

REVIEW ARTICLE

The Vast Biomedical Applications of Zinc Oxide Nanoparticles

Ismail G. Abd-Elmaqsoud^{1,2}, Hamad A. Elsaadawi², Amany I. Ahmed², Adel AbdelKhalek³,
Ahmed H. Arisha^{1,4*}

¹Department Animal Physiology and Biochemistry, Faculty of Veterinary Medicine, Badr University in Cairo (BUC), Badr City, Cairo, Egypt.

²Department of Biochemistry, Faculty of Veterinary Medicine, Zagazig University, Zagazig, 44511, Egypt.

³Department of Veterinary Medicine, Badr University in Cairo (BUC), Badr City, Cairo, Egypt.

⁴Department Physiology, Faculty of Veterinary Medicine, Zagazig University, 44511 Zagazig, Egypt.

* Corresponding author: vetahmedhamed@zu.edu.eg

Article History: Received: 21/06/2022 Received in revised form: 16/07/2022 Accepted: 25/07/2022

Abstract

Biomedical nanomaterials have gained a lot of attention in the last decade. They have emphasized various concerns due to their vast and significant biological properties and biomedical applications. Metal oxide nanoparticles offer a wide range of medicinal uses, including anticancer, drug/gene delivery, antibacterial, cell imaging, and biosensing, among others. Zinc oxide nanoparticles (ZnO-NPs) have been employed like a key material in a range of industries in last years, including medicine, cosmetics, concrete, antimicrobials, and textiles. the automotive industry, and cancer prevention. Anticancer and antibacterial effects are linked to the capability of ZnO-NPs to produce reactive oxygen species (ROS) and trigger cell programmed death (apoptosis). The following review articles summarize the different biomedical applications of ZnO nanoparticles.

Keywords: Nanoparticles; Zinc Oxide Nanoparticles; Antidiabetic; antibacterial.

Introduction

Nanoparticles and nanomaterials have a growing number of applications, and working with them has enabled a much deeper knowledge of biology [1, 2]. Zinc oxide nanoparticles (Zn-ONPs) are broadly utilized in a variety of sectors as much as their particular physical and chemical characters [3, 4]. Zn-ONPs also exhibit outstanding antibacterial, antimicrobial, and UV blocking capabilities. As a result, final fabrics containing Zn-ONPs in the textile industry display attractive UV and visible light resistance, antibacterial, and deodorant properties [5].

Zinc is a naturally occurring element that plays an important role in the metabolism of man, animals, and plants

[6]. Zinc is broadly recognized as an essential trace element that can be found in all body tissues, including bone, brain, muscles, and skin. Zinc is an essential component of many enzyme systems and plays an important role in protein and nucleic acid production, hematopoiesis, and neurogenesis [3, 4, 7, 8]. Nano-ZnO is more easily absorbed by the body due to small particle size of Zinc. As a result, nano-ZnO is widely used in the food industry. Zinc Oxide (ZnO) is also designated as a "GRAS" (Generally Recognized As Safe) approved by Food and Drug Administration (FDA) [9]. ZnO is broadly utilized in pharmaceutical, cosmetic, and medicinal applications and is acknowledged to be a valuable nutritional addition. Despite the fact that zinc oxide dust and fumes are typically

considered safe, breathing them should be avoided. [6] ZnO is found as the mineral zincite in the earth's crust, but most of it is synthesized for commercial use. ZnO is made safe and compatible with human skin by adding it to fabrics and surfaces that come into touch with flesh. Zn-ONPs are broadly used in variance of industries, including UV light-emitting devices [10, 11], ethanol gas sensors [12, 13], photocatalysts [14, 15], industries of pharmaceutical and cosmetics [16]. Non-toxic and self-cleaning are just a few of the benefits of ZnO-NPs [17], UV-blockers were used in sunscreens and a diversity of medical specialty practice because they are skin-friendly, antibacterial, and dermatologic associate degrees[18]. ZnO appears to have a high resistance to germs, and various studies have found that CaO, MgO, and ZnO have a significant antibacterial drug activity. [19] This is because of the creation of Reactive Oxygen Species (ROS) on the oxides' surfaces as summarized in Figure (1).

Various studies have utilized ZnO-NPs as a drug delivery mechanism in a variety of disorders. These nanoparticles have been shown to be capable of delivering drugs to a variety of cells and tissues [2]. ZnO-NPs have the probability to be employed as a bioimaging tool. Zn-ONPs have received a lot of attention in biomedical applications because of these features. Zn-ONPs, which have economic price and less toxic than other metal oxide NPs, have a broad range of medicinal practice, including anticancer, antimicrobial, and diabetic treatment; anti-inflammation; wound healing; and bioimaging [20-22]. Because of their great ultraviolet (UV)absorption and transparency for visible light, Zn-ONPs are efficient sunscreen agents [23, 24]. Other features, such as anticancer activity and antibacterial, have also been investigated as a result of their capacity to create (ROS) [25]. Zn-ONPs are effective

drug carrier systems in addition to their intrinsic biological features. Bulk ZnO has also been certified as GRAS material by the US Food and Drug Administration (FDA) and Zn-ONPs bigger than 100 nm are regarded biocompatible, indicating that they could be used for drug administration. [26]. Another metal NPs, for example as iron oxide NPs, have anticancer activity as well, although no well-defined antibacterial or UV-absorbing properties have been found. In addition, when make a comparison to other metal oxide NPs, Zn-ONPs are more affordable, biocompatible, and low poisonous, which further enhance its application potential [27, 28]

Biomedical Applications of ZnO Nanoparticles

As a recent form of Lower cost, less-toxic nanomaterial, Zn-ONPs have generated a lot of interest in biomedical domains like antioxidant, anticancer, antibacterial, anti-inflammatory activities, and anti-diabetic, as well as bioimaging and drug administration [9, 21]. This study summarizes some developments in the use of Zn-ONPs in biomedical applications. Zn-ONPs with a diameter of less than 100 nm are deemed biocompatible, which supports their biomedical applications and is a strong motivator for biomedical research as summarized in Figure (2) and with special emphasis to their mechanism of action as summarized in Table (1).

1. Anticancer Activity

Chemotherapy, radiation, and surgery have all been used to treat cancer in the past few decades. Cancer is a disease marked by uncontrolled malignant cell proliferation. Although all of these drugs appear to be efficient for killing cancer cells in theory, they all have a long list of negative side effects [29]. Zn-ONPs cause malignant cells to die without causing cytotoxicity in healthy cells [30, 31]. Before Zn-ONPs may be used in

medicine, a number of issues must be addressed, including a best understanding of the mechanism of their specific cytotoxic effect and the shortage appropriate biocompatible dispersion techniques [32]. ZnO-NPs have been found to have deleterious outcome on the vitality of primary human T cells at concentrations that are noxious to both gram positive and gram negative bacteria, according to a study mentioned by Reddy *et al.* [33]. According to many studies, these nanomaterials are innocuous to grown human dermal fibroblasts, but they are harmful to metastatic tumor cells [34] and vascular endothelial cells [35]. At the same time stimulating programmed cell death (apoptosis) in neural stem cells. Nanomaterial-based nanomedicine has nowadays showed the capacity to resolve these negative outcomes because of its high biocompatibility, cancer targeting, easily of surface functionalization, and drug distribution capacity. Adults require Zn^{2+} , and ZnO nanoparticles are regarded harmless in Living organism. Due to these benefits, Zn-ONPs can be used as biodegradable and biocompatible nanoplatfroms, as well as being investigated for cancer therapy [36, 37]. Zn-ONPs show anticancer action via causing the production of ROS as well as promoting apoptosis. Furthermore, the electrostatic characteristics of Zn-ONPs, which have been used for anticancer activity, are a helpful feature. Because of the chemisorbed neutral hydroxyl groups on their surface, Zn-ONPs have unique surface charge behavior. In an aqueous solution with a high pH, protons (H^+) flow away from the particle surface, generating a negative charge on surface with oxygen atoms that are only partially bonded (ZnO). At lower pH, protons from the surrounding environment are transported to the particle surface, resulting in a positive charge on surface ($ZnOH_2^+$) [9, 38]. Zn-ONPs have an isoelectric point of 9–10, under physiological parameters they have a high

positive surface charge [39]. Cancer cells, on the other hand, have substantial negative membrane potentials and on the surface of their membranes they have a high concentration of anionic phospholipids (phosphatidylserine) [40, 41]. Cancer cells interact with positive charge Zn-ONPs because of electrostatic interactions that boost cellular absorption, phagocytosis, and cytotoxicity [9].

1.2. Anticancer by Autophagy

Autophagy is a catabolic process which is strictly controlled that is triggered by many stressors such as damaged organelles, ROS, protein aggregation and anticancer drugs. Highly cellular damage can cause cancer cell apoptosis by cellular self-consumption and extending autophagy, which results in cell death [42, 43]. As a result, autophagy enhances cell viability, also activates death pathways in cancer cells making it a key event in nanoparticle induced cytotoxicity.

