples. Thereafter, the tubes were left to cool at room temperature and diluted with 1ml deionized water (30%). However, such tubes were diluted with deionized water till reach 25 ml. All the digests were filtered.

Analysis

All the digests (standard, digested and blanks) were absorbed by AAS (VARIAN, Australia, model AA240 FS) and examined for cadmium and copper residues.

Condition	Cadmium	Copper
Lamp wavelength (nm)	228.8	324.8
Lamp current (m/amp)	4	15
Fuel flow rate	1.2	1.00
Used gas	Argon	Argon
Measurement time (seconds)	4	4
Detection limit (ppb)	2-8	8-40

The concentration of cadmium, and copper was estimated according to the following equation:

$$C = R \times (D/W)$$

Where, C=concentration of cadmium and copper (mg/kg) wet weight. R=reading of digital scale of AAS. D= Dilution of prepared sample. W= Weight of the sample.

N.B. The concentration of each heavy metal in the blank solution was also calculated and subtracted from each analyzed sample.

3. RESULTS

The results recorded in table (1) and fig (1) revealed that the cadmium was detected in *C. lazera*, *O. niloticus*, *S. pilchardus* and *P. pagrus* with percentages of 52%, 36%, 32% and 20%, respectively.

Table (2) and fig (2) showed that the concentrations of cadmium (mg/Kg) in *C. lazera*, *O. niloticus*, *S. pilchardus* and *P. pagrus* ranged from 0.05 to 0.44,

0.03 to 0.28, 0.01 to 0.19 and 0.01 to 0.16 with an average of 0.25 ± 0.01 , 0.17 ± 0.01 , 0.09 ± 0.01 and 0.08 ± 0.01 , respectively.

Table 1 Incidence of Nile and marine fishes contaminated with cadmium (n=25).

Fish species	.No	%
lazera Clarias	13	52
Oreochromis niloticus	9	36
Sardina pilchardus	8	32
Pagrus pagrus	5	20
Total	35	35

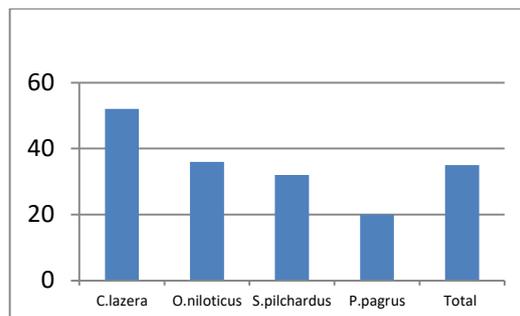


Fig. 1 Incidence of cadmium in the examined Nile and Marine fishes

Table 2: Statistical analytical results of cadmium residues (mg/Kg) in the examined samples of Nile and marine fishes (n=25)

Fish species	Min	Max	Mean \pm S.E
Lzera Clarias	0.05	0.44	^a 0.01 \pm 0.25
Oreochromis niloticus	0.03	0.28	^b 0.01 \pm 0.17
Sardina pilchardus	0.01	0.19	^c 0.01 \pm 0.09
Pagrus pagrus	0.01	0.16	^d 0.01 \pm 0.08

The difference between different letters in the same column were significant High significant differences (P<0.01)

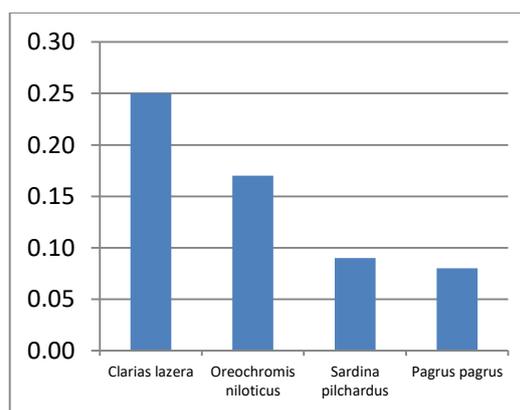


Fig. 2 Averages of cadmium residues (mg/Kg) in the examined Nile and marine fishes.

The results in table (3) and fig (3) indicated that the accepted samples of *C. lazera*, *O. niloticus*, *S. pilchardus* and *P. pagrus* according to their contents of cadmium were 52, 72, 76 and 84%, respectively.

Results given in table (4) and fig (4) showed that the copper residues in *C. lazera*, *O. niloticus*, *S. pilchardus* and *P. pagrus* were detected in all examined samples of fresh water and marine fishes.

Table 3: Validity of the examined fresh and marine fishes according to their cadmium residues (n=25).

