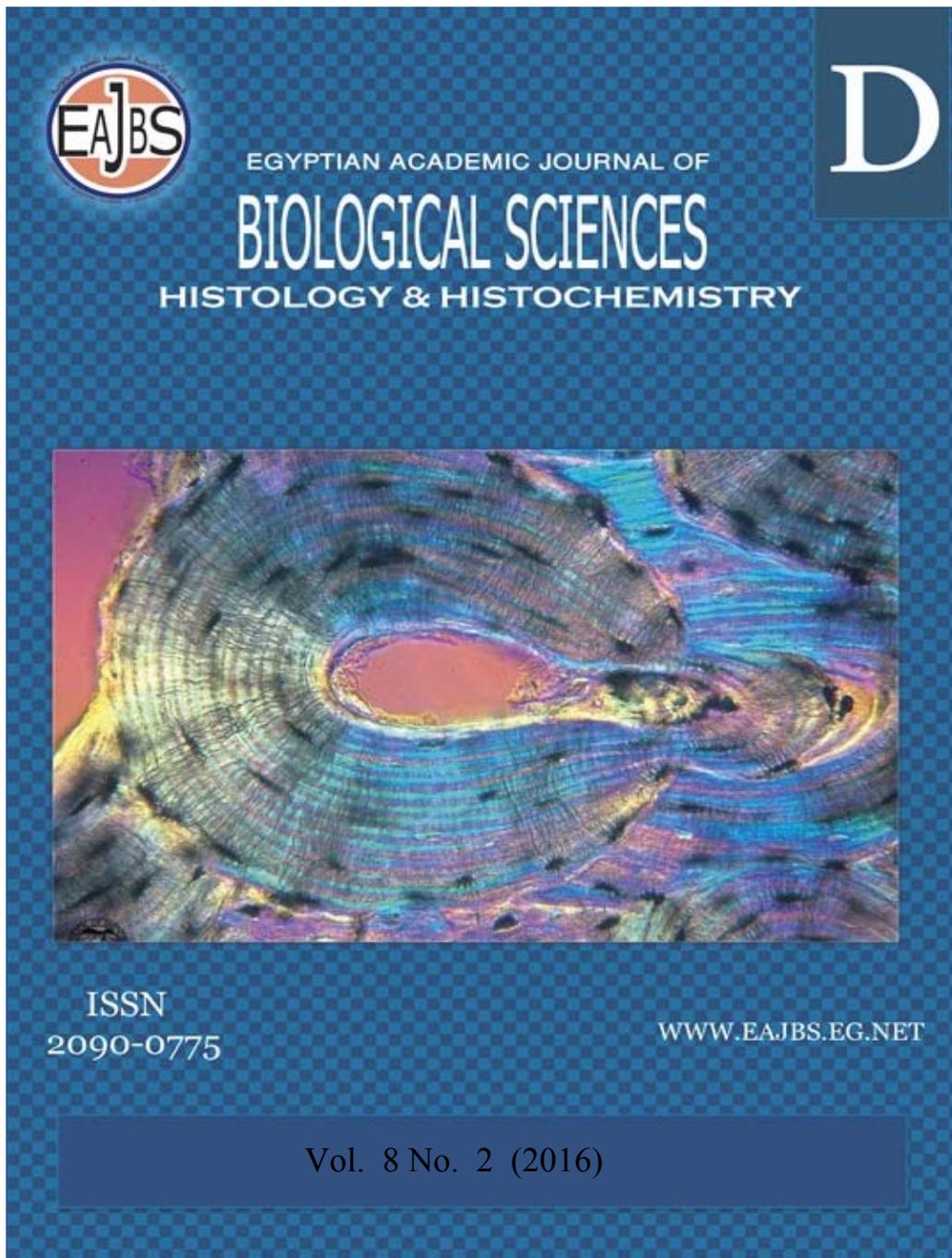


Provided for non-commercial research and education use.  
Not for reproduction, distribution or commercial use.



Egyptian Academic Journal of Biological Sciences is the official English language journal of the Egyptian Society for Biological Sciences ,Department of Entomology ,Faculty of Sciences Ain Shams University .

Histology& Histochemistry Journal include various morphological, anatomical, histological, histochemical, toxicological , physiological changes associated with individuals, and populations. In addition, the journal promotes research on biochemical and molecular-biological or environmental, toxicological and occupational aspects of pathology are requested as well as developmental and histological studies on light and electron microscopical level, or case reports..

[www.eajbs.eg.net](http://www.eajbs.eg.net)



## Impact of Induced Thyroxine and Carbimazole Vacillation on Liver of Female Rats

Heba A. Hashem<sup>1</sup>, Hagar El-Metwaly<sup>2</sup>, Yomn M. Mobarak<sup>3</sup>, Zohour N. Ibrahim<sup>3</sup>

1- Joint laboratory for food and water analysis, Directorate of health affairs, Port-Said, Egypt.

2- Faculty of Science, Al-Arish University, North-Sinai, Egypt.

3- Department of Zoology, Faculty of Science, Suez Canal University, Ismailia, Egypt.

E-mail: [heba\\_heroo@yahoo.com](mailto:heba_heroo@yahoo.com)

### ARTICLE INFO

#### Article History

Received:29/5/2016

Accepted:1/7/2016

#### Keywords:

Levothyroxine,  
Carbimazole,  
hyper- hypothyroidism  
physiology,  
histopathology,  
female rats

### ABSTRACT

The present study was designed to investigate the effects of levothyroxine and carbimazole on body weight gain, liver functions, and histology of female rats. Rats were divided into three groups (12 rats/group), with each group divided into two sub-groups (6 rats/group) based on the period of treatment for three and six weeks. At the end of the study, the rate of body weight gain of hyperthyroid rats exhibited a significant decrease after three and six weeks. The rate of body weight gain of hypothyroid female rats displayed a significant decrease after three weeks only. Liver to body weight ratio (L/BW) of hyperthyroid rats displayed a significant increase after three and six weeks. However, (L/BW) of hypothyroid rats manifested no significant change and a significant decrease, respectively after three and six weeks of study versus control. The rate of food consumption was different in the three groups and even in the same group at different times. The liver function tests were found changed in both hyper- and hypothyroid female rats. The alanine aminotransferase (ALT) and aspartate aminotransferase (AST) levels of both hyper- and hypothyroid rats displayed a significant increase after three and six weeks. Reduced glutathione (GSH) levels exhibited a significant decrease after three weeks and a significant increase after six weeks. On other hand, GSH of the hypothyroid rats displayed no significant change after three and six weeks. The present study also showed a histopathological damage in liver of either hyper- or hypothyroid rats. These results suggested that both hyper- and hypothyroidism induced adverse effects on liver physiology and histology in female rats.

### INTRODUCTION

Thyroid gland is a unique large endocrine gland being largest, superficially located in the neck region and being amenable to physical examination and biopsy (Zaidi *et al.*, 2004). The human's thyroid gland is a brownish-red organ consists of two lobes connected by an isthmus and weighs between 20 - 25 grams (Choksi *et al.*, 2003; Fox, 2008). It has a very rich blood supply and it requires iodine to function properly (Petersen, 2007). The gross structure of thyroid gland of laboratory animals is similar to that described for human (U.S. EPA, 1998). The thyroid gland secretes two iodine containing amine hormones; 3, 5, 3', 5' tetraiodo-L-thyronine (T4 or thyroxine) and 3, 5, 3' triiodo-L-thyronine (T3) (Diekman *et al.*, 2000).

Thyroid hormones (THs) influence the functions of all body organs and cells (Soliman, 2013). No other hormone affects such wide range of cells and tissues such as THs (Capuco, 2001). Animal and human studies indicated that THs play a role in the cardiovascular, nervous, immune, and reproductive system development and function (Krassas, 2000; Choksi *et al.*, 2003). In addition, they mediate several physiological processes, including embryonic development, cellular differentiation, metabolism, and the regulation of cell proliferation (Wu *et al.*, 2013). Also, they are responsible for regulation of oxygen consumption, thermogenesis and lipogenesis (Capuco, 2001).

