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The Optimum Conditions for Producing Fermented Milk Used in the Manufacture of Fayesh Bread

Elsayed A. Mahmoud^{1*}, Abd-Ellah A. Abd-Alla^{2#}, Elsayed K. Bakhiet³ and Mohamed A. Sorour¹

¹Food Science and Nutrition Department, Faculty of Agriculture, Sohag University, Sohag 82524, Egypt ²Dairy Science Department, Faculty of Agriculture, Sohag University, Sohag 82524, Egypt

³Botany and Microbiology Department, Faculty of Science, Al-Azhar University, Assiut, 71524, Egypt

> **F**AYESH is one of the most popular types of bread in Egyptian kitchen, which depends on the fermentation of boiled milk using cereals and legumes. In this study, the optimum conditions for fermented milk used in the manufacture of Fayesh were evaluated. The bacteria responsible for the milk fermentation were also isolated and identified. Ten grams of whole or crushed some cereals and legumes were added separately or incombination to 150 mL beilod milk and incubated at room temperature (traditional method) or at 30, 40 and 50 °C. The results illustrated that milk fermentation needs more time at room temperature than fermentation at 40 °C. At 40°C, the time of milk fermentation was the lowest compared to that at 30 and 50 °C. Regarding the type of grains, the fermentation time was the lowest using both whole lentil (7.5 hr) and crushed wheat green (7.5 hr),followed by whole faba bean (8.0 hr).When mixing the grains, the least time for fermentation was 6.5 hr, using lentil/wheat grain mixture, followed by fababean/green wheat (7.0 hr), fababean/wheat grain (7.0 hr), green wheat/wheat grain (7.0 h), and lentil/green wheat (7.25 hr).*Staphylococcus vitulinus* was isolated from faba beans, green wheat,and wheat grain. *Staphylococcus lentus* was isolated from chickpea and sesame, while *Enterococcus faecium* was also isolated from lentil.

Keywords: Fermented foods, Milk fermentation, Fayeshbread, Cereals, Legumes.

Introduction

Indigenous fermented food such as bread, cheese, and wine, have been prepared and consumed for thousands of years and are strongly linked to culture and tradition, especially in rural households and village communities (Madsen et al., 2001). Fermentation is cheap and the most economical technique of food production and preservation. Many types of indigenous fermented foods are made based on milk, raw products, fruits, cereals, grains, and vegetables using locally available raw materials and local knowledge (Joshi et al., 2012). The fermented foods are derived from substrates like roots, legumes, cereals, oilseeds, nuts, meat,

*Corresponding author: Email, <u>sayed@agr.sohag.edu.eg</u> #Email: <u>drabdellahhassan@gmail.com</u> Received:19/3/2020; accepted:21/5/2020 DOI: 10.21608/EJFS.2020.26172.1047 ©2020 National Information and Documentation Centre (NIDOC)

fish, milk, palm tree, sap, etc (Uzogara et al., 1990).

Fermented foods in the Arab countries and many others in the Middle East are very typical of those found in Egypt. In Arabic and other countries, some of them have different national names in this region. One or more of these foods participate and thus provide an essential source of digestible protein in daily meals. The fermented foods are analyzed concerning their traditional manufacturing processes. It includes sour milk, Carish, Mish, Laban Zeer, Kishk, Zabady, Shamsian bread, and certain Sudanese foods that have been fermented (El-Gendy, 1983).



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The preparation of many indigenous or traditional fermented foods and beverages remains today as a house art. They are produced in homes, villages, and small-scale industries (Bol and DeVos, 1997). Traditional fermentation processes are typically uncontrolled and dependent on microbes from the environment, fermentation substrate. region geography, topography. agricultural factors, and climate to initiate it, and as such, can result in products of low yield and variable quality (Holzapfel, 2002; Ross et al., 2002 and Methaet al., 2012). Fermentation in food processing is essentially the conversion of carbohydrates to alcohols and carbon dioxide or organic acids using yeasts, bacteria, or a combination thereof under anaerobic conditions. Many societies have considered traditional knowledge and skills, which is a critical component for successful fermentation activities. Additional overarching qualities deemed useful. The level of fermentation sophistication is highly related to the level of technological development and existing institutional support systems within a region or country (Rolle & Satin, 2002). Fermentation is spontaneous as a result of competitive microbial activities. Hence some strains are best adapted with rapid growth, thereby dominating others at stages of fermentation. This microbial accounts for the succession process, as reported by many workers (Mbata et al., 2009).

Most of the traditional fermented foods are solid substrate-based, where the substrates undergo the process of natural fermentation either naturally or by adding starter cultures. Fermented foods can be described as palatable and whole foods prepared from raw or heated raw materials. They are generally appreciated for attributes such as pleasant flavor, aroma, texture, and improved cooking and processing properties. Microorganisms, by their metabolic activities, contribute to the development of characteristic features such as taste, aroma, visual appearance, texture, shelf life, and safety. Enzymes indigenous to the raw materials may play a role in enhancing these characteristics (Hammes, 1990). Traditional foods are those produced by ethnic people using locally available raw materials to make edible products that are culturally and socially acceptable to the consumer (Tamang, 2010). The traditional art of fermentation practiced by the common man continues despite the scientific and technological revolution, though remains confined mainly to rural and tribal areas, due to the inaccessibility of commercially made products in the remote regions

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and their cost, and the sociocultural linkages of indigenous fermented products (Thakur et al., 2004). Fermentation food can take place if there is a suitable substrate, appropriate microorganism(s) either from nature or by inoculation of specific microorganisms, and the necessary environmental conditions for the fermentation to take place (Caplice and Fitzerald, 1999).

