Evaluation of Interleukin-9 Serum Level and Gene Polymorphism in A Sample of Systemic Lupus Erythematosus in Iraqi Female's Patients Marwah Mohammed Jasim¹, Dunya Fareed Salloom¹

Department of Biology, College of Science, University of Baghdad, Baghdad, Iraq

*Corresponding author: marwamohammed90n@gmail.com, mobile: +9647707373618

ABSTRACT

Background: Immunological gene and serum level for interleukin-9 rs 17317275 have been established to have linked to predisposition systemic lupus erythematosus (SLE) and its severity. SLE is a severe, systemic autoimmune disease characterized by autoantibody generation, complement activation, and immune complex deposition. In the pathophysiology of SLE, cytokines have a pleiotropic function. Recently, IL-9 was discovered to mediate strong anti-inflammatory effects in numerous cells or experimental autoimmune models.

Objective: This study aimed to determine the role of age, IL-9 serum level and genetic polymorphism, C-reactive protein (CRP), Anti-nuclear antibody (ANA) and Anti- double-stranded DNA (anti-dsDNA) to recognize SLE pathogenesis.

Materials and Methods: This case-control study was carried out in Baghdad Teaching Hospital and Typical Rheumatology Unit through the period from October 2021 to January 2022.103 Iraqi patients with SLE illness and 50 healthy Iraqis were included. Blood samples were taken. Serum IL-9 levels measured using the sandwich enzyme-linked immune-sorbent assay technology (ELISA) and cytokine genotyping by using allele-specific PCR technique

Results: SLE groups had greater levels of IL-9 than did healthy volunteer. Furthermore, both autoantibodies (ANA, Ant-dsDNA) were positive in SLE patients. The findings indicated that CRP was much higher in SLE subjects than those of healthy controls. On the other hand, the results showed that there was non-significance (p-value > 0.05) in age of studied groups. Concerning genotyping frequencies, the GG and AG genotyping were greater in the SLE group compared to the healthy subjects, while the AA genotyping frequency was significantly lower in the SLE group compared to the healthy people.

Conclusion: This study indicated the major roles of the serum level and genetic polymorphism of IL-9, CRP, ANA and anti-ds DNA in pathogenesis and severity of SLE.

Keywords: SLE, IL-9, Polymerase chain reaction, ANA.

INTRODUCTION

SLE is a complex and heterogeneous autoimmune condition that usually affects women throughout their reproductive years. It is caused by a breakdown in immunological tolerance and the interaction of SLE susceptibility genes with diverse environmental variables, which results in the creation of pathogenic autoantibodies ⁽¹⁾. SLE has an etiology that is unknown. Several potential factors have been identified and demonstrated to increase disease susceptibility or to stimulate the immune system resulting in an inflammatory response that eventually causes the illness to develop. Predisposition to SLE is influenced by (2) factors Autoantibody genetic generation. inflammation, and endorgan damage emerge from a complex combination of genetics, environment, and hormones that cause immunological dysregulation and the lss of self-antigen tolerance (3).

Besides, immune dysfunction, inflammation, and organ damage are all caused by cytokine abnormalities. Cytokine production is increased when both the innate and adaptive immune systems are disordered ⁽⁴⁾. They are small, soluble proteins made by immune system cells and mediate the activation or functional direction of neighboring cells by attaching to their surface receptors. Communication between immune cells is mediated by cytokines, which is essential for organizing defenses against infections ⁽⁵⁾. Secreted cytokines can be seen in the bloodstream, saliva, urine, as well as in the tissues of target organs such as the skin, kidneys, and synovia. Most of the cytokines promote inflammation, although some also have immunoregulatory or anti-inflammatory activities ⁽⁶⁾.

In SLE patients, the chronic inflammatory condition can cause mortality and morbidity. Many immunological lineages' development, maturation, and activation depend greatly on cytokines. These cytokines accelerate the inflammatory process in several organs, affect the immune cells' gene expression, and the pathophysiology of SLE is influenced by many cytokines⁽⁷⁾.

In this study, we aimed to understand the role of interleukin-9 (IL-9) in the pathogenicity of SLE. It is a 144-amino acid protein with an 18-amino acid secretory signal sequence that belongs to the gamma-chain cytokine family. It was first identified as one of a growing number of cytokines that play critical roles in the growth, survival, proliferation, and differentiation of various types of cells ⁽⁸⁾.