1.3. Anticancer by using Zn-ONPs in drug delivery.

The use of nanoparticles in focused drug delivery raises the potential for cancer treatment which are safer and more effective. By targeting specific areas on cancer cells, nanoparticle based drug delivery has the potential to lower the overall amount of medications used and, as a result, prevent undesirable side effects [9, 44]. Zn-ONPs are appealing owing to their lower toxicity and biodegradability when compared to other nanomaterials. The usage of Zn-ONPs in cancer medicine delivery has attracted a lot of interest. Drugs such as curcumin, doxorubicin, paclitaxel, and baicalin, as well as DNA fragments, may be loaded onto ZnO nanoparticles to demonstrate improved toxicity, solubility, and delivery into cancer cells compared to individual compounds [45-48]. Previous research has suggested that ROS and autophagy have a role in Zn-ONPs cytotoxicity. The regulative mechanisms that govern autophagy and ROS, on the other hand, have yet to be found. Zhang

et al. [49] explored the control mechanism of autophagy and the relationship between autophagy and ROS in lung epithelial cells mediated with Zn-ONPs.

1.4. Targeting Functionalization

Apart from specificity and localization, targeted nanoparticles (NPs) offer additional therapeutic benefits such as large payload, multidrug conjugation, facile discharge kinetics adjusting, selectiveness localization, and bypassing multidrug resistance mechanisms [50]. Many functionalization approaches for nanoparticle modification have been documented in conductive to improve the targeting effects and selectivity against cancer cells. Surface modification of Zn-ONPs increased their persistence and specificity against certain cancer cells even more. Surface modification of Zn-ONPs with a variety of biological substances, such as folic acid, proteins, peptides, nucleic acids, hyaluronan, and so on, is the focus of the research [51-55]. The anticancer activity of Zn-ONPs was unaffected by the biocompatible coating. However, the ability to target cancer cells was enhanced, and the protection against normal cells has been improved.

2. Antibacterial Activity

Zn-ONPs can be employed as an antibacterial material due to its remarkable properties, which include a large specific surface area and the ability to kill a wide range of pathogens. However, the antibacterial action of Zn-ONPs has just lately been discovered. In the formation of nanoscale systems and microscale for therapeutic uses, Zn-ONPs have accepted to be a potent drug. Although Zn-ONPs appear to have greater medical curative activity than microparticles, the exact mechanisms of action of drug have yet to be determined [56]. The Zn-ONPs have germicidal properties against both gram negative and gram positive bacteria [57]. They also have antimicrobial efficacy in the face of

spores that can withstand high temperatures and pressure. The pharmacological action of Zn-ONPs is dependent on their concentration and size; with crystalline structure and particle form have little effect. As a result, it is revealed, the greater the extent and concentration of nanoparticles, the greater the therapeutic pharmacological activity. The mechanism of Zn-ONPs' medical pharmacological action is still unknown in its totality. In their investigation, some researchers stated that the production of hydrogen peroxide is the primary issue with medical action, while others suggested that particle adherence on the microorganism surface owing to fixed forces could be other factor [32]. With increased particle dose, treatment period, and synthesis technology, nanoparticle efficacy will rise. Furthermore, the particulate size variability and surface area to volume ratio of green Zn-ONPs are undeniably the source of the significantly improved antibacterial activity revealed by this data. According to the researchers, unpracticed Zn-ONPs had the ability to be employed efficiently in food safety applications and agricultural, as well as to solve future medical challenges [58]. Gram negative bacteria such as *Escherichia coli* (*E. coli*) and Gram positive bacteria such as *Staphylococcus aureus* (*S. aureus*) are now the most commonly used model bacteria to investigate Zn-ONPs antibacterial activity [33, 59]. Some other Gram-negative bacteria such as *Vibrio cholerae* (*V. cholerae*) [60], *Pseudomonas aeruginosa* (*P. aeruginosa*) [61, 62], *Proteus vulgaris* (*P. vulgaris*) [63], and other Gram positive bacteria such as *Enterococcus faecalis* (*E. faecalis*) [64], and *Bacillus subtilis* (*B. subtilis*) [65] are also investigated. Jiang *et al.* [66] recorded that Zn-ONPs have antibacterial properties against *E. coli*. Zn-ONPs with mean range size of 30 nm were discovered to destroy cells by directly touching the membrane's phospholipid

bilayer and damaging membrane integrity, Zn-ONPs' ability to kill bacteria could be inhibited by the addition of radical scavengers like mannitol, vitamin E, and glutathione, possibly indicating that ROS generation was essential for Zn-ONPs' antibacterial effects. However, it did not appear that Zn^{2+} released from ZnONP suspensions had an antimicrobial impact. Reddy *et al.* [36] made Zn-ONPs with a diameter of 13 nm and tested their antibacterial activity against *E. coli* and *S. aureus* germs, according to the results, Zn-ONPs fully stopped the development of *S. aureus* at doses of less than 1 mM, but completely resisted the growth of *E. coli* at roughly 3.4 mM [33].

The Gram-negative bacterium *V. cholera* infects the intestine and causes cholera, a deadly diarrheal disease, primarily affects people in impoverished nations [60, 67]. Sarwar *et al.* [68] investigated the impact of Zn-ONPs on *Vibrio cholerae* (two cholera bacteria biotypes) (classical and El Tor) with the goal of developing nanomedicine against cholera. Zn-ONPs were found to be more effective in resisting the growth of the El Tor (N16961) biotype of *V. cholera*, which was linked to the formation of ROS. These outcomes would harm bacterial membranes, enhance permeabilization, and alter their shape significantly. In cholera toxin (CT) mouse models, Zn-ONPs exhibit antibacterial properties. Zn-ONPs have been found to have the ability to cause the CT secondary structure to gradually collapse and interact with CT through interfering with CT attachment to the GM1 gangliosides receptor [69]. The specific antibacterial mechanism of Zn-ONPs remains unknown. As a result, understanding it in depth provides a lot of theoretical and practical significance. We expect Zn-ONPs will be investigated as antibacterial agents in the future, such as lotions, creams, and mouthwashes. Also, it can be coated on a variety of substrates to inhibit

germ from adhering to, propagating through, and reproducing in biomedical application. The capacity of Zn-ONPs to cause oxidative stress also contributes to their antibacterial activity. The thiol group of respiratory enzymes interacts with Zn^{2+} ions produced by ZnO, limiting their function. Zn-ONPs have been shown to influence the cell membrane and lead to the generation of ROS. When bacteria contact with Zn-ONPs, they uptake Zn^{2+} , which suppresses respiratory enzymes, produces ROS, and free radicals, causing oxidative stress. ROS causes irreversible damage to bacterial membranes, DNA, and mitochondria, culminating in bacterial cell death [70]. Ghasemi and Jalal [71] reported that Zn-ONPs were evaluated for their effect on the efficacy of the traditional antibiotics ciprofloxacin and ceftazidime, also their mechanisms of action against resistance *Acinetobacter baumannii*, an opportunist bacterium that causes pneumonia and meningitis. The antibacterial activity of both medicines enhanced when presence of a sub-inhibitory concentration of Zn-ONPs, according to the findings. Combining antibiotics with Zn-ONPs boosted antibiotic absorption and converted bacterial cells from rods to cocci. The production of ROS and DNA alteration were also reported. These results suggest that Zn-ONPs and antibiotics can be used in combination to treat bacterial infections. It has also been found that Zn-ONPs improve the photosensitizer crystal violet's (CV) antibacterial activity [72].

Both Gram-negative and Gram-positive microbes exhibit antibacterial activity in response to Zn-ONPs [73]. Zhang *et al.* [74] investigated both dosage- and moment-dependent phenotypic bacterial reaction to Zn-ONPs using Surface enhanced Raman spectroscopy (SERS) method. Their findings revealed spectral change profiles that were both clear and informative. Significant changes were seen in smaller

dose limit rather than bigger dose limit, demonstrating a decrease in Zn-ONPs bioavailability as dosages were increased. Within 0.5 hour, rapid activity was established, and smaller doses and lengthy exposure times had similar outcome to large doses.

3. Antimicrobial Potential of Zn-ONPs

In order to overcome multi-drug resistance, Zn-ONPs have been researched for the creation of next-generation nano-antibiotics against pathogenic microbes [75, 76]. These nanoparticles exhibit distinctive morphology, particle size, crystallinity, and porosity characteristics [77]. Based on these features, Zn-ONPs have a broad spectrum of antimicrobial activity against a variety of microbes, including the *M13 bacteriophage*, *Pseudomonas aeruginosa*, *Escherichia coli*, *Staphylococcus aureus*, and *Pseudomonas aeruginosa* [78-81]. In both non-clinical and clinical settings, they can be used with antibiotics and inflammatory medications to increase antimicrobial effect against pathogenic microbes without antibiotic resistance [81, 82].