Sour dough starter is likely the oldest, being reliant on organisms present in the grain and local environment. These starters have fairly complex microbiological makeup, and the most notable including wild yeasts, lactobacillus, and Acetobacteria in symbiotic relationship referred to as a SCOBY (Scheirlinck et al., 2008) and (Arendt et al., 2007). They are usually maintained over long periods. For example, the Boudin Bakery in San Francisco has used the same starter dough for over 150 years. Mother dough often refers to a sourdough, and in this context, the term starter often refers to all or a piece of mother dough (Renneberg and Demain, 2007). However, mother dough may also apply to a firstgeneration yeast sponge (Esposito, 2003) so the process (Hui et al., 2006) used concerning the ingredients and fermentation times is essential to understanding yeast versus sourdough methods. A roughly synonymous term used in French baking is Chef (Calvel, 2001).Fayish bread is a traditional fermented food in Upper Egypt, which is produced at the household level and prepared during marriage ceremonies, religious and served with tea. Fermenting starter of Fayesh bread prepared from mixed cereals and legumes with milk. Relatively little information is available on the starter culture of Fayesh. In Upper Egypt, Fayesh bread manufactured using fermented milk by adding a cup of boiling milk to 2 table spoons of faba bean, lentil, greenwheat "Freekeh", chickpea, sesame and wheat placed in a jar, sleeve, wrapped in towel or cloth and leave in a warm dark place for two days or less. Therefore, this work aims to evaluate the optimal conditions for producing fermented milk using some cereals or legumes as a starter used in the manufacture of Fayesh bread, as well as isolating and identifying the microbial strains responsible for this fermentation.

Materials and Methods

Materials

• Cereals and legumes such as fababean (*Vicia faba*), lentil (*Lens culinaris*), green wheat "Freekeh" (*Triticum aestivum*),chickpea (*Cicer arietinum*), sesame (*Sesamum indicum*) and wheat

(*Triticum aestivum*) were obtained from local markets, Sohag, Egypt. Milk was obtained from animal production Farm, Faculty of Agriculture, Sohag University Sohag, Egypt. The average composition was 87.20, 3.55, 3.75. 4.75 and 0.75% for moisture, proteins, fat, carbohydrates and ash, respectively.

Methods

Fermentation process

The fermentation process was carried out by adding 10 g of faba bean, lentil, green wheat (Freekeh), chickpea, sesame, and wheat grains separately or in combination, as whole grains or crushed grains, to 150 mL of boiled milk after cooling in an airtight container. The containers were left to ferment by its microflora at room temperature (~ 28 °C, traditional method) or a 30, 40 and 50°C until the fermentation process was completed (milk separated into two layers;serum layer, and foamed fermented layer). The time of fermentation was measured and expressed as a hour.

Chemical composition of raw materials

Moisture, protein, fatand ash contents of cereals, legumes and milk were determined according to the methods of AOAC (2007). The total carbohydrate content was calculated by difference.

Microbiological analysis

The total bacterial counts were determined using the plate counts technique on a nutrient agar medium to enumerate different viable microorganism groups according to procedures by APHA (1976) and Difco (1984). The plates were incubated at 37 °C for 48 hr. All tests were performed in duplicate, expressing the results as log colony-forming units (CFU)/g of the product.

Microbial isolation and enumeration

Isolation and cultivation of bacteria

One gram was weighted from each fermented sample, added to 100 mL sterilized distilled water in 150 mL conical flask, and then stirred (bioMérieux Inc., Hazelwood, MO,USA) for 15 min at a speed of 200 rpm/min (Janssen et al., 2002). The stirred sample was also serially diluted to 10^{-2} , 10^{-3} , 10^{-4} , 10^{-5} , 10^{-6} and 10^{-7} , and 200 µL of each last three dilutions was used to inoculate each of triplicate plates at each dilution level to constitute a counting set of 9 plates. Inoculate were spread over the surface of the Nutrient-agar medium using sterile glass spreading rods (Davis et al., 2005). All plates were incubated at 30° C

for 2 days in sealed polyethylene bags (40- μ m film thickness). Morphologically distinct colonies were purified and maintained on nutrient agarslant at 4 °C.

Screening and characterization of strain identification Morphological and physiological test

The ability of the bacterial isolates from faba bean, lentil, green wheat (Freekeh), chickpea, sesame, and wheat to hydrolyze urea, arginine, and Polymyxin B resistance were examined. The morphological and physiological characteristics of the bacterial isolates were compared with the data from Bergey's Manual of Determinative Bacteriology (Holt et al., 1994).

Metabolic profiling

The Vitek 2 compact Automated Expert System (AES) (bioMérieux Inc., Hazelwood, MO,USA) was used for metabolic profiling. Using the Vitek 2 ID-GP card (bioMérieux), identification of Gram-positive cocci occurs through testing the organism's metabolic activity in 43 biochemical tests designed to measure carbon source utilization and enzymatic activity, the reference identification was determined with API Staph and API 20 Strep identification kits. Overall, the VITEK 2 GP correctly identified 96.5% of the isolates, including 2.2% low discrimination with the correct species listed. Misidentifications occurred at 3.3%, and no identifications occurred at 0.2%. The GP identification card is based on established biochemical methods and newly developed substrates (Atlas, 1993; Barros et al., 2001; Billeet al., 1992; Collins &Lawson, 2000 and Holt et al., 1994).

