

## Assessment of the Physiological Changes Induced by Sodium Nitrite, Annatto or Mono Sodium Glutamate in Male Albino Rats

Eman G.E. Helal, Rasha A.A. El-Sayed, Mariam S. El-Gamal

Department of Zoology, Faculty of Science (Girls), Al-Azhar University, Egypt

\*Corresponding author Eman Helal, emanhelal@hotmail.com, mobile:00201001025364, orcid.org/0000-0003-0527-7028

### ABSTRACT

**Background:** food additives are added to most junk and fast foods, especially those for kids. Sodium nitrite is an inorganic salt with widespread applications in the food industry as a color fixative and preservative in meat and fish. Annatto extract is a natural food color obtained from the outer coatings of the seeds of the Annatto tree (*Bixa orellana L.*). Monosodium glutamate (MSG), the sodium salt of amino acid glutamate, is a food additive that popularly used all over the world as “flavor enhancer”. **Aim of the work:** this study was aimed to determine the hazardous effects of sodium nitrite, annatto and monosodium glutamate on some physiological parameters in male albino rats. **Materials and methods:** this study had been done on forty male albino rats with an average body weight 100-145 g. The animals were divided into four groups; **Group 1:** control (untreated group), **Group 2:** sodium nitrite treated group, **Group 3:** annatto treated group and **Group 4:** monosodium glutamate treated group. Blood samples were collected, sera were separated and used for estimation of some biochemical parameters (liver enzymes, kidney functions, glucose, protein profile and lipid profile) and hormonal levels [testosterone, T3 (triiodothyronine) and T4 (thyroxine)]. **Results:** the biochemical results showed an increase in the activities of liver enzymes [aspartate aminotransferase (ASAT) and alanine aminotransferase (ALAT)], and the levels of glucose, kidney functions (urea, and creatinine), lipid profile [total cholesterol, triglycerides, low density lipoprotein (LDL-C)] and thyroid hormones [thyroxin (T4) and triiodothyronine (T3)] in all treated groups when compared to the control group. A drop in protein profile (total protein, albumin, globulin and A/G ratio), testosterone hormone and HDL level were observed in the treated groups as compared to the control rats. **Conclusion:** it could be concluded that some food additives like sodium nitrite, annatto, and monosodium glutamate have extreme effects on liver and kidney functions, protein and lipid profiles and also on thyroid and testosterone hormones. So, it is recommended to minimize the use of these additives to protect young children and mature people from these destructive effects.

**Keywords:** sodium nitrite, annatto (*Bixa orellana*), monosodium glutamate (umami), liver and kidney functions.

### INTRODUCTION

Humans are continuously exposed to different kinds of chemicals such as food additives. Many of these additives have been increasingly recognized as potentially hazardous to human health. Sodium nitrite is a food additive that is used, because of its role in inhibiting the growth of *Clostridium botulinum* spores in the refrigerated meats <sup>(1)</sup>. Meanwhile, large amounts of sodium nitrite can be toxic to animals, including humans. The cytotoxicity and detrimental effects of nitrite can be attributed to its oxidative properties <sup>(2)</sup>. The reactive nitrogen species that are produced by exposure to nitrite have many toxic effects including hepatotoxicity, nephrotoxicity and dysregulation of inflammatory responses and tissue injury <sup>(3)</sup>.

Food colorants may often be considered simply cosmetic in nature, but its role is very significant. Both food quality and flavor are closely associated with color. Annatto is a natural colorant that imparts colors ranging from yellow to red due to

the concentration of color compounds in the solution <sup>(4)</sup>. This pigment is obtained from the seed coat of the tropical shrub *Bixa orellana L.* This tree is native to tropical South America, where it has been a traditional ingredient of some foods for centuries <sup>(5)</sup>. Bixin and nor-bixin are the main pigments of annatto seeds that are carotenoids of huge importance in the food, pharmacological and cosmetic industries. In food industries, these natural pigments are used in cheeses, sausages, meats and candies <sup>(6)</sup>. Lately, FDA (Food and Drug Administration) of United States of America, classified annatto as a color additive exempt from certification that is safe for human consumption. Furthermore, many reports revealed that annatto is not carcinogenic neither maternally toxic nor embryotoxic <sup>(7)</sup>.