ZnO is being studied as a medication agent in formulations at the micro and Nano scale levels, although the specific mechanism of drug action is unknown, it has been claimed that the majority of reasons of cell ballooning are the rule of ROS generated on particle surfaces, zinc ion discharge, membrane malfunction, and nanoparticle acquisition area unit. Processing of Zn-ONPs at high temperatures has a significant impact on their medicinal action, whereas treatment at lower temperatures has a reduced impact [6]. Zn-ONPs are being investigated in conjunction with medical procedure ablation protocols. Nanoparticles will give antineoplastic drug specialized that shows a synergistic antineoplastic action in the presence of heat, and they may even be photographed

to obtain accuracy in medicinal assistance. Furthermore, lead to a stronger thermal impact on neoplasm ablation. Several studies have shown that understanding the molecular process underlying tumor-mediated nanoparticle ablation could aid in the development of nanoparticles with the right composition and characteristics to enhance the ablation property [83-85]. Mechanisms of antimicrobial actions of ZnO materials have been recorded in association with particular interactions based on their unique physicochemical properties of (a) Zn^{2+} ion release, (b) adsorption, (c) ROS generation [86, 87], and the intracellular responses in microorganisms of (d) energy metabolism inhibition; (e) lipid peroxidation, Nanomaterials 2021, 11, 263 8 of 35 and cell membrane damage; and (f) DNA replication disruption, and DNA break [88, 89]. Due to disruption of enzyme systems and interference with metabolic activities, the Zn^{2+} ions produced from ZnO-NPs/MPs cause an antimicrobial response in microorganisms [90, 91]. Additionally, ZnO-NPs/MPs have the ability to adsorb particles to the bio-membrane through a charge-charge interaction and generate ROS as photocatalysts when exposed to UV and visible light [77, 92-94]. Zn-ONPs/MPs with positively charged surfaces interact with microorganisms with negatively charged cell walls or bio-membranes [95]. After adsorption, they are ingested by the bacteria, causing cell integrity to be lost as a result of membrane or cell wall rupture. They also mediate oxidative stress due to lipid peroxidation, which causes DNA damage. Depending on their physicochemical properties, such as shape, particle size, and porosity, Zn-ONPs/MPs have a variable susceptibility to pathogenic bacteria based on the basic modes of action [77, 95-97]. Zn-ONPs, and occasionally MPs; have improved antimicrobial activity even toward dangerous viruses. They also offer wide-spectrum antimicrobial activity that can

be tailored for several treatment options and used to increase their suitability as commercial and clinical translation platforms when combined with other antibiotic medicines, metal oxide NPs/MPs or metal doping, and other biomaterials [98].

4. Anti-diabetic activity

Zinc has a key role in insulin storage, biosynthesis, and secretion; hence Zn-ONPs have been studied for their anti-diabetic properties. Zn-ONPs were found to have good anti-diabetic efficacy than ZnSO₄, as evidenced by enhanced insulin levels, glucose disposal, and zinc state when compared to ZnSO₄ [99]. Wahba *et al.* [100] recorded that ZnO-NPs efficiently corrected diabetes stimulated pancreatic disease, as evidenced by ultrastructural and structural improvements and biochemical normalization of serum insulin and blood glucose. In another experiments, ZnO-NPs were combined with the anti-diabetic medicines red sandalwood and vildagliptin to boost their effectiveness [101, 102]. By breaking down oligosaccharides and disaccharides, α -amylase enzyme secreted in the pancreas and α -glucosidase enzyme secreted in the intestine produced monosaccharide (glucose) [103]. Diabetes mellitus is a metabolic disorder caused by a defect in insulin secretion or activity. It is characterized by persistent hyperglycemia and ineffective carbohydrate metabolism [104]. As a result of insulin's poor performance, blood glucose levels in diabetes patients continue to be elevated. Therefore, it is essential to regulate blood glucose levels by blocking the enzymes α -glucosidase and α -amylase. There are numerous medications available today to suppress the α -amylase and α -glucosidase enzymes, but they also have some negative effects [105].

Ci-ZnO-NPs inhibit α -amylase and α -glucosidase enzymes, for α -amylase, the

proportion of inhibition ranged from 20% (20 g/ml) to 74 percent (100 g/ml), while for α -glucosidase, it ranged from 36% (20 g/ml) to 82 percent (100 g/ml). Compared to α -amylase, α -glucosidase had a higher rate of inhibition [106].

5. Antifungal activity

ZnO-NPs are also excellent antifungal agents in addition to their antibacterial properties. Surendra *et al.* [107]. synthesized ZnO-NPs from *M. oleifera*, which were harmful to two plant disease strains, *Sclerotium rolfsii* and *Alternaria saloni*. Gunalan *et al.* [108] investigated the antifungal action of Zn-ONPs in a variety of fungal strains and recorded that they were toxic to plants and foodstuff pathogens in the following order: *Rhizopus stolonifera* > *Aspergillus flavus* > *Aspergillus nidulans* > *Trichoderma harzianum*. As a result, the scientists speculated that ZnO-NPs could be useful in the agricultural and food industries. ZnO-NPs have a concentration dependent influence on *Candida albicans* viability, according to Lipovsky *et al.* [109]. ZnO-NPs (0.1 mg/ml) decreased *C. albicans* vitality by >95 percent. Adding visible light to ZnO-NPs increased yeast cell death even more.

6. Anti-Inflammatory activity

The effectiveness of nano-ZnO and bulk-ZnO was investigated by Ilves *et al.* [110]. based on ZnO's anti-inflammatory properties and found that only nano-ZnO could permeate the deepest layers of allergic skin. Nano-ZnO also suppressed local skin irritation and promoted the synthesis of IgE antibodies in the system. According to the authors, this outcome is the consequence of nonspecific reactions triggered by liberated Zn²⁺ impairing B cells' ability to produce IgE.

7. Treatment of a range of different skin conditions

Zinc Oxide is commonly found in products that used in treatment of a

variety of skin disorders such as diaper rash powder, barrier creams, hemimorphite cream, antimicrobial ointments, and anti-dandruff shampoos. It's also a component of tape (known as "zinc chemical compound tape"), which is utilized by sport manas a bandage to prevent soft tissue damage during exercises [111]. ZnO-NPs can be used in creams, ointments, and lotions to protect the skin against sunburn and other skin injuries induced by UV light. It's the most broad-range UVA and UVB reflector that the Bureau has certified for use as a sunscreen, and it's entirely photo-stable [112].

8. Drug delivery

Drug delivery has been proved as an effective instrument in the therapy of many diseases including cancer, among the various uses of nanotechnology [1, 2]. One of the most essential systems in medicine administrations is nanoparticles (NPs). In several researches, ZnO-NPs were employed for medication delivery in a variety of disorders [1, 2]. In a study, Yuan *et al.* [113] used ZnO quantum dots as a medicine administration device to target doxorubicin in HeLa cells. To improve the nanomaterial stability, they encased ZnO-NPs in chitosan. Their findings suggested that drug delivery technology could be utilized to efficiently deliver doxorubicin to cancer cells [113]. Another important feature of NPs' application is their usage as carriers for gene transport to various cells, specially malignant cells [2]. The usage of this technology for gene transfer comes with several benefits. For example, putting plasmid-containing genes on the surface of NPs could allow for safely and effectively gene delivery to the recipient tissues [1, 2].

9-Bio-imaging

Using this technique to distribute genes has several benefits. For example, plasmid encoded genes expressed on the

surface of NPs may ensure that genes are transported safely and efficiently to the receiving tissues [1, 22]. Different forms of ZnO nanostructures have been identified in last years, including NPs, nanotubes, nanorods, and nanorings. Researchers are interested in using ZnO-NPs as bio-imaging agents [1, 22]. This feature has a variety of biological and medicinal uses and can be used at various levels. For example, luminous ZnO-NPs, also known as ZnOQDs, may have favorable photophysical properties [1, 22].

ZnO is generally considered to be a safe compound. ZnO has been used in sunscreen goods and as a food ingredient in food packaging. As a result, ZnO-NPs' luminous characteristics could be exploited in a variety of biological and medicinal applications. [1, 2, 22].

