Biochemical Effect Of L-Carnitine Against Doxorubicin And Vancomycin Induced Lipid Disorders In Rats

Rehab Zaid¹, Youssef Shehata², Mohamed Dowidar², Haytham Abdallah Ali³, Aaser Mohamed Abdelazim³

Department of Biochemistry, Faculty of Vet. Medicine, Zagazig University, Zagazig, Egypt.

ABSTRACT

The ameliorative effect of L-carnitine on the lipid, liver and kidneys disorders induced by doxorubicin and vancomycin was examined. Ninety male albino rats were randomly divided into six groups (15) each; control, L-carnitine treated, Doxorubicin treated, vancomycin treated, doxorubicin + L-carnitine, Vancomycin + L-carnitine. Animals were sacrificed after 3, 10, 15 days. Blood and tissue samples were collected. Leptin, insulin, MDA, HDL, TC, TP and albumin levels, LDH and GGT activities were determined. mRNA expression levels of leptin, leptin receptors and GAPDH genes were determined in adipose tissues. There was a significant increase of serum insulin, leptin, MDA, HDL, TC levels, GGT and LDH activities and significant decrease of Albumin, TP levels in the groups treated with doxorubicin and Vancomycin. In conclusion; DOX and VAN had a bad effect on the lipid profile as they induced obesity, also; they increase liver enzymes and heart markers and increase the oxidative stress in the body tissues due their ability to increase the level of MDA. L-carnitine administration ameliorated the hazards effect of DOX and VAN on the lipid sate in the body.

INTRODUCTION

Leptin, a product of ob gene, is a hormone secreted mainly by adipocytes; it acts mainly as the most important regulator of energy balance (1). It has a role in regulating food intake and energy balance, affects thyroid and growth hormones, as hematopoiesis, bone formation and immune system (2). Leptin deficiency is characterized by hyperlipidemia, excessive storage of lipid in tissues such as liver and skeletal muscle, and insulin resistance. These defects are markedly improved by the administration of leptin in humans and rodents (3). Leptin receptors are found in many areas of the brain, including the hypothalamus. cerebellum. hippocampus, thalamus and choroid plexus (4); it is also expressed in peripheral tissues, like kidneys, liver, pancreas, adrenals, ovaries, hematopoietic stem cells, and skeletal muscle (5), This wide expression may imply the great role of leptin (6).

Doxorubicin (DOX) the anthracycline antibiotic obtained from *Streptomyces peucetius* has been used against breast and esophageal carcinomas, osteosarcoma, Kaposi's sarcoma, soft tissue sarcomas (7). The DOX-induced hyperlipidemia resulted from reduced lipid storage and utilization and reduction of mitochondrial oxidation of long chain fatty acids in kidney and heart (8).

Vancomycin is a glycopeptides antibiotic that has been used clinically for nearly 50 years as a penicillin alternative to treat methicillin resistant S. aureus (MRSA) infections. One of the major adverse effects of vancomycin is nephrotoxicity which has limited administration. Vancomycin mono therapy may cause nephrotoxicity at an incidence of 5 to 10%; while in combination aminoglycosides has increased the incidence of nephrotoxicity to 14 and as high as 35% (9). Records demonstrated a significant reduction in

serum leptin level in albino rats received vancomycin (10,11).

L-Carnitine is biosynthesized primarily in the liver and kidneys from the amino acids lysine (via trimethyl lysine) or methionine (12). The requirement of L-carnitine might exceed its natural production in certain circumstances like growth and pregnancy (13). There are many studies that reported the protective effect of Lcarnitine against doxorubicin (14,15).Doxorubicin (DOX) may cause adenosine 5'triphosphate (ATP) depletion by inhibition of carnitine palmitoyl transferase both mitochondrial outer and inner membrane (16). Administering L-carnitine can facilitate the transport of long chain fatty acids into the mitochondria and promotes fatty acid oxidation (17). In rats, administration of L-carnitine prevented DOX-induced histological metabolic damage (18).

This study was designed to investigate the protective effect of L-carnitine against DOX and vancomycine induced adverse effects on lipid metabolism.

MATERIALS AND METHODS

Drugs

Adricin (Doxorubicin HCl) from EIMC United pharmaceuticals, Badr city-Cairo-A.R.E. Each vial (5ml) contains Doxorubicin HCl 10 mg.

Rat were injected intraperitoneal (I/P) dose with adriamycin, 2.5 mg/kg B.wt. once/week for 4 successive weeks (19) and examined 3, 10 and 15 days post administration.

Vancomix from Sigmatec pharmaceutical industries, Egypt. One gm Vial contains Vancomycin HCL chromatographically purified equivalent to 1 gm vancomycin activity). Rat were injected I/P daily with vancomycin (1 gm/kg B.wt/every 12 hr for 7-

10 successive days. The dose per rat was calculated according to the surface area (20).

L-Carnitine from MEPACO Medifood Co. Enshas- Sharkeya. Egypt. Each 5 ml ampoule contains 1 gm L-Carnitine. Molecular Rat were injected I/P daily with L-carnitin (200 mg/kg B.wt) daily for 2 successive weeks (21).