Mast cells, NKT cells, Th2, Th17, Treg, and ILC2 cells were among the cells that generated it, although Th9 cells are the main ones ⁽⁹⁾.

The human IL-9 gene can be identified on chromosome 5 long arm ⁽¹⁰⁾. A wide spectrum of immunological functions has been attributed to IL-9 in promoting cellular and humoral immune responses ⁽¹¹⁾. IL-9 has been linked to a variety of disease pathogenic processes, most notably allergic disorders such as

asthma and atopic dermatitis. (12).

Its serum levels are raised in people with autoimmune diseases such as systemic lupus erythematosus ⁽¹³⁾ and rheumatoid arthritis⁽¹⁴⁻¹⁵⁾.

MATERIALS AND METHODS

From October 2021 to January 2022, a casecontrol study of 50 healthy control individuals with an average age of 41.09 ± 2.38 and 103 SLE female Iraqi volunteers with an average age of 35.74 ± 1.59 was done. C-RP levels in SLE and control group sera were measured using an automated technique (Roche, Germany). ANA and anti-dsDNA information was collected from Baghdad Teaching Hospital and Typical, Rheumatology Unit.

Using a commercial sandwich ELISA kit, serum from SLE patients and control groups were quantitatively examined for IL-9 serum level evaluation (Al-shkairate institution for medical supply, Jordan). The polymerase chain reaction (PCR) was used to identify the IL-9 gene in SLE patients.

The DNA was extracted from the 300 µl blood samples of all participants using the FavorPrep TM /Cultured cells Genomic DNA extraction small kit according to the manufacturer's procedure (FAVORGEN; USA).

The sample's purity and concentration were evaluated. The concentration ranged from 60 to 95 g/mL, while the purity ranged from 1.6 to 2. IL-9 genotyping was accomplished by electrophoresis on a 1.5% agarose gel at 100 V/cm2 for 30 minutes and staining with Red Safe dye (Intron Biotechnology Inc., Korea) using UV trans illuminator at 350 nm. The size of the PCR result was determined using a Universal DNA Ladder (Intron Biotechnology Inc., Korea).



Figure (1): Standard curve of IL-9 concentration (pg/ml)

Ethical approval: An approval of this study was obtained from University of Baghdad Academic and Ethical Committee. Informed consents of all patients were taken. This study was carried out in accordance with the World Medical Association Code of Ethics (Declaration of Helsinki) for studies involving humans.

Statistical Analysis

homogeneity, The linearity, and normal distribution tests for the research groups' IL-9 serum level, age, CRP, ANA, and anti-ds DNA were performed using IBM SPSS version 26.0 (IBM Corp., released 2019). Aside from the mean, standard error, and probability, the WinPepi program version 11.65 was used to compute the odd ratio, Pearson's chi-square test, and Fishers' exact probability of genotyping and allele frequency. Abramson (2011) was used to estimate genotyping and allele frequency probability. The Hardy-Weinberg online calculator was also employed. $P \le 0.05$ was significant when the p-value was adjusted using a website called false discovery rate.

RESULTS

SLE disease according to Age.

Table (1) showed that there was no significant difference (P>0.05) in age between the SLE group and the controls.

 Table (1): SLE group demographic statistics against control group

Groups	Age	mean	±	SE	Probability
	(Year	rs)			
Patients group	35.74	± 1.59			0.06
Control group	41.09	0 ± 2.38			

CRP level according studied groups

With reference to C-RP level our study documented that C-RP serum level was raised significantly (P <0.05) in SLE patient (11.61 \pm 1.31 mg/dl) in comparison with control (2.67 \pm 0.19 mg/dl) as shown in table (2).

Table (2)	: C-RP	levels	distribution	among	the	studied
orouns						

Groups	CRP level mean ± SE (mg/ dl)	Probability
Patients group	11.61 ± 1.31	0.000004
Control group	2.67 ± 0.19	

Autoantibodies (ANA and ant-ds DNA) frequency (%) in SLE group compared to controls

Ant-nuclear antibody (ANA), Ant-dsDNA were increased significantly (P<0.05) in SLE patients compared to the control group (ANA 43.2 % and Ant-dsDNA 31.3%) (Table 3).

