

Study of the single and combined genotoxic effects of chlorpyrifos and quercetin in *Saccharomyces cerevisiae*

Nada, H. A. Al-Twaty

Department of Biology, King Abdelaziz University

Abstract

The genotoxic effects of chlorpyrifos and quercetin either single or combined were tested in terms of their ability to induce reverse mutation, gene conversion and mitotic crossing over in *Saccharomyces cerevisiae* D7. The results indicated that all single and combined treatments induced reverse mutation, gene conversion and mitotic crossing over in *Saccharomyces cerevisiae* D7. Combined treatment was more effective than the single treatment of quercetin. The insecticide (chlorpyrifos and quercetin which are common flavonoids) proved to be mutagenic in *Saccharomyces cerevisiae*.

Introduction

In many genetic investigation the organophosphorus insecticides has been reported as a potent genotoxic agents (Abdallah *et al.* (1973); Villani *et al.* (1983); Nafei *et al.* (1984); Salam *et al.* (1984); and Mansour *et al.* (1988)). The induction of mitotic crossing over in diploid yeast *Saccharomyces cerevisiae* is strongly correlated with the mutagenic effects. These tests very sensitivity react with compounds which induce base-pair substitution as well as from-shift mutations. This system has revealed the genetic activity of large number of carcinogens, pesticides, radiation and many other chemical mutagens (Siebert and Elsenbrand, (1974); Zimmermann *et al.*, (1975); Altwaty, (1999); Anjaria and Rao, (2001) and Buschini *et al.*, (2003 and 2004)).

Quercetin is one of the most common flavonoids in plants, widely distributed in natural foods, consumed by humans in a range of 50 mg per day (Brown and Dietrich, 1979 and Caria *et al.*, 1995). Quercetin was shown to be mutagenic in the Ames assay (Bjeldames and Chang 1977; MacGregor and Jurd 1978; Brown and Dietrich 1979 and Rueff *et al.*, 1986). Also it has been shown to be carcinogenic in rats (Pamukcu *et al.*, 1980), to be

mutagenic in a variety of genotoxicity tests (Muller *et al.*, 1991 and Caria *et al.*, 1995). The conflicting inform-ations on the genotoxicity of chlorpyrifos and quercetin from previous studies reported in the literature led to us to study the genotoxic effects of both of them in single and combined forms.

Materials and methods

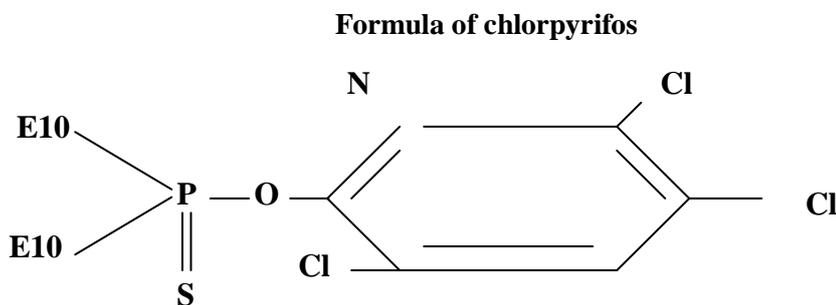
1- Yeast strain:

The D7 strain of *Saccharomyces cerevisiae* was used as a test organism (Courtesy of F. K. Zimmermann, Darmstad, Germany). This strains has the following genotype: ade2-40 / ade2-119, trp5-12 / trp5-27, ilv1-92 / ilv1-92. It is used for the simultaneous detection of induced reverse mutation, mitotic gene conversion, and mitotic crossing over (Zimmermann *et al.*, 1975).

2- Chemicals

a. The insecticide chlorpyrifos was obtained from Hanoo Agricultural, the sole agent in K.S.A.,P.O. Box.4894 Riyadh 114412. Manufactured by Chemac-Agriphar / Rue De Renory, 261B-41020 Ugree/Belgium.

Study of the single and combined genotoxic effects.....