Monosodium L-glutamate (MSG) is a common glutamic acid salt that contains 78% glutamic acid, 22% sodium salt and water <sup>(8)</sup>. MSG is the commonest food additive that has been used as a

flavor enhancer in the home as well as in food industry since 1907<sup>(9)</sup>.

Therefore, most of the canned and fast food as flavored chips, canned soups, prepared meals, marinated meats, bottled soy or oriental sauces, freezing foods and tested tuna containing variable concentrations of MSG<sup>(10)</sup>. In animals, higher doses of MSG were confirmed to be neurotoxic as it destructs neurons in the hypothalamic nuclei through their changes in the hypothalamic-pituitary-adrenal axis (HPA)<sup>(11)</sup>. Many findings denote that unbound glutamate dissociated from MSG may possibly act on certain receptors in the central or peripheral neurons, causing many histopathological changes<sup>(12)</sup>. Moreover, the excessive MSG administration may lead to damage of liver and kidney<sup>(13)</sup>.

So, the aim of this work is to investigate the hazardous effects of sodium nitrite, annatto and monosodium glutamate on some physiological parameters in male albino rats.

## MATERIALS AND METHODS

fourty young male albino rats (weighing 100-145 g) were used in this study. Animals were housed in stainless steel cages, fed on rat chew and offered water *ad libidum*. The animals were divided into four equal groups (10 rats each) as follows: **The first group:** the control untreated group, **the second group:** orally administered with sodium nitrite, NaNO<sub>2</sub> (0.1 mg/kg b.wt./ day), **the third group:** orally administered with annatto (0.065 mg/kg b.wt./day) and **the fourth group:** orally administered with monosodium glutamate MSG (15 mg/kg b.wt./day). Body weights were recorded every week. After 30 days of treatment, animals were weighed and then decapitated.

Blood samples were collected for biochemical parameters. Blood samples were centrifuged for 10 min. at 5000 rpm and supernatant sera were separated for analysis without storage or delay.

### Biochemical Examination

In the present study total protein (TP) and albumin concentration were estimated, then serum globulin concentrations were calculated according to the formula:

**Globulin (g/dl) = total protein (g/dl) –albumin (g/dl)**

Aspartate aminotransferase (ASAT), alanine aminotransferase (ALAT) activities, Creatinine, urea, glucose concentrations as well as lipid profile that including total cholesterol, triglycerides and high-density lipoprotein cholesterol (HDL-C) were also determined.

Concentrations of testosterone and thyroid hormones (T3 and T4) were measured. All parameters were estimated using **BioMerieux SA kits, France**.

The both ratios of serum albumin/ globulin and albumin/creatinine were determined. However, ratios of TC/HDL (risk factor 1) and LDL/HDL (risk factor 2) were also calculated after calculation of serum LDL-C (low-density lipoprotein cholesterol) and VLDL (very low-density lipoprotein cholesterol) using the **Friedwald's**<sup>(14)</sup> and **Norbert**<sup>(15)</sup> formulas, respectively as following:

**Friedewald's**<sup>(14)</sup> equation: **LDL (mg/dl) = TC - {HDL + [TG/5]}.**

**Norbert**<sup>(15)</sup> equation: **VLDL = TG/5**

### Statistical analysis

The results were expressed as Mean ± SEM of the mean. Data were analyzed by one way analysis of variance (ANOVA) and were performed using the Statistical Package (SPSS) program, version 20. The Bonferroni test was used as a method to compare significance between groups.

## RESULTS

**Body weight:** animals that received sodium nitrite has a highly significant decrease in body weight ( $p < 0.001$ ), while those administrated with monosodium glutamate showed a highly significant increase in body weight ratio ( $P < 0.001$ ) and the annatto group showed insignificant changes as compared to control rats (Table 1).

**Glucose level:** there was a highly noticeable increase in glucose level in all the treated groups ( $p < 0.001$ ) in comparison with the control group (Table 1).