Database development

The databases of the VITEK 2 identification products are constructed with large strain sets of well-characterized microorganisms tested under various culture conditions. These strains are derived from a variety of clinical and industrial sources as well as from the public (*e.g.*, ATCC) and university culture collections.

Analytical techniques

Test data from an unknown organism are compared to the respective database to determine a quantitative value for proximity to each of the database taxa. Each of the composite values is compared to the others to determine whether the data are sufficiently unique or close to one and more of the other database taxa. If a unique identification pattern is not recognized, a list of possible organisms is given, or the strain is determined to be outside the scope of the database.

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Statistical analysis

Data was analyzed by analysis of variance (MANOVA) using the GLM procedure with SAS software (2008). Significant differences were determined as $P \le 0.05$ using Duncan's multiple range tests.

Results and Discussion

Composition of cereals and legumes

The type of microflora in all food fermentations depend on the pH, water activity, temperature, salt concentration, and chemical composition of the food matrix (Blandino et al., 2003). Thus, the composition of cereals and legumes (faba bean, lentil, green wheat "Freekeh," chickpea, sesame, and wheat), used in the making of Egyptian Fayesh bread were determined (Table 1). Faba bean had the highest protein content (31.33%, P < 0.05), followed by lentils (26.44%, P < 0.05), chickpea (20.58%), and Sesame (18.58%), but wheat and green wheat had the lowest protein content (12.86 and 12.40%, P < 0.05), respectively. Carbohydrates content was higher in both green wheat and wheat (73.48 and 72.44 %) followed by chickpea, lentil, and faba bean, while was lower in sesame (22.92%). However, Sesame had the highest fat (46.83%) and ash (5.57%) content (P < 0.05)than other cereals and legumes.

These values were close to those mentioned by Carsanba et al. (2017) in green wheat, Nzikou et al. (2009) in sesame seeds, and Fišteš et al. (2014) in Chickpea. Additionally, the protein and carbohydrates content of lentil was in the normal range (26.6-30.4% and 48.3-61.0%, respectively), reported by Hefnawy (2011).

Raw materials and fermentation process

Initial experiments indicate that the use of sterilized cereals or legumes, using autoclave at 121°C for15 min, with boiled milk after cooling, had no positive effect on the fermentation process compared to the non-sterilized cereals or legumes. These results explained that the fermentation process was done by endoenzymes or the original microflora on the surface of cereals or legumes. These results in the same line with (Harlander, 1992) who reported that microbes responsible for the fermentation might be the natural microflora indigenously present on the substrate, or they may be added as starter cultures. The addition of milk to the cereal and legumes stimulates the endoenzyme into action and growth of microbes (Achi and Ukwuru, 2015). Figure 1 illustrates the success of fermentation of boiled milk using unspecialized grains by separating the milk into two layers; serum layer and foamed fermented layer.



Fig. 1. Images of fermentd milk by using unsterilized cereals or legumes.

- 1- Fermented milk using crushed green wheat "Freekeh"
- 2- Fermented milk using whole wheat
- 3- Fermented milk using crushed faba bean
- 4- Fermented milk using whole chickpea
- 5- Fermented milk using whole lentils

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Milk fermentation using whole cereals and legumes at different temperatures

Traditional Egyptian Fayesh bread is made by adding fermented milk using cereals or legumes at room temperature in a warm place, as a starter, to Fayesh bread dough. The effect of the incubation temperature on the time required to milk fermentation using whole cereals and legumes is presented in Table 1. In general, the period of milk fermentation at 40° C was the lowest, while fermentation at 30 °C was the longest (P < 0.05). This means the incubation at 40° C was more suitable for milk fermentation with whole cereals and legumes; except green wheat, milk fermentation was the lowest at 50° C (7.0 h).Regarding the type of grains, the fermentation period was the lowest with whole lentils and chickpea as (P < 0.05) followed by faba bean at 40 and 50° C, while the lowest was with lentils and chickpea followed by faba bean at 30° C.(P < 0.05).

Also, the incubation of green wheat "Freekeh" with chilled boiled milk at 30, 40 and 50° C, decrease the fermentation time to 9, 8 and 7 hr (P <0.05), respectively. There is a difference between the samples incubated on high-temperature degrees and those equipped with the normal room temperature. Using high temperature leads to accelerating the starter production compared with room temperature. On the contrary, using sesame and wheat grains in milk fermentation exhibited the longest fermentation period compared to other cereals and legumes at different temperatures (P < 0.05). The increased fermentation period with sesame may be due to the high fat content (Table 1). These results are agreement with those reported by Caplice and Fitzerald (1999). Environmental conditions, like temperature and moisture, need to be optimum for a specific fermentation, as well as the intrinsic factors of fermentation, including the pH, type of sugar, nutrients, availability or otherwise of oxygen, etc. The composition and quality of the raw materials, the microflora involved, the amount of water, type of raw material, and time also influence the fermentation. Also, the type of organisms that are found in association with each type of fermentation depends on ecological parameters like temperature, pH, inoculums, size, and nature of cereal and propagation time. Hence, fermenting organisms are not readily substrate-specific but are conditioned by the factors mentioned (Hammes et al., 2005).