Chlorpyrifos is an organophosphorus insecticide, its chemical is:

O.o-diethylo-3,5,6 trichloro-2 pyridyl phosphorothioate.

b. Quercetin: is one of the flavonoids in *Senna* spp (*Cassia*) (*Leguminosae*), obtained from Dr. Aisha Mohamed, Ali Khogli, Faculty of Science, King Abdelaziz University.

3- Medium

a. Complete medium

This medium was used for routine culture growth, it contains : peptone 5 mg/L, yeast extract 10 g/L, glucose 20 g/L and Agar 20 g/L

b. Minimal medium

The medium components have been described in detail by Zimmermann et al. (1975).

4- Testing assay:

- a- Three concentrations were prepared from chlorpyrifos, these concentrations were 1, 2, 5 μ l per ml media.
- b- The used concentration of quercetin was 5 μ l per ml media
- c- Combined treatment

The used concentration of chlorpyrifos and quercetin for combined treatment was 5 μ l/ml media

Treatment protocol:

1. 10 ml of liquid complete medium were inoculated with about 5×10^6 cells/ml in a 50 ml conical flask.
2. The culture was incubated on an orbital shaker water bath at 24 c for 6 hrs.
3. The sample of the cells was examined under the microscope, the proper culture must be in experimental phase (at least 90 % of the cells have buds).
4. Concentration series for treatment were inoculated with 1 ml sample cells and incubated at 28 c on a water bath shaker for 18 hrs.
5. After appropriate dilution, the cells were plated onto:
 - Complete medium with cycloheximide to detect mitotic crossing over
 - Synthetic complete medium without tryptophan to detect gene conversion
 - Synthetic complete medium without isoleucine to detect point mutation

Analysis and evaluation of the data

The frequencies of gene conversion, reverse mutation and mitotic crossing over were computed by dividing the number of revertant, revertant and mitotic crossing over colonies. The general consensus was

that the increase in an end point under investigation up to two folds or more of the mean of control frequency is biologically considered as a significant response (Brusick, 1980).

Results and discussion

The results in table (1) show the genetic activities in such chlorpyrifos in *Saccharomyces cerevisiae* D7. Chlorpyrifos exhibited moderate toxicity at the lower concentration which proportionally increased by increasing the treatment dose (1-5 $\mu\text{l/ml}$). Survival percentages ranged from 70 % at the lowest concentration (1 $\mu\text{l/ml}$) to 27 % at highest one (5 $\mu\text{l/ml}$). Weak positive mutagenic activity was obtained using the concentration 1 $\mu\text{l/ml}$ where the induced frequency of mitotic crossing over at the cycloheximide (Cyh) locus was 4.7 times the spontaneous frequency, while the same concentration showed negative results in the induction of gene conversion at the tryptophan-5 (Trp-5) locus and reversion at isoleucine (il) locus. Also, moderate mutagenic activity was obtained at the three loci under study when chlorpyrifos applied at the concentration 2 $\mu\text{l/ml}$ which resulted in mitotic gene conversion, reversion and mitotic crossing over in frequency 3.6, 4.1 and 9.6 times the spontaneous ones respectively. Chlorpyrifos as a mutagen proved to be more potent at the concentration 5 $\mu\text{l/ml}$ which caused 27 % survival and resulted in mitotic gene conversion, reversion and mitotic crossing over in frequencies 13.1, 13.2 and 20 times of control ones respectively. These results suggest the mutagenic effect of chlorpyrifos in the induction of conversion of revertant, revertant and mitotic crossing over in *Saccharomyces cerevisiae* strain D7. This is in agreement with the results obtained by many reports used pesticides in *Saccharo-*

