Study of Serum Levels of Visfatin Amongst Pre-Diabetic Obese Patients

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ABSTRACT

Background: Insulin resistance, pre-diabetes, and obesity have all been linked to excess adiposity. Proinflammatory adipokine visfatin is thought to play a key role in type 2 diabetes inflammation. **Objective:** This study aimed to estimate visfatin level among prediabetic obese patients and to observe and detect the interplay between visfatin, insulin resistance and obesity.

Patients and methods: 96 adult subjects were studied in case-control research at Internal Medicine Department and Clinical Pathology Department, Zagazig University Hospital. The study was carried out from January 2021 to November 2021. Subjects were divided into 3 groups: Group (1) included 24 healthy individuals as a control group, group (2) included 48 prediabetic individuals and group (3) that included 24 patients type 2 diabetes who never been treated in any of these. Serum visfatin was assessed in all participants.

Results: Diabetes mellitus (DM) patients had higher visfatin levels than those with impaired glucose tolerance (IGT) or impaired fasting glucose (IFG). Visfatin had a significant positive correlation with BMI, WC, FBS, PPS, HA1c, LDL, fasting insulin, and HOMA IR, while HDL had a significant negative correlation.

Conclusion: Visfatin levels were significantly linked to type 2 diabetes. HOMA-IR, fasting insulin, and BMI all showed a strong positive correlation with visfatin levels, suggesting that it may be a useful biomarker for detecting type 2 diabetes.

Keywords: Visfatin, Prediabetes.

INTRODUCTION

One of the most major risk factors related to prediabetes, insulin resistance as well as type 2 diabetes is obesity. Adipocyte-derived hormones, now known as adipokines, and various proteins and enzymes are all produced by adipose tissue, in addition to serving as a significant endocrine and metabolically active organ⁽¹⁾.

Adipocytokine visfatin, pre-B cell colonyenhancing factor (PBEF), is identified as a protein that is strongly produced then secreted by fat of viscera. It was isolated by Fukuhara et al.⁽²⁾ in 2005 from patients who had blood levels of visfatin correlated with obesity.

Biosynthesis of Nicotinamide Adenine Dinucleotide regulated by visfatin role as nicotinamide is phosphoribosyl transferase (NAD). Stress-induced energy metabolism is controlled by visfatin, which possesses anti-apoptotic effects. Activation of the immune system is also an important purpose for this compound $^{(3,4)}$.

It was previously hypothesized that visfatin, which is produced mostly by macrophages, would act as an insulin mimic because of its involvement in promoting triglyceride (TG) synthesis and glucose transfer. The novel adipokine, visfatin, plays an important part in the inflammatory process associated with type 2 diabetes ⁽⁵⁾. However, the results on visfatin levels are based on research using a non-specific test based on the Cterminus of visfatin, which led to significant inconsistencies. Visfatin concentrations in obese and healthy individuals are not known to be consistent ⁽⁶⁾. It has been suggested that increased visceral fat in these

patients induces a state of inflammation, which may lead to insulin resistance. Fukuhara and colleagues (2) have previously showed that visfatin may have a glucose-lowering effect, but the study was later withdrawn by the Editor of "Science" because of the controversy surrounding the findings ⁽⁷⁾.

Insulin resistance and other metabolic illnesses such as obesity and type 2 diabetes have been related to persistent low-grade inflammation characterize by aberrant cytokines over the years, and that these inflammatory markers may be directly linked to an increased risk of developing diabetes. Type 2 diabetes and insulin resistance are both closely associated with obesity, and previous research suggests that adipocytokines and inflammatory markers may play a role in their development ^(8, 9).

The goal of this case-control analytic investigation was to estimate visfatin level among prediabetic obese patients and to observe and detect the interplay between visfatin, insulin resistance and obesity potentially enhancing our knowledge of this controversial marker.

PATIENTS AND METHODS

96 adult subjects were studied in case-control research at Internal Medicine and Clinical Pathology Departments, Zagazig University Hospital. The study was carried out from January to November 2021.