Milk fermentation using crushed cereals and legumes at 40 $^{\circ}$ C and room temperatures

Based on the above results, 40°C was the best incubation temperature for milk fermentation using whole cereals and legumes, the use of crushed cereals and legumes in the fermentation of milk was compared between 40 °C and room temperature ('25-30°C). The time recorded for complete milk fermentation using crushed cereals and legumes was shown in Table 3. At 40°C, using whole faba beans and lentils in milk fermentation was better than crushed grains, while crushed wheat grains, sesame, chickpea, and wheat were the best (P < 0.05). Compared to room temperature, milk fermentation using crushed cereals and legumes was better than that at 40 °C. However, using crushed green wheat "Freekeh" was the best (7.5 hr) at 40 °C. The time of milk fermentation were 8.5, 8.5, 7.5, 8.5, 9.0 and 8.5 hr using faba beans, lentils, green wheat, lentils, sesame and wheat, respectively. At room temperature, the time for fermentation with crushed Faba beans and green wheat was the lowest (9.5 hr), while the fermentation period was the longest (P < 0.05) when using crushed lentils and sesame seeds (13-14 hr). These results agree with Achi and Ukwuru (2015) who reported that reduction of size through milling and sometimes the use of specific microorganisms and enzyme actions enable fermentation to start (Caplice and Fitzerald, 1999). The microflora which may be indigenously present on the substrate, or added as a starter culture (Blandino et al., 2003), or maybe present in or on the ingredients and utensils, or in the environment are selected through adaptation to the substrate and by adjusting the fermentation conditions (Soni and Sandhu, 1990).

Milk fermentation using the cereals and legumes alone or incombination at 40 °C and room temperature

Some peoples sometimes prepare the fermented milk for making Egyptian Fayesh bread using mixtures of two types of cereals or legumes to improve the fermentation process and decrease the fermentation period. Regarding the added quantity, the time of milk fermentation decreased on the addition of faba bean, lentil, chickpea, or wheat (P <0.05), up to 20 g both alone; the decreasing was more pronounced with lentil, from 8.0 to 7.5 hr (Table 3). On the contrary, the time of fermentation was not affected by the increase in the amount of sesame, while it increased with the increase of green wheat from 7.5 to 8.5 hr (P <0.05).

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Raw materials	Moisture (%)	Protein (%)	Fat	Total Carbohydrates (%)	Ash (%)
	7 4 4 4	21.22.0	(%)	56 504	2.00.0
Faba bean	7.44 ^d	31.33 a	1.45 ^d	56.70 ^d	3.08 °
Lentils	8.22 °	26.44 ^b	1.30 ^d	60.69°	3.35 b
"Greenwheat "Freekeh	10.20 ^b	12.40 ^e	2.09 °	73.48ª	1.83 ^d
Chick pea	6.68 °	20.58 °	5.80 ^b	63.67 ^b	3.57 ^b
Sesame	6.10 °	18.58 ^d	46.83 a	22.92 ^f	5.57 ª
Wheat grains	11.20 ª	12.86 °	1.90 ^d	72.44ª	1.60 ^f

TABLE 1. Chemical	composition of cere	als and legumes used	in milk fermentation.

Means (n=3) with the same small letters in the same column are not significantly different at P < 0.05.

TABLE 2. Effect of whole cereals and	egumes on the period of milk fermentation at differ	ent temperature.

Corressond logumes	Fermentation period (h) at different temperature			
Cerealsand legumes	30°C	40 °C	50 °C	
Whole faba beans	10.0 ^{Ad}	8.0 ^{Ce}	9.5 ^{Bb}	
Whole lentils	9.0 ^{Ae}	7.5 ^{Cd}	8.0 ^{By}	
Whole green wheat "Freekeh"	9.0 ^{Ae}	8.0 ^{Bc}	7.0 ^{Cd}	
Whole chickpea	11.5 ^{Ab}	9.0 ^{Cb}	10.0 ^{Ba}	
Whole sesame	11.0 Ac	10.0 ^{Ba}	10.0 ^{Ba}	
Whole wheat grains	12.0 ^{Aa}	10.0 ^{Ba}	10.0 ^{Ba}	

Means (n=3) with the same capital letters in the same rowor the same small letters in the same column are not significantly different at P < 0.05.

TABLE 3. Effect of crushed cereals and legumes on the	period of milk fermentation at 40	^o C and room temperature (25-30 ^o C).

	Fermentation time (h)			
Cereals and legumes	Room temperature	40 °C		
Crushed faba beans	9.5 ^{Ad}	8.5 ^{Bf}		
Crushed lentils	14.0 ^{Aa}	8.5 ^{Bf}		
Crushed green wheat "Freekeh"	9.5 ^A	7.5 ^{Bg}		
Crushed chickpea	10.0 ^A	8.5 ^{Bf}		
Crushed sesame	13.0 ^A	9.0 ^{Be}		
Crushed wheat grains	10.0 ^A	8.5 ^{Bf}		

Means (n=3) with the same capital letters in the same rowor the same small letters in the same column are not significantly different at P < 0.05.

When mixing seeds, there are significant differences (P > 0.05) in the time of fermentation, depending on the type of mixture and temperature of incubation. Using lentil/wheat lowers the time of milk fermentation (6.5 hr), followed by faba bean/green wheat, faba bean/wheat or green wheat/ wheat grains, lentil/sesame, chickpea/wheat, crushed faba bean/sesame or sesame/wheat grains (7.0, 7.0, 7.5, 7.5, 8.0 or 8.0 hr, respectively). The use of sesame alone or with wheat grains led to delaying the formation of the fermenting starter (P <0.05). This effect may be since sesame is rich in oil that may hinder the growth of microbes on

starter. On the other hand, the fermentation process was better at 40 °C than at room temperature (P < 0.05). The fermentation period ranged from 9.5 to 17 hr at room temperature, while it ranged from 6.5 to 9 hr at 40 °C. However, using 20 g of faba bean or green wheat showed the lowest time of fermentation at room temperature (9.5 hr) compared to other grains or their mixtures. This means that increasing the weight of cereals or legumes alone or in combination, > 10 g, does not reduce the time of fermentation at room temperature.

