



Detection of Heavy Metals Residues in Fish and Shellfish

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ABSTRACT

The present study was conducted on 90 random samples of fish of *Claris gariepinus*, *Oreochromis niloticus* and *Mugil cephalus* and shellfish of Oyster, Shrimp and Crab (15 of each) were collected from different fish markets in Kalyobia governorate, Egypt, for determination of their content of mercury, lead and cadmium. The obtained results recorded that, the mean value of concentrations with mg/kg of mercury; lead and cadmium in fish samples were 1.62 ± 0.15 , 1.29 ± 0.12 , 0.95 ± 0.08 ; 0.67 ± 0.09 , 0.53 ± 0.06 , 0.36 ± 0.05 and 0.25 ± 0.03 , 0.19 ± 0.01 , 0.14 ± 0.01 for *C. gariepinus*, *O. niloticus* and *M. cephalus*, respectively. Meanwhile, in shellfish samples they were 1.37 ± 0.16 , 1.14 ± 0.10 , 0.73 ± 0.09 ; 0.56 ± 0.07 , 0.48 ± 0.05 , 0.40 ± 0.04 and 0.22 ± 0.02 , 0.15 ± 0.01 , 0.12 ± 0.01 for oyster, shrimp and crab, respectively. Moreover, the results revealed that, the highest mercury contamination was in *Claris gariepinus* followed by oyster; *O. niloticus*; shrimp; *M. cephalus* and finally crab. Whereas the highest lead contaminations were in *C. gariepinus* followed by oyster; *O. niloticus*; shrimp; crab and finally *M. cephalus*. Moreover, for cadmium the highest ratio in in *C. gariepinus* followed by oyster; *O. niloticus*; shrimp; *M. cephalus* and finally crab.

Key words: Shellfish, Residues, mercury, cadmium, lea.

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1. INTRODUCTION

Fish and fish products are important part of a healthy diet due to many nutritional benefits as they contain high-quality protein, low cholesterol level, omega-3 fatty acids, minerals as well as vitamins. Also, their lower costs potentiate their nutritive values. A well-balanced diet includes a variety of fish and shellfish can contribute to heart health and children's growth and development. The pollution of aquatic environment with heavy

metals constitutes a public health hazard during recent years. Scientific researches have proven that, the presence of the residues in foods is harmful to human health which be associated with immune-suppression, hypersensitivity to chemical agents, breast cancer, reduce sperm count and infertility (Sharp, 1999). Some heavy metals such as mercury, cadmium and lead introduced into environmental water system may pose high

toxicities on the aquatic organisms (Ambreen *et al.*, 2015). Pollution enters fish and shell fishes through five main routes: via food or non-food particles, gills, oral consumption of water and the skin (Mitra *et al.*, 2012). In general, heavy metals are not biodegraded and therefore, their bioaccumulation in fish, oyster, mussels, sediments and other components of aquatic ecosystems have been reported from all over the world (Kumar and Singh, 2010). In addition, it appears that problem of heavy metals accumulation in aquatic organisms including fish needs continuous monitoring and surveillance owing to biomagnifying potential of toxic metals in human food chain (Kumar *et al.*, 2009). The accumulation of metals in fresh water fish and marine shellfish has been proposed as a more sensitive and specific indicator of environmental contamination (Kavun and Podgurskaya, 2009). It is important to investigate the levels of heavy metals in these organisms to assess whether the concentration is within the permissible level and will not pose any hazard to the consumers (Cid *et al.*, 2001). Scientific researches have proven that, the presence of heavy metals residues in foods is harmful to human health which be associated with immune-suppression, hypersensitivity to chemical agents, anaemia, chronic renal failure, encephalopathy, breast cancer, reduce sperm count and infertility (Rubin and Strayer ,2008 and Rhman *et al.* ,2011). Therefore, the present study was conducted to evaluate the contamination levels of fish and shellfish with heavy metals and their acceptability for human consumption.

2. MATERIAL AND METHODS

2.1. Collection of samples

A total of 90 random samples of fresh fish of *Claris gariepinus*, *Oreochromis niloticus* and *Mugil cephalus* (15 of each) and shellfish of Oyster, Shrimp and Crab (15 of each) were collected from different fish markets in Qaliobia governorate, Egypt, for determination of mercury, lead and cadmium levels.