Ethical approval:

All participants signed informed consent forms that were submitted to Zagazig University's



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Research Ethics Committee where the study was allowed (ZU-IRB#6649). We followed the World Medical Association's ethical code for human experimentation (Helsinki Declaration).

Inclusion criteria: Age from 30 to 50 years old irrespective of sex and obese patients (BMI >30) following the criteria for determining pre-diabetic symptoms [FPG (fasting plasma glucose) between 100 and 125 mg/dl (5.6 and 6.9 mmol/l), (IFG=impaired fasting glucose), PPG for two hours after a 75-g OGTT (oral glucose tolerance test) range: 140 to 199 mg/dl (7.8-11.0 mmol/l) (IGT= impaired glucose tolerance) and HA1c of 5.7–6.4% (39–47 mmol/mol)] and newly diagnosed diabetic patients without treatment.

Exclusion criteria: Unwilling patients, patients with type 1 DM, pregnant patients, renal disease, macrovascular diseases, liver diseases, drugs influencing insulin sensitivity (e.g., metformin, thiazolidinediones, etc.), and BMI < 30.

Subjects were classified into three groups:

Group 1: A healthy control group of twenty-four people (based on fasting blood glucose, oral glucose tolerance test and HbA1c) There were 10 men and 14 women in total. The mean age of the group was 42.41 \pm 4.9 years.

Group 2: Included 48 individuals had pre-diabetes (as determined by HbA1c and other markers of haemoglobin A1c and fasting glucose levels), 22 of them were males and 26 were females, who were subdivided to: **Group A:** 24 IGF subjects, 10 were males, while 14 were females. The average age of the group was 39.70 years old, and **group B:** 24 IGT subjects, males were 12 and ladies were 12, their average age was 41.29 ± 6.6 years.

Group 3: Involved 24 patients with type 2 diabetes mellitus never been treated in any of these. Males and ladies were 12 and 12 respectively. The average age of the group was 41.37 ± 5.6 years.

- 1- Full medical history: A detailed history was taken including personal history (age, sex and special history as smoking). History of medical diseases as hypertension and hyperlipidemia. Drug history, and past history of renal or hepatic diseases.
- 2- Full clinical examination was performed to rule out organic diseases for exclusion from the study. Blood pressure measurement. Anthropometric measurements (weighed in kilograms after voiding the bladder in light clothing without footwear). From the top of their heads to their soles, their height was measured in meters. Weighing in kilos divided by the square of the height was used to calculate the BMI.
- 3- Laboratory tests: CBC. 75-gram OGTT with

fasting plasma glucose and 2-hour postprandial glucose. HbA1c. Triglycerides, HDL, and LDL cholesterol levels in the blood. CRP. HOMA-IR, and enzyme-linked immunosorbent test was used to assess serum visfatin levels.

Statistical analysis

In order to analyze the data acquired, it was loaded into a computer and run via the Statistical Package of Social Sciences, version 20. (SPSS). Tables and graphs were used to present the findings. The Shapiro-Wilk test was used to examine the distribution properties of variables as well as the homogeneity of variance. The quantitative data was reported in the form of the mean, median, standard deviation, and confidence interval. The frequency and proportions of qualitative data were used to present the information. For quantitative independent data, the student's t test (T) and the Mann-Whitney test (MW) were employed to examine the data as needed. To examine qualitatively independent data, researchers employed the Pearson Chi-Square Test and the Chi-Square for Linear Trend ($\chi 2$). P value equals or less than 0.05 was considered significant.

RESULTS

Age was distributed among groups as 42.41 ± 4.9 , 39.70 \pm 6.3, 41.29 \pm 6.6 and 41.37 \pm 5.6 with nonsignificant difference among groups. Also, there was non-significant difference among groups regarding height, weight, WC and BMI. Additionally, there was non-significant difference regarding sex distribution, smoking or HTN (**Table 1**).