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Samples	Treatments	Fermentation time (h) at 40 °C	Fermentation time (h) at room temperature
	20 g faba bean	8.0 ^{Bb}	9.5 ^h
	10 g faba bean + 10 g green wheat "Freekeh"	7.0^{Bg}	12.0 ^f
	10 g faba bean + 10 g lentil	7.5 ^{Bd}	12.0 ^{Af}
Faba bean	10 g faba bean + 10 g chickpea	7.45 ^{Be}	13.0 ^{Ae}
	10 g faba bean + 10 g sesame	8.0 ^{Bb}	12.0 ^{Af}
	10 g faba bean+ 10 g wheat	7.0^{Bg}	13.0 ^{Ae}
	20 g lentil	7.5 ^{Bd}	14.0 ^{Ad}
	10 g lentil + 10 g green wheat "Freekeh"	7.25 ^{Bf}	15.0 Ac
Lentil	10 g lentil + 10 g chick pea	8.0 ^{Bi}	17.0 ^{Aa}
	10 g lentil + 10 g sesame	7.5 ^{Bd}	16.5 ^{Ab}
	10 g lentil + 10 g wheat	6.5 ^{Bh}	14.0 ^{Ad}
	20 g green wheat "Freekeh"	8.0 ^{Bc}	9.5 ^{Ah}
Green wheat	10 g green wheat "Freekeh" +10 g chickpea	8.5 ^{Bb}	10.0 Ag
"Freekeh"	10 g green wheat "Freekeh" +10 g sesame	8.5 ^{Bb}	14.0 ^{Ad}
	10 g green wheat "Freekeh" + 10 g wheat	7.0^{Bg}	12.0 ^{Af}
	20 g chickpea	9.0 ^{Ba}	10.0 Ag
Chickpea	10 g chickpea + 10 g sesame	8.5 ^{Bb}	13.0 ^{Ae}
	10 g chickpea + 10 g wheat	7.5 ^{Bd}	14.0 ^{Ad}
Sesame	20 g sesame	9.0 ^{Ba}	13.0 ^{Ae}
sesallie	10g sesame + 10 g wheat	8.0 ^{Bc}	15.0 Ac
Wheat	20 g wheat	9.0 ^{Ba}	10.0 Ag

TABLE 4. The effect of the cereals and legumes alone (20 g) or incombinatios on the period of milk fermentation at 40 °C and room temperature (25-30°C.

Means (n=3) with the same capital letters in the same rowor the same small letters in the same column are not significantly different at P < 0.05.

Total bacterial countof fermented milk

Table 5 shows the total bacterial count of fermented milk using cereals and legumes. The total bacterial count was the highest in fermented milk with whole wheat grains (5.70 \log_{10} CFU/g) but was the lowest in fermented with sesame (4.07 \log_{10} CFU/g). The remaining treatments had

mediumtotal bacterial count, which ranged between 4.23 to 4.84 \log_{10} CFU/g.These results are in the same line with Harlander (1992) who reported that microbes responsible for the fermentation might be the natural microflora indigenously present on the substrate, or they may be added as starter cultures.

TABLE 5. Total bacterial count of milk fermented using	g cereals and legumes at 40	°C.
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Cereals and legumes	Total bacterial count (Log ₁₀ cfu/g)
Fababeans	4.23 °
Lentils	4.69 ^b
Green wheat "Freekeh"	4.65 ^b
Chickpea	4.84 ^b
Sesame	4.07 °
Wheat grains	5.70 ª

Means (n=3) with the same small letters in the same column are not significantly different at P < 0.05.