Experimental animals: Ninety mature male albino rats (weight mg 120 ± 20 gm, 6 month old) were used. They were obtained from the Laboratory Animal unit, Faculty of Veterinary Medicine, Zagazig University. The animals were clinically healthy, kept under hygienic conditions, housed in metal cages with hard wood shavings as bedding. They were maintained on balanced ration composed of barley, milk, green fodder and water *ad-libitum*. The animals were accommodated at the laboratory condition for two weeks before being experimented.

Experimental design: The rats were acclimatized for 2 weeks and randomly divided into six groups of 15 rats at each. First group served as control and didn't receive any treatment, Group II treated with L-carnitine, Group III treated with Doxorubicin, Group IV treated with Vancomix. Group V treated DOX L-carnitine. Group VI treated with Vancomix and L-carnitine. The rats were kept for 3, 10 and 15 days after last treatments of each group then were sacrificed and samples were collected.

Sampling: Blood samples were collected into two clean, dry, sterile, and labeled centrifuge tubes, the first tube contained sodium fluoride for determination of blood glucose whereas, the second tube contained no anticoagulants where blood used for separation of serum for determination of total proteins, albumin, total cholesterol, HDLc, GGT, MDA, LDH and leptin.

Tissue sampling: Adipose tissue was immediately removed from each animal after sacrificing and they are washed out from contaminated blood with normal saline weighted and immediately kept in liquid nitrogen until be used for determination of gene expression of *Ob* gene.

Statistical analysis: The obtained data were analyzed and graphically represented using the statistical package for social science (SPSS, 18.0 software, 2011) for obtaining means and standard error. Duncan's test was used for making a multiple comparisons among the groups for testing the inter-grouping homogeneity (22).

RESULTS

Effect of different treatments on body gain

Body weight was significantly increased in the Vancomycin and doxorubicin treated groups while there was none significant changed in L-carnitine treated groups compared with the control.

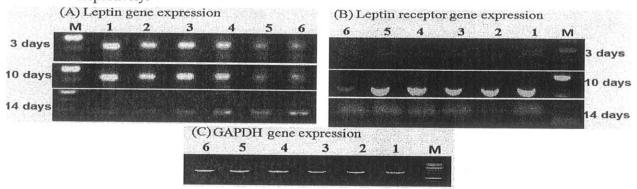
Table 1. Weight gain in albino rats after 3, 10 and 15 day post treatment

Group	Weight gain (grams)			
Отопр	3 day	10 day	15 day	
Control	128.00 ± 3.14^{D}	128.00 ± 3.14 D	128.00± 3.14 ^D	
L- Carnitine	126.75 ± 5.15^{D}	$125.75 \pm 5.16^{\mathrm{D}}$	125.15± 5.13 D	
DOX	$172.50 \pm 4.78^{\text{ B}}$	199.25± 11.37 B	197.50± 15.92 B	
Vancomix	206.25± 9.87 ^A	215.00± 8.42 A	217.50± 9.24 A	
DOX + L	149.75b± 11.52 ^C	154.00± 11.19 ^C	155.00± 11.75 ^C	
Vancomix + L	167.50± 8.54 B	203.50± 10.44 B	204.75 ± 10.37 B	

Means in the same raw that carry different subscripts are significant at P<0.05

Molecular Biological determinations

Figure (1): The electophorteic photograph of mRNA expression levels in A; leptin B; leptin receptors and C; GAPDH genes in the adipose tissues of rats; the product sizes are 195, 171, 455 bps respectively.



N.B M; 100bp-1000bp DNA-ladder, 1; Male rats (control)2; Male rats treated with L-carnitine, 3; Male rats treated with doxorubicin, 4; Male rats treated with Vancomycin, 5; Male rats treated with doxorubicin and L-carnitine and 6; Male rats treated with Vancomycin and L-carnitine.

Figure 1 represents a decrease in the level of gene expression of leptin and leptin receptors in the groups treated with DOX and Vancomycin and this decrease was ameliorated by supplementation with L-carnitine. GAPDH control gene show almost stable pattern in all groups.

Biochemical findings

Our results represents that there were a significant decrease in the level of serum

leptin, insulin, total protein and albumin in the groups treated with DOX and vancomycin, this decrease was improved by treatment with L-carnitine that can ameliorates their effect. Whereas the levels of MDA, LDH, TC, HDL-c and GGT were significantly increased due to DOX and Vancomycin treatment and L-carnitine succeeded to restore this increase.