Fish species	MRL (mg/Kg)*	Accepted samples		Unaccepted samples	
		No.	%	No.	%
<i>C. lazera</i>	0.05	13	52	12	48
<i>O.niloticus</i>	0.05	18	72	7	28
<i>S.pilchardus</i>	0.05	19	76	6	24
<i>P.pagrus</i>	0.05	21	84	4	16
Total (100)		71	71	29	29

* Maximum Residual Limit stipulated by Egyptian Organization for Standardization "EOS" (2010).

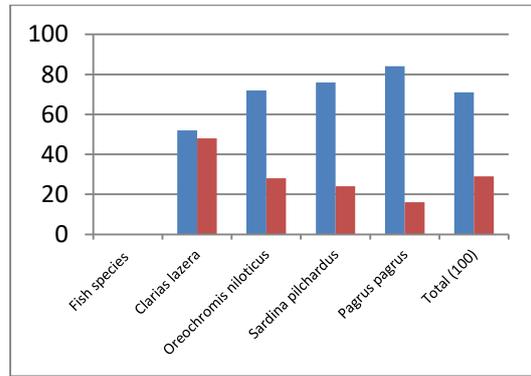


Fig 3 Incidence of accepted and unaccepted fish samples based on their levels of cadmium.

Table 4 Incidence of fresh and marine fishes contaminated with copper residues (n=25).

Fish species	No.	%
<i>C. lazera</i>	25	100
<i>O.niloticus</i>	25	100
<i>S.pilchardus</i>	25	100
<i>P.pagrus</i>	25	100
Total	100	100

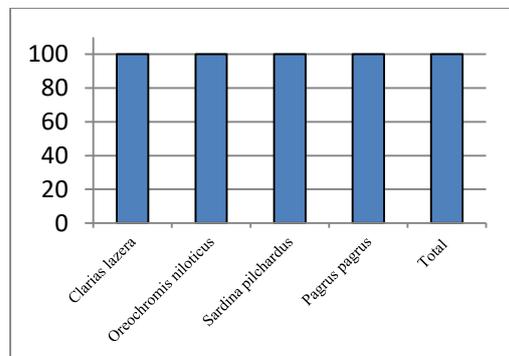


Fig. 4 Incidence of copper in the examined fresh and marine fishes.

Table (5) and fig (5) showed that the concentrations of copper residues (mg/Kg) in the examined samples of *C. lazera*, *O. niloticus*, *S. pilchardus* and *P. pagrus* ranged from 1.58 to 4.76, 1.30 to 4.12, 0.89 to 3.25 and 0.74 to 2.91 with averages of 3.05 ± 0.27 , 2.62 ± 0.19 , 2.10 ± 0.13 and 1.73 ± 0.08 , respectively.

The results achieved in table (6) and fig (6) showed that all examined samples of fresh water and marine fishes were accepted based on their levels of copper.

4. DISCUSSION

The impacts of the heavy metals of primary concern are mainly cadmium and copper because of their known toxicity to human as well as their health hazard as lethal, sublethal, acute and chronic toxicity (Levensen and Barnard, 1988).

Table 5 Statistical analytical results of copper residues (mg/Kg) in the examined samples of fresh and marine fishes (n=25).

Fish species	Min	Max	Mean \pm S.E
<i>C. lazera</i>	1.58	4.76	3.05 ± 0.27^a
<i>O.niloticus</i>	1.3	4.12	2.62 ± 0.19^b
<i>S.pilchardus</i>	0.89	3.35	2.10 ± 0.13^c
<i>P.pagrus</i>	0.74	2.91	1.73 ± 0.08^d

The difference between different letters in the same column were significant High significant differences ($P < 0.01$)

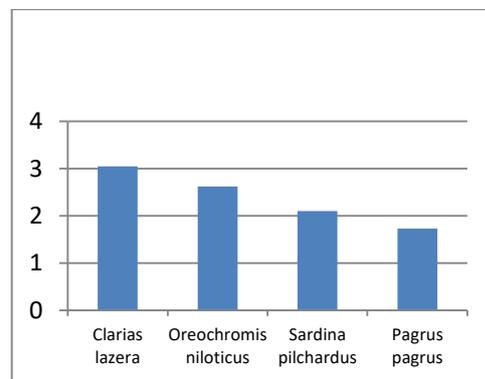


Fig. 5 Averages of copper residues (mg/Kg) in the examined Nile and marine fishes.

Table 6 Validity of the examined fresh and marine fishes according to their copper residues (n=25).