No.		Samples	Faba bean	Lentils	Green- wheat "Freekeh"	Chick- pea	Sesame	Wheat grains
	Test	Isolates	Staph. vitulinus	Enterococ- cus faecium	Staph. vitulinus	Staph. lentus	Staph. lentus	Staph. vitulinus
	D-Amygdalin	1	-Ve	-Ve	-Ve	-Ve	-Ve	-Ve
	phosphatidylinositol	l phospholipase c	-Ve	-Ve	-Ve	-Ve	-Ve	-Ve
	D-Xylose		+Ve	+Ve	+Ve	+Ve	+Ve	+Ve
	Arginine dihydrola	se	-Ve	+Ve	-Ve	+Ve	+Ve	-Ve
	Beta galactosidase		-Ve	+Ve	-Ve	+Ve	+Ve	-Ve
	Alpha glucosidase		-Ve	-Ve	-Ve	-Ve	-Ve	-Ve
	Ala-Phe-pro-arylan	nidase	-Ve	-Ve	-Ve	-Ve	-Ve	-Ve
	Cyclodextrin		-Ve	-Ve	-Ve	-Ve	-Ve	-Ve
	L-aspartic acid aryl	lamidase	-Ve	-Ve	-Ve	-Ve	-Ve	-Ve
	beta glucopyranosi	dase	-Ve	-Ve	-Ve	-Ve	-Ve	-Ve
	Alpha-mannosidase	e	-Ve	-Ve	-Ve	-Ve	-Ve	-Ve
	phosphatase		+Ve	-Ve	+Ve	+Ve	+Ve	+Ve
	leucinearylamidase	;	-Ve	-Ve	-Ve	-Ve	-Ve	-Ve
	l-prolinearylamidas	se	-Ve	-Ve	-Ve	-Ve	-Ve	-Ve
	Beta-Glucorinidase	e (0.018mg)	-Ve	-Ve	-Ve	-Ve	-Ve	-Ve
	Alpha-Galactosidas	se	-Ve	+Ve	-Ve	+Ve	+Ve	-Ve
	l-pyrrolidonyl-aryla	amidase	-Ve	+Ve	-Ve	-Ve	-Ve	-Ve
	Beta-Glucorinidase	e (0.0378 mg)	-Ve	-Ve	-Ve	-Ve	-Ve	-Ve
	Alanine arylamidas		-Ve	+Ve	-Ve	+Ve	+Ve	-Ve
	Tyrosine arylamida	ise	-Ve	-Ve	-Ve	-Ve	-Ve	-Ve
	D-Sorbitol		-Ve	+Ve	-Ve	+Ve	+Ve	-Ve
	Urease		-Ve	-Ve	-Ve	-Ve	-Ve	-Ve
	Polymyxin B resist	ance	-Ve	+Ve	-Ve	-Ve	-Ve	-Ve
	D-galactose		-Ve	+Ve	-Ve	+Ve	+Ve	-Ve
	D-ribose		+Ve	+Ve	+Ve	+Ve	+Ve	+Ve
	l-lactate alkalizatio	n	-Ve	+Ve	-Ve	+Ve	+Ve	-Ve
	Lactose		-Ve	+Ve	-Ve	-Ve	-Ve	-Ve
	N-acetyl-glucosam	ine	-Ve	+Ve	-Ve	+Ve	+Ve	-Ve
	D-Maltose		-Ve	+Ve	-Ve	+Ve	+Ve	-Ve
	Bacitracin resistance	ce	+Ve	+Ve	+Ve	+Ve	+Ve	+Ve
	Novobiocin resistar	nce	-Ve	+Ve	-Ve	+Ve	+Ve	-Ve
	Growth in 6.5 % N	aCl	+Ve	+Ve	+Ve	+Ve	+Ve	+Ve
	D-Mannitol		+Ve	+Ve	+Ve	+Ve	+Ve	+Ve
	D-Mannose		+Ve	+Ve	+Ve	+Ve	+Ve	+Ve
	Methyl B-D-glucor	oyranoside	+Ve	+Ve	+Ve	+Ve	+Ve	+Ve
	pullulan		-Ve	-Ve	-Ve	-Ve	-Ve	-Ve
	D- Raffinose		-Ve	+Ve	-Ve	+Ve	+Ve	-Ve
	O 129 resistance (c	comp. vibrio.)	-Ve	+Ve	-Ve	+Ve	+Ve	-Ve
	Salicin		+Ve	+Ve	+Ve	+Ve	+Ve	+Ve
	Sucrose		+Ve	+Ve	+Ve	+Ve	+Ve	+Ve
	D- Trehalose		+Ve	+Ve	+Ve	+Ve	+Ve	+Ve
	Arginine dihydrola	se	-Ve	+Ve	-Ve	-Ve	-Ve	-Ve
	optochin test resista		+Ve	+Ve	+Ve	+Ve	+Ve	+Ve

TABLE 6. Biochemical characterization of bacteria isolated from fermented milk using cereals and legumes.

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Isolation and characterization of bacteria isolated from fermented milk

From each fermented sample, one bacterial strain, which is the cause of success fermentation, was isolated and identified by Vitek 2 compact Automated Expert System (AES) was used for metabolic profiling. Using the Vitek 2 ID-GP card The morphological and physiological characteristics of the bacterial isolates were compared with the data from Bergey's Manual of Determinative Bacteriology (Holt et al., 1994). The results exhibit that Staphylococcus vitulinus isolated from faba bean, green wheat "Freekeh and wheat, Enterococcus faecium isolated from lentils, and Staphylococcus lentus isolated from chickpea and sesame. Staphylococcus vitulinus and S. pulvereri were first described as novel Staphylococcus spp. in 1994 (Webster et al., 1994) cited in Nam et al. (2012). Staphylococcus lentus is a member of the Staphylococcus sciuri group, which are Gram-positive, oxidase-positive, and coagulase-negative cocci, Staphylococcus *lentus* F1142, was isolated from doenjang, which is a Korean soybean paste (Nam et al., 2012). Enterococci, especially Enterococcus faecalis and Enterococcus faecium, have been studied in Gaborone, Botswana for the occurrence, and antibiotic resistance of milk, beef, and chicken. Enterococci were isolated from these sources using API 20 Strep kits with bile esculin agar (Chingwaru et al., 2003).

Conclusion

Fayesh is one of the most famous Egyptian breads which have extended to all Arab kitchens. It mainly depends on using fermented milk using cereals and legumes at room temperature (~28 °C). we can conclude that 40 °C was the optimum temperature for milk fermentation using whole or crushed cereals or legumes. The best cereals and legumes (whole or crushed) for milk fermentation are faba bean, lentil, green wheat, and chickpea. Increasing the added amount of cereals or legumes, > to 10 g, or mixing them does not significantly improve the time of milk fermentation (P < 0.05). Staphylococcus vitulinus, Staphylococcus lentusor Enterococcus faecium, which have been isolated from fermented milk, may have been responsible for the fermentation process. In the future, we need other studies to activate these strains and adding them as starter cultures, as well as to study the optimal conditions for these cultures and the properties of the resultant product.