The nanotoxicological effect of ZNO-NPs

For instance, ZnO, which is typically thought of as a water insoluble compound, was demonstrated to exert toxicity through the release of Zn-ions in the case of ZnO nanoparticles, and it was found that the cellular uptake pathways of NPs were dependent on the size, shape, and surface characteristics of nanoscale particles. Nanotoxicologists have acknowledged the need for improved knowledge about the molecular interactions of NP with biological systems in order to better predict the potential toxicity of novel nanomaterials and ensure the safe and sustainable development of nanotechnology as knowledge on the mechanisms and behavior of NP toxicity in the environment and organisms has grown [114]. Given that NP toxicity has been shown to depend on interactions with organic matter in the environment, anions and cations, pH levels, and other environmental factors, environmental nanotoxicology is moving in the direction

of increasing the relevance of testing conditions (e.g., media composition, exposure concentrations, and duration) to those relevant to the environment and organism physiology. The safety evaluation of nanomaterials created for use in biotechnology, environmental bioremediation, wastewater treatment, agriculture, and nanomedicine are another significant area of nanotoxicology that has gained traction in last years. In order to ensure effective nano innovation across

a wide range of applications and to enable the safe-by-design approach in nanotechnology, nanotoxicology plays a significant role not only in the risk assessment of purposefully or inadvertently created nanomaterials [114] The long-term toxicity of ZnO-NPs to microalgae was evaluated by Aravantinou *et al.* [115] utilizing a simulated natural water treatment system with a semi-continuous supply of NPs.

Table 1. Biomedical application of ZnO-NPs.

Biomedical Applications of ZnO Nanoparticle	Mechanism of action	Reference
1-Anticancer Activity	In human liver cancer cell (HepG2) →↑ p53, Bax (mRNA and protein) and ↓ caspase-3, DNA Fragmentation, bcl-2. ↑ ROS →↑ cytoplasmic trigger of calcium ions and [30, 31] ↑ reaction with the cytoplasmic membrane → loss of membrane integrity →↑ calcium exit via membrane channels.	[30, 31]
2-Antibacterial Activity	The oxidation of fatty acids →↑ lipid peroxides chain reaction → disruption of plasma and organelle membranes → cell death.	[32]
3-Antimicrobial of ZnO-NPs	Potential Cell swelling due to →↑ROS, zinc oxide and membrane dysfunction.	[6]
4- Antidiabetic activity	↑ phosphorylation of insulin receptor B-subunit, phosphatidyl inositol 3-kinase (PI3-K) and stimulate protein kinase B (PKB) → regulation glucose metabolism .	[99]
5- Antifungal activity	↑ surface → oxygen species →↑ disruption of the membrane Pathogen die .	[108]
6-Anti-Inflammatory activity	Reduced Zn ²⁺ ion → in Blood, Lymph node and spleen →↑ Abs →↑ B-cell driven IgE antibody production.	[110]
8-Drug delivery	Drug administration has proved as an effective instrument in the therapy of many diseases including cancer, among the various uses of nanotechnology.	[1, 2]
9-Bio-imaging	This feature has a variety of biological and medicinal uses and can be used at various levels. For example, luminous Zn-ONPs, also known as ZnO-QDs, may have favorable photophysical properties.	[1, 22].

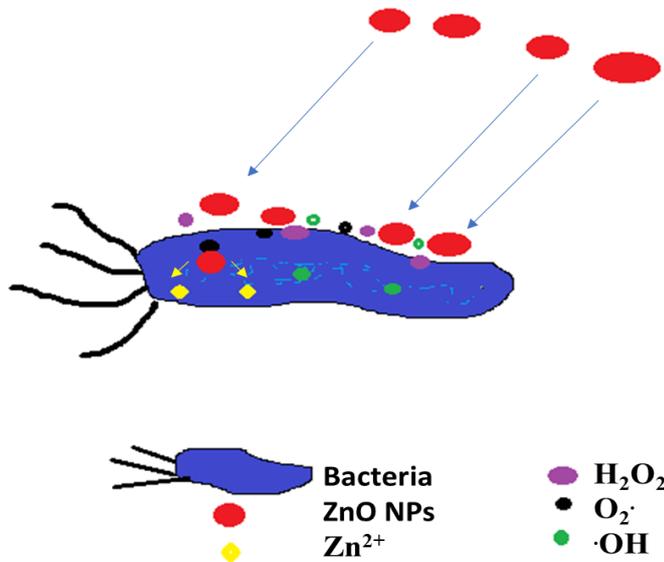


Figure 1: Schematic diagram of antibacterial activity of Zn-ONPs.

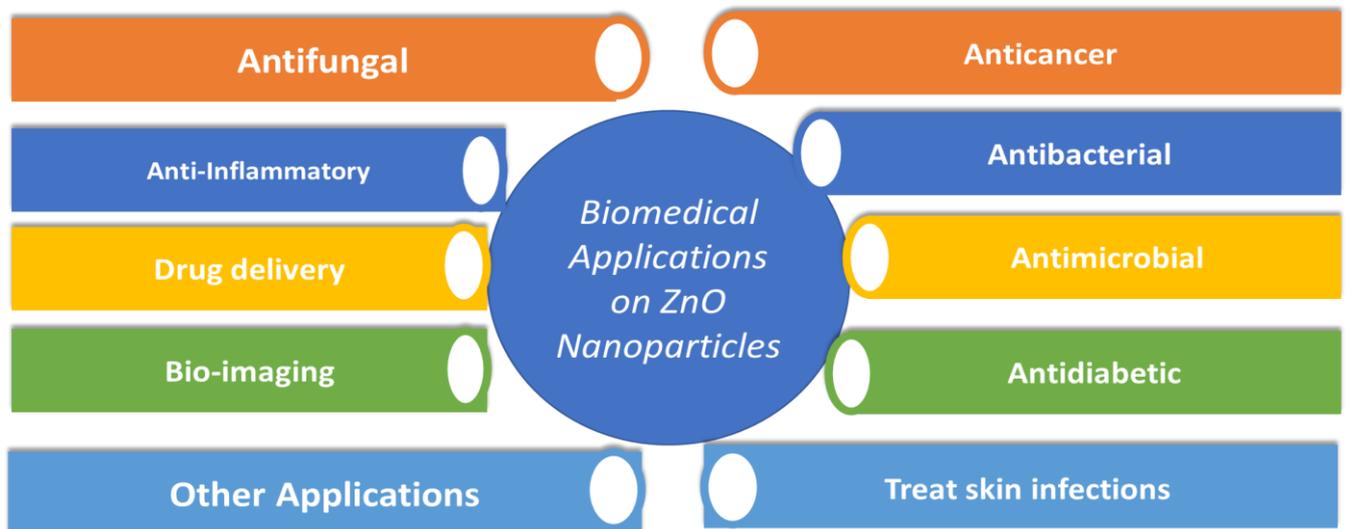


Figure 2: Biomedical application of ZnO Nanoparticles.

Conclusions

ZnO-NPs are known for their ability to create ROS and trigger apoptosis, like other metal oxide NPs, offer great medicinal potential. ZnO-NPs are useful as anticancer, antibacterial, and antifungal agents because of their properties. When loaded and given with other medicinal regimens, ZnO-NPs have been shown to have synergistic benefits.

Targeted drug delivery and clinical diagnostics are becoming more common as ZnO-NPs are a type of nanomaterial that has a lot of applications in medical

purposes and green technologies, and they can be safely manufactured at a cheap cost.

Conflict of interest

All the authors have no conflict of interest to declare.

References

[1] Asharani, P.; Wu, Y.L.; Gong, Z. and Valiyaveetil, S. (2008): Toxicity of silver nanoparticles in zebrafish models. *Nanotechnology*, 19(25): 255102.
 [2] Taylor, E. and Webster, T.J. (2011): Reducing infections through