Fish species	MRL (mg/Kg)*	Accepted samples		Unaccepted samples	
		No.	%	No.	%
<i>C. lazera</i>	20	25	100	0	0
<i>O.niloticus</i>	20	25	100	0	0
<i>S.pilchardus</i>	20	25	100	0	0
<i>P.pagrus</i>	20	25	100	0	0
Total (100)		100	100	0	0

Maximum Residual Limit stipulated by Food Stuffs Cosmetics and Disinfectant Act (2007)

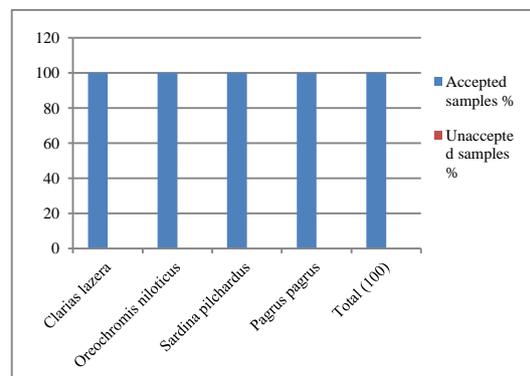


Fig. 6 Incidence of accepted and unaccepted fish samples based on their levels of copper residues

Cadmium was detected in *C. lazera*, *O. niloticus*, *S. pilchardus* and *P. pagrus* with percentages of 52%, 36%, 32% and 20%, respectively (Table 1 & Fig 1). Generally, the cadmium concentrations were recorded at highest levels in the examined samples of *C. lazera*

followed by *O. niloticus*, *S. pilchardus* and *P. pagrus* (Table 2 & Fig 2). The difference between different letters in the same column were significant, high significant differences ($p < 0.01$) as a result of type of product also high significant difference ($p < 0.01$) appeared between such examined samples due to their origin. Moreover, non-significant differences appeared as a result of interaction between types of products and their origin.

Nearly similar results were recorded by Sohsah (2009) who recorded that the mean value of cadmium in *C. lazera* ranged from 0.05 to 0.37 mg/kg with an average of 0.20 ± 0.03 mg/kg in large size while in *T. nilotica* ranged from 0.03 ± 0.26 mg/kg and Abou-Donia (1990) who reported that such mean value was 0.208 mg/kg.

Lower results were recorded by El-Said (2016) who recorded that the mean value of cadmium *T. nilotica* and *C. lazera* were 0.051 ± 0.011 mg/kg and 0.049 ± 0.012 mg/kg, respectively. Also, El-Nahas (2015) who recorded those values were 0.10 ± 0.01 mg/kg and 0.14 ± 0.02 mg/kg and Gawish and Hosni (2017) who reported that such mean value was 0.049 mg/kg in sardine and 0.036 mg/kg in morgan.

Higher results were reported by Seddek et al. (1996) who reported that, the mean value of cadmium in *T. nilotica* ranged from 0.01 to 0.62 mg/kg with an average of 0.26 mg/kg, while much value in *C. lazera* ranged from 0.03 to 0.48 mg/kg with an average of 0.33 mg/kg, Latif (2018) who reported that the mean value of cadmium in sardine was 0.15 ± 0.01 mg/kg, Mehoul et al. (2019) who recorded that the mean value of cadmium in sardine was 0.55 ± 0.14 mg/kg and Hadeed et al. (2017) who recorded that such mean value in *P. pagrus* to be 0.14 ± 0.02 mg/kg.

Furthermore, table (3) and fig (3) revealed that the accepted samples of *C. lazera*, *O. niloticus*, *S. pilchardus* and *P. pagrus* were 52%, 36%, 32% and 20%, respectively according to EOS (2010) which stated that the maximum residual limit of cadmium should not exceed than 0.05 (mg/kg) in fresh and marine water fishes.

The main sources of cadmium contamination are operations in industry and mining also the using in phosphate in fertilization process that can contaminate fish Kaoud and El-Dahshan (2010).

The toxicity of cadmium originated by its ability complete with other essential elements for binding sites, furthermore, its ability to interfere with sulfhydryl groups that are important for protein and enzymes function as cadmium can block these groups (Alan and Miller, 1998). Cadmium is naturally found in earth crust and in environment with minute can cause cancer at high level, the production of some metals as zinc, aluminium, steel and iron production are main sources of cadmium contamination as well as the using of phosphate as fertilizer (United States Environment Protection Agency, 2003).