In comparison to the IGT and IFG groups, the DM group had significantly higher FBS and PPS, as well as a significantly higher HA1c. While the control group had significantly lower cholesterol and LDL, as well as significantly higher HDL. Regarding TG, the IGT and IFG groups had significantly higher levels, while the control group and the DM group had significantly lower levels. HOMA IR was substantially greater in the DM group than in the IGT or IFG groups or in the control group compared to the fasting insulin levels (**Table 2**).

Visfatin levels were considerably greater in the DM group than in the IGT group and the IFG group than in the control group (**Table 3**).

Non-significant area under curve with cutoff >1.21 with sensitivity 66.7% and specificity 55.0% regarding prediabetes group. (**Table 4, figure 1**).

Significant area under curve with cutoff >2.45 with sensitivity 88.2% and specificity 83.3% regarding type 2 DM group. (Table 5, figure 2).

There was significant positive correlation as regards visfatin with BMI, WC, FBS, PPS, HA1c and LDL. There was a strong negative association between fasting insulin and HOMA IR and HDL cholesterol (**Table 6**).

			Control group (N =24)	IFG group (N=24)	IGT group (N=24)	Diabetic group (N=24)	F	Р
Age (Years)			42.41±4.9	39.70±6.3	41.29±6.6	41.37±5.6	0.844	0.473
Height (Meters			1.58 ± 0.11	1.60 ± 0.08	1.61 ± 0.12	1.64 ± 0.07	1.592	0.197
Weight (Kilogr	rams)		95.78±6.45	97.04±10.8	94.16±10.44	96.82±7.9	0.502	0.682
Waist circumfe (CM)	erence		98.66±7.69	99.62±9.5	102.0±7.9	103.20±7.9	1.525	0.213
BMI (kg / m ²)			38.50±4.23	37.82±4.79	36.40±4.55	35.91±3.91	1.814	0.150
	Female	Ν	14	14	12	12		
Cender		%	58.3%	58.3%	50.0%	50.0%		
Gender	Male	Ν	10	10	12	12	0.67	0.88
		%	41.7%	41.7%	50.0%	50.0%		
Smoking	NO	Ν	20	19	20	18		
	no	%	83.3%	79.2%	83.3%	75.0%		
Shloking	YES	Ν	4	5	4	6	0.72	0.86
		%	16.7%	20.8%	16.7%	25.0%		
	NO	Ν	21	18	20	22		
Hypertension	NU	%	87.5%	75.0%	83.3%	91.7%		
	Voc	Ν	3	6	4	2	2.76	0.42
	1 65	%	12.5%	25.0%	16.7%	8.3%		
N%		Ν	24	24	24	24		
		100.0%	100.0%	100.0%	100.0%			

Table (1): The study group's age and sex composition among studied patients

Table (2): LAB data distribution among studied groups

	Control group	IFG group	IGT group	Diabetic group	F	Р
FBS (mg/dl)	97.91±6.80	111.0±7.55	114.25 ± 6.01	141.33±12.63*	118.8	0.00**
PPS (mg/dl)	116.87 ± 7.82	134.33±3.26	171.5±16.02	206.41±18.26	230.7	0.00**
HBA1c (%)	5.46 ± 0.43	6.01±0.22#	6.05±0.22#	7.70±0.68*	118.8	0.00**
Cholesterol(mg/dl)	177.87 ± 20.2	202.75±13.6#	206.45±13.3#	199.20±18.9#	18.0	0.00**
LDL(mg/dl)	$103.50{\pm}10.16$	115.54±7.77#	118.16±9.33#	120.75±18.24#	9.594	0.00**
HDL(mg/dl)	56.58±2.94*	34.95±4.12	31.75±2.92	48.41±4.69	261.9	0.00**
TG(mg/dl)	95.54±5.42@	254.0±19.3#	263.91±18.2#	124.83 ± 23.76	555.7	0.00**
CRP	$0.68 \pm 0.18@$	2.27±0.75#	2.21±0.69#	1.46 ± 0.47	27.7	0.00**
Fasting insulin (mIU/ML)	21.25±3.27	28.79±3.71#	27.58±4.3#	37.62±6.30*	52.714	0.00**
HOMA_IR	1.24 ± 0.25	1.82 ± 0.37	2.52±0.41	3.15±0.53	100.82	0.00**