- nanotechnology and nanoparticles. *International journal of nanomedicine*, 6: 1463.
- [3] Ruszkiewicz, J.A.; Pinkas, A.; Ferrer, B.; Peres, T.V.; Tsatsakis, A. and Aschner, M. (2017): Neurotoxic effect of active ingredients in sunscreen products, a contemporary review. *Toxicology reports*, 4: 245-59.
- [4] Smijs, T.G. and Pavel, S. (2011): Titanium dioxide and zinc oxide nanoparticles in sunscreens: focus on their safety and effectiveness. *Nanotechnology, science and applications*, 4: 95.
- [5] Hatamie, A.; Khan, A.; Golabi, M.; Turner, A.P.; Beni, V.; Mak, W.C.; Sadollahkhani, A.; Alnoor, H.; Zargar, B. and Bano, S. (2015): Zinc oxide nanostructure-modified textile and its application to biosensing, photocatalysis, and as antibacterial material. *Langmuir*, 31(39): 10913-21.
- [6] Mirzaei, H. and Darroudi, M. (2017): Zinc oxide nanoparticles: Biological synthesis and biomedical applications. *Ceramics International*, 43(1): 907-14.
- [7] Kołodziejczak-Radzimska, A. and Jesionowski, T. (2014): Zinc oxide—from synthesis to application: a review. *Materials*, 7(4): 2833-81.
- [8] Sahoo, S.; Maiti, M.; Ganguly, A.; Jacob George, J. and Bhowmick, A.K. (2007): Effect of zinc oxide nanoparticles as cure activator on the properties of natural rubber and nitrile rubber. *Journal of applied polymer science*, 105(4): 2407-15.
- [9] Rasmussen, J.W.; Martinez, E.; Louka, P. and Wingett, D.G. (2010): Zinc oxide nanoparticles for selective destruction of tumor cells and potential for drug delivery applications. *Expert opinion on drug delivery*, 7(9): 1063-77.
- [10] Izu, N.; Shimada, K.; Akamatsu, T.; Itoh, T.; Shin, W.; Shiraishi, K. and Usui, T. (2014): Polyol synthesis of Al-doped ZnO spherical nanoparticles and their UV-vis-NIR absorption properties. *Ceramics International*, 40(6): 8775-81.
- [11] Rajalakshmi, M.; Sohila, S.; Ramya, S.; Divakar, R.; Ghosh, C. and Kalavathi, S. (2012): Blue green and UV emitting ZnO nanoparticles synthesized through a non-aqueous route. *Optical Materials*, 34(8): 1241-5.
- [12] Guo, J. and Peng, C. (2015): Synthesis of ZnO nanoparticles with a novel combustion method and their C₂H₅OH gas sensing properties. *Ceramics International*, 41(2): 2180-6.
- [13] Xie, J.; Cao, Y.; Jia, D.; Li, Y. and Wang, Y. (2016): Solid-state synthesis of Y-doped ZnO nanoparticles with selective-detection gas-sensing performance. *Ceramics International*, 42(1): 90-6.
- [14] Deng, Z.; Chen, M.; Gu, G. and Wu, L. (2008): A facile method to fabricate ZnO hollow spheres and their photocatalytic property. *The journal of physical chemistry B*, 112(1): 16-22.
- [15] Yang, S.J. and Park, C.R. (2007): Facile preparation of monodisperse ZnO quantum dots with high quality photoluminescence characteristics. *Nanotechnology*, 19(3): 035609.
- [16] Lu, P.-J.; Huang, S.-C.; Chen, Y.-P.; Chiueh, L.-C. and Shih, D.Y.-C. (2015): Analysis of titanium dioxide and zinc oxide nanoparticles in cosmetics. *Journal of food and drug analysis*, 23(3): 587-94.
- [17] Shirgholami, M.A.; Nazari, A. and Mirjalili, M. (2015): Statistical optimization of self-cleaning technology and color reduction in wool fabric by nano zinc oxide and eco-friendly cross-linker. *Clean Technologies and Environmental Policy*, 17(4): 905-19.
- [18] Krishnakumar, T.; Jayaprakash, R.; Pinna, N.; Singh, V.; Mehta, B. and Phani, A. (2009): Microwave-assisted synthesis and characterization of flower shaped zinc oxide nanostructures. *Materials Letters*, 63(2): 242-5.
- [19] Padmavathy, N. and Vijayaraghavan, R. (2008): Enhanced bioactivity of ZnO nanoparticles—an antimicrobial study. *Science and technology of advanced materials*.

- [20] Mishra, P.K.; Mishra, H.; Ekielski, A.; Talegaonkar, S. and Vaidya, B. (2017): Zinc oxide nanoparticles: a promising nanomaterial for biomedical applications. *Drug discovery today*, 22(12): 1825-34.
- [21] Xiong, H.M. (2013): ZnO nanoparticles applied to bioimaging and drug delivery. *Advanced Materials*, 25(37): 5329-35.
- [22] Zhang, Z.-Y. and Xiong, H.-M. (2015): Photoluminescent ZnO nanoparticles and their biological applications. *Materials*, 8(6): 3101-27.
- [23] Sruthi, S.; Millot, N. and Mohanan, P. (2017): Zinc oxide nanoparticles mediated cytotoxicity, mitochondrial membrane potential and level of antioxidants in presence of melatonin. *International journal of biological macromolecules*, 103: 808-18.
- [24] Vijayakumar, S.; Vaseeharan, B.; Malaikozhundan, B. and Shobiya, M. (2016): *Laurus nobilis* leaf extract mediated green synthesis of ZnO nanoparticles: characterization and biomedical applications. *Biomedicine & Pharmacotherapy*, 84: 1213-22.
- [25] Condello, M.; De Berardis, B.; Ammendolia, M.G.; Barone, F.; Condello, G.; Degan, P. and Meschini, S. (2016): ZnO nanoparticle tracking from uptake to genotoxic damage in human colon carcinoma cells. *Toxicology in Vitro*, 35: 169-79.
- [26] Hanley, C.; Layne, J.; Punnoose, A.; Reddy, K.; Coombs, I.; Coombs, A.; Feris, K. and Wingett, D. (2008): Preferential killing of cancer cells and activated human T cells using ZnO nanoparticles. *Nanotechnology*, 19(29): 295103.
- [27] Aksoy, B.; Atakan, N.; Aksoy, H.M.; Tezel, G.G.; Renda, N.; Özkara, H.A. and Önder, E. (2010): Effectiveness of topical zinc oxide application on hypertrophic scar development in rabbits. *Burns*, 36(7): 1027-35.
- [28] Shokri, N. and Javar, H. (2015): Comparison of calcium phosphate and zinc oxide nanoparticles as dermal penetration enhancers for albumin. *Indian journal of pharmaceutical sciences*, 77(6): 694.
- [29] Sharma, H.; Kumar, K.; Choudhary, C.; Mishra, P.K. and Vaidya, B. (2016): Development and characterization of metal oxide nanoparticles for the delivery of anticancer drug. *Artificial cells, nanomedicine, and biotechnology*, 44(2): 672-9.
- [30] Akhtar, M.J.; Ahamed, M.; Kumar, S.; Khan, M.M.; Ahmad, J. and Alrokayan, S.A. (2012): Zinc oxide nanoparticles selectively induce apoptosis in human cancer cells through reactive oxygen species. *International journal of nanomedicine*, 7: 845.
- [31] Taccola, L.; Raffa, V.; Riggio, C.; Vittorio, O.; Iorio, M.C.; Vanacore, R.; Pietrabissa, A. and Cuschieri, A. (2011): Zinc oxide nanoparticles as selective killers of proliferating cells. *International journal of nanomedicine*, 6: 1129.
- [32] Premanathan, M.; Karthikeyan, K.; Jeyasubramanian, K. and Manivannan, G. (2011): Selective toxicity of ZnO nanoparticles toward Gram-positive bacteria and cancer cells by apoptosis through lipid peroxidation. *Nanomedicine: Nanotechnology, Biology and Medicine*, 7(2): 184-92.
- [33] Reddy, K.M.; Feris, K.; Bell, J.; Wingett, D.G.; Hanley, C. and Punnoose, A. (2007): Selective toxicity of zinc oxide nanoparticles to prokaryotic and eukaryotic systems. *Applied physics letters*, 90(21): 213902.
- [34] Ivask, A.; Titma, T.; Visnapuu, M.; Vija, H.; Kakinen, A.; Sihtmae, M.; Pokhrel, S.; Madler, L.; Heinlaan, M. and Kisand, V. (2015): Toxicity of 11 metal oxide nanoparticles to three mammalian cell types in vitro. *Current topics in medicinal chemistry*, 15(18): 1914-29.
- [35] Gojova, A.; Guo, B.; Kota, R.S.; Rutledge, J.C.; Kennedy, I.M. and Barakat, A.I. (2007): Induction of inflammation in vascular endothelial cells by metal oxide nanoparticles: effect