Thyroid dysfunctions are considered as some of the most important endocrinopathies both in human and in veterinary medicine (Shi *et al.*, 2002; Rijnberk *et al.*, 2003). When T4 and T3 levels are too low or too high, the hypothalamus and pituitary glands start to regulate their production (Hadley, 1996). Hypothyroidism is resulted usually from damage, removal, or inhibition of the function of the thyroid gland (Braverman and Utiger, 2005; Daniels & Dayan, 2006; Mitrou *et al.*, 2011). On the other hand, hyperthyroidism is defined as the clinical syndrome of hyper-metabolism resulting from increased freeT4 and/or free T3 serum levels (Braverman and Utiger, 2005; Saraji *et al.*, 2012). Therefore, the purpose of this study was to compare the effects of both hyper- and hypothyroidism on body weight gain, liver weight, food consumption level, glutathione reduced (GSH) level, liver physiology and histology of female rats.

## MATERIALS AND METHODS

### Animals

36 healthy female albino rats weighing between 120-140g were

obtained from a colony at the animal house of Abu-Rawash, Giza, Egypt. The female rats were maintained under constant light, humidity, and temperature in the animal house of the faculty of pharmacy - Sinai University, Al-Arish, North-Sinai, Egypt. Rats were allowed free access of food and water *ad libitum*, and were acclimatized a week before the starting of the experiment. The rats were divided into three groups (12 rats/group), then each group was further divided into two sub-groups (6 rats/group) based on the period of treatment, three and six weeks as follows:

**Group 1 (Euthyroid):** served as a control group for the other two groups and received distilled water orally.

**Group 2 (Hyperthyroid):** received thyroxine + distilled water orally.

**Group 3 (Hypothyroid):** received carbimazole + distilled water orally.

### Chemicals and Reagents

All reagents used for the present study were of the analytical grade. Thyroxine in the form of commercial tablets (Eltroxin) was obtained from GlaxoSmithKline GmbH, Germany. Carbimazole in the form of commercial tablets (Carbimazole) was obtained from Chemical Industries Development (CID). T3 and T4 ELISA kits were purchased from Biocheck, Inc Company, Foster City, U.S.A. Glutathione reduced kit was purchased from Biodiagnostics Company, Dokki, Egypt. Kits for quantitative determination of Alanine aminotransferase (ALT), Aspartate aminotransferase (AST), albumin and total protein (T.Potein) were purchased from Spinreact Company, Ctra.Santa Coloma, Spain.

### Experimental Protocol

In the present study, 600 µg/kg of thyroxine and 1.35 mg/kg of carbimazole were orally administrated to female rats for induction of hyper- and hypothyroidism, respectively for three and six weeks. These doses were selected

after Chakrabarti *et al.* (2007), Paget and Barnes (1964). The body weight of each animal as well as food consumption rate of each group was recorded at the start and on alternate days until the end of experiments. At the end, the female rats were weighed, fasted for 12-14 hrs, anaesthetized with di-ethyl ether and dissected. Then, blood was drawn by a ventricular cardiac puncture. For the estimation of GSH, 1ml of the fresh blood samples was transferred into test tubes that contained an anti-coagulant agent (EDTA). The rest of the blood was transferred into test tubes without any additives where it was further centrifuged at a speed of 3,000 rpm for 10 min and the clear serum was collected in sterilized, properly labeled plastic tubes and frozen at -20°C till use. Serum was used for determination of THs and liver functions. Thyroid hormones were assayed by the quantitative method using ELISA kit. ALT, AST, Albumin and T.protein were assayed by the methods of Reitman and Frankel (1957), Gendler (1984) and Koller (1984), respectively.

#### **Histological Techniques**

For histological study, rat livers were removed, washed in saline, weighed, and small pieces were fixed in aqueous Bouin's solution for 24 hours (Usenko *et al.*, 1998). After fixation, they were washed several times in 70% ethanol then dehydrated in ascending grade of ethyl alcohol concentrations and cleared using xylene. Then, liver sections were stained with hematoxylin and counter stained with eosin for routine histological study using the light microscope at (X 160 & X 640).

#### **Statistical Analysis**

The statistical software package SPSS for windows version 19.0 (SPSS Inc., Chicago, IL, USA) was used for data analysis. Data of the present study were subjected to descriptive statistics

and the results presented as Means  $\pm$  Standard Error of Mean. Data were analyzed using one-way analysis of variance (ANOVA), followed by post hoc multiple comparisons using Sheffe's multiple comparison, with  $P < 0.05$  accepted as statistically significant (Mano *et al.*, 1994; Okon *et al.*, 2013).

## **RESULTS**

### **The Body Weight Gain and liver weight**

The rate of body weight gain of hyperthyroid rats exhibited a significant ( $P < 0.001$ ,  $P < 0.004$  vs. control) decrease after three and six weeks, respectively. Also, the rate of body weight gain of hypothyroid rats exhibited a significant ( $P < 0.000$  vs. control) decrease after three weeks. However, the rate of body weight gain of hypothyroid rats showed no significant change ( $P > 0.05$  vs. control) after six weeks (Table.1). The liver to body weight ratio (L/BW) of hyperthyroid rats displayed a significant ( $P < 0.000$  vs. control) increase after three and six weeks of study. On the other hand, (L/BW) of hypothyroid rats manifested no significant ( $P > 0.05$  vs. control) change after three weeks. However, the liver to body weight ratio of hypothyroid rats showed a significant ( $P < 0.03$  vs. control) decrease after six weeks (Table.1).

### **The Level of Food Consumption**

The rate of food consumption was changed in the three groups and even in the same group but at different times. The rate of food consumption of hyperthyroid rats exhibited a significant ( $P < 0.006$ ,  $P < 0.000$ ,  $P < 0.000$ ,  $P < 0.002$ ,  $P < 0.000$ , and  $P < 0.001$  vs. control) increase after the first, second, third, fourth, fifth, and sixth week of treatments, respectively.

Table 1: Mean values of body weight gain (BWG) and liver to body weight ratio (L/BW) of the control (CON), hyperthyroid (HYR) and hypothyroid (HYO) female rats after three, and six weeks of treatment with distilled water, thyroxine, and carbimazole, respectively.

Parameter	CON-3	HYR-3	HYO-3	CON-6	HYR-6	HYO-6
BWG (g)	30±1.0	24±0.6**	14±1.0***	47±1.8	32±3.2**	42±2.5°
Change%	20%	14%	9%	31%	19%	26%
L/BW (g)	3.3±0.1	4.4±0.2***	3.3±0.1°	3.2±0.1	4.1±0.2***	2.6±0.1*
Change%	----	33%	0%	----	28%	-19%

Values are expressed as Mean ± (SE), (n= 6/group).

Key: °, P > 0.05; \*, P < 0.05; \*\*, P < 0.01; \*\*\*, P < 0.001 versus control group using Scheffe's test; one way ANOVA.

On the other hand, the rate of food consumption of hypothyroid rats showed no significant (P > 0.05 vs. control) change in the rate of food consumption after the first week of treatments. However, the rate of food consumption

of hypothyroid rats showed a significant (P < 0.02, P < 0.04, P < 0.01, P < 0.05, and P < 0.001 vs. control) decrease after the second, third, fourth, fifth, and sixth week of the treatments, respectively (Table 2).

Table 2: Mean values of food consumption of control (CON), hyperthyroid (HYR) and hypothyroid (HYO) female rats recorded from the first to sixth week of treatment with distilled water, thyroxine, and carbimazole, respectively.