Table	(3):	Autoantibodies	frequency	(%)	among
studied	group	os			

Groups	ANA freque	ency (%)	Probability
	Positive	Negative	
Patients	41 (43.2)	62 (56.8)	1.85 x 10 ⁻⁷
group			
Control	0 (0.0)	50 (100.0)	
group			
	Anti- dsDN		
	(%)		
	Positive	Negative	
Patients	32 (31.3)	71 (68.9)	0.000009
group			
Control	0 (0.0)	50 (100.0)	
group			

IL-9 serum level among studied groups

IL-9 serum levels were considerably higher (P< 0.05) in SLE patients compared to the control group (16.44 0.19 vs. 7.13 0.26 Pg/ml) as seen in table (4).

Table (4): The distribution of IL-9 serum levels in the

 SLE group against the control group

Groups	IL-9 mean \pm SE (Dg/ml)	Probability
Patients group	3E(Pg/III) 16.44 ± 0.19	7.54 x 10 ⁻⁴⁷
Control group	7.13 ± 0.26	

IL9 gene polymorphisms rs 17317275 in studied groups

Regarding genotyping and allele frequencies, table (5A) showed that SLE group had a higher prevalence of the GG, AG/G allele (Genotyping /allele) than the control group. The AA/A allele (Genotyping /allele) was less common in SLE patients compared to controls.

Table (5A): The	genotyping and	l alleles frequ	lencies of
IL9 gene polymor	phisms rs 1731	7275 in studi	ed groups

IL9 gene poly	IL9 gene polymorphisms rs 17317275 in studied groups					
Genotyping	Patients	group	Control	group		
of	frequency	/(%)	frequency	(%)		
rs17317275	Observed	Expected	Observed	Expected		
GG	22	32.10	5 (10.0)	8 (16.0)		
	(21.36)	(31.16)				
AG	71	50.80	30	24		
	(68.93)	(49.32)	(60.0)	(48.0)		
AA	10	20.10	15	18		
	(9.71)	(19.51)	(30.0)	(36.0)		
	103	103	50	50		
Total	(100.0)	(100.0)	(100.0)	(100.0)		
<i>P</i> -HWE	0.0001		0.0771 N	S		

Furthermore, table (5B) showed that the GG, AG/ G (Genotyping /allele) had higher odds ratio (OR) values (2.44 and 1.48, respectively), indicating that it could be considered a risk factor, whereas the AA/ A (Genotyping /allele) had a lower OR value (0.25), indicating that this genotyping and allele had a protective property.

8	·····, p·····, o ····				
Genotyping of	Patients group	Control group	OR (95% CI)	EF or	Fisher's exact
rs17317275	frequency (%)	frequency (%)		PF %	probability
GG	22 (21.36)	5 (10.0)	2.44 (0.87 - 6.83)	12.6	0.113 NS
AG	71 (68.93)	30 (60.0)	1.48 (0.74 – 2.97)	22.3	0.281 NS
AA	10 (9.71)	15 (30.0)	0.25 (0.10 - 0.61)	22.5	2.0 x 10 ⁻³
Total	103 (100.0)	50 (100.0)			
Alleles frequenci	es				
G	115 (56)	40 (40)	1.90 (1.17 – 3.07)	26.4	0.01
Α	91 (44)	60 (60)	0.53 (0.33 – 0.86)	28.3	

Table (5B): The genotyping, odd ratio and alleles frequencies of IL9 gene polymorphisms rs 17317275 in studied groups

IL-9 serum level and genotyping

Table (6) showed a significant increase in IL-9 level GG, AG of patients' group compared to control groups ($16.28 \pm 0.16^{\text{A}}$ vs. $6.34 \pm 0.51^{\text{A}}$ and $16.68 \pm 0.23^{\text{A}}$ vs. $6.62 \pm 0.30^{\text{A}}$, respectively). These results were reported for the first time.