- of particle composition. Environmental health perspectives, 115(3): 403-9.
- [36] Martínez-Carmona, M.; Gun'Ko, Y. and Vallet-Regí, M. (2018): ZnO nanostructures for drug delivery and theranostic applications. *Nanomaterials*, 8(4): 268.
- [37] Zhang, Y.; R Nayak, T.; Hong, H. and Cai, W. (2013): Biomedical applications of zinc oxide nanomaterials. *Current molecular medicine*, 13(10): 1633-45.
- [38] Qu, F. and Morais, P. (2001): The pH dependence of the surface charge density in oxide-based semiconductor nanoparticles immersed in aqueous solution. *IEEE transactions on magnetics*, 37(4): 2654-6.
- [39] Degen, A. and Kosec, M. (2000): Effect of pH and impurities on the surface charge of zinc oxide in aqueous solution. *Journal of the European Ceramic Society*, 20(6): 667-73.
- [40] Abercrombie, M. and Ambrose, E. (1962): The surface properties of cancer cells: a review. *Cancer research*, 22(5 Part 1): 525-48.
- [41] Vallabhapurapu, S.D.; Blanco, V.M.; Sulaiman, M.K.; Vallabhapurapu, S.L.; Chu, Z.; Franco, R.S. and Qi, X. (2015): Variation in human cancer cell external phosphatidylserine is regulated by flippase activity and intracellular calcium. *Oncotarget*, 6(33): 34375.
- [42] Hackenberg, S.; Scherzed, A.; Gohla, A.; Technau, A.; Froelich, K.; Ginzkey, C.; Koehler, C.; Burghartz, M.; Hagen, R. and Kleinsasser, N. (2014): Nanoparticle-induced photocatalytic head and neck squamous cell carcinoma cell death is associated with autophagy. *Nanomedicine*, 9(1): 21-33.
- [43] Yu, K.-N.; Yoon, T.-J.; Minai-Tehrani, A.; Kim, J.-E.; Park, S.J.; Jeong, M.S.; Ha, S.-W.; Lee, J.-K.; Kim, J.S. and Cho, M.-H. (2013): Zinc oxide nanoparticle induced autophagic cell death and mitochondrial damage via reactive oxygen species generation. *Toxicology in Vitro*, 27(4): 1187-95.
- [44] Erathodiyil, N. and Ying, J.Y. (2011): Functionalization of inorganic nanoparticles for bioimaging applications. *Accounts of chemical research*, 44(10): 925-35.
- [45] Ghaffari, S.-B.; Sarrafzadeh, M.-H.; Fakhroueian, Z.; Shahriari, S. and Khorramizadeh, M.R. (2017): Functionalization of ZnO nanoparticles by 3-mercaptopropionic acid for aqueous curcumin delivery: Synthesis, characterization, and anticancer assessment. *Materials Science and Engineering: C*, 79: 465-72.
- [46] Li, Y.; Zhang, C.; Liu, L.; Gong, Y.; Xie, Y. and Cao, Y. (2018): The effects of baicalein or baicalin on the colloidal stability of ZnO nanoparticles (NPs) and toxicity of NPs to Caco-2 cells. *toxicology Mechanisms and Methods*, 28(3): 167-76.
- [47] Puvvada, N.; Rajput, S.; Kumar, B.; Sarkar, S.; Konar, S.; Brunt, K.R.; Rao, R.R.; Mazumdar, A.; Das, S.K. and Basu, R. (2015): Novel ZnO hollow-nanocarriers containing paclitaxel targeting folate-receptors in a malignant pH-microenvironment for effective monitoring and promoting breast tumor regression. *Scientific reports*, 5(1): 1-15.
- [48] Wang, J.; Lee, J.S.; Kim, D. and Zhu, L. (2017): Exploration of zinc oxide nanoparticles as a multitarget and multifunctional anticancer nanomedicine. *ACS applied materials & interfaces*, 9(46): 39971-84.
- [49] Zhang, J.; Qin, X.; Wang, B.; Xu, G.; Qin, Z.; Wang, J.; Wu, L.; Ju, X.; Bose, D.D. and Qiu, F. (2017): Zinc oxide nanoparticles harness autophagy to induce cell death in lung epithelial cells. *Cell death & disease*, 8(7): e2954-e.
- [50] Kamaly, N.; Xiao, Z.; Valencia, P.M.; Radovic-Moreno, A.F. and Farokhzad, O.C. (2012): Targeted polymeric therapeutic nanoparticles: design, development and clinical translation. *Chemical Society Reviews*, 41(7): 2971-3010.

- [51] Han, Z.; Wang, X.; Heng, C.; Han, Q.; Cai, S.; Li, J.; Qi, C.; Liang, W.; Yang, R. and Wang, C. (2015): Synergistically enhanced photocatalytic and chemotherapeutic effects of aptamer-functionalized ZnO nanoparticles towards cancer cells. *Physical Chemistry Chemical Physics*, 17(33): 21576-82.
- [52] Kc, B.; Paudel, S.N.; Rayamajhi, S.; Karna, D.; Adhikari, S.; Shrestha, B.G. and Bisht, G. (2016): Enhanced preferential cytotoxicity through surface modification: synthesis, characterization and comparative in vitro evaluation of TritonX-100 modified and unmodified zinc oxide nanoparticles in human breast cancer cell (MDA-MB-231). *Chemistry Central Journal*, 10(1): 1-10.
- [53] Ma, Y.-Y.; Ding, H. and Xiong, H.-M. (2015): Folic acid functionalized ZnO quantum dots for targeted cancer cell imaging. *Nanotechnology*, 26(30): 305702.
- [54] Namvar, F.; Azizi, S.; Rahman, H.S.; Mohamad, R.; Rasedee, A.; Soltani, M. and Rahim, R.A. (2016): Green synthesis, characterization, and anticancer activity of hyaluronan/zinc oxide nanocomposite. *OncoTargets and therapy*, 9: 4549.
- [55] Othman, B.A.; Greenwood, C.; Abuelela, A.F.; Bharath, A.A.; Chen, S.; Theodorou, I.; Douglas, T.; Uchida, M.; Ryan, M. and Merzaban, J.S. (2016): Correlative light-electron microscopy shows RGD-targeted ZnO nanoparticles dissolve in the intracellular environment of triple negative breast cancer cells and cause apoptosis with intratumor heterogeneity. *Advanced Healthcare Materials*, 5(11): 1310-25.
- [56] Huang, Z.; Zheng, X.; Yan, D.; Yin, G.; Liao, X.; Kang, Y.; Yao, Y.; Huang, D. and Hao, B. (2008): Toxicological effect of ZnO nanoparticles based on bacteria. *Langmuir*, 24(8): 4140-4.
- [57] Limbach, L.K.; Wick, P.; Manser, P.; Grass, R.N.; Bruinink, A. and Stark, W.J. (2007): Exposure of engineered nanoparticles to human lung epithelial cells: influence of chemical composition and catalytic activity on oxidative stress. *Environmental science & technology*, 41(11): 4158-63.
- [58] Sangeetha, G.; Rajeshwari, S. and Venkatesh, R. (2011): Green synthesis of zinc oxide nanoparticles by aloe barbadensis miller leaf extract: Structure and optical properties. *Materials Research Bulletin*, 46(12): 2560-6.
- [59] Dutta, R.K.; Nenavathu, B.P.; Gangishetty, M.K. and Reddy, A. (2013): Antibacterial effect of chronic exposure of low concentration ZnO nanoparticles on E. coli. *Journal of Environmental Science and Health, Part A*, 48(8): 871-8.
- [60] Chatterjee, T.; Chakraborti, S.; Joshi, P.; Singh, S.P.; Gupta, V. and Chakrabarti, P. (2010): The effect of zinc oxide nanoparticles on the structure of the periplasmic domain of the Vibrio cholerae ToxR protein. *The FEBS journal*, 277(20): 4184-94.
- [61] Mahendra, C.; Murali, M.; Manasa, G.; Ponnamma, P.; Abhilash, M.; Lakshmeesha, T.; Satish, A.; Amruthesh, K. and Sudarshana, M. (2017): Antibacterial and antimitotic potential of bio-fabricated zinc oxide nanoparticles of Cochlospermum religiosum (L.). *Microbial pathogenesis*, 110: 620-9.
- [62] Singh, B.N.; Rawat, A.K.S.; Khan, W.; Naqvi, A.H. and Singh, B.R. (2014): Biosynthesis of stable antioxidant ZnO nanoparticles by Pseudomonas aeruginosa rhamnolipids. *PLoS One*, 9(9): e106937.
- [63] Ishwarya, R.; Vaseeharan, B.; Kalyani, S.; Banumathi, B.; Govindarajan, M.; Alharbi, N.S.; Kadaikunnan, S.; Al-Anbr, M.N.; Khaled, J.M. and Benelli, G. (2018): Facile green synthesis of zinc oxide nanoparticles using Ulva lactuca seaweed extract and evaluation of their photocatalytic, antibiofilm and insecticidal activity. *Journal of Photochemistry and Photobiology B: Biology*, 178: 249-58.