2.2. Determination of heavy metals

All collected samples were examined for determination of heavy metals (mercury, lead and cadmium) levels on the basis of wet weight (mg/Kg).

2.2.1. *Washing procedures following (Lars, 2003).*

2.2.2. *Digestion technique following (Staniskiene et al., 2006).*

2.2.3. *Preparation of blank and standard solutions following (Andreji et al., 2005).*

2.2.4. *Quantitative determination of heavy metals:*

Absorbency of mercury, lead and cadmium was directly recorded from the digital scale and their concentrations were calculated according to the following equation: $C=R \times (D/W)$. Where, C= Concentration of the element (wet weight) R= Reading of digital scale of AAS.D= Dilution of the prepared sample. W= Weight of the sample.

N. B. The concentration of each element in the blank solution was also calculated and subtracted from each analysed sample.

3. RESULTS

The obtained results in Table (1), revealed that, the minimum and the maximum mercury concentrations "mg/Kg " in the examined samples of *C. gariepinus*, *O. niloticus* and *M. cephalus* were ranged from 0.38 to 2.45; 0.31 to 2.07 and 0.22 to 1.51 with mean values of 1.62 ± 0.15 ; 1.29 ± 0.12 and 0.95 ± 0.08 , respectively. Meanwhile, in shellfish samples of oyster, shrimp and crab were ranged from 0.35 to 2.24; 0.26 to 1.85 and 0.17 to 1.33 with mean values of 1.37 ± 0.16 ; 1.14 ± 0.10 and 0.73 ± 0.09 , respectively. Moreover, 73.3% of *C. gariepinus*; 66.7% of *O. niloticus*; 46.7% of *M. cephalus*; 9 oyster (60 %); 53.3% of shrimp and 40% of crab, were unaccepted, as they were exceeded the maximum permissible limit of mercury in fish and shellfish that should not exceed 0.50 mg/kg (EOS, 2010).

Results in Table (2) declared that, the minimum and the maximum lead concentrations "mg/Kg " in the examined samples of *C. gariepinus*, *O. niloticus* and *M. cephalus* were ranged from 0.12 to 1.04; 0.09 to 0.82 and 0.05 to 0.60 with mean values of 0.67 ± 0.09 ; 0.53 ± 0.06 and 0.36 ± 0.05 , respectively. Meanwhile, for shellfish samples oyster, shrimp and crab were ranged from 0.10 to 0.89; 0.08 to 0.74 and 0.07 to 0.68 with

mean values of 0.56 ± 0.07 ; 0.48 ± 0.05 and 0.40 ± 0.04 , respectively. In addition; 42 samples, 10 *C. gariepinus* (66.7%); 8 *O. niloticus* (53.3%); 4 *M. cephalus* (26.7%); 8 oyster (53.3%); 7 shrimp (46.7%) and 5 crabs (33.3%), were unaccepted, as they were exceeded the maximum permissible limit of lead in fish and shellfish that should not exceed 0.30 mg/kg (EOS, 2010).

As shown in Table (3) results revealed that, the minimum and the maximum cadmium concentrations "mg/Kg " in the examined samples of fish were ranged from 0.02 to 0.53; 0.02 to 0.37 and 0.01 to 0.24 with a mean value of 0.25 ± 0.03 ; 0.19 ± 0.01 and 0.14 ± 0.01 in *C. gariepinus*, *O. niloticus* and *M. cephalus*, respectively. Also, in shellfish samples of oyster, shrimp and crab were ranged from 0.02 to 0.41; 0.01 to 0.30 and 0.01 to 0.19 respectively, with mean values of 0.22 ± 0.02 ; 0.15 ± 0.01 and 0.12 ± 0.01 , respectively. Moreover; 27 samples, 7 *C. gariepinus* (46.7%); 5 *O. niloticus* (33.3%); 2 *M. cephalus* (13.3%); 6 oyster (40%); 4 shrimp (26.7%) and 3 crabs (20%), were unaccepted, as they were exceeded the maximum permissible limit of cadmium in fish and shellfish that should not exceed 0.05 mg/kg (EOS, 2010).