Food (g)	CON	HYR	HYO
1st week	76±1.5	88±3.5**	74±1.4°
Change %	----	16%	-3%
2nd week	80±1.8	105±3.4***	70±1.0*
Change %	----	31%	-14%
3rd week	86±2.3	112±4.9***	73±2.0*
Change %	----	30%	-15%
4th week	95±3.8	117±1.6**	78±4.8**
Change %	----	23%	-18%
5th week	90±3.6	119±3.5***	77±3.3*
Change %	----	32%	-14%
6th week	98±2.6	118±1.7***	77±4.4***
Change %	----	20%	-21%

Values are expressed as Mean ± (SE), (n= 6/group).

Key: °, P > 0.05, P > 0.05; \*, P < 0.05; \*\*, P < 0.01; \*\*\*, P < 0.001 versus control group using Scheffe test; one way ANOVA.

### The Level of Thyroid Hormones

The level of T3 in hyperthyroid female rats showed a significant (P < 0.02 and P < 0.000 vs. control) increase after three and six weeks, respectively. The level of T4 in hyperthyroid female rats also showed a significant (P < 0.001 and P < 0.000 vs. control) increase after three and six weeks, respectively. However, the level of T3 in hypothyroid female rats manifested a significant (P < 0.007 and P < 0.000 vs. control) decrease after three and six weeks, respectively. The level of T4 in hypothyroid female rats also showed a significant (P < 0.05

and P < 0.000 vs. control) decrease after three and six weeks, respectively.

### The Liver Function Tests

After three weeks, ALT and AST levels of hyperthyroid female rats showed a significant (P < 0.005 and P < 0.003 vs. control) increase, respectively. However, albumin and T. protein levels showed no significant (P > 0.05 vs. control) change after the same period. The level of ALT and AST of hypothyroid female rats exhibited a significant (P < 0.03 vs. control) increase after three weeks. However, albumin and T. protein levels showed no significant (P

> 0.05 vs. control) change in albumin and T. protein levels after the same period. After six weeks, ALT and AST levels of hyperthyroid female rats showed a significant ( $P < 0.000$  and  $P < 0.001$  vs. control) increase, respectively. However, albumin and T. protein levels showed a significant ( $P > 0.02$  and  $P < 0.006$  vs. control) decrease after the same period.

The level of ALT and AST of hypothyroid female rats exhibited a significant ( $P < 0.000$  and  $P < 0.004$  vs. control) increase after three weeks. The level of albumin and T. protein also showed a significant ( $P > 0.02$  and  $P < 0.03$  vs. control) increase after the same period (Table 3).

Table 3: Mean values of T3, T4, ALT, AST, Albumin, T. protein, and GSH levels of control (CON), hyperthyroid (HYR) and hypothyroid (HYO) female rats after three and six weeks of treatment with distilled water, thyroxine, and carbimazole, respectively.

Parameter	CON-3	HYR-3	HYO-3	CON-6	HYR-6	HYO-6
T3 (ng/ml)	2.4±0.3	3.8±0.5*	0.7±0.1**	2.2±0.3	5.7±0.7***	0.3±0.9*
Change%	----	58%	-70%	----	159%	-88%
T4 (µg/dl)	7.3±0.4	11.9±0.8***	4.5±0.8*	6.8±0.6	18.5±1.1***	2.0±0.2***
Change%	----	63%	-38%	----	172%	-71%
ALT (U/L)	18±1.9	32±2.0**	29±3.6*	18±1.8	42±2.4***	35±2.3***
Change%	----	78%	61%	----	133%	94%
AST (U/L)	25±2.2	39±2.6**	36±2.4*	24±3.0	46±3.5***	42±2.6**
Change%	----	56%	44%	----	92%	75%
Albumin (g/dl)	4.1±0.2	3.8±0.2°	4.6±0.2°	4.4±0.2	3.2±0.2*	5.2±0.3*
Change%	----	-7%	12%	----	-27%	18%
T. Protein (g/dl)	6.2±0.4	5.9±0.2°	6.1±0.3°	6.3±0.2	4.9±0.3**	7.4±0.3*
Change%	----	-5%	-2%	----	-22%	17%
GSH (mg/dl)	22±2.3	12±1.3*	18±2.5	19±2.4	29±2.7*	21±2.4°
Change%	----	-45%	-18%	----	53%	11%

Values are expressed as Mean ± (SE), (n= 6/group).

Key: °,  $P > 0.05$ ; \*,  $P < 0.05$ ; \*\*,  $P < 0.01$ ; \*\*\*,  $P < 0.001$  versus control group using Scheffe test; one way ANOVA.

### The Level of Reduced Glutathione

The level of GSH of hyperthyroid rats exhibited a significant ( $P < 0.003$  vs. control) decrease after three weeks. However, hyperthyroid rats showed a significant ( $P < 0.05$  vs. control) increase after six weeks. On the other hand, the level of GSH of hypothyroid rats displayed non-significant change ( $P > 0.05$  vs. control) after neither three nor six weeks (Table 3).

### The Histopathology of Liver

The examination of histological sections of liver from control group showed normal central veins, portal triads, and a normal structure and a regular arrangement of hepatocytes with clearly visible nuclei and hepatocytes. On the other hand, the histopathological examination of liver in both hyper- and

hypothyroid rats revealed some of pathological alterations. Both levothyroxine and carbimazole treatments caused liver injury and hepatocytes destruction parallel to significant increase in liver function indices. Liver sections of both hyper- and hypothyroid rats showed disorganized hepatocyte cords and the normal lobular architecture disappeared in some areas (Fig.1).

The liver tissue of hyperthyroid rats manifested a fusion of some portal triads with the central vein of the liver lobule with presence of increased inflammatory cell infiltration. Also, there was a lobular inflammatory infiltrate in the vicinity of the enlarged and edematous portal tracts. Moreover, the hyperthyroid rats also showed different degrees of

intracanalicular bile plugs with enlarged hepatocytes mainly centri-lobular intrahepatocytic cholestasis. The intrahepatic cholestasis and mild inflammatory infiltrates were indicators of acute liver injury. The hyperthyroid rats also displayed some degree of fatty infiltration in liver tissue, steatosis. There was a vacuolar degeneration as a result of the mixed pattern of small and large lipid droplets in hepatocytes. In some hepatocytes, the nucleus was squeezed into displaced rim of cytoplasm of the fat vacuole.

On the other hand, the liver tissue of hypothyroid female rats showed cellular discontinuity with loss of hepatocytes radial distribution, hyperplasia and irregular arrangement of hepatocytes. There was also a focal necrosis within liver parenchyma; a cluster of inflammatory cells which marked the site of necrotic hepatocyte. An extensive portal tract fibrosis with a moderately dense infiltrate of mononuclear inflammatory cells was noticed. The spaces between hepatic plates were increased in some areas with progression to fibrosis (Fig. 2 ).

## DISCUSSION

During the experiments, rats from control and treated groups remained alive for their respective periods of treatment. Levothyroxine caused a significant rise in T4, T3 levels when compared with control group, confirming a hyperthyroid state. On the contrast, carbimazole caused a significant fall in T3, T4 levels when compared with control group, confirming a hypothyroid state. The general condition of both treated rats was compared with control rats throughout the study. Control rats appeared healthy, active and showed normal behavior throughout the period of experimental study. On the contrast, the hyperthyroid female rats appeared strange, hyperactive, and nervous especially during administration by gastric tube.

They always displayed fast movement in the cage and had increased appetite with decreased body weight gain. At the same time, the hypothyroid female rats appeared ill-looking and suffered from depression as they had a diminished activity and increased sleep periods. They had a sluggish behavior and exhibited decreased response to external stimuli. They also suffered from loss of appetite with reduction in weight gain in the first three weeks of the study.