**Protein profile:** the present study showed that there was highly significant decrease in the total protein, albumin (except annatto) and globulin levels ( $p < 0.001$ ) in all treated groups as compared to control. Meanwhile, the treated groups recorded insignificant changes in albumin/globulin ratio as compared to the control group (Table 2).

**Liver functions:** ASAT and ALAT activities revealed a highly significant increase among the treated groups in contrast to the control group ( $p < 0.001$ ) (Table 3).

**Lipid profile:** there was a highly significant increases in total cholesterol, triglycerides, LDL and VLDL levels ( $p < 0.001$ ) and a highly significant decrease in HDL in the treated groups ( $p < 0.001$ ) as compared to control group.

Meanwhile, annatto showed a significant increase in TC/HDL & LDL/HDL ratios ( $p < 0.05$ ), while  $\text{NaNO}_2$  and MSG groups showed a highly significant increase ( $p < 0.001$ ) in both ratios in compare to control values (Table 4).

**Kidney functions:** there was a significant increase in creatinine value in  $\text{NaNO}_2$  group ( $p < 0.05$ ), while there was no significant changes in annatto and MSG groups, whenever urea level

revealed a highly significant increase in all treated groups ( $p < 0.001$ ), in comparison to the control group (Table 5).

**Hormones:** all treated groups revealed a highly significant decline in testosterone ( $p < 0.001$ ), while there was another highly significant increase in T3 and T4 hormones levels ( $p < 0.001$ ) as compared to the control group (Table 6).

**Table (1): Percentage of body weight change and glucose level in control,  $\text{NaNO}_2$ , annatto and MSG treated animals.**

| Parameter        | Control         | $\text{NaNO}_2$     | Annatto              | MSG                  |
|------------------|-----------------|---------------------|----------------------|----------------------|
| % of body weight | $35.34 \pm 0.3$ | $17.69 \pm 2^{**}$  | $30.81 \pm 2$        | $49.31 \pm 2.2^{**}$ |
| Glucose (mg/dl)  | $66.6 \pm 1.3$  | $85.7 \pm 1.5^{**}$ | $72.9 \pm 1.03^{**}$ | $77.2 \pm 1.07^{**}$ |

Values represent mean  $\pm$ SE (standard error). ( $P^* < 0.05$ ,  $P^{**} < 0.001$  as compared to control group).

**Table (2): Serum total protein (g/dl), albumin (g/dl), globulin, albumin/globulin ratio and albumin/creatinine ratio in control,  $\text{NaNO}_2$ , annatto and MSG treated animals.**

| Groups              | Control         | Sodium nitrite       | Anatto               | Mono sodium glutamate |
|---------------------|-----------------|----------------------|----------------------|-----------------------|
| Total Protein(g/dl) | $6.28 \pm 0.4$  | $4.09 \pm 0.2^{**}$  | $4.67 \pm 0.19^{**}$ | $4.36 \pm 0.18^{**}$  |
| Albumin(g/dl)       | $3.86 \pm 0.29$ | $2.19 \pm 0.23^{**}$ | $3.28 \pm 0.21$      | $2.76 \pm 0.19^{**}$  |
| Globulin (g/dl)     | $2.43 \pm 0.1$  | $1.89 \pm 0.05^{**}$ | $1.39 \pm 0.04^{**}$ | $1.6 \pm 0.03^{**}$   |
| Albumin/Globulin    | $1.58 \pm 0.08$ | $1.17 \pm 0.14$      | $2.38 \pm 0.22$      | $1.72 \pm 0.14$       |
| Albumin/creatinine  | $7.9 \pm 1.1$   | $3.4 \pm 0.7^{**}$   | $5.9 \pm 0.7^*$      | $4.7 \pm 0.6^{**}$    |

Values represent mean  $\pm$ SE (standard error). ( $P^* < 0.05$ ,  $P^{**} < 0.001$  as compared to control group).