myces cerevisiae, El-Adawy *et al.* (1998); Salam *et al.* (1993 and 1995); Ahmed *et al.* (1999) and Al-twaty (1999). The results shown in table (2), showed the genetic activities of quercetin. Its positive indications of mutagenic activity were obtained at the concentration of 5 $\mu\text{l/ml}$ of quercetin, where the induced frequency of mitotic gene conversion and mitotic crossing over was 5.1 and 4.7 times the control ones respectively. Also, the same concentration showed a strong activity at the (ilv) locus, causing revertants in a frequency 11.9 times the control level. Meanwhile, the combined treatment of quercetin and chlorpyrifos, showed a cumulative mutagenic effects when compared with quercetin alone which resulted in mitotic gene conversion, revertant and crossing over in frequencies at 11.7, 11.2 and 5.4 times as the control levels, respectively. This result suggests that quercetin and the combined treatment with chlorpyrifos showed mutagenic effect in induction of revertant, revertant and mitotic crossing over in *Saccharomyces cerevisiae* D7. This is in agreement with Caria *et al.* (1995), who reported that quercetin induced micronuclei in human lymphocytes. Moreover, quercetin was shown to be mutagenic in the Ames assay (Brown and Dietrich, 1979 and Rueff *et al.*, 1986). Induced safe our sheep (SoS) functions (Rueff *et al.*, 1992). The results of the present study show that chlorpyrifos (organophosphorus insecticides) and quercetin (one of the common flavonoids in plants) were capable to induce the three genetic end points and to reveal genetic activity at the three loci under study. Chlorpyrifos showed obviously high genotoxicity to strain D7 of *Saccharomyces cerevisiae* when compared with quercetin. Moreover, the treatment of chlorpyrifos and quercetin was slightly lower as compared with chlorpyrifos alone.

Study of the single and combined genotoxic effects.....

Table (1): Response of *Saccharomyces cerevisiae* D7 to the treatment with different concentrations of chlorpyrifos

Con. M/ml	Number of cells	Convertant			Revertant			Mut Freq	T/C	D. of Act.
		Mut Freq	T/C	D. of Act.	Mut Freq	T/C	D. of Act.			
Control	17084	14.1 (24)	1	-	11.7 (20)	1	-	16.4 (28)	1	-
1 MI	10930	25.6 (28)	1.8	-	21.9 (24)	1.8	-	76.8 (84)	4.7	+
2 MI	7570	52.6 (40)	3.6	+	47.5 (36)	4.1	+	158.5 (120)	9.6	+
5 MI	4642	189.6 (88)	13.1	++	155.2 (72)	13.2	++	336.2 (156)	20	++

Key: Con = Concentration Mut = Mutation
 C = Control value T = Treatment value
 + = 2 – 10 control level ++ = > 10 control level - = non significant
 D. of Act = Degree of activity, numbers between parenthese represents actual concoly counts

Table (2): Response of *Saccharomyces cerevisiae* D7 to the treatment with quercetin alone on combined with chlorpyrifos

Con. M/ml	Number of cells	Convertant			Revertant			Mut Freq	T/C	D. of Act.
		Mut Freq	T/C	D. of Act.	Mut Freq	T/C	D. of Act.			
Control	17084	14.1 (24)	1	-	11.7 (20)	1	-	16.4 (28)	1	-
Ch. 5 MI/ml	4642	189 (88)	13.1	++	155.2 (72)	31.2	++	336.2 (156)	20	++
Quer 5 MI/ml	9482	72.8 (69)	5.1	+	139.2 (132)	11.9	++	77 (73)	4.7	+
Com. Tr.	9197	166.3 (153)	11.7	++	155.2 (72)	11.2	++	89.2 (82)	5.4	++

Key: Con = Concentration Mut = Mutation
 C = Control value T = Treatment value
 + = 2 – 10 control level ++ = > 10 control level - = non significant
 Ch. = chlorpyrifos quer. = quercetin com Tr. = combined treatment
 D. of Act = Degree of activity, numbers between parenthese represents actual colony counts