Table (1): Mean values of mercury concentrations "mg/Kg" in the examined samples of fish and shellfish (n=15).

Fish and shellfish species	Min.	Max.	Mean \pm SEM*	Unaccepted Samples	
				Maximum Residual Limit (mg/Kg) **	No. %
<u>Fish:</u>					
<i>C. Gariepinus</i>	0.38	2.45	1.62 \pm 0.15	11	73.3
<i>O. Niloticus</i>	0.31	2.07	1.29 \pm 0.12	10	66.7
<i>M. Cephalus</i>	0.22	1.51	0.95 \pm 0.08	7	46.7
<u>Shellfish:</u>					
Oyster	0.35	2.24	1.37 \pm 0.16	9	60
Shrimp	0.26	1.85	1.14 \pm 0.10	8	53.3
Crab	0.17	1.33	0.73 \pm 0.09	6	40

SEM* = standard error of mean

** Maximum Residual Limit of mercury (0.50 mg/Kg) stipulated by Egyptian Organization of Standardization "EOS" (2010).

Table (2): Mean values of lead concentrations "mg/Kg" in the examined samples of fish and shellfish(n=15).

Fish and shellfish species	Min.	Max.	Mean \pm SEM*	Unaccepted Samples	
				Maximum Residual Limit (mg/Kg) **	No. %
<u>Fish:</u>					
<i>C. Gariepinus</i>	0.12	1.04	0.67 \pm 0.09	10	66.7
<i>O. Niloticus</i>	0.09	0.82	0.53 \pm 0.06	8	53.3
<i>M. Cephalus</i>	0.05	0.60	0.36 \pm 0.05	4	26.7
<u>Shellfish:</u>					
Oyster	0.10	0.89	0.56 \pm 0.07	8	53.3
Shrimp	0.08	0.74	0.48 \pm 0.05	7	46.7
Crab	0.07	0.68	0.40 \pm 0.04	5	33.3

S.E.M* = standard error of mean

** Maximum Residual Limit of lead (0.30 mg/Kg) stipulated by Egyptian Organization of Standardization "EOS" (2010).

Table (3): Mean values of cadmium concentrations "mg/Kg" in the examined samples of fish and shellfish (n=15).

Fish and shellfish species	Min.	Max.	Mean \pm SEM*	Unaccepted Samples Maximum Residual Limit (mg/Kg) **	
				No.	%
<u>Fish:</u>					
<i>C. Gariepinus</i>	0.02	0.53	0.25 \pm 0.03	7	46.7
<i>O. Niloticus</i>	0.02	0.37	0.19 \pm 0.01	5	33.3
<i>M. Cephalus</i>	0.01	0.24	0.14 \pm 0.01	2	13.3
<u>Shellfish:</u>					
Oyster	0.02	0.41	0.22 \pm 0.02	6	40
Shrimp	0.01	0.30	0.15 \pm 0.01	4	26.7
Crab	0.01	0.19	0.12 \pm 0.01	3	20

SEM* = standard error of mean

** Maximum Residual Limit of cadmium (0.05 mg/Kg) stipulated by Egyptian Organization of Standardization "EOS" (2010).

4. DISCUSSION

The recorded results for mercury concentrations revealed that, the highest mercury contamination was in *C. gariepinus* followed by oyster; *O. niloticus*; shrimp; *M. cephalus* and finally crab. The results for mercury concentrations in fish samples were nearly similar to that obtained by Love *et al.* (2003) which was with mean level was 1.10mg/kg. But, they were disagreed with those of Stancheva *et al.* (2013) who found the mean mercury residual level in muscle samples of *M. cephalus* fish collected from two different Black sea areas- Varna Lake and Nesebar were (0.08 \pm 0.01 and 0.05 \pm 0.01) ppm, respectively and with those of and reji *et al.* (2005) who detected higher levels (2.85 \pm 1.22 μ g/kg, wet weight). Meanwhile, for shellfish samples, the results came in accordance with those