Table (3): Visfatin among studied groups

	Control group	IFG group	IGT group	Diabetic group	F	Р
Visfatin (ng/ml)	0.77±0.16	2.10±0.55#	2.13±0.59#	7.60±1.51*	57.16	0.00**

 Table (4): Visfatin cutoff regarding pre-diabetic

Amon Cutoff		р	95% Confide	ence Interval	Congitivity	Specificity	
Area	Cuton	r	Lower Bound	Upper Bound	Sensitivity	specificity	
0.631	>1.21	0.071	0.502	0.760	66.7%	55.0%	

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Figure (1): ROC Curve for visfatin cutoff regard pre-diabetic

Table (5): Visfatin cutoff regarding Type 2 DM

A moo	Cutoff	D voluo	Asymptotic 95% Confidence Interval		
Area	Cuton	r value	Lower Bound	Upper Bound	
.961	>2.45	0.001*	88.2%	83.3%	





		Visfatin
1 33	r	-0.039
Age	Р	0.709
Waist	r	0.201*
circumference	Р	0.049
DM	r	0.231*
DIVII	Р	0.029
CDD	r	-0.111
SDF	Р	0.284
DPD	r	-0.016
DBP	Р	0.876
FDC	r	0.712**
FDS	Р	0.000
DDC	r	0.651**
115	Р	0.000
HRA1C	r	0.719**
IIDAIC	Р	0.000
Cholesterol	r	0.040
	Р	0.699
I DI	r	0.272**
	Р	0.007
HDI	r	-0.203*
IDL	Р	0.047
ТС	r	-0.155
10	Р	0.091
CRP	r	0.007
	Р	0.948
Fasting	r	0.641**
insulin	Р	0.000
HOMA ID	r	0.693**
	Р	0.000

 Table (7): Correlation

DISCUSSION

It is reported in studies that visfatin plasma level is involved in pathogenesis of different metabolic disorders and multiple studies have been conducted around this issue. Increasing the plasma level of visfatin was reported in individuals with obesity, GDM (gestational diabetes mellitus) and insulin resistance and in IFG (Impaired fasting glucose) patients ⁽¹⁰⁾.

Age was distributed among groups as 42.41 ± 4.9 years, 39.70 ± 6.3 years, 41.29 ± 6.6 years and 41.37 ± 5.6 years with non-significant difference among groups. Also, there was non-significant difference among groups regarding height, weight, WC, BMI, sex distribution, smoking and HTN. Our findings are in conformity with the findings of a previous study by **Gunduz** *et al.* ⁽¹¹⁾ as evidenced by the fact that their research involved 63 participants, 40 of whom had type 2 diabetes and 23 of whom did not. There were 44 women and 19 men in all (70% female to 30% male).

In terms of age, gender, and BMI, there were no statistically significant differences between the T2DM patient and control groups (P>0.05).

Our study showed that in comparison to the IGT and IFG groups, the DM group had significantly higher FBS and PPS, and HA1c. While, the control group had significantly lower cholesterol and LDL, but significantly higher HDL. Regarding TG, the IGT and IFG groups had significantly higher levels, while the control group and the DM group had significantly lower levels. HOMA IR was substantially greater in the DM group than in the IGT or IFG groups or in the control group compared to the fasting insulin levels. Our results are supported by study of Hetta et al. ⁽⁶⁾ as they reported that there was a significant difference in the mean of cholesterol, HDL and TG (Triglycerides) between T2D patients and healthy controls. In the study of Sheta et al. (12), obese subjects had higher serum triglyceride and lower HDL-C measurements than those of control subjects.