**Table (3): ALAT and ASAT activities in control,  $\text{NaNO}_2$ , annatto and MSG treated animals.**

| Groups     | Control          | Sodium nitrite        | Anatto                | Mono sodium glutamate |
|------------|------------------|-----------------------|-----------------------|-----------------------|
| ALAT (U/l) | $69.8 \pm 0.88$  | $88.5 \pm 0.9^{**}$   | $81.8 \pm 1.06^{**}$  | $84.1 \pm 0.6^{**}$   |
| ASAT (U/l) | $262.7 \pm 1.06$ | $282.6 \pm 1.16^{**}$ | $277.3 \pm 0.33^{**}$ | $280.6 \pm 1.06^{**}$ |

Values represent mean  $\pm$ SE (standard error). ( $P^* < 0.05$ ,  $P^{**} < 0.001$  as compared to control group).

**Table (4): Changes in total cholesterol (TC), triglyceride (TG), HDL-C, LDL-C, vLDL-C, LDL/HDL ratio and TC/HDL ratio in control,  $\text{NaNO}_2$ , annatto and MSG treated animals.**

| Groups                   | Control          | Sodium nitrite        | Anatto                | Mono sodium glutamate |
|--------------------------|------------------|-----------------------|-----------------------|-----------------------|
| Total Cholesterol(mg/dl) | $55.02 \pm 1.2$  | $76.37 \pm 1.02^{**}$ | $67.44 \pm 1.14^{**}$ | $68.92 \pm 0.51^{**}$ |
| Triglycerides(mg/dl)     | $49.6 \pm 0.8$   | $85.05 \pm 0.7^{**}$  | $86.75 \pm 1^{**}$    | $82.24 \pm 1.02^{**}$ |
| HDL-C (mg/dl)            | $37.976 \pm 1.2$ | $19.19 \pm 1.14^{**}$ | $29.12 \pm 1.1^{**}$  | $24.09 \pm 1.1^{**}$  |
| LDL-C (mg/dl)            | $7 \pm 1.5$      | $40.17 \pm 3.4^{**}$  | $22.17 \pm 2.15^{**}$ | $28.39 \pm 2.7^{**}$  |
| vLDL (mg/dl)             | $9.84 \pm 0.37$  | $17 \pm 0.5^{**}$     | $16.15 \pm 0.5^{**}$  | $16.45 \pm 0.5^{**}$  |
| LDL/HDL                  | $0.178 \pm 0.02$ | $2.15 \pm 0.29^{**}$  | $0.77 \pm 0.08^*$     | $1.21 \pm 0.18^{**}$  |
| TC/HDL                   | $1.44 \pm 0.02$  | $4.06 \pm 0.36^{**}$  | $2.33 \pm 0.12^*$     | $2.91 \pm 0.24^{**}$  |

Values represent mean  $\pm$ SE (standard error). ( $P^* < 0.05$ ,  $P^{**} < 0.001$  as compared to control group).

**Table (5): Serum creatinine and urea levels in control,  $\text{NaNO}_2$ , annatto and MSG treated animals.**

| Groups           | Control         | Sodium nitrite        | Anatto               | Mono sodium glutamate |
|------------------|-----------------|-----------------------|----------------------|-----------------------|
| Creatinine(mg/l) | $0.51 \pm 0.04$ | $0.68 \pm 0.06^*$     | $0.56 \pm 0.04$      | $0.61 \pm 0.04$       |
| Urea(mg/dl)      | $34.44 \pm 0.6$ | $44.39 \pm 1.01^{**}$ | $41.59 \pm 0.8^{**}$ | $43.45 \pm 1.08^{**}$ |

Values represent mean  $\pm$ SE (standard error). (P\* $<$ 0.05, P\*\* $<$ 0.001 as compared to control group).

**Table (6): Serum Testosterone, T3 and T4 levels in control, NaNO<sub>2</sub>, annatto and MSG treated animals.**

| Groups              | Control          | Sodium nitrite     | Anatto             | Mono sodium glutamate |
|---------------------|------------------|--------------------|--------------------|-----------------------|
| Testosterone(ng/dl) | 60.6 $\pm$ 1     | 45.48 $\pm$ 0.9**  | 48.03 $\pm$ 1.2**  | 46.87 $\pm$ 0.76**    |
| T3(ng/dl)           | 94.41 $\pm$ 1.1  | 118.97 $\pm$ 1.7** | 104.6 $\pm$ 1.5**  | 110 $\pm$ 1.7**       |
| T4(ng/dl)           | 5.578 $\pm$ 0.19 | 24.19 $\pm$ 1.17** | 12.54 $\pm$ 0.88** | 17.48 $\pm$ 1.19**    |

Values represent mean  $\pm$ SE (standard error). (P\* $<$ 0.05, P\*\* $<$ 0.001 as compared to control group).