obtained by Balfour *et al.*, (2012) who reported that the mercury levels were varied from 86.67 \pm 2.66 –98.60 \pm 12.16%. (μ g g⁻¹). But disagreed with those of Falusi and Olanipekun (2007) who detected mercury in shellfish samples with lower concentrations (0.665mg/kg). Moreover, such variations in mercury concentration among the examined fish and fish products may be attributed to variation in feeding habits, and type of sea fish (Morgan, 1999). Mercury is highly toxic to human health; it is a particular threat to the development of child in utero and early life. Methyl mercury affects the kidneys and also the central nervous system, particularly during development, as it crosses both the blood –brain barrier and placenta (Clarkson, 2002).

The poisoning with lead in human leading to brain dysfunction impairment in learning abilities as it affects 3 main neurotransmission systems: dopaminergic, cholinergic and glutaminergic so, it causes neurodegenerative disease. Also, it affects humeral immunity (perhaps through interference with macrophage function), may or may not affect cellular immunity and increases the frequency of chromosomal aberrations (Xu *et al.*, 2009). The obtained results for lead concentrations revealed that, the highest lead contamination was in *C. gariepinus* followed by oyster; *O. niloticus*; shrimp; crab and finally *M. Cephalus*. The results for lead concentrations in fish samples were nearly similar to that recorded by Ayeloja *et al.* (2014) who mentioned that, the mean residual level of lead in muscle samples of *Tilapia nilotica* and Catfish collected from Eleyele reservoir Ibadan Oyo State South-western, Nigeria, were (0.393 ± 0.04 and 0.463 ± 0.12) ppm and Badr *et al.* (2014) who evaluated the mean value of lead in muscle samples of *O. niloticus* fish collected from two areas in River Nile which were (0.662 ± 0.058 ppm and 0.833 ± 0.057) ppm. Meanwhile, the results were disagreed with those of Olusola *et al.*, (2012) and Stancheva *et al.*, (2013) who detected lead in fish samples with lower concentrations which were 0.039 ± 0.004 ppm and 0.07 ± 0.01 and 0.05 ± 0.002 ppm for them, respectively. And with those of Turkmen *et al.*, (2011) who detected higher levels (0.63 ± 0.17 ppm) and with those of Tajiri *et al.*, (2011) and Adeosun *et al.*, (2015) who failed to detect it in all examined samples of fish. In addition, for shellfish samples, the results came in accordance with those obtained by Bat *et al.*, (2013); Olgunoğlu *et al.*, (2015) and Olgunoğlu and Olgunoğlu (2016)

which were $0.291-0.491 \mu\text{g. g}^{-1}$ wet weight, $0.43 \mu\text{g/g}$, and $0.02 \pm 0.01 \text{ mg/kg}$ for them, respectively. But, these results came in contrast to those of Falusi and Olanipekun (2007) who detected lead in shellfish samples with lower concentrations which were 0.195 mg/kg ; and those of Vazquez-Boucard *et al.*, (2014) who detected higher levels (7.2 to $9.9 \mu\text{g/g}$) and those of Olowu *et al.*, (2010) who failed to detect it in all examined samples of shellfish.

The contamination of the environment with heavy metals associated directly with public health hazard due to bioaccumulation and bio magnification nature in food chain (Hussain *et al.*, 2012). The obtained results for cadmium concentrations cleared that, the highest cadmium contamination was in *C. gariepinus* followed by oyster; *O. niloticus*; shrimp; *M. cephalus* and finally crab. The results for cadmium concentrations in fish samples were nearly like that recorded by Olusola *et al* (2012); Ayeloja *et al.*, (2014) and Badr *et al.*, (2014) which were 0.020 ± 0.006 ppm, 0.123 ± 0.04 ppm and 0.024 ± 0.005 ppm, respectively. But these results were disagreed with those of Tajiri *et al.* (2011); Stancheva *et al.*, (2013) and Shreadah *et al.*, (2015) who detected cadmium in fish samples with lower concentrations ($0.011 \mu\text{g. g}^{-1}$, 0.012 ± 0.002 and 0.08 ppm) for them, respectively and with those of Turkmen *et al.*, (2011) who detected higher levels (0.49 ± 0.05 ppm) and with those of Adeosun *et al.*, (2015) who failed to detect it in all examined samples of fish. Meanwhile, for shellfish samples, the results came in accordance with those obtained by Olgunoğlu *et al.*, (2015) and Olgunoğlu and Olgunoğlu