For the DM group, fasting insulin levels were substantially higher than for the IGT group, the IFG group and the control group, according to the results of our research. It was found that the HOMA IR of the DM group was significantly greater than the IGT group, IFG group, and finally the control group. Patients with type 2 diabetes were found to have greater FBG (165.5 vs.87, P 0.001) and HOMA-IR (2.84) measurements, as reported by **Nezhadali and colleagues**⁽¹³⁾.

In the study in our hands, visfatin was significantly higher among DM group (7.60±2.51 ng/ml P= 0.00) than IGT group (2.13±0.89 ng/ml, P= 0.00), IFG group (2.10±0.75 ng/ml P= 0.00) and control group $(0.77 \pm 0.26 \text{ ng/ml P} = 0.001)$. Our results are supported by study of Hetta et al. ⁽⁶⁾ as they showed a high significant difference in the mean serum level of visfatin. Study in Iraq showed higher visfatin and IR in type 2 diabetic patients but have failed to show any difference in visfatin level between diabetic and control group⁽¹⁴⁾ while another study displayed a lower concentration of visfatin in T2DM and metabolic syndrome patients compared to controls in Vienna of Austria⁽¹⁵⁾. Moreover, study detecting visfatin level in human saliva was introduced as a biomarker in type 2 diabetic and pre-diabetic population ⁽¹⁶⁾. In contrary to our results, study of Nezhadali et al. (13) concluded that visfatin level did not differ between diabetic and nondiabetic group and this rejects the hypothesis that visfatin could be a potential biomarker for diagnosis of type 2 diabetes in their population. Also, in contrast to our results, Telejko et al. (17) suggested that plasma visfatin level did not differ in woman with gestational diabetes and normal glucose tolerance.

Our results showed that using ROC curve for visfatin cutoff regarding pre-diabetic group, there was non-significant area under curve with cutoff >1.21

with sensitivity 66.7% and specificity 55.0%. As regards ROC curve for visfatin cutoff regard DM, there was significant area under curve with cutoff >2.45 with sensitivity 88.2% and specificity 83.3%. Visfatin showed significant positive correlation with BMI, WC, FBS, PPS, HA1C, LDL, fasting insulin and HOMA IR while was significantly negatively correlated with HDL. Our results are supported by study of Naithani et al. ⁽³⁾ as they reported that significant correlation was seen between visfatin and fasting plasma glucose (p<0.01), cholesterol (p<0.01), triglyceride (p<0.01), insulin (p<0.01) and HOMAIR (p<0.01). Positive correlations between visfatin and HDL cholesterol can support the hypothesis that visfatin has a protective effect and association with lipoprotein metabolism that was also found in Indonesia, Asian Indians, Caucasian subject and Taiwan^(18, 19).

CONCLUSION

There is a significant association between visfatin level and type 2 DM. Visfatin level was significantly positively correlated with BMI, fasting insulin and HOMA-IR suggesting that it can play a role in pathogenesis of type 2 DM, and also could be a potential biomarker for diagnosis of type 2 DM.

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References:

- 1. Galic S, Oakhill J, Steinberg G (2010): Adipose tissue as an endocrine organ. Mol Cell Endocrinol., 316 (2): 129-139.
- 2. Fukuhara A, Matsuda M, Nishizawa M *et al.* (2005): Visfatin: a protein secreted by visceral fat that mimics the effects of insulin. Science, 307: 426-33.
- **3.** Naithani M, Saha S, Mathew A *et al.* (2020): A Cross-Sectional Study to Assess and Correlate Serum Levels of Visfatin with Insulin Resistance amongst Newly Diagnosed Diabetes Mellitus, Metabolic Syndrome and Pre-Diabetes Patients. Recent Adv Biol Med., 6 (4): 17989-93.
- **4.** Dakroub A, Nasser A, Younis S *et al.* (2020): Visfatin: A Possible Role in Cardiovasculo-Metabolic Disorders. Cells, 9 (11): 2444-48.
- **5.** Liu S, Qiao S, Yuan J *et al.* (2009): Association of plasma visfatin levels with inflammation, atherosclerosis and acute coronary syndromes (ACS) in humans. Clinical Endocrinology, 71: 202-206.
- 6. Hetta H, Ez-Eldeen M, Mohamed G *et al.* (2018): Visfatin serum levels in obese type 2 diabetic patients: relation to proinflammatory cytokines and insulin resistance. Egypt J Immunol., 25 (2): 141-151.
- 7. Chang Y, Chang D, Lin K *et al.* (2011): Visfatin in overweight/obesity, type 2 diabetes mellitus, insulin