References

1. **Abdallah, M. D.; Zaazou, M. H.; Ali, A. M. and Rizkallah, M. R. (1973):** Cholinesterase and aliesterase activity of different stages in the life cycle of organophosphorus resistant and susceptible *Spodoptera littoralis* (Boised). Bull. Ent. Soc. Egypt. Econ. Ser. 7: 222-228.
2. **Ahed, E.S.; Asal, N. M. and Baeshen, A. (1999):** Study of the single and combined genotoxic effects of furadan and lead in *Saccharomyces cerevisiae*. Alex. J. Agric. Res. 44 (1): 153-170.
3. **Altwyty, N.H. (1999):** Genetic toxicity of insecticide chlorcyrin in *Saccharomyces cerevisiae*. Delta. J. Sci. 23 (1): 523-263.
4. **Anjaria, K. B. and Rao, B. S., (2001):** Effect of caffeine on the gentoxic effects of gamma radiation on 4- NQO in diploid yeast. J Environ pathol Toxicol Oncol., 20 (1): 39-45.
5. **Bjeldanes, L. F. and Chang, G. W. (1977):** Mutagenic activity of quercetin and related compounds, Science, 197, 577-263.
6. **Brown, J. P. and Dietrich, P. S. (1977):** Mutagenic of plant flavonols in the

- salmonella/mammalian microsome test. Activation of flavonol glycosides by mixed glycosidases from rat faecal bacteria and other sources. *Mutation Res.*, 66, 223-240.
7. **Brusick, D. (1980):** Principles of Genetic Toxicology. Plenum press, New York, p. 2790.
 8. **Buuschini, A.; Poli, P. and Rossi, C. (2003):** *Saccharomyces cerevisiae* as an eukaryotic cell model to assess cytotoxicity and genotoxicity of three anticancer anthraquinone. *Mutagenesis*. 18 (1): 25-36.
 9. **Buschini, A.; Carboni, P.; Furlini, M.; Poki, P. and Tossi, C. (2004):** Sodium hypochlorite, chlorine dioxide and peracetic acid induced genotoxicity detected by the Comet assay and *Saccharomyces cerevisiae* D7 tests. *Mutagenesis*. 2004 19 (2): 157-162.
 10. **Caria, H.; Chaveca, T.; Laires, A. and Rueff, J. (1995):** Genotoxicity of quercetin in the micro nucleus assay in mouse bone marrow erythrocytes, human lymphocytes, V 79 cell line and identification of kinetochore- containing (CREST staining) micronuclei in human lymphocytes. *Mut. Res.* 343:85-94.
 11. **El- Adawy, R. A.; Abd El-Naby, W. N.; Hassanein, S. H.; Shawky, A. S. H. and salam, A. Z. El-Abidin (1988):** Mutagenic potentiality of triazophos, sumithion, fenpropathrin and amitraz in yeast, *Saccharomyces cerevisiae* XLX Annual Conf. Soc. Genet., Egypt, pp. 125-123.
 12. **Mansour, S. A.; Hassan, A. H. M.; Awad, A. A. M. and Salam, A. Z. El-Abidin (1988):** Allozyme polymorphism in *Drosophila*: Induction of polymorphism in a homozygous enzyme strain under the effect of the three different organophosphorus compounds. Proc. 2nd Conf. Agric. Develop. Res., Ain Shams University, Cairo, 1: 237-250.
 13. **Mac Gregor, J. T. and Jurd. L. (1978):** Mutagenicity of plant flavonoids: structural requirements for mutagenic activity in *Salmonella typhimurium*, *Mutation Res.*, 54,297-309.
 14. **Muller, L. P. Kasper and Madle, S. (1991):** The quality of genotoxicity testing of drugs. Experiences of a regulatory agency with new and old compounds, *Mutagenesis* 6, 143-149.