- [64] Divya, M.; Vaseeharan, B.; Abinaya, M.; Vijayakumar, S.; Govindarajan, M.; Alharbi, N.S.; Kadaikunnan, S.; Khaled, J.M. and Benelli, G. (2018): Biopolymer gelatin-coated zinc oxide nanoparticles showed high antibacterial, antibiofilm and anti-angiogenic activity. *Journal of Photochemistry and Photobiology B: Biology*, 178: 211-8.
- [65] Hsueh, Y.-H.; Ke, W.-J.; Hsieh, C.-T.; Lin, K.-S.; Tzou, D.-Y. and Chiang, C.-L. (2015): ZnO nanoparticles affect *Bacillus subtilis* cell growth and biofilm formation. *PloS one*, 10(6): e0128457.
- [66] Jiang, Y.; Zhang, L.; Wen, D. and Ding, Y. (2016): Role of physical and chemical interactions in the antibacterial behavior of ZnO nanoparticles against *E. coli*. *Materials Science and Engineering: C*, 69: 1361-6.
- [67] Salem, W.; Leitner, D.R.; Zingl, F.G.; Schratter, G.; Prassl, R.; Goessler, W.; Reidl, J. and Schild, S. (2015): Antibacterial activity of silver and zinc nanoparticles against *Vibrio cholerae* and enterotoxigenic *Escherichia coli*. *International Journal of Medical Microbiology*, 305(1): 85-95.
- [68] Sarwar, S.; Chakraborti, S.; Bera, S.; Sheikh, I.A.; Hoque, K.M. and Chakrabarti, P. (2016): The antimicrobial activity of ZnO nanoparticles against *Vibrio cholerae*: Variation in response depends on biotype. *Nanomedicine: Nanotechnology, Biology and Medicine*, 12(6): 1499-509.
- [69] Sarwar, S.; Ali, A.; Pal, M. and Chakrabarti, P. (2017): Zinc oxide nanoparticles provide anti-cholera activity by disrupting the interaction of cholera toxin with the human GM1 receptor. *Journal of Biological Chemistry*, 292(44): 18303-11.
- [70] Dwivedi, S.; Wahab, R.; Khan, F.; Mishra, Y.K.; Musarrat, J. and Al-Khedhairi, A.A. (2014): Reactive oxygen species mediated bacterial biofilm inhibition via zinc oxide nanoparticles and their statistical determination. *PloS one*, 9(11): e111289.
- [71] Ghasemi, F. and Jalal, R. (2016): Antimicrobial action of zinc oxide nanoparticles in combination with ciprofloxacin and ceftazidime against multidrug-resistant *Acinetobacter baumannii*. *Journal of global antimicrobial resistance*, 6: 118-22.
- [72] Sehmi, S.K.; Noimark, S.; Bear, J.C.; Peveler, W.J.; Bovis, M.; Allan, E.; MacRobert, A.J. and Parkin, I.P. (2015): Lethal photosensitisation of *Staphylococcus aureus* and *Escherichia coli* using crystal violet and zinc oxide-encapsulated polyurethane. *Journal of Materials Chemistry B*, 3(31): 6490-500.
- [73] Manzoor, U.; Siddique, S.; Ahmed, R.; Noreen, Z.; Bokhari, H. and Ahmad, I. (2016): Antibacterial, structural and optical characterization of mechano-chemically prepared ZnO nanoparticles. *PLoS One*, 11(5): e0154704.
- [74] Zhang, B.; Cui, L. and Zhang, K. (2016): Dosage- and time-dependent antibacterial effect of zinc oxide nanoparticles determined by a highly uniform SERS negating undesired spectral variation. *Analytical and Bioanalytical Chemistry*, 408(14): 3853-65.
- [75] Makabenta, J.M.V.; Nabawy, A.; Li, C.-H.; Schmidt-Malan, S.; Patel, R. and Rotello, V.M. (2021): Nanomaterial-based therapeutics for antibiotic-resistant bacterial infections. *Nature Reviews Microbiology*, 19(1): 23-36.
- [76] Muzammil, S.; Hayat, S.; Fakhar-El-Alam, M.; Aslam, B.; Siddique, M.H.; Nisar, M.A.; Saqalein, M.; Atif, M.; Sarwar, A. and Khurshid, A. (2018): Nanoantibiotics: Future nanotechnologies to combat antibiotic resistance. *Frontiers in Bioscience-Elite*, 10(2): 352-74.
- [77] da Silva, B.L.; Abuçafy, M.P.; Manaia, E.B.; Junior, J.A.O.; Chiari-Andréo, B.G.; Pietro, R.C.R. and Chiavacci, L.A. (2019): Relationship between structure and antimicrobial activity of

- zinc oxide nanoparticles: An overview. *International journal of nanomedicine*, 14: 9395.
- [78] Dizaj, S.M.; Lotfipour, F.; Barzegar-Jalali, M.; Zarrintan, M.H. and Adibkia, K. (2014): Antimicrobial activity of the metals and metal oxide nanoparticles. *Materials Science and Engineering: C*, 44: 278-84.
- [79] Jin, S.-E. and Jin, H.-E. (2019): Synthesis, characterization, and three-dimensional structure generation of zinc oxide-based nanomedicine for biomedical applications. *Pharmaceutics*, 11(11): 575.
- [80] Naveed Ul Haq, A.; Nadhman, A.; Ullah, I.; Mustafa, G.; Yasinzai, M. and Khan, I. (2017): Synthesis approaches of zinc oxide nanoparticles: the dilemma of ecotoxicity. *Journal of Nanomaterials*, 2017.
- [81] Sánchez-López, E.; Gomes, D.; Esteruelas, G.; Bonilla, L.; Lopez-Machado, A.L.; Galindo, R.; Cano, A.; Espina, M.; Ettcheto, M. and Camins, A. (2020): Metal-based nanoparticles as antimicrobial agents: an overview. *Nanomaterials*, 10(2): 292.
- [82] Van Giau, V.; An, S.S.A. and Hulme, J. (2019): Recent advances in the treatment of pathogenic infections using antibiotics and nano-drug delivery vehicles. *Drug design, development and therapy*, 13: 327.
- [83] Ramamurthy, C.; Sampath, K.; Arunkumar, P.; Kumar, M.S.; Sujatha, V.; Premkumar, K. and Thirunavukkarasu, C. (2013): Green synthesis and characterization of selenium nanoparticles and its augmented cytotoxicity with doxorubicin on cancer cells. *Bioprocess and biosystems engineering*, 36(8): 1131-9.
- [84] Wang, X.; Chen, H.; Zheng, Y.; Ma, M.; Chen, Y.; Zhang, K.; Zeng, D. and Shi, J. (2013): Au-nanoparticle coated mesoporous silica nanocapsule-based multifunctional platform for ultrasound mediated imaging, cytolysis and tumor ablation. *Biomaterials*, 34(8): 2057-68.
- [85] Zhang, H.; Shan, Y. and Dong, L. (2014): A comparison of TiO₂ and ZnO nanoparticles as photosensitizers in photodynamic therapy for cancer. *Journal of biomedical nanotechnology*, 10(8): 1450-7.
- [86] Espitia, P.J.P.; Soares, N.d.F.F.; Coimbra, J.S.d.R.; de Andrade, N.J.; Cruz, R.S. and Medeiros, E.A.A. (2012): Zinc oxide nanoparticles: synthesis, antimicrobial activity and food packaging applications. *Food and bioprocess technology*, 5(5): 1447-64.
- [87] Sirelkhatim, A.; Mahmud, S.; Seeni, A.; Kaus, N.H.M.; Ann, L.C.; Bakhori, S.K.M.; Hasan, H. and Mohamad, D. (2015): Review on zinc oxide nanoparticles: antibacterial activity and toxicity mechanism. *Nano-micro letters*, 7(3): 219-42.
- [88] Cheeseman, S.; Christofferson, A.J.; Kariuki, R.; Cozzolino, D.; Daeneke, T.; Crawford, R.J.; Truong, V.K.; Chapman, J. and Elbourne, A. (2020): Antimicrobial metal nanomaterials: from passive to stimuli-activated applications. *Advanced Science*, 7(10): 1902913.
- [89] Lemire, J.A.; Harrison, J.J. and Turner, R.J. (2013): Antimicrobial activity of metals: mechanisms, molecular targets and applications. *Nature Reviews Microbiology*, 11(6): 371-84.
- [90] Ali, A.; Phull, A.-R. and Zia, M. (2018): Elemental zinc to zinc nanoparticles: Is ZnO NPs crucial for life? Synthesis, toxicological, and environmental concerns. *Nanotechnology Reviews*, 7(5): 413-41.
- [91] Pasquet, J.; Chevalier, Y.; Pelletier, J.; Couval, E.; Bouvier, D. and Bolzinger, M.-A. (2014): The contribution of zinc ions to the antimicrobial activity of zinc oxide. *Colloids and Surfaces A: Physicochemical and Engineering Aspects*, 457: 263-74.
- [92] Adhikari, S.; Gupta, R.; Surin, A.; Kumar, T.S.; Chakraborty, S.; Sarkar, D. and Madras, G. (2016): Visible light assisted improved photocatalytic activity of combustion synthesized spongy-ZnO