(2016) which were $144.12 \pm 20.42 \mu\text{g/g}$ and $0.50 \pm 0.02 \text{mg/kg}$, respectively. But, they were disagreed with those of Ayas and Ozogul (2011) who detected cadmium in shellfish samples with lower concentrations with 0.4 mg/kg and with those of Falusi and Olanipekun (2007) who determined the cadmium concentrations crab which were 3.375 mg/kg ; Olowu *et al.*, (2010) who investigated the level of cadmium in crabs and prawns and found that, the mean concentration of cadmium was $1.66 \pm 1.82 \mu\text{g/g}$ and $0.07 \pm 0.08 \mu\text{g/g}$ respectively, and Balfouret *et al.*, (2012) who found the cadmium level in shrimp were varied from 95.53 ± 0.60 - $95.77 \pm 2.06\%$.

Finally, the present study proved that the highest contamination for fish was in *C. gariepinus* and the lowest one was in *M. cephalus* but for shellfish the highest contamination was in *oyster* and the lowest one was in *crab*. So, these have a public health importance as the heavy metal residual levels (mercury; lead and cadmium) in them, might be exceeded the recommended safe permissible limits stipulated by Egyptian Organization for Standardization (EOS, 2010) and must be controlled to prevent or minimize them and improve the sanitary status of fish and shellfish.

5. REFERENCES

- Adeosun, F.I.; Akinyemi, A.A.; Taiwo, I.O.; Omoike, A. and Ayorinde, B.J.O. 2015. The effects of heavy metals concentration on some commercial fish in Ogun River, Opeji, Ogun state, Nigeria. *Afr. J. Environ. Sci. Technol.*, 9(4): 365-370.
- Ambreen, F.; Javed, M. and Batool, U. 2015. Tissue specific heavy metals uptake in economically important fish, *Cyprinus carpio* at acute exposure of metals mixtures. *Pakistan J. Zool.*, 47: 399-407.
- Andreji, J.; Stranai, Z.; Massonyl, P. and Valent, M. 2005. Concentration of selected metal in muscle of various fish species. *J. Environ. Sci. Heal.* 40 (4):899-912.
- Ayas, D. and Ozogul, Y. 2011. The chemical composition of carapace meat of sexually mature blue crab (*Callinectes sapidus*, RATHBUN 1896) in the Mersin Bay. *Int. J. Food Sci. Tech.*, 5 (3):262-269.
- Ayeloja, A.A.; George, F.O.A.; Shorinmade, A.Y; Jimoh, W.A; Afolabi, Q.O. and Olawepo, K.D. 2014. Heavy metal concentration in selected fish species from El-Eyele reservoir Ibadan Oyo state south-western Nigeria. *Afr. J. Environ. Sci. Tech.*, 8(7): 422-427.
- Badr, A.M; Mahana, N.A. and Eissa, A. 2014. Assessment of Heavy metals levels in water and their toxicity in some tissues of Nile Tilapia (*Oreochromis niloticus*) in River Nile Basin at Greater Cairo, Egypt. *Glob Vet.*, 13(4): 432-443.
- Balfour, S.; Badrie, N.; Yen, I.C. and Chatergoon, L. 2012. Seasonal influence and heavy metal analysis in marine Shrimp (*Penaeus* spp.) sold in Trinidad, West Indies. *J. Food Research*, 1(1): 193-199.
- Bat, L.; Şahin, F.; Sezgin, M.; Üstün, F.; Baki, O.G. and Öztekin, H.C. 2013. Heavy metals in edible