Genotyping	IL-9 level mean \pm SE (pg/ml)		Probability		
	Patients	Control			
GG	$16.28\pm0.16^{\rm A}$	6.34 ± 0.51^{A}	6.54 x 10 ⁻¹⁵		
GA	16.68 ± 0.23^{A}	6.62 ± 0.30^{A}	6.09 x 10 ⁻²⁸		
AA	$15.65 \pm 0.89^{\text{A}}$	$7.87\pm0.43^{\rm A}$	2.09 x 10 ⁻⁸		
The similar letters referred to non-significant differences					

Table (6): The genotyping and alleles frequencies in studied groups

CRP and genotyping

Table (7) showed an increasing in CRP in AG and AA genotyping of patients compared to control group. While GG showed non-significant differences in patients compared to control group. In addition, non-significant differences between genotype in respect to CRP level (similar letters referred to non-significant differences).

Table (7): CRP distribution in SLE according to the genotyping of the studied groups

Genotyping	CRP level mean \pm SE (mg/dl)		Probability
	Patients	Control	
GG	$10.76 \pm 2.28^{\rm A}$	$3.17\pm0.35^{\rm A}$	0.116 NS
AG	$11.25 \pm 1.57^{\text{A}}$	$2.51\pm0.25^{\rm A}$	0.0004
AA	15.81 ± 5.83^{A}	$2.81\pm0.39^{\rm A}$	0.011
The similar letters referred	to non-significant differen	ces	

Autoantibodies (ANA and ant-dsDNA) and genotyping

In this study, these genotyping results were published for the first time showed ANA increasing in the frequency percentage of AG 63.4% and GG 22.0% as shown in table (8A)

Table (8A): ANA frequency distribution in SLE according to the genotyping of the studie	d groups
---	----------

Genotyping	ANA frequence	Probability			
	Patients		Control		
	Positive	Negative	Positive	Negative	
GG	9 (22.0)	13 (21.0)	0 (0.0)	5 (10.0)	0.08 NS
AG	26 (63.4)	45 (72.6)	0 (0.0)	30 (60.0)	1.2 x 10 ⁻⁴
AA	6 (14.6)	4 (6.5)	0 (0.0)	15 (30.0)	0.001
Probability	0 365 NS		-		

The result of ant-ds DNA showed increasing in the frequency percentage of AG genotyping (78.1%). While GG showed non- significant improvement in level of ant-dsDNA in patients compared to control (P>0.05) as shown in table (8B).

Table (8B): Ant-dsDNA fr	equency distribution in SLI	E according to the genoty	ping of the studied groups
--------------------------	-----------------------------	---------------------------	----------------------------

Genotyping	dsDNA frequence	Probability			
	Patients		Control		
	Positive	Negative	Positive	Negative	
GG	6 (18.8)	16 (22.5)	0 (0.0)	5 (10.0)	0.067 NS
AG	25 (78.1)	46 (64.8)	0 (0.0)	30 (60.0)	1.8 x 10 ⁻⁴
AA	1 (3.1)	9 (12.7)	0 (0.0)	15 (30.0)	0.211 NS
Probability	0.248 NS		-		

DISCUSSION

The present findings aligned with those of a prior inquiry. There were no statistically significant variations in age between SLE patients and controls (p>0.05) ⁽¹⁶⁾. Recent research has confirmed previous findings that SLE may arise at any age ⁽¹⁷⁾. In around 10-20% of cases, it begins before the age of 16. Although it is more prevalent in women of reproductive age, it can begin at any age. However, it is more common in the first five years of life, and especially in the next ten years ⁽¹⁸⁾.

Several studies have demonstrated that serum CRP levels increase as the duration of SLE disease increases, and this study revealed that CRP levels in SLE patients are greater than in the control group. C-reactive protein is an important and sensitive diagnostic for predicting lupus diseases. CRP is a nonspecific measure of systemic inflammation that is higher in patients with active SLE. These findings support the first use of these assays to examine individuals with suspected inflammation ⁽¹⁹⁾.

Antibodies to numerous nuclear antigens are the immunological characteristic of SLE and are seen in more than 95% of patients. As a result of its great sensitivity and ability to predict the clinical start of SLE, ANA testing is commonly employed for SLE early diagnosis ⁽²⁰⁾. Furthermore, the current investigation found that all patients were positive for these autoantibodies, indicating that autoantibodies have diagnostic value as early indicators of SLE development ⁽²¹⁾. It has been proposed that these antibody deposits in the kidney may induce lupus nephritis ⁽²²⁾. Anti-dsDNA IgG is a highly specific illness marker in SLE that may be used to explore the relationship between autoimmune disease-specific pathogenic autoantibodies and IgG isotype ⁽²³⁾.