According to Rajab *et al.* (2015), TH disorders led to emotional and behavioral disturbances and impair patients' everyday life. There is an association between thyroid function and psychiatric disorders particularly mood disorders (Hage and Azar, 2012). According to Marian *et al.*, 2009, hyperthyroidism might be associated with various psychiatric symptoms, such as emotional disturbances, irritability, restlessness and anxiety. However, Brown *et al.* (2005) suggested that, depression is one of the major symptoms associated with hypothyroidism.

The present hyperthyroid rats showed a significant reduction in body weight gain despite the significant increase in food consumption when compared with control rats after three and six weeks. This agrees with Ajayi *et al.* (2013) who suggested that hyperthyroid group displayed a reduction in body weight gain. Thyroid hormones caused a negative energy balance due to the increased metabolic rate & energy expenditure so; there was always a hungry feeling (Costanzo 2010; Ganong, 2010). The present hypothyroid rats displayed a very high significant reduction in body weight gain when compared with control rats after the first three weeks. After the fourth week, there was no significant change in body weight gain of hypothyroid rats when compared with control rats. This agrees with Wang *et al.* (2000) who suggested that, the body weight gain of hypothyroid rats per

week was very low (or even negative) during the whole experimental period and their body growth was practically arrested. This confirms the fact that normal concentrations of THs are required for growth to adulthood and any deficiency in THs can cause arrest in growth (Sharma *et al.*, 2013). Conversely, Perveen *et al.* (2012) displayed that there is a marked weight gain after treatment with carbimazole where weight continued to rise with time.

The thyroid status also affected (L/BW), the present (L/BW) of hyperthyroid rats showed a very high significant increase when compared with control after three and six weeks. This result is in accordance with Ajayi and Akhigbe (2012) who suggested that liver of hyperthyroid group manifested increased weight than control group. The increase in liver weight of hyperthyroid rats could be related to the increase in the rate of absorption of carbohydrates from the gastrointestinal tract (Laycock and Wise, 1983). Malik and Hodgson, (2002) also suggested the usage of T<sub>3</sub> as a hepatic growth factor as it induces hepatocyte proliferation and increases liver mass. On the other hand, the present (L/BW) of hypothyroid rats displayed no significant change after three weeks and a significant decrease after six weeks when compared with control. This result disagrees with Ajayi and Akhigbe (2012) who suggested that liver of hypothyroid group manifested increased weight than control group. In fact, T<sub>3</sub> mediates several physiological processes, including embryonic development, cellular differentiation, metabolism, and the regulation of cell proliferation (Wu *et al.*, 2013).

The present results revealed significant increase in ALT and AST enzymes activity in the serum of both hyperthyroid and hypothyroid rats when compared with control rats after three and six weeks. The increase in ALT and AST levels was higher in the

hyperthyroid rats than hypothyroid rats. The present result agrees with Messarah *et al.* (2011) who suggested an elevation in ALT and AST enzymes in the serum of hyperthyroid group. Also, Christ-Crain *et al.* (2004) suggested the elevation of ALT and AST levels in overt hypothyroidism. The present results disagrees with Khan *et al.* (1999) who found a non-significant increase in AST and no change in ALT in hyperthyroid rats.

The increase in circulating liver enzymes of hyper- and hypothyroid rats might be either due to their increased synthesis and secretion, or to diminished catabolism (Christ-Crain *et al.*, 2004). Malik *et al.* (2000) suggested that increased metabolism in hepatocytes of hyperthyroid rats might lead in turn to increased ALT and AST levels. According to Christ-Crain *et al.* (2004), hypothyroidism is associated with a decreased metabolic rate and a diminished catabolism of AST and ALT. Ajayi and Akhigbe (2012) suggested that the damage which might happen to the hepatocytes of liver of hypothyroid rats leads to leakage of enzymes from the cells.

The present albumin and T.protein levels were significantly reduced in hyperthyroid rats when compared with control rats, only after six weeks. These results agree with the findings of Jalal *et al.* (2010) and Kondaveeti *et al.* (2014). Conversely, the hypothyroid rats showed a significant increase in both albumin and T.protein levels when compared with control rats, only after six weeks. These results agree with Kondaveeti *et al.* (2014). TH is known to promote albumin catabolism (Koga *et al.*, 2009). Larsen and Davies (2002) suggested that TH promotes albumin metabolism so, the mean concentration of albumin and total protein in serum were lower in hyperthyroid patients than the hypothyroid patients. Kondaveeti *et al.* (2014) suggested that, the increased

albumin levels in hypothyroid patients along with non-clinical hyperglycemia is attributed to the deterioration of protein metabolism which further decreases turnover of proteins and increase of half-life of proteins.

The oxidative stress status was determined by the study of one of the most important antioxidants, GSH. The present GSH level of hyperthyroid rats showed a significant decrease when compared with the control rats after three weeks. The present decrease in GSH content in hyperthyroid rats reflects its consuming through the oxidative stress. This result agrees with Kumar *et al.* (2004) who suggested that GSH level was decreased significantly in the hyperthyroid group when compared with the healthy control group. Glutathione depletion might be due to its exhaustion in the oxidative stress as the increased metabolism in response to hyperthyroidism caused oxidative damage to certain organs, including liver, heart and muscles (Venditti and Di-Meo, 2006). This cellular damage occurred when the balance of oxidants and antioxidants in the body is disturbed, and the antioxidant system of the body is not able to neutralize these oxidants. The increase in oxidative stress caused peroxidation of lipids, damage to proteins and DNA fragmentation (Rapozzi *et al.*, 1999).

After six weeks, the GSH levels in hyperthyroid rats were significantly increased and slightly reached above the normal range when compared with the three weeks hyperthyroid rats and control rats. Blagojević *et al.* (1998) suggested that the antioxidant defense system is an endogenous, dynamic system incorporated in homeostatic regulation lead by internal regulatory signals. So, the body might increase the production of GSH as a response to the increased oxidative stress.

On the other hand, the present hypothyroid rats displayed no significant

change in GSH levels when compared with control rats after three and six weeks. This result agrees with Petrulea *et al.* (2012) who reported that GSH levels in the hypothyroid rats showed no significant difference from the control rats and disagrees with Sarandol *et al.* (2005) and Erdamar *et al.* (2008) who suggested an increased production of reactive oxygen species in hypothyroidism. There is a complex relationship between TH levels and oxidative stress, but the general principle is that elevated TH levels (hyperthyroidism) induce oxidative stress, whereas reduced THs levels (hypothyroidism) result in non-detectable to mild oxidative stress (Villanueva *et al.*, 2013). Cano-Europa *et al.* (2011) also suggested that a hypothyroidism-induced hypometabolic state protects against oxidative damage caused by toxins.

The present histological liver sections from control rats showed a normal liver structure including normal central vein, portal triads, and hepatocytes. The histological liver sections from both hyper- and hypothyroid rats showed a hepatic injury and hepatocytes lysis. A lobular inflammatory infiltrate of polymorphonuclear leucocytes was also found in the liver of hyperthyroid rats (fig. 1). This result agrees with Ajayi and Akhigbe (2012) who suggested that, there was a hepatic damage in the hyperthyroid rats and this damage was observed as a rise in ALT and AST levels. Saraji *et al.* (2012) also suggested that moderate parenchymal inflammation and edema were found in hyperthyroid rats. Moreover, Cano-Europa *et al.* (2010) suggested that hypothyroidism caused cell damage in the liver. The mechanism of hepatic injury appears to be relative to hypoxia in the perivenular regions, due to an increase in hepatic oxygen demand without an appropriate increase in hepatic blood flow (Malik and Hodgson, 2002). In addition to hepatic

injury, the present hyperthyroid liver sections showed different degrees of intrahepatocytic and extrahepatocytic cholestasis, mainly centrilobular (fig.2). This result agrees with Shen *et al.* (2010) suggested that, liver biopsy revealed characteristic features of intrahepatic cholestasis with mild inflammatory infiltrates indicative of acute liver injury. Cholestasis might result either from a functional defect in bile formation at the level of the hepatocyte or from an impairment in the secretion of bile and/or obstruction of flow at the bile duct level (Trauner *et al.*, 2007).