Table 2. Biochemical determinations in albino rats treated with Doxorubicin and Vancomycin after 3, 10, 15 days respectively

	Time	Leptin	Insulin	MDA	LDH	HDL	Cholesterol	T. protein	Albumin	GGT
Group	(Day)	(ng/ml)	(ng/ml)	(lm/lomu)	(U/I)	(mg/dl)	(mg/dl)	(g/dl)	(U/I)	(I/I)
	3	6.5 ± 0.25^{a}	99.6± 3.8 ^a	$8.1 \pm 1.45 a$	869.2±25°	49.2± 0.4°	121.2± 0.3°	5.4 ± 0.18^{a}	5.29 ± 0.12^{a}	1± 0.14 ^b
Control	10	6.5 ± 0.2^{a}	97.9 ± 3.7^{a}	$8.1 \pm 1.4^{\circ}$	869.2± 25.0ab	$49.2\pm0.4^{\circ}$	121.2± 0.3 ^d	5.4± 0.18 ^d	5.29± 0.12a	1±0.14 ^b
	15	6.5 ± 0.2^{a}	98.5 ± 3.6^{a}	8.1 ± 1.4^{c}	869.2± 25 ^{cd}	$49.2\pm0.4^{\circ}$	121.2± 0.3 ^d	5.4 ± 0.18^{a}	5.29 ± 0.12^{a}	1± 0.14 ^{bc}
	3	6.4 ± 0.3^{a}	99.0±7.2ª	1.6 ± 0.9^{b}	407±77 ^d	43.5 ± 0.6^{d}	90.9± 4°	5.3 ± 0.14^{a}	4.5 ± 0.25^{b}	0.7 ± 0.2^{b}
L- Carnitine	10	6.3 ± 0.2^{a}	97.1 ± 6.4^{a}	1.6 ± 0.9^{d}	407±77.7 c	43.5 ± 0.6^{f}	90.9± 4°	5.3 ± 0.14^{a}	4.5 ± 0.25^{b}	0.7 ± 0.20^{b}
	15	6.35 ± 0.2^{a}	99.0±6.5ª	1.6 ± 0.9^{d}	407±77.7°	43.5 ± 0.6^{d}	90.9± 4e	5.3 ± 0.14^{a}	4.5 ± 0.25^{b}	$0.7 \pm 0.20^{\circ}$
	3	8.2 ± 0.14^{b}	101.7 ± 8^{c}	10.8 ± 1.1^{a}	1485.5 ± 64^{a}	64.2 ± 0.38^{a}	166.4 ± 0.6^{3}	4.2 ± 0.14^{c}	2.3 ± 0.01^{d}	4.4± 2.2ab
DOX	10	8.1 ± 0.12^{b}	107.3 ± 8^{c}	29.8 ± 0.8^{a}	1057 ± 32.7^{ab}	70.2 ± 0.3^{b}	176.7 ± 0.8^{a}	4.1 ± 0.26^{b}	2 ± 0.00^{e}	8 ± 2.48^{a}
	15	8.01 ± 0.2^{b}	101 ± 8^{c}	13.9 ± 2.3^{b}	1451.5±135.1 ^a	65.8 ± 0.7^{a}	168.8 ± 1^{a}	$4.2\pm0.16^{\circ}$	$3.48\pm0.16^{\circ}$	3.2 ± 0.58^{a}
	3	5.98± 0.33 ^b	79.7 ± 4.2^{d}	9.3 ± 1.3^{a}	1345.5 ± 83^{a}	61.9 ± 0.39^{a}	130.3 ± 3.4^{b}	4.7 ± 0.23^{b}	2.7± 0.25 ^d	5.7 ± 2.2^{a}
Vancomix	10	5.9 ± 0.3^{b}	80.1 ± 4.8^{d}	16.0 ± 0.8^{b}	1111.7 ± 50.7^{a}	67.7 ± 0.7^{b}	143.9 ± 1.8^{b}	4.8 ± 0.27^{ab}	2.7 ± 0.19^{d}	7.5 ± 1.93^{a}
	15	5.9 ± 0.2^{b}	77.7± 4.9 ^d	19.7 ± 0.7^{d}	1020 ± 40^{bc}	63.6± 0.3a	$138.5\pm0.6^{\circ}$	4.4± 0.07 ^{bc}	2.8 ± 0.10^{d}	2.1 ± 0.45^{ab}
	3	$6.31\pm0.3a^{b}$	99.4±3.9°	3.2 ± 0.9^{b}	1226.5± 76 ^{ab}	50.8± 0.27°	127.9± 0.9bc	4.9 ± 0.17^{ab}	4.0± 0°	1.5 ± 0.2^{b}
DOX+L	10	6.3± 0.23a ^b	97.1±4.8°	$6.7\pm0.2^{\circ}$	886± 133.57ab	59.6± 0.5°	$134.5\pm 1.6^{\circ}$	4.8 ± 0.23^{ab}	3.5± 0.08°	2.5 ± 0.78^{b}
	15	$6.2\pm0.13a^{b}$	98.5±4.6°	5.4± 0.4 ^{cd}	1197.7±35.7 ^{ab}	52.8± 1.6 ^b	125 ± 1.3^{b}	4.8 ± 0.20^{b}	4.1 ± 0.01^{b}	1.4± 0.67bc
	3	6.18±0.27 ^{ab}	96.3 ± 2.6^{b}	2.1 ± 0.6^{b}	952± 49 ^{bc}	53.8± 1.6 ^b	101.5 ± 2.4^{d}	4.1± 0.11°	3.8± 0.22°	1.13 ± 0.4^{b}
Vancomix + L	10	$6.12 \pm 0.2ab$	96.8 ± 2.8^{b}	$6.5\pm0.63^{\circ}$	868.75±89.4 ^b	53.837±0.7 ^d	133.10± 1.9°	4.64± 0.21 ^b	3.25 ± 0.25^{c}	3.4± 0.82 ^b
	15	6.27±0.25ab 96.4± 2.5 ^b	96.4 ± 2.5^{b}	7.9± 2.7°	898±151.37 ^{de}	52.1 ± 1.13^{b}	126.15 ± 2^{d}	4.73± 0.12bc	3.24± 0.12 ^{cd}	0.6 ± 0.22^{c}
7	, L	1.00					The state of the s		The same of the sa	

Means in the same raw that carry different subscripts are significant at P<0.05.