## DISCUSSION

The goal of this study was to assess the side effects of treatment with three types of food additives (sodium nitrite, annatto, and monosodium glutamate) on some physiological parameters in male albino rats. The increase in MSG may be due to the palatability of food and disrupting the hypothalamic signaling cascade of leptin action which cause the link between monosodium glutamate and obesity and its effect on energy balance <sup>(16)</sup>. However, annatto and sodium nitrite groups recorded a significant decrease that may be related to the reduction of food utilization as reported by Grand and Butlar <sup>(17)</sup> or may be due to the increased catabolic processes in the body as reported by Til *et al.* <sup>(18)</sup>. Also, many researchers recorded a reduction in body weight as a result of colorants supplementation. <sup>(19)</sup>

The concentrations of total protein and albumin in the serum can be used as indicators for the state of the liver and differentiation between different types of liver damage <sup>(20)</sup>.

A drop in total protein, albumin and globulin levels in NaNO<sub>2</sub>, annatto and MSG-treated groups were determined by our results. Yousef *et al.* <sup>(21)</sup> indicated an inhibitory effect of some food additives on the biosynthesis of protein and albumin which in turn reflects that the liver is unable to perform its functions. This may be attributed to decrease protein synthesis or especially albumin through its effect on the liver by inhibiting oxidative phosphorylation process as reported by Anthony *et al.* <sup>(22)</sup> or due to the alternation of synthetic function of the liver by MSG.

Treatment with NaNO<sub>2</sub>, annatto, and MSG cause an increase in glucose levels in the blood. Similar results were obtained from Hassan *et al.* <sup>(23)</sup> who reported an increase in glucose level of NaNO<sub>2</sub> treated groups as a result of glucogenesis, and glucose shift from tissue to blood or an impairment of glucose level mobilization. Furthermore, nitroso-compounds can alter the antioxidant system causing a disturbance in the metabolic process leading to hyperglycemia <sup>(24)</sup>.

ALAT and ASAT are used as important biomarkers for the detection of the hepatotoxic effect of different materials on the liver. ALAT and ASAT activities were significant increased (p $<$ 0.001) in all treated groups that may be related to hepatotoxicity or destruction of the liver cells as reported by Ibrahim *et al.* <sup>(25)</sup> and Egbuonu *et al.* <sup>(26)</sup>. Our results are also in agreement with Poli *et al.* <sup>(27)</sup> who showed a dissociation of MSG to free glutamate which produces toxic ammonium ions (NH<sub>4</sub><sup>+</sup>). Thus, the possible ammonium ion overload may occur as a result of an increased level of glutamate following MSG intake could damage the liver, consequently releasing the ALT enzyme that may lead to its elevation.

Data of the present work revealed a highly significant increase in serum total cholesterol, triglycerides, LDL and vLDL while HDL concentration in all treated rats showed a reduction in its level when treated with sodium nitrite, annatto or monosodium glutamate. These results run parallel with Ati *et al.* <sup>(28)</sup> and Hassan *et al.* <sup>(21)</sup>. This elevation in cholesterol may be attributed to the blockage of liver bile ducts, causing reduction or cessation of its secretion to the duodenum. Consequently, it appeared in the serum causing cholestasis <sup>(29)</sup>. Hence, the increased level of serum cholesterol noted here in rats exposed to the NaNO<sub>2</sub> could be attributed to the peroxidation of cell membrane lipids as reported by Standberg <sup>(30)</sup> and Beaupre and Schiffman <sup>(31)</sup>. The higher plasma TG could be linked to the increased number of vLDL particles that associated with visceral fat area in obese individuals as happened in MSG <sup>(32)</sup>. Low HDL-C attributed to high plasma TG that is linked to vLDL metabolism. In plasma, vLDL can exchange TG for CE (Cholesterol Ester) with HDL, a process mediated by cholesterol ester transfer protein (CETP) <sup>(33)</sup>. The exchange of lipids between these two lipoproteins leads to the production of TG-rich HDL particles <sup>(34)</sup>.