References

- Achi, O. K. and Ukwuru, M. (2015) Cereal-Based Fermented Foods of Africa as Functional Foods. *International Journal of Microbiology and Application*, 2 (4),71-83.
- AOAC (2007) Official Methods of Analysis of the Association of Official Analytical Chemists.18th ed., Association of Official Analytical Chemists, Washington, DC., USA.
- APHA (1976) American public Healthy Association of Methods for the Microbiological Examination of Foods. Speck, M. L. ed., Washington, D. C., USA.
- Arendt, E.K., Ryan, L.A.M. and Dal Bello, F. (2007) Impact of sourdough on the texture of bread. *Food Microbiology*, 24, 165-174.
- Atlas, R.A. (1993) Handbook of Microbiological Media. CRC Press, Ann Arbor.
- Barros. R.R., Carvalho, G.S., Peralta, J.M. and Facklam, R.R. (2001)Teixeira LM. Phenotypic and Genotypic Characterization of Pediococcus Strains Isolated from Human Clinical Sources. *Journal Clinical Microbiology*, **39**, 1241-1246.
- Bergey's (1994) Manual of Determinative Bacteriology, 9th ed.; J.G. Holt et al.: Williams & amp; Wilkins: Baltimore, MD, USA, 175–190,Eds.
- Bille, J., Catimel, B., Bannerman, E., Jacquet, C., Yersin, M.N., Caniaux, I., Monget, D.andRocour,t J. (1992) API Listeria, a New and Promising One-Day System to Identify Listeria Isolates. *Applied* and Environmental Microbiology, 58, 1857-1860.
- Blandino, A., Al-Aseeri, M.E., Pandiella, S.S., Cantero, D. and Webb, C. (2003) Cereal-based fermented foods and beverages. *Food Research International*, 36(6),527–543.
- Bol, J., and De Vos, W.M. (1997) Fermented foods: an overview. In: J. Green (Ed.), *Biotechnological Innovations in Food Processing* (pp. 45–76) Oxford: Butterworth-Heinemann.
- Calvel, R. (2001) *The Taste of Bread*. Gaithersburg, Md: Aspen Publishers.
- Caplice, E. and Fitzgerald, G.F. (1999) Food fermentations: Role of microorganisms in food production and preservation. *International Journal* of Food Microbiology, **50**, 131–149.
- Carsanba,E., Akca, I. and Timur, M. (2017) Examination of firik produced in hatay region in terms of nutritional aspect. *GIDA*,42 (6), 726-730.

Egypt. J. Food Sci. 48, No.1 (2020)

- Chingwaru W, Mpuchane S.F. andGashe B.A.(2003) *Enterococcus faecalis* and *Enterococcus faecium* isolates from milk, beef, and chickenand their antibiotic resistance. *Journal Food Protection*, **66** (6), 931-936.
- Collins, M.D. and Lawson, P.A. (2000) The genus Abiotrophia (Kawamura et al.) is not monophyletic:proposal of Granulicatella gen. nov.,Granulicatellaadiacens comb nov., Granulicatellaelegans comb. nov. and Granulicatellabalaenopterae comb. nov... International Journal of Systematic and Evolutionary Microbiology, 50, 365-369.
- Davis, K.E., Joseph, S. J., and Janssen P. H. (2005) Effects of growth medium, inoculums size, and incubation time on the culture ability and isolation of soil bacteria. *Applied Environmental Microbiology Journal*, **71**,826-834.
- Difco-Manual (1984) Dehydrated culture media and reagents microbiological and clinical laboratory procedures, Pub-Difco-Lab-Detroits Michigan, USA.
- El-Gindy, S.M. (1983) Fermented foods of Egypt and Middle East. *Journal of Food Protection*, 46, 358-367.
- Esposito, M.A. (2003) Ciao Italia in Tuscany: traditional recipes from one of Italy's most famous regions.94.
- Fišteš,A., Došenovic,T., Rakic, D., Pajin, B., Šereš, Z., Simovic,Š. and Loncarevic, I. (2014) Statistical analysis of the basic chemical composition of whole grain flour of different cereal grains. *ActaUniversitatis Sapientiae-Alimentaria*, 7,45-53.
- Hammes, A., Willnow, T.E., Andreassen, T.K., Spoelgen, R., Raila, J., Metzger, J., Schweigert, F.J., Luppa, P.B., Nykjaer, A., Huebner, N. and Schultz, H. (2005) Role of Endocytosis in Cellular Uptake of Sex Steroids. *Cell*, **122**, 751–762.
- Hammes, W.P. (1990) Bacterial starter cultures in food production. *Food Biotechnology*, 4, 383–397.
- Harlander, S.K. (1992) Genetic Improvement of Microbial Starter Culture. In: *Application of Biotechnology to Traditional Fermented Foods*. Report of an Ad-Hoc Panel of the Board on Science and Technology for Intl Dev. National Acad. Press. Washington, D.C: 20-26.
- Hefnawy, H.T.M. (2011) Effect of processing methods on nutritional composition and anti-nutritional factors in lentils (*Lens culinaris*). *Annals of Agricultural Sciences*, 56, 61-65.
- Egypt. J. Food Sci. 48, No.1(2020)