The present histological section of hyperthyroid rats also displayed a fatty change of the liver tissue, microvesicular steatosis, in which nucleus is squeezed into the displaced rim of cytoplasm about the fat vacuole (Fig. 2). This result agrees with the finding of Upadhyay *et al.* (2004) who suggested that, in most cases of hyperthyroidism and liver dysfunction without heart failure, liver histology demonstrates some degree of fatty infiltration, cytoplasmic vacuolization, nuclear irregularity, and hyperchromatism in hepatocytes.

The present hypothyroid female rats suffered from liver injury, necrosis, and fibrosis (Figs.1&2). Areas of centrilobular necrosis were observed in the present liver histological sections of hypothyroid rats. The extracellular factors inducing cell death (such as Tumor necrosis factor) increased causing damage to liver cells through the activation of an external pathway of apoptosis (Saraji *et al.*, 2012). Karadeniz *et al.* (2008) suggested that kupffer cells (macrophages) activation and infiltration contributed to tissue injury. Liver fibrosis is a common consequence of chronic liver injury (Albanis and Friedman, 2001) and it is defined as the abnormal accumulation of extracellular matrix in the liver. Cano-Europa *et al.* (2010) suggested that methimazole (anti-thyroid drug) caused cell damage in the liver of

hypothyroid objects, whereas hypothyroidism caused by thyroidectomy does not cause hepatic-cell damage. So, it seems that the main cause of the present damage of liver of female rats wasn't the hypothyroidism itself but the method of hypothyroidism induction as long as the anti-thyroid drug-induced hypothyroidism can cause cellular damage.

## REFERENCES

- Ajayi AF and Akhigbe RE. (2012). Implication of altered thyroid state on liver function. *Thyroid Research and Practice* Vol. 9 Issue 3, September-December .
- Ajayi AF, Akhigbe RE, Ajayi LO. (2013). Hypothalamic-pituitary-ovarian Axis in Thyroid Dysfunction. *West Indian Med J*; 62 (9): 835.
- Albanis E and Friedman SL. (2001). Hepatic fibrosis. Pathogenesis and principles of therapy. *Clin. Liver Dis.*; 5:315-34.
- Blagojević D, Buzadžić B, Korać B, Saičić ZS, Radojičić R, Spasić MB, Petrović VM. (1998). Seasonal changes in the antioxidative defense in ground squirrels (*Citellus citellus*): possible role of GSH-Px. *J Environ Pathol Toxicol Oncol* 17: 241-250.
- Braverman L, Utiger R.(2005). Introduction to hypothyroidism in Werner & Ingbar's. *The Thyroid: a fundamental and clinical text*. Ninth Edition, Eds: L Braverman and R Utiger. Pus: Lippincott Williams & Wilkins. Chapter 46, pps 697-9.
- Brown BT, Bonello R, and Pollard H. (2005). The biopsychosocial model and hypothyroidism. *Chiropr Osteopat*. 2005; 13: 5.
- Cano-Europa E, Blas-Valdivia V, Lopez-Galindo GE, Franco-Colin M, Pineda-Reynoso M, Hernandez-Garcia A, Ortiz-Butron R. (2010). Methimazole-induced hypothyroidism causes alteration of the REDOX environment, oxidative stress, and hepatic damage; events not caused by

- hypothyroidism itself. *Ann Hepatol.* Jan-Mar; 9(1):80-8.
- Capuco AV., Wood DL., Elsasser TH., Kahl S, Erdman RA., Van Tassell CP, lefcourt A, Piperova LS. (2001). The effect of somatotropin on thyroid hormones and cytokines in lactating dairy cows during ad libitum and restricted feed intake, *J. Dairy. Sci.*, 82, 2430-2439.
- Chakrabarti S, guria S, samanta I, Das M. (2007). Thyroid dysfunction modulates glucoregulatory mechanism in rat. *Indian Journal of experimental biology.* Vol. 45, June, pp. 549-553.
- Choksi NY, Jahnke GD, Hilaire C St., and Shelby M (2003). Role of Thyroid Hormones in Human and Laboratory Animal Reproductive Health. *Birth Defects Res. (Part B)* 68:479–491.
- Christ-Crain M, Meier C, Puder J, Staub J J, Huber P R, Keller U, Müller B. (2004). Changes in Liver Function correlate with the Improvement of Lipid Profile after Restoration of Euthyroidism in Patients with Subclinical Hypothyroidism. *EXCLI Journal*; 3:1-9.
- Costanzo LS. (2010). *Endocrine Physiology In: Physiology*, 4Ed, 9: 401 – 409, Saunders, Inc., an affiliate of Elsevier, Inc.
- Daniels E G, Dayan C. (2006). Hypothyroidism, oetiology and presentation in *Fast Facts: Thyroid Disorders*, Pub: Health Press Ltd, UK. Chapter 4, pps 69-78.
- Diekman MJ, Angheliescu N, Endert E, Bakker O, Wiersinga WM. (2000). Changes in plasma low-density lipoprotein (LDL)-and high-density lipoprotein cholesterol in hypo- and hyperthyroid patients are related to changes in free thyroxine, not to polymorphisms in LDL receptor or cholesterol ester transfer protein genes. *J Clin Endocrinol Metab.*; 85: 1857-1862.
- Erdamar H, Demirci H, Yaman H, Erbil MK, Yakar T, Sancak B, Elbeg S, Biberoglu G, Yetkin I. (2008). The effect of hypothyroidism, hyperthyroidism, and their treatment on parameters of oxidative stress and antioxidant status. *Clin Chem Lab Med*; 46: 1004-1010.
- Fox CS, Pencina MJ, D'Agostino RB, Murabito JM, Seely EW and Pearce EN. (2008). Relations of thyroid function to body weight. *Arch. Intern. Med.*, 168(6): 587-592.
- Ganong WF. (2010). The thyroid gland. In: *Review of Medical Physiology.* 23<sup>rd</sup> edition. By Kim E. Barrett, Susan M. Barman, Scott Boitano, Heddwen L. Brooks; McGraw Hill company. Chapter 20: 310-312.
- Gendler S. (1984). Uric acid. Kaplan A *et al.* *Clin Chem.* The C.V. Mosby Co. St Louis. Toronto. Princeton; 1268-1273 and 425.
- Hadley ME. (1996). *Endocrinology*, 4th ed. Prentice Hall, New Jersey; pp. 290-313
- Hage MP and Azar ST. (2012). The Link between Thyroid Function and Depression. *Journal of Thyroid Research*, Volume 2012, Article ID 590648, 8 pages.
- Jalal NA, Al-Samarrai AHM, Al-Tikriti KA. (2010). Biochemical changes in patints with hyperthyroidism. *Tikrit Journal of Pure Science* Vol.15 No.1.
- Karadeniz G, Acikgoz S, Tekin I O, Tascýlar O, Gun B D, CömertI M. (2008). Oxidized low-density lipoprotein accumulation is associated with liver fibrosis in experimental cholestasis; 64:531-40.
- Khan TM, Malik S, Diju IU. (2010). Correlation between plasma thyroid hormones and liver enzymes level in thyrotoxic cases and controls in hazara. *J Ayub Med Coll Abbottabad* 22 (2).
- Koga M, Murai J, Saito H, Matsumoto S, Kasayama S. (2009). Effects of thyroid hormone on serum glycated albumin levels: study on non-diabetic