DISCUSSION

In the present study we tended to evaluate the effect of doxorubicin and Vancomycin on the lipid state. Our results showed an increase in body weight in case of treatment with doxorubicin that may be due to leptin resistance which is related to the development of insulin resistance (23). Hyperinsulinemia and insulin resistance has been demonstrated and obesity is associated with a marked increase in circulating leptin concentration (24). The increase in the body weight till reaches obesity in case of treatment with vancomycin was in agreement with (25) who reported that Vancomycin caused reducing of leptin in circulation. In the same line the genetic defect of leptin has the ability to induce severe obese phenotypic traits (26).

Leptin is a main hormone that conveys signal containing information of energy storage in the body and it functions to suppress energy intake as a response of adequate energy requirement (27). In our study we relived the effect of some antibiotics as doxorubicin and vancomycin on leptin level with antioxidant as L-carnitine on the leptin level and their effect on body weight and the occurrence of obesity which happened after treatment of the infection as in case of treatment with vancomycin after operation which named intensive care obesity and also obesity which happened after treatment of cancer when the treatment with doxorubicin.

Our results showed that, there is an increase in leptin level after treatment with doxorubicin, which agrees with (28), and this may be due to increase synthesis of leptin mRNA and serum leptin level in obese individuals as detected in table 2 and figure 1 when compared to non-obese individuals which brings into hypothesis of leptin resistance (29).

Leptin resistance is related to the development of insulin resistance in individuals with type II diabetes (24). Our results showed an increase in insulin level in groups treated with doxorubicin. This results agreed with (30) who mentioned that insulin resistance

syndrome, have been linked to an increased risk of developing cancer.

Oxidative stress leads to tissue damage and has been linked to impairment of insulin action and β -cell function with the resultant development of type II diabetes (31,32).

Our result presented that, there were an increase level of LDH as also reported by (33,34), they attributed that to the inflammatory cells forming granulomatous lesions fibrosis periportal were detected doxorubicin administration which has been shown to induce accumulation of inflammatory cells, associated with increased activity of LDH, indicating hepatic damage. These results also was in agreement with (35) who mentioned that heart tissue injury induced by doxorubicin in rats was indicated by elevated level of the marker enzymes such as serum LDH and CPK. The increase of LDH level in serum and extracellular fluid suggests an increase leakage of this enzyme from mitochondria as a result of toxicity induced by treatment with doxorubicin.

Our result also revealed an increase in the activities of gamma glutamyl transferase (GGT) in groups treated with doxorubicin and this in agreement with (36) that doxorubicin induce liver disorders manifested by an increase liver enzymes (GGT) (37).

Serum albumin is the most important member of export proteins. Export proteins are synthesized on polyribosomes bound to the rough endoplasmic reticulum of the hepatocyte. In contrast, proteins destined for intracellular use are synthesized on free rather than polyribosomes (38). In the relation to the effect of doxorubicin and Vancomycin on plasma proteins and albumin. Our results showed that doxorubicin afforded a significant decrease in serum total proteins four weeks post treatment which is represented by a significant decrease in serum albumin. L-carnitine and combination with doxorubicin elicited a significant increase in serum total proteins compared with the groups doxorubicin alone and normal control group respectively. Other investigation by showed that rats that received doxorubicin had

low plasma albumin by losing protein in urine consistent with hyaline droplets presented in capsular space and tubular lumen. The mechanism may associate with alteration of glomerular capillary permeability due to sieving defect.

Malondialdehyde is a metabolite derived from lipid peroxidation which has been widely used as an indicator of oxidative stress (40). The measurement of this MDA provides a convenient index of lipid peroxidation (41). body develops several endogenous antioxidant systems to deal with the production of reactive oxygen species. Antioxidants act as radical scavengers, inhibit lipid peroxidation and other free radical mediated processes, and there by protect human body from several diseases attributed to the reactions of radicals (42,43). The present study demonstrated that treatment with DOX increased MDA (an index of lipid peroxidation). These results correlate well with previous studies. DOX treatment was shown to induce peroxidative alterations in tissues which were evident by significant elevation in MDA production in the rat's heart, kidneys and liver tissues (44.45). Doxorubicin either alone or given in combination with L-carnitine enhanced lipid peroxidation measured as malondialdehyde (MDA) along the course of the experiment when compared with normal control group. Doxorubicin induced marked biochemical alterations characteristic of cardiac toxicity including enhanced lipid peroxidation as measured by malondialdehyde (MDA). Furthermore, it has been demonstrated that 20 mg/kg single dose of DOX resulted in renal lipid peroxidation and protein oxidation at 10th day of DOX injection in rats (46,47).