Higher levels of IL-9 in the serum of SLE patients compared to healthy controls, particularly in active SLE patients, may stimulate the inflammatory process in SLE ⁽²⁴⁾. Furthermore, the IL-9 pathway might be seen as a possible method for treating several types of inflammatory illnesses ⁽²⁵⁾. Since IL-9 plays a role in allergic processes such as asthma by promoting the proliferation of mast cells and T cells, it has been linked to immunopathological disorders ⁽²⁶⁾. According to one of these recent studies, IL-9 serum levels raised dramatically in rheumatoid arthritis and type 2 diabetes patients ^(15, 27).

The genotyping and allele frequencies in the research groups revealed that T2DM group had a greater AA, AG/A (genotyping/allele) than healthy persons. T2DM was lower in the GG/ G (genotyping/ allele) group than in the control group. Furthermore, in T2DM group, the elevated OR value of AA, AG/A (genotyping/allele) was deemed as a risk factor, particularly in genetically predisposed people. While the greatest OR value of GG/ G (genotyping/allele) in healthy persons was predicted to be a protective factor, this was not the case. ⁽²⁷⁾. Other study indicate that IL-9

SNP rs1313970720 showed associations (positive and negative) with Bacterial vaginitis (BV) patients and may be having a role in the mechanism of etiopathogenic of BV in the samples of Iraqi patients ⁽²⁸⁾. The HLA-DR3 link with SLE SNP, rs2187668, is considerably a higher influence on the propensity to manufacture autoantibodies ⁽²⁹⁾.

CONCLUSION

In conclusion, this study indicated the major roles of serum level and genetic polymorphism of IL-9, CRP, ANA and anti-ds DNA in pathogenesis and severity of SLE.

REFERENCES

- 1. Oparina N, Martínez-Bueno M, Alarcón-Riquelme M (2019): 'An update on the genetics of systemic lupus erythematosus', Current Opinion in Rheumatology, 31 (6): 659–668.
- 2. Furlong G (2019): Assessment of cases of systemic Lupus erythematosus- A clinical study. International Journal of Dermatology, Venereology and Leprosy Sciences, 2 (1): 04–06.
- **3.** Moulton V, Suarez-fueyo A, Meidan E *et al.* (2018): Sex hormones in acquired immunity and autoimmune disease. Cellular Perspective, 23 (7): 615–35.
- 4. Rojas M, Rodríguez Y, Leon K *et al.* (2018): Cytokines and Inflammatory Mediators in Systemic Lupus Erythematosus. EMJ Rheumatol ., 5 (1): 83–92.
- 5. François C, Laurent A (2018): Targeting interferons and their pathways in systemic lupus erythematosus. Autoimmun. Rev., 17(1): 44–52.
- 6. Idborg H, Oke V (2021): Cytokines as biomarkers in systemic lupus erythematosus: Value for diagnosis and drug therapy. Int J Mol Sci., 22 (21): 1–15.
- 7. Ruchakorn N, Ngamjanyaporn P, Suangtamai T *et al.* (2019): Performance of cytokine models in predicting SLE activity. Arthritis Res Ther., 21 (1): 1–11
- 8. Dantas A, Marques C, Da Rocha Junior L *et al.* (2015): Increased serum interleukin-9 levels in rheumatoid arthritis and systemic lupus erythematosus: Pathogenic role or just an epiphenomenon? Dis Markers, 6:18758630
- 9. Rojas-Zuleta W, Sanchez E (2017):IL-9: Function, sources, and detection. Methods Mol Biol., 1585: 21–35.
- **10. Hassan I, Kadim A, Sultan A (2020):** Study of some immunological indicators of interleukin-9 in Rheumatoid arthritis of Iraqi Patients. J Phys Conf Ser., 1: 1660.
- **11.** Xiong T, Attar M, Gnirck A *et al.* (2020): Interleukin-9 protects from early podocyte injury and progressive glomerulosclerosis in Adriamycin-induced nephropathy. Kidney Int ., 98 (3): 615–29.
- **12.** Yao W, Tepper R, Kaplan M (2011): Predisposition to the development of IL-9–secreting T cells in atopic infants. J Allergy Clin Immunol., 128 (6): 1357–60.
- **13.** Ouyang H, Shi Y, Liu Z *et al.* (2013): Increased Interleukin-9 and CD4+ IL-9+ T cells in patients with systemic lupus erythematosus. Mol Med Rep., 7: 1031–7.
- 14. Kar S, Gupta R, Malhotra R *et al.* (2021): Interleukin-9 Facilitates Osteoclastogenesis in Rheumatoid Arthritis. Int J Mol Sci., 22 (19): 10397.
- 15. Ali S, Salloom D (2020): A novel finding of il-9 genetic