- subjects. *Diabetes Res Clin Pract.* May; 84 (2):163-167.
- Koller A. (1984). Total serum protein. Kaplan A *et al.* *Clin Chem. The C.V.* Mosby Co. St Louis. Toronto. Princeton; 1316-1324 and 418.
- Kondaveeti SB, Prakash B, Shaker IA. (2014). Estimation of glycated albumin levels in various thyroid disorders. *Int J Cur Res Rev*, May / 6 (09).
- Krassas GE. (2000). Thyroid disease and female reproduction. *Fertil Steril* 74: 1063-1070.
- Kumar KM, Bobby Z, Selvaraj N, Kumar Das A, Chandra Koner B, Sen SK, Ramesh R, Ranganathan P. (2004). Possible link between glycated hemoglobin and lipid peroxidation in hyperthyroidism. *Clin Chim Acta.* Apr; 342(1-2):187-92.
- Larsen P, Davies T. (2002). Hypothyroidism and thyroiditis. In Williams Textbook of Endocrinology. 10<sup>th</sup> ed. Larsen PR, Kronenberg HM, Melmed S, Polonsky KS, Eds. Maryland Heights, Missouri, Saunders Elsevier, p. 423–455.
- Laycock JF, Wise PH. (1983). The thyroid. In: of cardiac bioenergetics: role Essential Endocrinology. Second Edn, Oxford University Press, Oxford, pp: 193-230.
- Malik R, Hodgson H. (2002). The relationship between the thyroid gland and the liver. *Q J Med*, 95:559-69.
- Mano T, Kawakubo A, Yamamoto M. (1994). Glucose and insulin metabolism in patients with hyperthyroidism due to grave's disease. *Nagoya J. Med. Sci.*, 57: 61-68.
- Marian G, Nica AE, Ionescu BE, and Ghinea D. (2009). Hyperthyroidism—cause of depression and psychosis: a case report. *J Med Life.* Nov 15; 2(4): 440–442.
- Messarah M, Saoudi M, Boumendjel A, Boulakoud MS, Feki AE. (2011). Oxidative stress induced by thyroid dysfunction in rat erythrocytes and heart. *Environ Toxicol Pharmacol.*, 31(1):33-41.
- Mitrou P, Raptis SA, Dimitriadis G. (2011). Thyroid disease in older people. *Maturitas*; V.70, I. 1:P. 5-9.
- Okon UA, Nku CO, Udobang JA, Uwah AF. (2013). Comparative Effect of Carbimazole, Glycine Max and Citrus Sinensis on Serum Electrolytes and Urea. *RJPBCS*, 4(2): 395.
- Paget GE and Barnes JM. (1964). Toxicity Tests in Evaluation of Drug Activities Pharmacometrics, D. R. Laurence and A. L. Bacharach, Eds., London and New York: Academic Press.
- Perveen K, Rafique M, Rukhsana N and Khan N. (2012). Comparison of body weight, absolute and relative weight of pituitary gland in carbimazole and carbimazole plus thyroxin treated male albino. *Pakistan J. Pharmacology*, .29(1): 17-23.
- Petrulea M, Muresan A, Duncea I. (2012). Oxidative stress and antioxidant status in hypo- and hyperthyroidism. In *The Antioxidant Enzyme*, Chapter 8, pp. 197-236. Ed M.A. El-Missiry. Croatia: Intech Open Access Publisher.
- Rajab NMA, Ukropina M, Cakic-Milosevic M. (2015). Histological and ultrastructural alterations of rat thyroid gland after short-term treatment with high doses of thyroid hormones. *Saudi Journal of Biological Sciences*.
- Rapozzi V, Comelli M, Mavelli I, Sentjurc M, Schara M, Perissin L. (1999). Melatonin and oxidative damage in rats liver induced by the prooxidant antitumor drug, adriamycin. *In Vivo*; 13(1): 45-50.
- Reitman S, Frankel S. (1957). A colorimetric method for the determination of serum glutamic oxaloacetic and glutamic pyruvic transaminases. *American journal of clinical pathology*, 28: 56-63.

- Rijnberk A, Kooistra HS, Jan-Mol A. (2003). Endocrine diseases in dogs and cats: similarities and differences with endocrine diseases in humans. *Growth Horm. IGF Res.*, 13: S158-S164.
- Saraji A A, Doroudian M, Soezi M, Eydi A, Olia P B A, Bagheri H. (2012). The effect of hyperthyroidism on the levels of liver enzymes in adult male Wistar rats. *Journal of Paramedical Sciences (JPS) Autumn Vol.3, No.4.*
- Sarandol E, Tas S, Dirican M, Serdar Z. (2005). Oxidative stress and serum paraoxonase activity in experimental hypothyroidism: effect of vitamin E supplementation. *Cell Biochem Funct*; 23:1-8.
- Sharma AK, Arya R, Mehta R, Sharma R, Sharma AK. (2013). Hypothyroidism and cardiovascular disease: factors, mechanism and future perspectives, *Curr Med Chem.* 20(35):4411-8.
- Shen C, Zhao C-Y, Liu F, Wang Y-D, Yu J. (2010). Acute-on-chronic liver failure due to thiamazole in a patient with hyperthyroidism and trilogy of Fallot: case report. *BMC Gastroenterology*, 10:93.
- Shi Y, Ritchie JWA, Taylor PM. (2002). Complex regulation of thyroid hormone action: multiple opportunities for pharmacological intervention. *Pharmacol. Therap.*, 94: 235-251.
- Soliman G Z A. (2013). Effects of Hyperthyroidism on Lipid Profile, adiponectin and liver function tests in male albino rat. *Indian journal of applied research.*, 3: Issue: 9.
- Trauner M, Fickert P, Wagner M. (2007). MDR3 (ABCB4) defects: a paradigm for the genetics of adult cholestatic syndromes. *Semin Liver Dis.* 27:77-98.
- Upadhyay G, Singh R, Kumar A, Kumar S, Kapoor A, Godbole MM. (2004). Severe hyperthyroidism induces mitochondria-mediated apoptosis in rat liver. *Hepatology*, 39(4): 1120-30.
- U.S. EPA (United States Environmental Protection Agency). (1998). Assessment of thyroid follicular cell tumors. Washington, DC: EPA Document No. DC EPA/630/R-97/002.
- Usenko VS, Lepekhn EA, Kornilovska IN, Lyzogubov VV, Apostolov EO, Ralets IS, Witt M. (1998). Immunohistochemical study of fibronectin and thyroglobulin in the thyroid gland of female rats after exposure to radioactive iodine. *Anat Rec. Dec*; 252(4):600-7.
- Venditti P, Di-Meo S. (2006). Thyroid hormone-induced oxidative stress. *Cell Mol Life Sci.*; 63(4): 414-34.
- Villanueva I, Alva-Sánchez C, and Pacheco-Rosado J. (2013). The Role of Thyroid Hormones as Inductors of Oxidative Stress and Neurodegeneration. Hindawi Publishing Corporation, *Oxidative Medicine and Cellular Longevity* Volume, Article ID 218145, 15 pages.
- Wang JL, Chinookoswong N, Yin S, Shi ZQ. (2000). Calorigenic actions of leptin are additive to, but not dependent on, those of thyroid hormones. *A J Physiol Endocrinol Metab* 279: E1278-1285.
- Wu SM, Cheng WL, Lin CD, Lin KH. (2013). Thyroid hormone actions in liver cancer. *Cell Mol Life Sci.*, 70(11):1915-36.
- Zaidi TM, Khan AA, Hasan BM and Faruqi AN. (2004). Carbimazole Induced Thyroid Histopathology in Albino Rats during Development. *J. Anat. Soc. India* 53 (2) 14-17.