Vancomycin carries the potential to exacerbrate metabolic disorders by increasing adiposity and body mass index when it used to treatment the gut microbial or as probiotic (48, 49).

Several investigators have suggested that oxygen free radicals are considered to be important mediators of gentamicin-induced nephrotoxicity (50). The genetic defect of

leptin-deficient ob/ob mice, which causes a severe obese phenotype, is associated with increased sensitivity to roinflammatory monocyte/macrophage-activating stimuli and impairment of phagocytic functions, as well as reduced T-cell function (26).

Our results showed decrease in leptin level in rats treated with VAN and this in agreement with (10) who reported that that one week of VAN treatment led to a significant reduction in serum leptin levels. Despite a 41% reduction in serum leptin levels by VAN. Also, (25) mentioned that VAN may be able to affect a biphasic protection of the myocardium by reducing the level of leptin in the circulation. As demonstrated in the current study, decreased leptin levels in the circulation decreases the myocardium's susceptibility to acute injury from ischemia/reperfusion while other studies have showed that reduced leptin signaling through blockade of the leptin receptor results decreased chronic cardiac hypertrophy. Administration of VAN in treatment of a case respiratory failure resulted in decreasing the frequency of in blood glucose measurement and improve glycemic status (51).

Our result showed an increase level of LDH and this agrees with (52). In cased of anosocomial infection I/V VAN 1000 mg every 12 hours. The increase in LDH level may be due to patient has hematologic abnormalities and developed leukocytosis, eosinophilia, normocytic anemia; schistocytes present on a peripheral blood sear (53).

Gamma-glutamyl transferase (GGT) is shown to be an independent risk factor for the mortality and morbidity of cardiovascular diseases in recent epidemiological and clinical studies. In addition, several prospective studies reported that baseline serum concentration was an independent risk factor for the development of coronary artery disease (CAD), diabetes mellitus, stroke hypertension. GGT plays an important role in antioxidant defense systems. Elevated GGT levels could be a marker of oxidative stress and sub clinical inflammation (54, 55). Some epidemiological studies also suggest that higher

GGT serum levels is associated with development of CVD risk factors, including diabetes, hypertension, and the metabolic syndrome. GGT may play a role in early diagnosis of metabolic syndrome with a high predictive value for both metabolic syndrome and cardiovascular disease presence (56). But VAN administration in combination with L carnitine resulted in decrease in GGT level. This result agrees with (35) who used VAN in with combination L carnitine as chemotherapy for lymphoblastic acute leukemia.

Our results showed reduction in albumin level and this agreed with (57) who said that serum albumin concentration was significantly lowered in burns patients when treated with VAN.

VAN could enhance cellular ATP concentrations and stimulate oxygen consumption, supporting the role of VAN as a stimulant of oxidative phosphorylation and the free radical production. Vancomycin induced free radical injury may be generated directly or indirectly (58).This destructive peroxidation leads to breakdown of membrane structure and function. Further decomposition of per oxidized lipids yields a wide variety of end products, including MDA. There was a significant increase of MDA concentration in renal tissue of rats treated with vancomycin; suggesting the involvement of oxidative stressinduced nephrotoxicity. This result was agreed with reports by (59,60).

Our results showed increase level of cholesterol in case of VAN treatment and this agrees with (61) who used vancomycin in treatment of cytotoxicity of human glial cell. But in case of vancomycin and L-carnitine showed decrease cholesterol level. Also there were an increase in HDL plasma and this agrees with (62) in treatment of gut microbial. But L-carnitine when given in combination with vancomycin makes improve HDL level but still more than normal.

Conclusion

Both doxorubicin and vancomycin have a bad effect on the lipid sate as they increase leptin and insulin receptors inducing obesity in experimental animals, also; they increase liver enzymes and heart markers as well as increase the oxidative stress in the body tissues due their ability to increase the level of MDA in the serum of the experimental animals. L-carnitine administration ameliorates the hazards effect of DOX and VAN on the lipid sate in the body.

REFERENCES

- 1. Kelesidis T and Mantzoros CS (2006): The emerging role of leptin in humans. Pediatr Endocrinol Rev. 3 (3): 239-48.
- Banks W (2003): Is obesity a disease of the blood-brain barrier? Physiological, pathological, and evolutionary considerations. Current Pharmaceutical Design. 9: 801-9.
- 3. Gibson WT, Farooqi IS, Moreau M DePaoli AM, Lawrence E, O'Rahilly S, and Trussell RA (2004): Congenital leptin deficiency due to homozygosity for the 133G mutation: report of another case and evaluation of response to four years of leptin therapy. J. Clin. Endocrinol. Metab., 89: 4821-4826.
- 4. Steiner RA (1997): Lords and ladies leapin' on leptin. Endocrinology, 137: 4533-4535.
- Matsuda J, Yokota I, Iida M, Murakami T, Naito E and Ito M (1997): Serum leptin concentration in cord blood: relationship to birth weight and gender. J. Clin. Endocrinol. Metab., 82: 1642-4.
- 6. Tritos N and Mantzoros CS (1997): Leptin: Its role in obesity and beyond. Diabetologia, 40:1371-9.
- 7. Quiles JL, Huertas JR, Battino M, Mataix J and Ramirez-Tortosa MC (2002):