polymorphism (rs17317275) and serum level in rheumatoid arthritis patients infected with CMV. Plant Arch.., 20: 582–7.

- **16. Osman E, El Nazar S, Maharem D** *et al.* (2021): Relation between vitamin d level and cyclin-dependent kinase-1 gene expression in egyptian patients with lupus nephritis and their impact on disease activity. Indian J Nephrol., 31 (2): 163.
- **17. Gorial F, Mahmood Z, Al Obaidi S (2018):** Body Composition in Iraqi Women With Systemic Lupus Erythematosus. Glob J Health Sci., 11 (1): 63.
- Illescas-Montes R, Corona-Castro C, Melguizo-Rodríguez L et al. (2019): Infectious processes and systemic lupus erythematosus. Immunology, 158 (3): 153–60.
- **19. Sbong S, Feldman M (2015):** Frequency and causes of C-reactive protein and erythrocyte sedimentation rate disagreements in adults. Int J Rheum Dis., 18 (1): 29–32.
- 20. Leuchten N, Hoyer A, Brinks R et al. (2018): Performance of Antinuclear Antibodies for Classifying Systemic Lupus Erythematosus: A Systematic Literature Review and Meta-Regression of Diagnostic Data. Arthritis Care Res., 70 (3): 428–38
- 21. Abbas A, Melconian A, Ad'Hiah A (2019): Autoantibody Profile in Systemic Lupus Erythematosus Patients. <u>Autoantibody Profile in Systemic Lupus</u> Erythematosus Patients - IOPscience
- 22. Fu S, Dai C, Zhao Z et al. (2015): Anti-dsDNA Antibodies are one of the many autoantibodies in systemic lupus erythematosus. <u>Anti-dsDNA Antibodies</u> are one of the many autoantibodies in systemic lupus

erythematosus - PubMed (nih.gov)

- **23.** Böröcz K, Simon D, Erdő-Bonyár S *et al.* (2021): Relationship between natural and infection-induced antibodies in systemic autoimmune diseases (SAD): SLE, SSc and RA. Clin Exp Immunol., 203 (1): 32–40.
- 24. Su N, Lu Y, Kong S (2013): Increased Interleukin-9 and CD4+IL-9+ T cells in patients with Systemic lupus erythematosus. Ouyang, Han Shi, Yongbing Liu, Zhichun Feng, Sheng Li, LingyuMolecular Med Reports, 7 (3): 1031–7.
- 25. Chakraborty S, Kubatzky K, Mitra D (2019): An update on interleukin-9: From its cellular source and signal transduction to its role in immunopathogenesis. https://pubmed.ncbi.nlm.nih.gov/31035677
- **26.** Schmitt E, Bopp T (2017): Discovery and initial characterization of Th9 cells: the early years. In: Seminars in immunopathology. Springer, Pp: 5–10.
- 27. Mohammed H, Salloom D(2021): Evaluation of interleukin-9 serum level and gene polymorphism in a sample of Iraqi type 2 diabetic mellitus patients. Meta Gene, 27: 100845.
- 28. Kadim A, Sultan A, Hassan I (2019): A genetic polymorphism of interleukin 9 gene (rs2069882) in rheumatoid arthritis of iraqi patients. Biochem Cell Arch., 19 (2): 3613–6.
- **29.** Chung S, Taylor K, Graham R *et al.* (2011): Differential genetic associations for systemic lupus erythematosus based on anti-dsDNA autoantibody production. https://journals.plos.org/plosgenetics/article?id=10.1371 /journal.