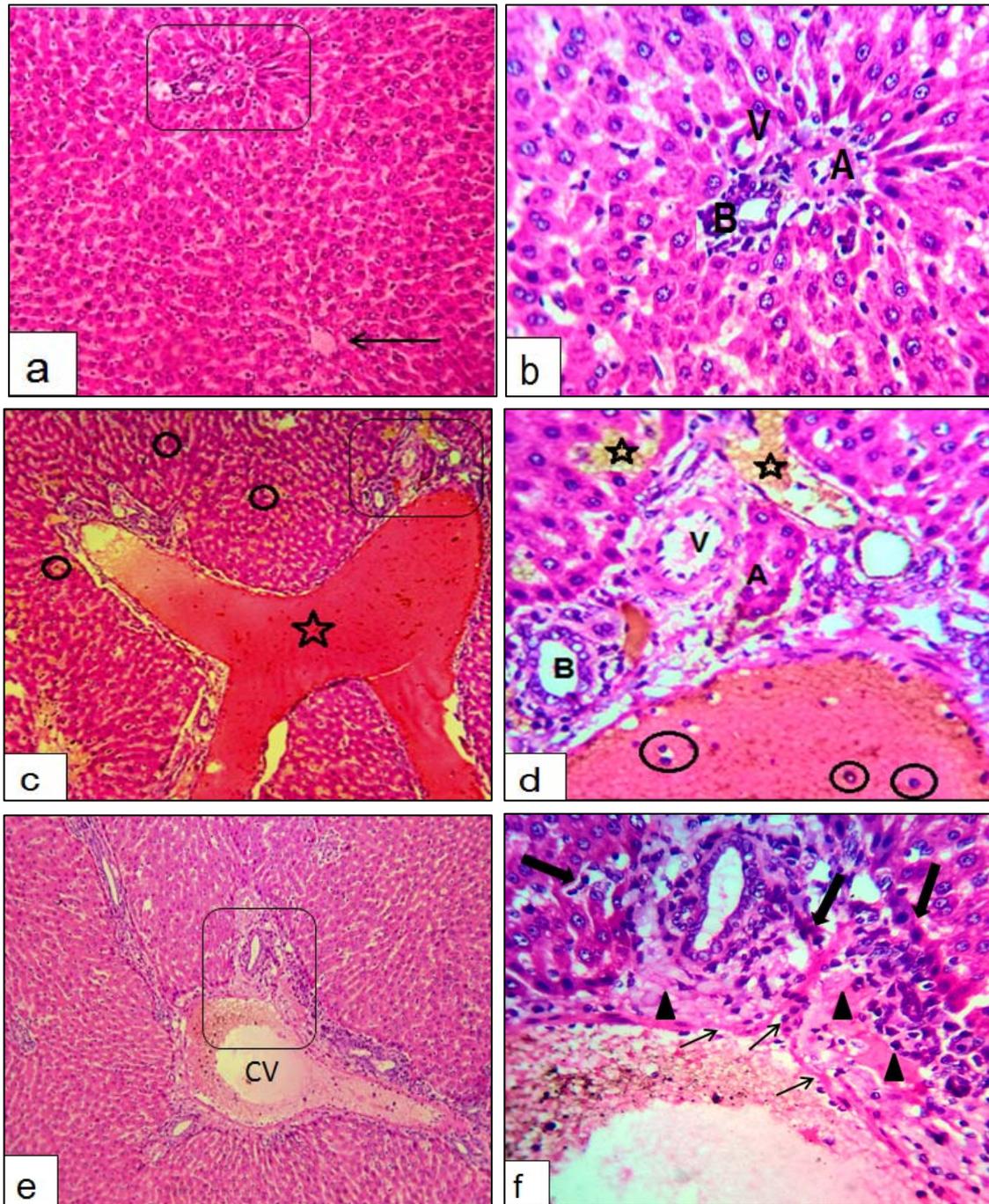


Fig.1: a) Section of liver tissue from a control rat after 6 weeks of treatment with dist. water only, showing a portal triad (square) and a central vein (thin arrow) (X 160).  
 b) High magnification of the portal triad of Figure 1a to show the portal vein (V), hepatic artery (A) and the bile duct (B) (X 640).  
 c) Section of liver tissue from a hyperthyroid rat after 6 weeks of treatment with thyroxine, showing the collapse of the liver parenchyma in which a number of portal triads collapsed together, the edematous vein (star), bile accumulation (circle) (X 160).  
 d) High magnification of one of the collapsed portal triad of Figure 1c to show the damage in the wall of portal tracts, portal vein (V), portal artery (A), and bile duct (B) (X 640).  
 e) Section of liver tissue from a hypothyroid rat after 6 weeks of treatment with carbimazole, showing the collapse of some portal triads together, the central vein (CV) (X 160).  
 f) High magnification of one of the portal triad of Figure 1e magnified to show the damaged wall of central vein (thin arrows) and pyknotic nuclei (thick arrows) of necrotic tissue (triangles) (X 640).