- Antioxidant nutrients and adriamycin toxicity. Toxicol., 180: 79-95.
- 8. Bizzi A, Ceriani L, Gerundino M, Spina A, Tacconi M and Veneroni E (1983):
 Adriamycin causes hyperlipemia as a consequence of nephrotoxicity. Toxicol. Lett., 18: 291-300.
- 9. Eli L (1999): Biochemical mode of action of vancomycin and history of global clinical vancomycin use. Interscience Conference on Antimicrobial Agents and Chemotherapy. 26-29; 39:780..
- 10.Lam V, Su J, Koprowski S, Hsu A, Tweddell JS, Rafiee P, Gross GJ, Salzman NH and Baker JE (2012): Intestinal microbiota determine severity of myocardial infarction in rats. FASEB J. Article, 26 (4): 1727-35.
- 11.McCafferty K, Byrne C and Yaqoob M (2012): Intestinal microbiota determine severity of myocardial infarction in rats. FASEB J. 26: 4388.
- 12.Liedtke AJ, Nellis SH, Whitesell LF and Mahar CQ (1982): Metabolic and mechanical effects using L- and D-carnitine in working swine hearts. Am. J. Physiol., 243 (5): 691-697.
- 13.Cederblad G, Niklasson A, Rydgren B, Albertsson-Wikland K and Olegård R (2008): Carnitine in Maternal and Neonatal Plasma. Acta Paediatrica, 74 (4): 500-504.
- 14.Tufekci O, Gunes D, Ozoul C, Koiatan E, Altun Z., Yilma O, Aktafl S, Erbayraktar Z, Kirkim G, Mutafolu K, Soylu A, Serbetgiolu B, Giineri EA and Olgun N (2009): Evaluation of the effect of acetyl L-carnitine on experimental cisplatin nephrotoxicity. Chemotherapy, 55:451-459.
- 15.Yürekli Y, Unak P, Yenisey C, Ertay T, Muftuler FZ and Medine E (2011): L-Carnitine protection against cisplatin nephrotoxicity in rats: comparison with amifostin using quantitative renal Tc 99m DMSA uptake. Molecular Imaging and Radionuclide Therapy, 20 (1):1 6.

- 16.Kaslfi K, Israel M, Sweatman TW, Seshadri R and Cook GA (1990): Inhibition of mitochondrial carnitine palmitoyltransferases by adriamycin and adriamycin analogues. Biochem. Pharmacol., 40: 1441 1448.
- 17. Wauters M, Considine R and Gaal L. (2000): Human leptin: From an adipocyte hormone to an endocrine mediator. Eur. J. Endocrinol., 143: 293-311.
- 18.Sayed MM, Sharawy SM, Shouman SA and Osman AM (1999): Reversal of doxorubicin-induced cardiac metabolic damage by L-carnitine. Pharmacol. Res., 39: 289 295.
- 19.Yueh-Chiao Y, Hui-Chih-Tai T, Wen-Lieng L, Li-Chuan W, Kuo-Yang W, Hui-Chun L and Tsun-Jui L (2007): Protection by doxycycline against doxorubicin-induced oxidative stress and apoptosis in mouse testes. Biochemical Pharmacology, 74 (7): 969-980.
- 20.Paget G and Barnes JM (1964): Evaluation of drug activities: Pharmacometric. Ed. Laurance and Becharach, Vol. I, Academic Press, New York.
- 21.Boonsanit D, Kanchanapangka S and Buranakarl C (2006): L-carnitine ameliorates doxorubicin-induced nephritic syndrome in rats. Nephrology, 11: 313-320.
- 22.Dagogo JS, Fanelli C, Paramore D, Brothers J and Landt M (1996): Plasma leptin and insulin relationships in obese and nonobese humans. Diabetes, 45: 695-698.
- 23.Duncan DB (1995): Multiple rang and multiple F test. Biometrics 11:1-42.
- 24.Chu NF, Spiegelman D., Rifai N, Hotamisligil GS and Rimm EB (2000): Glycemic status and soluble tumor necrosis factor receptor levels in relation to plasma leption concentrations among normal weight and overweight US men. Int. J. Obes. Relat. Metab. Disord., 24: 1085-92.
- 25.Purdham DM, Rajapurohitam V, Zeidan A, Huang C, Gross GJ and Karmazyn M (2008): A neutralizing leptin receptor