Chronic toxicity with cadmium is manifested by bone disorders, fractures, impaired renal function with protein urea, phosphorus and calcium regulation is affected and (Itai-Itai) or (Ouch-Ouch) disease that include joints pain and bone disorders (Friberg and Elinder, 1985 and Kagamimori et al., 1986). Cadmium causes hypertension and renal damage (Nishiya-Ma et al., 1986), testicular atrophy, renal damage, cancer prostate and nausea are symptoms of

cadmium toxicity (Kikuchi et al., 2002). Cadmium accumulates in brain causing damage in neurons cells, strogial death through the deplete of glutathione (Imetal., 2006).

The copper residues in *C. lazera*, *O. niloticus*, *S. pilchardus* and *P. pagrus* were detected in all examined samples of fresh water and marine fishes as shown in table (4) and fig (4) by a percentage of contamination (100%) in all examined samples.

It is evident from the results recorded in table (5) and fig (5) that the copper residual level (mg/kg) in *C. lazera*, *O. niloticus*, *S. pilchardus* and *P. pagrus* ranged from 1.58 to 4.76, 1.30 to 4.12, 0.89 to 3.35 and 0.74 to 2.91 with an average of 3.05 ± 0.27 , 2.62 ± 0.19 , 2.10 ± 0.13 and 1.73 ± 0.08 , respectively. The difference between different letters in the same column were significant, high significant differences ($p < 0.01$) as a result of type of product also high significant difference ($p < 0.01$) appeared between such examined samples due to their origin. Moreover, non-significant differences appeared as a result of interaction between types of products and their origin.

Nearly similar results was recorded by EL-said (2016) who found that the mean value of copper in *O. niloticus* and *C. lazera* were 2.49 ± 0.224 and 2.331 ± 0.3257 mg/kg, Al-Weher (2008) who reported that such value in Tilapia was 2.90 ± 0.3 mg/kg and Kaoud and El-Dahshan (2010) who found that such value was 2.54 ± 0.05 mg/kg in *O. niloticus*.

Higher results were recorded by Sohsah (2009) who recorded that such mean value ranged from 2.04 to 5.91 mg/kg and 1.76 to 5.13 mg/kg with an average of 4.26 ± 0.45 and 3.51 ± 0.37 mg/kg in *C. lazera* and *T. nilotica*, Olatunji and Osibanjo (2012) who recorded that such value was 3.30 ± 0.79 mg/kg in Tilapia and Ilgar (2016), who recorded that such value was 4.17 mg/kg in sardine.

Lower results were recorded by Adeosun et al. (2015) who found that the mean value of copper in *O. niloticus* was 0.015 mg/kg, Badr (2014) who found that such value was 0.264 ± 0.023 ppm in *O. niloticus* and Ayeloja et al. (2014) who recorded that such value was 0.23 ± 0.11 and 0.108 ± 0.03 mg/kg in *T. nilotica* and *C. lazera*, respectively.

The main sources of copper contamination are the using of air fallout and effluents as mining from industry, the using of copper sulphate in eradication killing of snails in water and the use of copper compounds such as pesticides, algicides and fungicides in soil treatment (Ali, 2010).

The chronic contamination with copper causes haemolytic anemia and even death, Jaundice, hypertension, diarrhoea and vomiting are symptoms of high dose with copper. Wilson's diseases is genetic disorder caused by high copper dose and recognized by damage of neuron that leads to neurological disorders, cirrhosis, odema, ascitis and Fleischer ring is a golden ring on the cornea formed from copper (Gossel and Bricher, 1990).

The results recorded in table (6) and fig (6) showed that all the examined samples of fresh and marine water fishes were accepted based on their levels of copper by a percentage of (100%) where, the don't exceed the permissible limit of copper according to Food Stuffs Cosmetics and Disinfectant Act (2007) which stated that the maximum residual limit of copper should not exceed than 20 (mg/kg).

5. CONCLUSION

The current study proved that there are great variations in the levels of cadmium and copper in the examined samples of fish. In addition, the examined samples were significantly polluted with high levels of toxic metals. The continuous consumption of these contaminated fish may result in public health hazard through progressive irreversible accumulation of such toxic pollutants in the human body.