Our results demonstrated an increase in serum creatinine and urea concentrations in MSG, annatto or NaNO<sub>2</sub> groups that is in agreement with

Piacenza *et al.* <sup>(33)</sup> and Vinodini *et al.* <sup>(36)</sup>. This may be related to changes in kidney convoluted tubules cell lining as well as in Bowman's corpuscles <sup>(37)</sup>. Furthermore, El-sheikh and Khalil <sup>(38)</sup> observed that there is an elevation in kidney functions parameters after administration of MSG, these impairments could be attributed to the changes in the threshold of tubular reabsorption, renal blood flow and glomerular filtration rate (GFR).

There is a decrease in testosterone hormone in the treated groups as compared with control. The decrease in testosterone hormone in the MSG-treated group agrees with Burde *et al.* <sup>(39)</sup> and Bodnár *et al.* <sup>(40)</sup> that may be resulted from disruption of the hypothalamic-pituitary-testes regulatory axis that controls testosterone production by testicular Leydig cells. This proposition is supported by the reports of previous authors who stated that administration of monosodium glutamate destroyed neurons of the hypothalamus in rats and mice that disrupt the hypothalamic-pituitary-testes regulatory axis, and these results also agree with Ochiogu *et al.* <sup>(41)</sup>. Our study showed an increase in the thyroid hormones (T3 & T4) in the treated groups compared to the control. This effect could be attributed to the chemical structure of NaNO<sub>2</sub> that can compete with thyroxine – binding globulin leading to its deficiency and to hyperthyroidism by feed – back mechanism <sup>(42)</sup>. These changes in thyroid hormones could also be resulted from alteration in the pituitary – thyroid axis as a consequence of the stressing effect of the chemical component; this was in accordance with El-Saadaney <sup>(43)</sup>. Food additives can markedly alter the endocrine function of thyroid gland leading to hyper function. This might play a role in children hyperactivity probably through affecting higher centers in the brain <sup>(44)</sup>.

## REFERENCES

- 1. Milkowski A, Garg HK, Coughlin JR *et al.* (2010):** Nutritional epidemiology in the context of nitric oxide biology: a risk-benefit evaluation for dietary nitrite and nitrate. *Nitric Oxide*, 22: 110-9.
- 2. De Saint Blanquat G, Fritsch P, Cazottes C (1983):** Effects of dietary nitrite and nitrate on experimentally-induced inflammation in the rat. *Int. J Tissue React.*, 5: 173-80.
- 3. Paik DC, Saborio DV, Oropeza R *et al.* (2001):** The epidemiological enigma of gastric cancer rates in the US: was grandmother's sausage the cause. *Int J Epidemiol.*, 30: 181-2.
- 4. Nobre BP, Mendes RL, Queiroz EM *et al.* (2006):** Supercritical carbon dioxide extraction of pigments

from *Bixaorellana* seeds (experiments and modeling). *Brazilian J Chemical Engineering*, 23 (2): 251–258.

- 5. Chuyen HV, Ngoc Hoi NT, Eun JB (2012):** Improvement of bixin extraction yield and extraction quality from annatto seed by modification and combination of different extraction methods. *International J Food Science Technology*, 47: 1333–1338.

- 6. Furtado M, Dyes M (2003):** food industry chooses natural dyes. *Chemicals and Derivatives Magazine*, 421: 1–10.

- 7. EFSA (2016):** The safety of annatto extracts (E 160b) as a food additive. *EFSA Journal*; 14(8):4544.

- 8. Samuels S (1999):** The toxicity/safety of MSG: a study in suppression of information. *Account Res.*; 6: 259-310.

- 9. Ikeda K (2002)** New seasonings. *Chem Senses*; 27: 847-849.

- 10.Bojanić V, Bojanić Z, Najman S *et al.* (2009):** Diltiazem prevention of toxic effects of monosodium glutamate on ovaries in rats. *Gen Physiol Biophys.*; 28: 149-154.