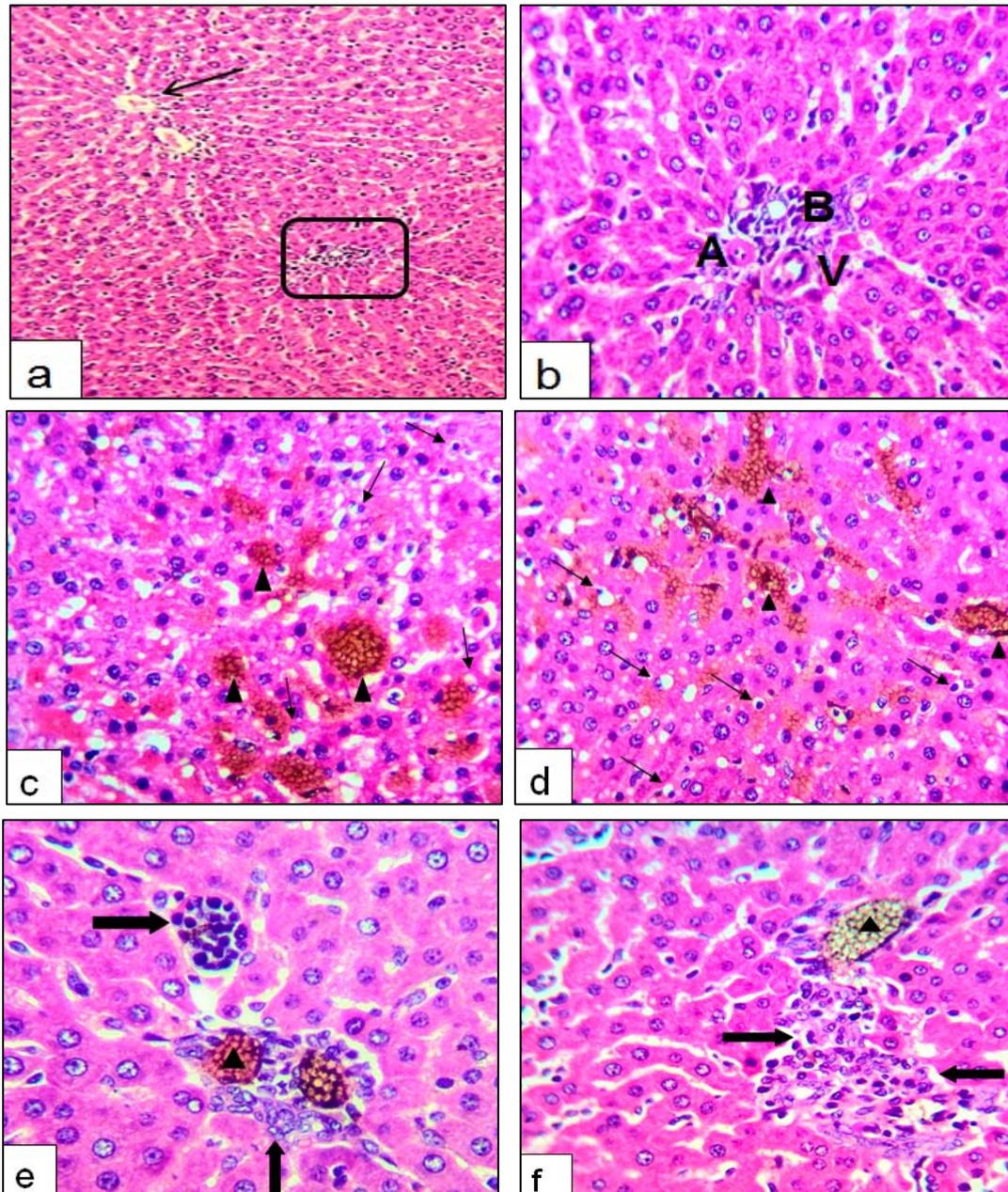


Fig. 2: a) Section of liver tissue from a control rat after 6 weeks of treatment with dist. water only, showing a portal triad (square) and a central vein (arrow) (X 160).  
 b) High magnification of the portal triad of Figure 2a to show the portal vein, hepatic artery and the bile duct. There is a normal appearance of the hepatic cells are normal (X 640).  
 c) Section of liver tissue from a hyperthyroid rat after 6 weeks of treatment with thyroxine, showing the fatty change of the liver tissue (microvesicular steatosis) (thin arrows). In some of cells, the nucleus is squeezed into the displaced rim of cytoplasm about the fat vacuole. There are also liver congestion and bile accumulation (cholestasis) (triangles) (X 640).  
 d) Another section of liver tissue from a hyperthyroid rat after 6 weeks of treatment with thyroxine, showing showing the fatty change of the liver tissue (microvesicular steatosis) (thin arrows). In some of cells, the nucleus is squeezed into the displaced rim of cytoplasm about the fat vacuole. There are also liver congestion and bile accumulation (triangles) (X 640).  
 e) Section of liver tissue from a hypothyroid rat after 6 weeks of treatment with carbimazole, showing a centizonal necrosis with an aggregation of the pycnotic nuclei of lysed hepatocytes (thick arrow) and bile accumulation (CV) (X 640).  
 f) Another section of liver tissue from a hypothyroid rat after 6 weeks of treatment with carbimazole, showing the bile accumulation (triangles) and the pyknotic nuclei of the necrotic cells (thin arrows) (X 640).

## ARABIC SUMMARY

## التأثير المعاكس للثيروكسين والكاربيمازول على الكبد في إناث الجرذان

هبة أحمد هاشم محمود<sup>١</sup>، هاجر المتولى<sup>٢</sup>، يمن مبارك<sup>٣</sup>، زهور نبيل<sup>٣</sup>

١- مجمع المعامل، مديرية الشؤون الصحية، بورسعيد، مصر

٢- كلية العلوم، جامعة العريش، شمال سيناء، مصر

٣- كلية العلوم، جامعة قناة السويس، الاسماعيلية، مصر

استهدفت الدراسة الحالية التحقق من بعض التأثيرات الفسيولوجية والهيستولوجية لإثنين من المواد الكيميائية وهما ( الثيروكسين - الكاربيمازول ) على وظائف الجسم المختلفة وتركيب الأنسجة في إناث الجرذان. في نهاية الدراسة أظهرت إناث الجرذان التي تعانى من فرط نشاط الغدة الدرقية إنخفاض ملحوظ في معدل زيادة وزن الجسم عند مقارنتها بالمجموعة الضابطة بعد ثلاثة وستة أسابيع من الدراسة. أظهرت أيضا إناث الجرذان التي تعانى من قصور في الغدة الدرقية إنخفاض ملحوظ في معدل زيادة وزن الجسم بعد ثلاثة أسابيع ولم يوجد فرق ملحوظ في معدل زيادة وزن الجسم بعد ستة أسابيع من الدراسة عند مقارنتها بالمجموعة الضابطة. كما أظهرت إناث الجرذان التي تعانى من كلا من فرط نشاط الغدة الدرقية وقصور في الغدة الدرقية زيادة في الوزن النسبي للغدة الدرقية عند مقارنتها بالمجموعة الضابطة بعد ثلاثة وستة أسابيع من الدراسة. وأتضح أيضا أن الوزن النسبي للكبد في إناث الجرذان التي تعانى من فرط نشاط الغدة الدرقية قد زاد بطريقة ملحوظة عند مقارنته مع المجموعة الضابطة بعد ثلاثة وستة أسابيع من الدراسة وفي الناحية الأخرى لم يتغير الوزن النسبي للكبد في إناث الجرذان التي تعانى من فرط نشاط الغدة الدرقية عند مقارنتها بالمجموعة الضابطة بعد ثلاثة وستة أسابيع من الدراسة. كما وجد أيضا إختلاف في درجة حرارة الجسم عند قياسها من خلال فتحة الشرج فأظهرت إناث الجرذان التي تعانى من فرط نشاط الغدة الدرقية إرتفاع ملحوظ في قيم درجة حرارة الجسم عند مقارنتها بالمجموعة الضابطة بعد ثلاثة وستة أسابيع من الدراسة وعلى العكس فقد أظهرت إناث الجرذان التي تعانى من قصور في الغدة الدرقية إنخفاض ملحوظ في قيم درجة حرارة الجسم عند مقارنتها بالمجموعة الضابطة بعد ثلاثة وستة أسابيع من الدراسة. كما تبين أن معدل إستهلاك الطعام كان مختلفا بين المجموعات الثلاثة وأحيانا كان مختلفا في نفس المجموعة ولكن في أوقات مختلفة.

أظهرت أيضا إناث الجرذان التي تعانى من فرط نشاط الغدة الدرقية إرتفاع ملحوظ في مستوى هرموني T3 و T4 في الدم عند مقارنتها بالمجموعة الضابطة بعد ثلاثة وستة أسابيع من الدراسة. وعلى العكس فإنناث الجرذان التي تعانى من قصور في الغدة الدرقية عانت من إنخفاض ملحوظ في مستوى هرموني T3 و T4 في الدم عند مقارنتها بالمجموعة الضابطة بعد ثلاثة وستة أسابيع من الدراسة. أيضا مستويات ALT و AST قد إزدادت بشكل ملحوظ في إناث الجرذان التي تعانى من كلا من فرط نشاط وقصور الغدة الدرقية عند مقارنتها بالمجموعة الضابطة بعد ثلاثة وستة أسابيع من الدراسة. أما عن مستويات كلا من Albumin و T.Protein فلم تتغير في كلتا المجموعتين عند مقارنتهم بالمجموعة الضابطة بعد ثلاثة أسابيع من الدراسة. ولكن بعد ستة أسابيع من الدراسة عانت إناث الجرذان المصابة بفرط نشاط الغدة الدرقية من إنخفاض ملحوظ في مستويات كلا من Albumin و T.Protein وعلى العكس إناث الجرذان المصابة بقصور في الغدة الدرقية قد عانت من إرتفاع ملحوظ في مستويات كلا من Albumin و T.Protein عند مقارنتهم بالمجموعة الضابطة. ومن ناحية أخرى فقد أظهرت إناث الجرذان المصابة بفرط نشاط الغدة الدرقية إنخفاض ملحوظ في مستوى GSH في الدم بعد ثلاثة أسابيع مع إرتفاع مستواه في بالدم بعد ستة أسابيع عند مقارنتها بالمجموعة الضابطة. كما لم تظهر إناث الجرذان المصابة بقصور في الغدة الدرقية أى تغيير ملحوظ في مستوى GSH في الدم بعد ثلاثة وستة أسابيع من الدراسة عند مقارنتها بالمجموعة الضابطة. أظهر الفحص الميكروسكوبي لقطاعات الغدة الدرقية والكبد أيضا تضرر أنسجتهما. وتشير كل هذه النتائج السابقة إلى أن كلا من فرط نشاط وقصور الغدة الدرقية قد تسبب في آثار خطيرة على وظائف الجسم في إناث الفئران.