REFERENCES

1. A.P.H.A (2002): American Public Health Association.
2. Abou-Donia, M.A (1990): Selected devised techniques for detection fish contaminants, Ph, Docter. Agriculatoin. Ain – Shams, University.
3. Adeosun, F.I., Akinyemi, A.A., Taiwo, I.O., Omoike, A., Ayorinde, B.J.O. et al., (2015): The effects of heavy metals concentration on some commercial fish in Ogun River, opeji, Ogun, Nigeria. *J. Environmental Science Technology*. 9(4), 365-370.
4. Alan, L. and Miller, N.D. (1998): Technical Advisor, Thor Research, Inc. Senior Editor *Alternative Medicine Review*. *Alternative Medicine Review* 3 (3): 193.
5. Ali, K.H.I.A. (2010): Assessment of some Environmental pollution in beni-suief Governorate, Ph. D., Faculty veterinary medicine, Beni-suief University, Egypt
6. Al-weher, S.M. (2008): Levels of heavy metal cadmium, copper and zinc in three fish species collected from Northern Jordan Valley, Jordan. *Jordan Journal of Biological Sciences*, 1(1):41-46.
7. Ayelaja, A. A., George, F.O.A., Shorinmade, A.Y., Jimoh, W.A., Afolabi, Q.O and Olawepo, K.D et al. (2014): Heavy metal concentration in selected fish species from Eleyele reservoir Ibadan oyo state South-Western Nigeria. *African journal Environmental science technology* 8(7):422-427.
8. Badr, A.M., Mahana, N.A., Eissa, A. (2014): Assessment of heavy metals levels in water and their toxicity in some tissue of Nile Tilapia in River Nile Basin at Greater Cairo, Egypt. *Global Veterinaria* 13 (4): 432-443.
9. Court, R.Road, M., Bootle, Merseyide, et al. (2011): Cadmium and "you":L20 7Hs, United Kingdom: Health and Safety executive March 2010. Retrieved. January 29, 2011.
10. Egyptian Organization for Standardization "EOS", (2010): Maximum Levels for certain contaminants in foodstuffs. No 7136/2010. Egyptian Standards, Ministry of Industry, Egypt.
11. Elhussien, M. E., Adwork, B.A. (2018): Determination of heavy metals in fish and water of White Nile during watery diarrhea out break from June to July 2017, Gezira Aba-sudan. *Science Journal Analytical Chemistry*.6 (1):1-6.
12. El-Nahas, S.B. (2015): Heavy metal residues in freshwater fish. Thesis, Master of Veterinary Medicine, (Meat Hygiene). Faculty Veterinary Medicine, Benha University, Egypt.
13. El-said, Marwa A.S.M. (2016): Heavy metal residues in imported and local fishes in Egypt. Master Thesis of Veterinary Medicine, Faculty Veterinary Medicine, BeniSuef University, Egypt.
14. Emin. O.O.; Hayati, Y.; Yasar, E.; Cevik, T.A. and Gunfer, T. (2010): The Effects of Copper Sulfate on Liver Histology and Biochemical Parameters of Term Ross Broiler Chicks. *Bio Trace Element Research*. 133 (3): 335-341.
15. Food Stuffs Cosmetics and Disinfectant Act, (2007): Regulations relating to metals in foodstuffs (No.1518). Food Government Gazette, Johannesburg, South Africa.
16. Friberg, L., Elinder, C.G. (1985): Cadmium and compounds In: *Encyclopedia of occupational health and safety* third Edn. (Second impression).
17. Gawish, M., Hosni, Inas M. (2017): Assessment of some heavy metals in the economic fishes in Hail market, Kingdom of Saudi Arabia. *International Journal Current Research in Biosciences and Plant Biology*, 4:7-15.
18. Gossel, T.A., Bricker, J.D (1990): Metal in principles of clinical toxicology second Edn .,Raven press .NewYork,162-192.
19. Hadeed, Halima A., Aljetawi, A .A. Alhemmalii., E. M., et al. (2017): Determination of some essential and toxic heavy metals in liver tissue of four marine fish species from cost of Misurata, Libya. First National Conference on Marine and Ground water pollution (first NCMGP 2017).
20. Ilgar, R., (2016): Canakkale Bogazi Ve cvresi Ekosisteminiin cografi Acidan Incelemes, Basilmam is , Doktora Tezi , Istanbul Universitesi, Deniz in Bilimlirive Is , letmeciligi , Enstitusu , Istanbul.
21. Im, J.Y., Paik, S.G., Han, P. L., et al. (2006): Cadmium – induced astroglial death proceeds via glutathione depletion *Journal. Neuroscience. Research*, 83 (2):301.
22. Kagamimori, S.,Williams , W.R ., Watanable , M. et al . (1986): GMP levels in chronic cadmium disease and osteoarthritis. *British. journal.Exp.Path.*, 67-517.
23. Kaoud, H.A., EL-Dahshan, A.R. (2010): Bioaccumulation and histopathological alterations of the heavy metals in Oreochromis niloticus fish .*Nature and science*, 8 (4):147-156.
24. Kikkuchi, Y., Nomiya, T., Kumagai, N., Uemura, T., Omar, K.et al. (2002): Cadmium concentration in current Japanese Food Beverages. *Journal Occupational Health*, 44:240-247.
25. Lars, J., (2003): Hazards of heavy metal contamination. *British Medicine. Bullet. Journal*. 68: 167-182.
26. Latif, H.M.A. (2018): Detection of some Heavy metals in fresh and salted fish. Thesis, Master of Veterinary Medicine, Faculty Veterinary Medicine, Benha University, Egypt.
27. Levensen, H., Barnard, W.D. (1988): Wastes in Marine Environment. Hemisphere Publishing Corporation. Cambridge New York London. Chapter 6, 123– 136.
28. Mehoul, Feta.,Bouayad Leila.,Hammoudi, A., AyadiOurda., RegadFifi et al. (2019): Evaluation of the heavy metals (Mercury, lead and cadmium) contamination of sardine (*Sardina pilchardus*) and sword fish(*Xiphias gladius*) fished in three Algerian coasts. *Journal of Veterinary World*, vol 2 p7-11.
29. Nishiyama, S.,Nakamura, K., Konish, Y. et al .(1986): Blood Pressure and sodium and potassium excretion ion in a cadmium treated male rats." *Environment. Research*, 40, 357 – 364.
30. Oguz, Emin.,O., Yuksel, Hayatia ., Yassar, E., Cevik,T.A .,et al . (2010): The effects of copper sulphate on liver Histology and Biochemical parameters of term Ross Broiler chicks. *Biological trace element Research*, 133 (3) 335-341.
31. Olaifa, F., Olaifa, A., Adelaja ,A., Owolabi, A et al. (2004): Heavy metal contamination of *Clarias gariepinus* from a lake and fish farm in Ibadan , Nigeria .*African Journal of Biomedical Research*, 7: 145-148.
32. Olatunji O.S., Osiban Jo, O. (2012): Comparative assessment of some heavy metals in some inland freshwater fish species from River Niger and River osara in North central Nigeria. *International. Journal Environmental Science*, 2(3): 1842-1851.
33. Olmedo, P., Pla, A., hernandez, A.F., Barbier, F., Ayouni, L., Gill, F. et al. (2013): *Environment International*, 59:63-72.