- 11.Seo HJ1, Ham HD, Jin HY *et al.* (2010):** Chronic administration of monosodium glutamate under chronic variable stress impaired hypothalamic-pituitary adrenal axis function in rats. *Korean J Physiol Pharmacol.*; 14: 213-21.

- 12.Iamsaard S, Sukhorum W, Samrid R *et al.* (2014):** The sensitivity of male rat reproductive organs to monosodium glutamate. *Acta Medica Acad.*, 43: 3-9.

- 13.Ortiz GG, Bitzer-Quinter OK, Beas Zárate C *et al.* (2006):** Monosodium glutamate-induced damage in liver and kidney: a morphological and biochemical approach. *Biomed Pharmacother.*; 60: 86-91. .

- 14.Friedewald WT, Levy RI, Fredrickson DS, *et al* (1972):** Estimation of the concentration of low-density lipoprotein cholesterol in plasma, without use of the preparative ultracentrifuge. *Clin. Chem.*18:499-502 (Cited in: *Clin. Chem.*, 1999; 36:15-19).

- 15.Norbert, W. T. (1995):** Clinical guide to laboratory tests. 3rd ed. Saunders W. B., Company, Philadelphia.

- 16.He K, Du S, Xun P *et al.* (2011):** Consumption of Monosodium Glutamate in Relation to Incidence of Overweight in Chinese Adults: China Health and Nutrition Survey (CHNS)". *The American Journal of Clinical Nutrition*; 93(6): 1328-36.

- 17.Grant D and Butler WH (1989):** Chronic toxicity of sodium nitrite in male F344 rat. *Food and Chem Toxicol.*; 27 (9): 565-571.

- 18.Til P, Falke HE, Kuper CF *et al.* (1998):** Evaluation of the oral toxicity of potassium nitrite in a 13 week drinking water study in rats. *Fd Chem Toxic.*; 26 (10): 851 – 859.

- 19.Abou El – Zahab HSH, El – Khyat ZA, Awadallah R *et al.* (1997):** Physiological effects of some synthetic food coloring additives on rats. *Boll Chim Farm.*; 136 (10): 615-627.

- 20.Naganna B, Srivastava LM and Moudgil KD (1989):** Textbook of Biochemistry and Human Biology,

2<sup>nd</sup> Edition, Prentice Hall of India Private Ltd., New-Delhi; 59-61.

**21. Hassan HA and Yousef MI (2010):** Ameliorating effect on chicory (*Cichorium intybus L.*)-supplemented diet against nitrosamine precursors-induced liver injury and oxidative stress in male rats. *Food Chem. Toxicol.*; 48 (8–9): 2163–2169.

**22. Anthony ML, Gartland KP, Beddell CR *et al.* (1994):** Studies on the biochemical toxicology of uranyl nitrate in the rat. *Arch. Toxicol.*; 68 (1): 43 – 53.

**23. Hassan HA, El-Agmy SM, Gaur RL *et al.* (2009):** In vivo evidence of hepato- and reno-protective effect of garlic oil against sodium nitrite-induced oxidative stress. *Inter J Biol Sci.*, 5(3):249- 255.

**24. Wiechetek M, Garwacki S, Karlik W *et al.* (1992):** Effect of nitrite on ureagenesis and carbohydrate metabolism in isolated rat hepatocytes. *Arch Environ. Contamin. Toxicol.*; 24:375-380.

**25. Ibrahim MA, El-Nashar E, Saad S (2010):** The Possible Ultra Structural Ameliorative Effect of Taurine in Rat's Liver Treated with Monosodium Glutamate (MSG). *The Open Hepatology Journal*; 2: 1- 9.

**26. Egbonu C, Cemaluk K and Osakwe ON (2011):** Effects of high monosodium glutamate on some serum markers of lipid status in male Wistar rats. *Journal of Medicine and Medical Sciences*; 2(1): 653-656.

**27. Poli G, Albano E and Dianzani M U (1990):** "Lipid Peroxidation and Covalent Binding in the Early Function Impairment of Golgi Apparatus by Carbon Tetrachloride. *Chem Biol. Interact.*; 8:1-10.