- Holt, J.G., Krieg, N.R., Sneath, P.H.A., Staley, J.T. and Williams S.T. (1994) Bergey's Manual of Determinative Bacteriology, 9th ed. Williams and Wilkins, Baltimore, Maryland.
- Holzapfel, W.H. (2002) Appropriate starter culture technologies for small-scale fermentation in developing countries. *International Journal of Food Microbiology*, **75**, 197–212.
- Holzapfel, W.H., (1997) Use of starter cultures in fermentation on a household scale. *Food Control*, 8: 241–258.
- Hui, Y.H., Cork, H.,Leyn, I.D., Nip W.K. and Cross, N.A. (2006) Bakery products: science and technology. Oxford: Blackwell. 551.
- Janssen, P.H., Yates, P.S. Grinton, B.E., Taylor, P.M. and Sait, M. (2002) Improved culture ability of soil bacteria and isolation in pure culture of novel membersof the divisions Acidobacteria, Actinobacteria, Proteobacteria, and Verrucomicrobia. *Applied Environmental Microbiology Journal*, 68, 2391-2396.
- Joshi, V.K., Garg, V., and Abrol, G.S. (2012) Indigenous fermented foods. In: Joshi, V.K. and Singh, R.S. (Eds.). *Food Biotechnology: Principles and Practices.* IK Publishing House, New Delhi, pp. 337–373.
- Khachatourians, G. G. (1994) Food Biotechnology: Microorganisms. New York: Wiley-Interscience, 799– 813.
- Madsen, K., Cornish, A., Soper, P., Mckaigney, C., Jijon, H., Yachimec, C., Doyle, J, Jewell, L and Desimone, C.(2001) Probiotic bacteria enhance murine and human intestinal epithehal barrier function. *Gastroenterology*, **121**(3), 580-591.
- Mbata, T.I., Ikenebomeh, M. J. and Alaneme, J.C. (2009) Studies on the microbiological, nutrient composition and antinutritional contents of fermented maize flour fortified with bambara groundnut (*Vigna subterranean L*). *African Journal* of Food Science, **3** (6), 156 – 171.
- Metha, B., Kamal-Eldin A. and Iwanski, R.Z. (2012) Fermentation effects on food properties. CRC, Boca Raton, 385. *Microbiology*, **71**:826-834.
- Nam, Y.D., Chung, W.H., Seo, M.J. and Lim, S.I. (2012) Draft genome sequence of *Staphylococcus vitulinus* F1028, a strain isolated from a block of fermented soybean. *Journal of Bacteriology*,**194** (21), 5961–5962.

- Nzikou, M., Matos, L. Bouanga, G. K., Kalou, C. B., Ndangui, N. P., Pambou, G., Kimbonguila, A., Silou, T. Linder, M. and Desobry, S. (2009) Chemical composition on the seeds and oil of sesame (*Sesamum indicum L.*) grown in Congo-Brazzaville. *Advance Journal of Food Science and Technology*, 1, 6-11.
- Reinhart, P. (1998) Crust & Crumb: Master Formulas for Serious Bakers. Berkeley, Calif: Ten Speed Press. p. 38.
- Renneberg, R. and Demain, A. L. (2007) *Biotechnology for Beginners*. New York and London: Academic Press.
- Rolle, R. and Satin, M. (2002) Basic requirements for the transfer of fermentation technologies to developing countries. *International Journal of Food Microbiology*, **75**, 181–187.
- Ross, R.P., Morgan, S. and Hill, C. (2002) Preservation fermentation: past, present and future. *International Journal of Food Microbiology*, 79, 3–16.
- SAS (2008) The SAS system for Windows, release 9.2.SAS Institute, Cary, N.C. USA.
- Scheirlinck, I., Van Der M. R. and Van Schoor, A. (2008) Taxonomic structure and stability of the bacterial community in Belgian sourdough ecosystems as assessed by culture and population fingerprinting. *Applied and Environmental Microbiology*,74 (8), 2414–2423.

- Soni, S.K. and Sandhu, D.K. (1990) Indian fermented foods: Microbiological and Biochemical aspects. *Indian Journal of Microbiology*, **30** (2), 135–137.
- Tamang, J.P. (2010) Diversity of fermented foods. In: Tamang, J.P. and Kailasapathy, K. (Ed.). *Fermented Foods and Beverages of the World*. CRC Press, Taylor &Franci Groups, Boca Raton, FL, p: 41–84.
- Thakur, N., Savitri and Bhalla, T.C. (2004) Characterization of some traditional foods and beverages of Himachal Pradesh. *Indian Journal Traditional Knowledge*, 3 (3), 325-335
- Uzogara, S.G., Agu, L.N., Uzogara, E.O. (1990). A review of traditional fermented foods, condiments and beverages in Nigeria: Their benefits and possible problems. *Ecology of Food* and Nutrition, 24,267-288.
- Young, L. and Cauvain, S. P. (2007) Technology of Bread Making. Berlin: Springer. p. 88.
- Webster, J.A.(1994) Identification of the Staphylococcus sciuri sp ecies group with EcoRI fragments containing rRNA sequences and description of Staphylococcus vitulus sp. nov. International Journal of Systematic Bacteriology, 44, 454 – 460.
- Xu, Y., Thomas, M., and Bhardwa, H. (2014) Chemical composition, functional properties and microstructural characteristics of three kabuli chickpea (*Cicer arietinum L.*) as affected by different cooking method. *International Journal of Food Science and Technology*, **49**, 1215–1223