- towards dye degradation and bacterial inactivation. RSC Advances, 6(83): 80086-98.
- [93] Kaliraj, L.; Ahn, J.C.; Rupa, E.J.; Abid, S.; Lu, J. and Yang, D.C. (2019): Synthesis of panos extract mediated ZnO nano-flowers as photocatalyst for industrial dye degradation by UV illumination. Journal of Photochemistry and Photobiology B: Biology, 199: 111588.
- [94] Pelgrift, R.Y. and Friedman, A.J. (2013): Nanotechnology as a therapeutic tool to combat microbial resistance. Advanced drug delivery reviews, 65(13-14): 1803-15.
- [95] Hajipour, M.J.; Fromm, K.M.; Ashkarran, A.A.; de Aberasturi, D.J.; de Larramendi, I.R.; Rojo, T.; Serpooshan, V.; Parak, W.J. and Mahmoudi, M. (2012): Antibacterial properties of nanoparticles. Trends in biotechnology, 30(10): 499-511.
- [96] Jin, S.-E.; Jin, J.E.; Hwang, W. and Hong, S.W. (2019): Photocatalytic antibacterial application of zinc oxide nanoparticles and self-assembled networks under dual UV irradiation for enhanced disinfection. International journal of nanomedicine, 14: 1737.
- [97] Sharma, S.; Kumar, K.; Thakur, N.; Chauhan, S. and Chauhan, M. (2020): The effect of shape and size of ZnO nanoparticles on their antimicrobial and photocatalytic activities: a green approach. Bulletin of Materials Science, 43(1): 1-10.
- [98] Jin, S.-E. and Jin, H.-E. (2021): Antimicrobial activity of zinc oxide nano/microparticles and their combinations against pathogenic microorganisms for biomedical applications: From physicochemical characteristics to pharmacological aspects. Nanomaterials, 11(2): 263.
- [99] Nazarizadeh, A. and Asri-Rezaie, S. (2016): Comparative study of antidiabetic activity and oxidative stress induced by zinc oxide nanoparticles and zinc sulfate in diabetic rats. AAPS PharmSciTech, 17(4): 834-43.
- [100] Wahba, N.S.; Shaban, S.F.; Kattaia, A.A. and Kandeel, S.A. (2016): Efficacy of zinc oxide nanoparticles in attenuating pancreatic damage in a rat model of streptozotocin-induced diabetes. Ultrastructural pathology, 40(6): 358-73.
- [101] El-Gharbawy, R.M.; Emara, A.M. and Abu-Risha, S.E.-S. (2016): Zinc oxide nanoparticles and a standard antidiabetic drug restore the function and structure of beta cells in Type-2 diabetes. Biomedicine & Pharmacotherapy, 84: 810-20.
- [102] Kitture, R.; Chordiya, K.; Gaware, S.; Ghosh, S.; More, P.A.; Kulkarni, P.; Chopade, B.A. and Kale, S. (2015): ZnO nanoparticles-red sandalwood conjugate: a promising anti-diabetic agent. Journal of nanoscience and nanotechnology, 15(6): 4046-51.
- [103] Balan, K.; Qing, W.; Wang, Y.; Liu, X.; Palvannan, T.; Wang, Y.; Ma, F. and Zhang, Y. (2016): Antidiabetic activity of silver nanoparticles from green synthesis using Lonicera japonica leaf extract. Rsc Advances, 6(46): 40162-8.
- [104] Rehana, D.; Mahendiran, D.; Kumar, R.S. and Rahiman, A.K. (2017): In vitro antioxidant and antidiabetic activities of zinc oxide nanoparticles synthesized using different plant extracts. Bioprocess and biosystems engineering, 40(6): 943-57.
- [105] Kim, K.-T.; Rioux, L.-E. and Turgeon, S.L. (2014): Alpha-amylase and alpha-glucosidase inhibition is differentially modulated by fucoidan obtained from Fucus vesiculosus and Ascophyllum nodosum. Phytochemistry, 98: 27-33.
- [106] Vinotha, V.; Iswarya, A.; Thaya, R.; Govindarajan, M.; Alharbi, N.S.; Kadaikunnan, S.; Khaled, J.M.; Al-Anbr, M.N. and Vaseeharan, B. (2019): Synthesis of ZnO nanoparticles using insulin-rich leaf extract: Anti-diabetic, antibiofilm and anti-oxidant properties.

- Journal of Photochemistry and Photobiology B: Biology, 197: 111541.
- [107] Surendra, T.; Roopan, S.M.; Al-Dhabi, N.A.; Arasu, M.V.; Sarkar, G. and Suthindhiran, K. (2016): Vegetable peel waste for the production of ZnO nanoparticles and its toxicological efficiency, antifungal, hemolytic, and antibacterial activities. *Nanoscale research letters*, 11(1): 1-10.
- [108] Gunalan, S.; Sivaraj, R. and Rajendran, V. (2012): Green synthesized ZnO nanoparticles against bacterial and fungal pathogens. *Progress in Natural Science: Materials International*, 22(6): 693-700.
- [109] Lipovsky, A.; Nitzan, Y.; Gedanken, A. and Lubart, R. (2011): Antifungal activity of ZnO nanoparticles—the role of ROS mediated cell injury. *Nanotechnology*, 22(10): 105101.
- [110] Ilves, M.; Palomäki, J.; Vippola, M.; Lehto, M.; Savolainen, K.; Savinko, T. and Alenius, H. (2014): Topically applied ZnO nanoparticles suppress allergen induced skin inflammation but induce vigorous IgE production in the atopic dermatitis mouse model. *Particle and fibre toxicology*, 11(1): 1-12.
- [111] Hughes, G. and McLean, N. (1988): Zinc oxide tape: a useful dressing for the recalcitrant finger-tip and soft-tissue injury. *Emergency Medicine Journal*, 5(4): 223-7.
- [112] Mitchnick, M.A.; Fairhurst, D. and Pinnell, S.R. (1999): Microfine zinc oxide (Z-cote) as a photostable UVA/UVB sunblock agent. *Journal of the American Academy of Dermatology*, 40(1): 85-90.
- [113] Yuan, Q.; Hein, S. and Misra, R. (2010): New generation of chitosan-encapsulated ZnO quantum dots loaded with drug: synthesis, characterization and in vitro drug delivery response. *Acta biomaterialia*, 6(7): 2732-9.
- [114] Kahru, A. and Mortimer, M. *Advances in Nanotoxicology: Towards Enhanced Environmental and Physiological Relevance and Molecular Mechanisms*. MDPI; 2021. p. 919.
- [115] Aravantinou, A.F.; Andreou, F. and Manariotis, I.D. (2020): Long-term toxicity of ZnO nanoparticles on *Scenedesmus rubescens* cultivated in semi-batch mode. *Nanomaterials*, 10(11): 2262.

الملخص العربي

التطبيقات الطبية الحيوية الواسعة لجسيمات أكسيد الزنك النانوية

- اسماعيل غريب عبدالمقصود^{1,2}، حمد أمين السعداوي²، أماني أحمد إبراهيم²، عادل عبدالخالق³، أحمد حامد عريشة^{1,4}
- 1 – قسم فسيولوجيا الحيوان والكيمياء الحيوية، كلية الطب البيطري، جامعة بدر بالقاهرة، مدينة بدر، القاهرة، مصر.
- 2- قسم الكيمياء الحيوية، كلية الطب البيطري، جامعة الزقازيق، 44511، الزقازيق، مصر.
- 3- كلية الطب البيطري، جامعة بدر بالقاهرة، مدينة بدر، القاهرة، مصر.
- 4- قسم علم وظائف الأعضاء، كلية الطب البيطري، جامعة الزقازيق، 44511 الزقازيق، مصر.

اكتسبت المواد النانوية الطبية الحيوية الكثير من الاهتمام مؤخرًا. لقد أكدوا على مخاوف مختلفة بسبب خصائصهم البيولوجية الواسعة والهامة والتطبيقات الطبية الحيوية. تقدم الجسيمات النانوية لأكسيد المعادن مجموعة واسعة من الاستخدامات الطبية، بما في ذلك مضادات السرطان، وتوصيل الأدوية / الجينات، ومضادات الجراثيم، وتصوير الخلايا، والاستشعار الحيوي، من بين أمور أخرى. الزنك عنصر طبيعي يلعب دورًا مهمًا في عملية التمثيل الغذائي للإنسان والحيوان والنبات. يُعرف الزنك على نطاق واسع بأنه عنصر أساسي يمكن العثور عليه في جميع أنسجة الجسم، بما في ذلك العظام والدماغ والعضلات والجلد. يعد الزنك مكونًا أساسيًا للعديد من أنظمة الإنزيمات ويلعب دورًا مهمًا في إنتاج البروتين والحمض النووي، وتكوين الدم، وتكوين الخلايا العصبية. تم استخدام جزيئات أكسيد الزنك النانوية (ZnO-NPs) كمادة رئيسية في مجموعة من الصناعات في السنوات الأخيرة، بما في ذلك الأدوية ومستحضرات التجميل والخرسانة ومضادات الميكروبات والمنسوجات وصناعة السيارات والوقاية من السرطان. ترتبط التأثيرات المضادة للسرطان والبكتيريا بقدرة Zn-ONPs على توليد أنواع الأكسجين التفاعلية (ROS) وتحفيز الموت المبرمج للخلايا (موت الخلايا المبرمج). تلخص هذه المقالة المرجعية التطبيقات الطبية الحيوية المختلفة للجسيمات النانوية ZnO.