**28. Ati KA, Ati S, Mohammed AM and Saad CA *et al.* (2009):** Response of broiler chicks to dietary monosodium glutamate. *Pakistan Vet J.*; 29(4): 165-168.

**29. Gomaa GMA (1995):** Protective effect of phospholipids and some vitamins against insecticides intoxication in male rats. PhD Thesis, Dep. Zool. Fac. Sci., Ain Shams University.

**30. Standberg A S (1977):** Nitrate and nitrite supply and metabolism in man. *Nutr. Abs. Revs.*; Ser (A) 47: 1119.

**31. Beaupre S R and Schiffman FJ (1994):** Rush hemolysis, a bile cell hemolytic anemia associated with volatile liquid nitrite use. *Arch Fam Med.*; 3: 545 –548.

**32. Okasi M, Usui S, Ishigami M (2005):** Identification of unique lipoprotein subclasses for visceral obesity by component analysis of cholesterol profile in high performance liquid chromatography. *Arterioscler. Thromb Vasc Bio.*; 125:578- 584.

**33. Guerin M, Le Goff W, Lassel TS *et al.* (2001):** Atherogenic role of elevated CE transfer from HDL to VLDL (1) and dense LDL in type 2 diabetes impact of

the degree of triglyceridemia. *Arterioscler. Thromb Vasc Biol.*; 21: 282–288.

**34. Huesca GC, Carreon TE, Nepomuceno MT *et al.* (2004):** Contribution of cholesteryl ester transfer protein and lecithin cholesterol acyltransferase to HDL size distribution. *Endocr. Res.*; 33: 403–415.

**35. Piacenza F, Malavolta M, Cipriano C *et al.* (2009):** L-Arginine normalizes NOS activity and zinc-MT homeostasis in the kidney of mice chronically exposed to inorganic mercury. *Toxicol.*; 189:200–205.

**36. Vinodini NA, Nayanatara AK, Ramaswamy C *et al.* (2010):** Study on evaluation of monosodium glutamate-induced oxidative damage on renal tissue on adult wistar rats. *Journal of Chinese clinical medicine*; 5: 3.

**37. El-Demerdash F M, Yousef M I, Kedwany F S *et al.* (2005):** Cadmium-induced changes in lipid peroxidation, blood hematology, biochemical parameters and semen quality of male rats: protective role of vitamin E and B-carotene. *Food Chem. Toxic*; 42: 1563-71.

**38. El-Sheikh NM and Khalil FA. (2011):** L-Arginine and L-glutamine as immune nutrients and modulating agents for oxidative stress and toxicity induced by sodium nitrite in rats. *Food and Chemical Toxicology*; 49: 758–762.

**39. Burde RM, Schainker B and Kayes J (1971):** Acute effect of oral and subcutaneous administration of monosodium glutamate on the arcuate nucleus of the hypothalamus in mice and rats. *Nature*; 233: 58 – 60.

**40. Bodnar I, Gooz P, Okamura H *et al.* (2001):** Effect of neonatal treatment with monosodium glutamate on dopaminergic and L-DOPA-ergic neurons of the medial basal hypothalamus and on prolactin and MSH secretion of rats. *Brain Research Bulletin*; 55: 767-774.

**41. Ochiogu IS, Ogwu D, Uchendu CN (2015):** Serum Luteinizing hormone, testosterone and total cholesterol levels, Libido and testicular histomorphology of male West African Dwarf goat orally or subcutaneously treated with monosodium L. glutamate. *Pol. J. Food Nutr. Sci.*; 60 (5):253-260.

**42. Gold E and Vladutin A (1994):** Latrogenic hyperthyroidism of long duration in an individual with thyroxin – binding globulin deficiency. *Clin Chem.*, 40 (12): 2323–2324.

**43. El-Saadany SS (1991):** Biochemical effect of chocolate colouring and flavoring like substances on thyroid function and protein biosynthesis. *Die Nahrung*; 4: 335–43.

**44. Helal EGE (2001):** Progressive effects of the interaction of Sodium nitrite and sunset yellow on different physiological parameters in albino rats. *The Egypt J Hospit Med.*, 2: 23–46.