resistance, metabolic syndrome and cardiovascular diseases: a meta-analysis and systemic review. Diabetes/Metab Res Rev., 27 (6): 515–27.

- **8.** Dev N, Marcus S (2012): High sensitive C-reactive protein, an independent and early novel inflammatory marker in healthy obese women. Biomedical Research, 12: 1-5.
- **9.** Yu L, Li Y, Du C *et al.* (2019): Pattern Recognition Receptor-Mediated Chronic Inflammation in the Development and Progression of Obesity-Related Metabolic Diseases. Mediators of Inflammation, 19: 126-131.
- **10.** Kabir F, Jahan F, Khan I *et al.* (2015): Increased concentration of circulating visfatin associates with post challenged hyperglycaemia and insulin resistance in IGT subjects. J Taibah Univ Med Sci., 10 (4): 481-487.
- **11.** Gunduz F, Yildirmak S, Temizel M *et al.* (2011): Serum visfatin and fetuin-a levels and glycemic control in patients with obese type 2 diabetes mellitus. Diabetes & Metabolism Journal, 35 (5): 523-528.
- **12.** Sheta Y, Elgohary E, Sharaf S (2012): Visfatin Between Fact and Fiction; A Marker af Obesity or a New Player in the Pathogenesis of Type 2 Diabetes Mellitus. Journal of Asian Scientific Research, 2 (12): 949-53.
- **13.** Nezhadali M, Mahdavi M, Saatian M *et al.* (2021): Association of Visfatin with Blood Glucose, Insulin Resistance and Body Mass Index in Patients with Type 2 Diabetes/Pre-Diabetes. Journal of Fasa University of Medical Sciences, 11 (3): 3984-3992.
- **14.** El Samahi M, Ismail N, Matter R *et al.* (2017): Study of Visfatin Level in Type 1 Diabetic Children and Adolescents. Open Access Maced J Med Sci., 5 (3): 299-304.
- **15.** Schindler K, Vila G, Hoppichler F *et al.* (2012): The Impact of Type 2 Diabetes on Circulating Adipokines in Patients with Metabolic Syndrome. Obes Facts, 5: 270–276.
- **16.** Srinivasan M, Meadows M, Maxwell L (2018): Assessment of Salivary Adipokines Resistin, Visfatin, and Ghrelin as Type 2 Diabetes Mellitus Biomarkers. Biochem Res Int., 18: 746-52.
- **17.** Telejko B, Kuzmicki M, Zonenberg A *et al.* (2009): Visfatin in gestational diabetes: Serum level and mRNA expression in fat and placental tissue. Diabetes Research and Clinical Practice, 84 (1): 68-75.
- **18.** Budiyati R, Lukito W, Wijaya A (2010): Correlation Between Visfatin, Insulin Resistance (Homeostasis Model Assessmentof Insulin Resistance), Inflammation (High Sensitivity C-Reactive Protein) and HDL Cholesterol Concentration in Individuals with Visceral Obesity. Med J Indones., 2 (1): 61-5.
- **19.** Wang P, van Greevenbroek M, Bouwman F *et al.* (2007). The circulating PBEF/NAMPT/visfatin level is associ-ated with a beneficial blood lipid profile. Eur J Appl Physio., 454: 971-75.