- antibody mitigates hypertrophy and hemodynamic dysfunction in the postinfarcted rat heart. Am. J. Physiol. Heart Circ. Physiol. 295: H441–H446.
- 26. Faggioni R, Feingold KR and Grunfeld C. (2001): Leptin regulation of the immune response and the immunodeficiency of malnutrition. FASEB J., 15: 2565-71.
- 27.Lloyd RV, Jin L, Tsumanuma I, Vidal S, Kovacs K and Horvath E (2001): Leptin and leptin receptor in anterior pituitary function. Pituitary, 4 (1-2): 33-47.
- 28.Usuki K, Okazaki R, Iki S, Muramatsu M, Yamaguchi Y, Totsuka Y and Urabe A. (1998): Serum leptin levels during cancer chemotherapy. Ann Hematol., 77: 191-192.
- 29.Emilsson V, Arch JR, de Groot RP, Lister CA and Cawthorne MA (1999): Leptin treatment increases suppressors of cytokine signaling in central and peripheral tissues. FEBS Lett.455 (1-2): 170-4.
- 30.Hewish M, Chau I and Cunningham D. (2009): Insulin-like growth factor 1 receptor targeted therapeutics: novel compounds and novel treatment strategies for cancer medicine. Recent Patents on Anti-Cancer Drug Discovery, 4: 54-72
- 31.Evans JL, Goldfine ID, Maddux BA and Grodsky GM (2002): Oxidative stress and stress-activated signaling pathways: a unifying hypothesis of type 2 diabetes. Endocr. Rev., 23: 599-622.
- 32.Halliwell B (1994): Free radicals, antioxidants, and human disease: Curiosity, cause, or consequence? Lancet, 344: 721-724.
- 33.Saad SY, Najjar TA and Al-Rikabi AC. (2001): The preventive role of deferoxamine against acute doxorubicininduced cardiac, renal and hepatic toxicity in rats. Pharmacol. Res., 43: 211-218.
- 34.Deepa PR and Varalakshmi P (2003):
 Protective effect of low molecular weight heparin on oxidative injury and cellular abnormalities in adriamycin-induced

- cardiac and hepatic toxicity. Chemico-Biological Interactions, 146: 201-210.
- 35.Koti BC, Vishwanathswamy AH, Wagawade J and Thippeswamy AH (2009): Cardioprotective effect of lipistat against doxorubicin induced myocardial toxicity in albino rats. Indian J. Exp. Biol., 47: 41-46.
- 36.Del Barco S, Colomer R, Calvo L, Tusquets I, Adrover E, Sánchez P, Rifa J, De la Haba J and Virizuela JA (2009): Nonpegylated liposomal doxorubicin combined with gemcitabine as first-line treatment for metastatic or locally advanced breast cancer. Final results of a phase I/II trial. Breast Cancer Res. Treat., 116:351-358.
- 37. Von Hoff DD, Rozencweig M and Piccart M (1982): The cardiotoxicity of anticancer agents. Semin Oncol., 9: 23-33.
- 38. Podolsky DK and Isselbacher KJ (1991):
 Derangements of hepatic metabolism. In:
 Wilson, J.D., Braunwald, E., Isselbacher,
 K.J. Petersdorf, R.G., Martin, J.B., Fauci,
 A.S., Root, R.K. (eds), Harrison's Principle
 of Internal Medicine. McGraw- Hill, New
 York. pp. 1311-1317.
- 39. Weening JJ and Rennkle HG (1983): Glomerular permeability and polyanion adriamycin nephrosis in the rat. Kidney Int., 24: 152-159.
- 40.Lepage G, Munoz G, Champagne J and Roy CC (1991): Preparative steps for the accurate measurement of malondialdehyde by high-performance liquid chromatography. Anal. Biochem., 197: 277-283.
- 41.Osman NN, Hussein EM and Farag MF. (2009): Black tea (Camellia sinensis) role in modulating antioxidant enzymes and biochemichal changes in y-irradiated rats. Egypt. J. Rad. Sci. Applic., 22 (1): 287 304.
- 42.Repetto MG and Llesuy SF (2002): Antioxidant properties of natural compounds used in popular medicine for

- gastric ulcers. Brazil. J. Med. and Biol. Res., 35: 523-534.
- 43.Dokmeci D, Akpolat M, Aydogdu N, Doganay L and Turan FN (2005): Carnitine inhibits ethanol-induced gastric mucosal injury in rats. Pharmaco. Reports, 57: 481-488.
- 44.Luo XP, Evrovsky Y, Cole D, Trines J, Benson LN and Lechotary DC (1997):
 Doxorubicin-induced acute changes in cytotoxic aldehydes, antioxidant status and cardiac function in the rat. Biochem. Biophy. Acta., 1360: 45-52.
- 45.Cao Y, Kennedy R and Klimberg VS. (1999): Glutamine protects against doxorubicin-induced cardiotoxicity. J. Surg. Res., 85: 178- 182.
- 46. Arafa HM, Abd-Ellah MF and Hafez HF (2005): Abatemant by naringenin of doxorubicin-induced cardiac toxicity in rats. J. Egypt. Nat. Cancer Inst., 17(4): 491-300.
- 47. Yagmurca M, Erdoganb IL, Irazc M, Songurd A, Ucare M and Fadilliogluf E (2004): Caffeic acid phenethyl ester as a protective agent against doxorubicin nephrotoxicity in rats. Clinica Chimica Acta, 348:27-34.
- 48.Cho I, Yamanishi S, Cox L, Methé BA, Zavadil J, Li K, Gao Z, Mahana D, Raju K, Teitler I, Li H, Alekseyenko AV and Blaser MJ (2012): Antibiotics in early life alter the murine colonic microbiome and adiposity. Nature, 488: 621-626.
- 49.Trasande L, Blustein J, Liu M, Corwin E, Cox LM and Blaser MJ (2013): Infant antibiotic exposures and early-life body mass. Int. J. Obes. (Lond). 37 (1): 16-23.
- 50.Ali BH (2003): Agents ameliorating or augmenting experimental gentamicin nephrotoxicity: some recent research. Food Chem. Toxicol. 41: 1447–1452.
- 51.Trissel L (2003): Handbook of Injectable Drugs. 12th ed Bethesda, MD, American Society of Health-Systems Pharmacists.