33. Patterson, J., Rnjitha, G. (2009): Qualities of commercially and experimentally sun-dried fin fish. *African Journal of food science*, 3:299-302.
34. Rhman, N.H.A., Backhiet, A.O., Adam, S.E.I .et al. (2011): Toxic effects of various dietary levels of combined cadmium chloride and zinc chloride on male wistar rats. *Journal pharmacology toxicology*, 6:76-81.
35. Rita, C.M., Catrina, V., Simone, M.,Maria delourdes, P. Julio, P. et al. (2009): Nephrotoxicity effects of the Wood preservative chromium copper arsenate on mice: Histopathological and quantitative approaches *Journal Trace Elements in Medicine Biological.*, 23(3): 224 – 230.
36. Seddek, A.S.H; Salem, D.A.;El-Sawi , N.M, Zaky, Z.M .et al (1996): Cadmium, lead, Nickel, Copper, Manganese and fluorine level sin River Nile Fish. *Assuit Veterinary Medicine Journal*, 34:95.
37. Shivakumar,C.,Thippeswamy .B.,Tejaswikumar., M., Prashanthakumar, S. et al . (2014): Bioaccumulation of heavy metals and its effect on organs of edible fishes located in Bhandra River, Karnatka. *Intern. Journal Research Fisheries &Aquaculture*, 4, 90-98.
38. Sohsah, Madiha A.M. (2009): Studies on some heavy metal residues in freshwater fish with special reference to water environmental pollution. Ph. D. Thesis, Faculty Veterinary Medicine, Benha University, Egypt.
39. Staniskiene, B.; Matusevicius, P.; Budreckiene, P., Skibniewska, K.A. et al. (2006): Distribution of heavy metals in tissues of freshwater fish in Lithuania. *Polish Journal Environmental Studies*, 15(4): 585-591.
40. United Stated Environmental Protection Agency (U.S.EPA) (2003): pollution Inventory data on discharges of metals to water. The Environment Agency privacy policy.