 15. **Nafei, H. A.; Hassan, A. M.; Mansour, S. A. and Salam A. Z. El-Abidin (1984):** The mutagenicity of organophosphorus insecticides in *D. Melanogaster*. Proc. 2nd Mediterranean Conf. Genet. Cairo, 717-725.
 16. **Pamukcu, A. M.; Yalciner. S.; Hatcher, J. F.; Bryan, G. T. (1980):** Quercetin, a rat intestinal and bladder carcinogen present in bracken fern (*Pteridium aquilinum*), *Cancer Res.*, 40, 3468-3472.
 17. **Rueff, J.; Laires, A.; Borba, H.; Chaveca, T. and Halpern, M. (1986):** Genetic Toxicology of flavonoids: the role of metabolic conditions in the induction of reverse mutation, SOS functions and sister-chromatid exchanges. *Mutagenesis*, 1, 179-183.
 18. **Rueff, J.; Laires, A.; Gasper, J.; Borba, H. and Rodrigues, A. (1992):** Oxygen species and the genotoxicity of quercetin, *Mutation Res.*, 89,69-74.
 19. **Salam A. Z. El-Abidin and Pinsker, W. (1981):** Effect of selection for resistance to organophosphorus insecticides on two esterase loci in *D. melanogaster*. *Genetica*, 55: 11-14.
 20. **Salam A. Z. El-Abidin; Hassan, A. H. M.; Nafei, H. A. and Mansour, S. A. M. (1984):** Isozyme polymorphism in *Drosophila* 5. The effect of two organophosphorus compounds on the gene frequency and the repair system. Proc. 2nd Mediterranean Conf. Genet. 11:701-716.
 21. **Salam A. Z. El-Abidin; Ebtissam, H. A. Hussein; Hanaiya, A. El-Itriby; Wagida, A. Anwar and Mansour, S. A. (1993):** The mutagenicity of Gramoxone (paraquat) on different eukaryotic systems. *Mut. Res.*, 319: 89-101.
 22. **Salam A. Z. El-Abidin; De-Hondt, H. A.; Fahmy, M. T.; Soussa, S. F.; Elnagar, T. F. K. and El-Din Ahmed, E. S. (1995):** The mutagenicity of Nudrin and Meothrin on two different eukaryotic systems *Drosophila* and yeast. *Annals. Agric. Sci.*, Ain Shams Univ., Cairo. 40 (2): 737-751.
 23. **Siebert, D. and Elsenbrand, G. (1974):** Induction of mitotic gene conversion in *Saccharomyces cerevisiae* by N-Nitrosated pesticides. *Mut. Res.*, 22: 121-126.
 24. **Villani, F.; Whhite, G. B.; Curtis, C. F. and Miles, S. J. (1983):** Inheritance and activity of some esterases associated with organophosphate resistance in mosquitoes of the complex of *Culex pipiens* L. (Diptera: Culicidae). *Bull. Ent. Res.*, 73: 153-170.
 25. **Zimmermann, F. K.; Kern, R. and Resenberger, H. (1975):** A yeast strain for simultaneous detection of induced mitotic crossing over. Mitotic gene conversion and reverse mutation. *Mut. Res.* 28: 381-388.

دراسة مفردة ومشاركة للسمية الوراثية للكلوربيرفوس والكيورستين فى فطر خميرة الخباز السلالة D7

ندى حسن على التواتى

قسم علوم الأحياء – جامعة الملك عبدالعزيز

تمت دراسة التأثير السمى الوراثى للكلوربيرفوس والكيورستين فى معاملة مفردة ومشاركة لكل منهما لمعرفة مقدرتهما على إحداث كل من الطفرة المرتدة والتحول الجينى والعبور الوراثى الجسمى على سلالة D7 لفطر خميرة الخباز وأوضحت النتائج المتحصل عليها أن كل المعاملات المفردة والمشاركة استحدثت طفرة مرتدة وتحول جينى وعبور وراثى جسمى فى سلالة D7 لفطر خميرة الخباز. المعاملات المشتركة كان لها تأثير أكبر من المعاملات المفردة للكيورستين. وقد أوضحت النتائج المتحصل عليها أن كلاً من المبيد الحشرى كلوربيرفوس وكيورستين (وهو الفلافوتيدات المعروفة) والمستخدم فى مقارنة الحشرات الموجودة على النباتات لهما تأثير طفرى على فطر خميرة الخباز.