- tissues of the brown shrimp *Crangon crangon* (Linnaeus, 1758) from the Southern Black Sea (Turkey) J. Black Sea/Mediterranean Environment, 19(1): 70-81.
- Cid, B. P.; Boia, C.; Pombo, L. and Rebelo, E. 2001. Determination of trace metals in fish species of the Ria de Aveiro (Portugal) by electrothermal atomic absorption spectrometry. Food Chem., 75: 93-100.
- Clarkson, T.W. 2002. The three modern faces of mercury. Environmental Health Prospect, 110(1):11-32.
- Court, R. Road, M.; Bootle, Merseyside. 2011. "Cadmium and you": L20 7HS, United Kingdom: Health and safety Executive. March 2010. Retrieved January 29, 2011.
- EOS" Egyptian Organization for Standardization "2010. Maximum Levels for certain contaminants in foodstuffs. No 7136/2010. Egyptian Standards, Ministry of Industry, Egypt.
- EL-Sayed, A.E; EL-Ayyat, M. S; Nasr, E.S and Khater, Z.Z.K. 2011. Assessment of heavy metals in water, sediment and fish tissues from Sharkia Province, Egypt. Egypt.J. Aquat. Biol.&Fish., 15 (2): 125-144.
- Falusi B.A. and Olanipekun E.O. 2007. Bioconcentration factors of heavy metals in tropical crab (*carcinus* sp) from River Aponwe, Ado-Ekiti, Nigeria. J. Appl. Sci. Environ. Manage., 11(4): 51 – 54.
- Gress, L. and Lord, J. 2002. Return and recycling of used high intensity bulbs for recycling and closed-loop mercury control. In mercury in products, Processes, Waste and the Environment: Eliminating, reducing and Managing risks from Non-Combustion Sources (ed. Oppelt, E.T.O, proceedings and Summary Reports EPA.
- Healey, N. 2009. Lead toxicity, vulnerable subpopulations and emergency preparedness. Radiatprot Dosimetry., 134(3-4): 143-151.
- Hozhabri, S.; White, F.; Rahbor, M.H.; Agboatwalla, M. and Luby, S. 2004. Elevated blood levels among children in a fishing community, Karachi, Pakistan. Arch Environ. Health, 59 (1):37-41.
- Hussain, R.T.; Ebraheem, M.K. and Moker, H.M. 2012. Assessment of Heavy Metals (Cd, Pb and Zn) contents in Livers of Chicken available in the local markets of Basrah City, Iraq. Bas. J.Vet. Res., 11(1): 43–51.
- Kavun, V. Y. and Podgurskaya, O. V. 2009. Adaptation strategy of bivalve *Modiolus modiolus* from upwelling regions of the Kuril Islands shelf (Sea of Okhotsk) to heavy metal effects. Cont. Shelf Res., 29: 1597-1604.
- Kumar, P.; Prasad, Y.; Patra, A.K.; Ranjan, R.; Patra, R.C.; Swarup, D. and Singh, S.P. 2009. Ascorbic acid, garlic extract and taurine alleviate cadmium-induced oxidative stress in freshwater catfish (*Clarias batrachus*). The Sci. Total Environ., 407: 5024– 5030.
- Kumar, P. and Singh, A. 2010. Cadmium toxicity in fish: An overview. GERF Bulletin of Biosciences, 1(1): 41-47.