- 52.Fleming P and Marik PE (2011): The DRESS syndrome: the great clinical mimicker. Pharmacotherapy, 31 (3): 332.
- 53.Schnetzke U, Bossert T, Scholl S, Freesmeyer M, Hochhaus A and La Rosée P (2011): Drug-induced lymphadenopathy with eosinophilia and renal failure mimicking lymphoma disease: dramatic onset of DRESS syndrome associated with antibiotic treatment. Ann Hematol., 90 (11):1353-1355.
- 54 Jousilahti P, Rastenyte D and Tuomilehto J (2000): Serum γ-glutamyl transferase, self-reported alcohol drinking, and the risk of stroke. Stroke., 31: 1851-1855.
- 55.Lee DH, Jacobs DRJr, Gross M, Kiefe CI, Roseman J, Lewis CE and Steffes M (2003): Gamma-glutamyl transferase is a predictor of incident diabetes and hypertension: the Coronary Artery Risk Development in Young Adults (CARDIA) Study. Clin. Chem., 49: 1358-1366.
- 56.Devers MC, Campbell S, Shaw J, Zimmet P and Simmons D (2008): Should liver function tests be included in definitions of metabolic syndrome? Evidence from the association between liver function tests, components of metabolic syndrome and prevalent cardiovascular disease. Diabetic Medicine, 25 (5): 523–529.
- 57.Dolton M, Xu H, Cheong E, Maitz P, Kennedy P, Gottlieb T, Buono E and McLachlan AJ (2010): Vancomycin pharmacokinetics in patients with severe burn injuries. Burns, 6 (4):469-476.
- 58.King DW and Smith MA. (2004): Proliferative responses observed following vancomycin treatment in renal proximal tubule epithelial cells. Toxicol In-vitro, 18:797-803.
- 59.Faruk O, Koyuncu AM, Fehmi O, Ozden C, Ramazan YH and Efkanevue CM (2005): In-vivo evidences suggesting the role of oxidative stress in pathogenesis of vancomycin-induced nephrotoxicity: Protection by erdosteine. Toxicology, 215 (3):227-233.

- 60.Cetin H, Olgar S, Oktem F, Ciris M and Atay A (2007): Novel evidence suggesting an anti-oxidant property for erythropoietin on vancomycin-induced nephrotoxicity in a rat model. Clinical and Experimental Pharmacology and Physiology. 34 (11): 1181-1185.
- 61. Sofian ZM, Abdullah JM, Rahim AA, Shafee SS, Mustafa Z and Abdul Razak S (2007): Cytotoxicity evaluation of
- vancomycin and its complex with betacyclodextrin on human glial cell line. Pakistan Journal of Pharmaceutical Sciences, 25 (4): 831-837.
- 62. Holmes E, Li JV, Athanasiou T, Ashrafian H and Nicholson JK (2011): Understanding the role of gut microbiomehost metabolic signal disruption in health and disease. Trends in Microbiology, 19: 349-359.

الملخص العربي

التأثير الكيميائى للكرياتينين ضد التأثير الضار لكل من

الدوكسيروبسين والفانكومايسين فى الفئران

رحاب زايد ، يوسف محمد على شحاته ، محمد فهمى دويدار ، هيثم عبد الله على ، آسر محمد عبد العظيم قسم الكيمياء الحيوية – كلية الطب البيطرى- جامعة الزقازيق

أجريت هذه الدراسة على الفئران لدراسة التأثير المضاد لمادة الكارنيتين على الاضرار النتاجة عن إستخدام كل من الدوكسير وبسين والفانكو مايسين على الكبد ومستوى الدهون والكلية.

استخدم في هذه الدراسة عدد ٩٠ فأر قسموا لست مجموعات متساوية (١٥ فأر لكل مجموعة) وبعد المعاملات تم ذبح الفئران بعد ٣، ٥، ١٥ يوم من إنتهاء المعاملات.

اسفرت النتائج عن نقص في مسبة هرمون الليبتن والأنسولين في الدم وكذلك نقص مستوى البروتينات الكلية والألبيومين مع زيادة في مسبة الممالونالديهايد واللاكتيك ديهيروجينيز والجاما جلوتاميل ترانسفيريز والكلوستيرول الكلي والعالى الكثافة ولكن استخدام الكارنيتين كان له من الاثر من تعديل في المستويات المضطربة وعودتها للنسب الطبيعية مما يدل على فوائد الكارنيتين في مجابهة التأثير الضار لكل من الدوكسيروبسين والفانكومايسين.