- Lars, J. 2003. Hazards of heavy metal contamination. *British Med. Bull* 68: 167-182.
- Love, J. L.; Rush, G.M. and Grath, H. 2003. Total mercury and methyl mercury levels in some New Zealand commercial marine fish species. *Food Addit. Contam.*, 20:37-45.
- Manahan S.E. 1992. *Toxicological chemistry*, 2nd Ed. Lewis publishers Inc. Boca Raon, Ann. Arbor, London, Tokyo.
- Mitra, A; Barua, P.; Zaman, S. and Banerjee, K. 2012. Analysis of trace metals in commercially important crustaceans collected from UNESCO protected world heritage site of Indian Sundarbans. *Turk. J. Fish. Aquat. Sci.*, 12: 53-66.
- Morgan, J.N. 1999. Effect of processing of heavy metal content of foods. *Adv. Exp. Med. Biol.* 459:195-211.
- Olgunoğlu, M.P.; Olgunoğlu, I.A. and Bayhan, Y.K. 2015. Heavy metal concentrations (Cd, Pb, Cu, Zn, Fe) in Giant Red Shrimp (*Aristaeomorpha foliacea* Risso 1827) from the Mediterranean Sea. *Pol. J. Environ. Stud.*, 24 (2): 631-635.
- Olgunoğlu, M. P. and Olgunoğlu, J. A. 2016. Heavy metal contents in Blue Swimming Crab from the North eastern Mediterranean Sea, Mersin Bay, Turkey. *Pol. J. Environ. Stud.*, 25(5): 2233-2237.
- Olowu; Ayejuyo; Adejoroi ; Adewuyi; Osundiya ; Onwordi; Yusuf and Owolabi. 2010. Determination of heavy metals in Crab and Prawn in Ojo Rivers Lagos, Nigeria. *Coden Ecjhae J. Chemistry*, 7(2): 526-530.
- Olusola, A.V.; Folashade, P.A. and Ayoade, O.I. 2012. Heavy Metal (Lead, Cadmium) and Antibiotic (Tetracycline and Chloramphenicol) Residues in Fresh and Frozen Fish Types (*Clarias gariepinus*, *Oreochromis niloticus*) in Ibadan, Oyo State, Nigeria. *Pakistan Journal of Biological Sciences*, 15: 895-899.
- Rhman, N.H. A; Bakhiet A.O. and Adam, S.E.I. 2011. Toxic effects of various dietary levels of combined cadmium chloride and zinc chloride on male wistar rats. *J. Pharmacol. Toxicol.*, 6: 76-81.
- Rubin, R. and Strayer, D.S. 2008. *Environmental and nutritional pathology. Rubin, pathology: Clinicopathological foundations of Medicine*, 5th edition. Lippincott Williams & Wilkins. ISBN 0781795168.
- Sharp, M. 1999. Towards sustainable pesticides. *J. Environ. Monit.*, 1: 33-36.
- Shibamoto, T. and Bjeldanes, L.F. 1993. *Introduction to food toxicology*. Academic press Inc Harcourt Brace and Company. New York. Food Science and Technology, International series.
- Shreadah, M.A., Abdel Fattah, Laila.M. and Fahmy, M.A. 2015. Heavy Metals in Some Fish Species and Bivalves from the Mediterranean Coast of Egypt. *J. Environ. Prot.* 6: 1-9.
- Stancheva, M.; Makedonski, L. and Petrova, E. 2013. Determination of heavy metals (Pb, Cd, As and Hg) in Black sea grey mullet

- (Mugilcephalus). *Bulg.J. Agric. Sci.*, 1: 30-34.
- Staniskiène, B.; Matusevicius, P.; Budreckiène, P. and Skibniewska, K.A. 2006. Distribution of heavy metals in tissues of freshwater fish in Lithuania. *Polish J. Environ. Studies*, 15(4): 585-591.
- Tajiri, A.; Daisy Pontes, D.; Sassahara, M.; Rodrigues, M.S. and Cavalcanti, C.A. 2011. Determination of presence and quantification of cadmium lead and copper in Nile tilapia (*Oreochromis niloticus*) fillets obtained from three cold storage plants in the state of Parana, Brazil. *Food Science and Technology (Campinas) .Ciênc. Tecnol. Aliment.* ,31 (2):
- Turkmen, M.; Turkmen, A. and Tepe, Y. 2011. Comparison of metals in tissues of fish From Paradeniz Lagoon in the Coastal Area of Northern East Mediterranean *Bull Environ. Contam. Toxicol.*, 87:381–385.
- Vázquez-Boucard C, Anguiano-Vega G, Mercier L, Rojas del Castillo E. 2014. Pesticide residues, heavy metals, and DNA damage in sentinel oysters *Crassostrea gigas* from Sinaloa and Sonora, Mexico. *J. Toxicol. Environ. Health A.* , 77(4):169-176.
- Xu, J.; Yan, H.C.; Yang, B.; Tong, L.S.; Zou, Y.X. and Tian, Y. 2009. Effects of lead exposure on hippocampal metabotropic glutamate receptor subtype 3 and 7 in developmental rats. *J. Negat. Results Biomed.*, 20: 8-15.