# Comparative study of the functional coupling of human chorionic gonadotropin and ca<sup>2+</sup> activated potassium channels on myometrial contractility in different trimesters of pregnancy

Zeinab Abdel Hafeez Alrefaie<sup>1</sup>, Lobna Abdel Aal Kassem<sup>2</sup> and Nahed Salah-Aldin<sup>2</sup> <sup>1</sup>Lecturer, <sup>2</sup>Assistant Professor, Department of Physiology, Kasr Al Aini -Faculty of Medicine, Cairo University

## ABSTRACT

Human chorionic gonadotropin, hCG, is central to the regulation of uterine quiescence during pregnancy. The present study was designed to investigate if the relaxant effect of hCG would vary between the different trimesters of pregnancy, whether this effect is linked functionally to the  $Ca^{2+}$  activated potassium channels,  $BK_{Ca}$ , and if this link would differ between the different stages of pregnancy. Methods: 32 female rats were used in the present study, 8 non-pregnant, 8 pregnant at day 4-5, 8 pregnant at day 11-12 and 8 pregnant at day 20-21, representing the 3 trimesters of pregnancy. Frequency and amplitude of isometric contractions were recorded from uterine strips from each group. These recordings were obtained either from spontaneous contractions, subgroup 1; contractions after addition of 20 iu hCG, subgroup 2; contractions after addition of 10 mM  $BK_{Ca}$  channels blocker, tetraethyl ammonium TEA, subgroup 3 and contractions after addition of both drugs, subgroup 4. Results: Our results showed that frequency and amplitude of spontaneous uterine contractions were significantly decreased in all trimesters of pregnancy when compared to non-pregnant group. The results also revealed that hCG significantly reduced frequency and amplitude of contraction in all groups and this reduction was more significant in the  $2^{nd}$  and  $3^{rd}$  trimesters of pregnancy. Both parameters of contraction were significantly increased by TEA, while adding both drugs did not alter frequency or amplitude of contraction in both non-pregnant and 1<sup>st</sup> trimester pregnant strips, but significantly decreased both of them in  $2^{nd}$  and  $3^{rd}$  trimesters pregnant strips. Conclusion: Our findings outlined that hCG exerted a more significant uterorelaxant effect in the late stages of pregnancy and that activation of  $BK_{Ca}$  channels may explain this relaxant effect.

*Key words: Myometrial contraction – rat – human chorionic gonadotropin – calcium activated potassium channels.* 

#### INTRODUCTION

An increased knowledge of the molecular and physiological mechanisms implicated in the control

of myometrial excitability and contractility is essential to the development of effective tocolytic that be agents can used in management of preterm labor which is largest cause of perinatal the

morbidity and mortality, particularly in developing countries.

Myometrial tissue is spontaneously myogenic and can generate slow wave activity as well as simple and complex action potentials. Changes in wave form of myometrial electrical activity and transition from quiescence to excitability at term, suggests extensive fluidity and remodeling of ion channels contributing to the dramatically altered excitability towards time of labor<sup>(1)</sup>.

Human chorionic gonadotropin [hCG], is a hormone produced primarily by the placenta to stimulate progesterone production by the corpus luteum in the early pregnancy<sup>(2)</sup>. It is a member of a glycoprotein hormone family that consists of leutinising hormone [LH], follicle stimulating [FSH], hormone and thyroid stimulating hormone [TSH]. The members of this family are heterodimers, with similar  $\alpha$ -subunits and dissimilar  $\beta$ -subunits<sup>(3)</sup>. hCG and LH have similar biological actions and bind to a common receptor, which is a member of a G-protein coupled family of receptors<sup>(4)</sup>. The hCG/LH receptors are present in the endometrial and myometrial layers of the uterus, fallopian tubes, gonads, fetal membranes, deciduas, placenta and umbilical cord<sup>(5)</sup>.

Unlike the previous belief that human myometrium is an indirect target of hCG via ovarian steroid hormones, studies have shown that it is a direct target of hCG regulation as the myometrium contains hCG receptors and exogenous hCG was found to directly increase myometrial hyperplasia and hypertrophy<sup>(6)</sup>. hCG also inhibited oxytocin stimulated myometrial contraction<sup>(7)</sup>. The importance for the enhancement of fertility, successful implantation and survival of the conceptus in early gestation is recognized, however, studies conducted worldwide in recent years indicate that hCG may also play a significant role in maintaining pregnancy well after the first trimester raising the question about its potential future role for tocolysis in human preterm labor<sup>(8)</sup>.

 $K^+$  channels have a widespread distribution in virtually all cell types. Their main function is to dampen cellular excitability by maintaining the cell membrane potential close to the equilibrium potential of  $K^+$  ions. By the generation of an outward  $K^+$ current, these channels produce hyperpolarization or repolarization thereby terminating or rendering action potential generation and ultimately contraction less likely<sup>(9)</sup>.

Ca<sup>2+</sup> activated potassium channels are a class of potassium channels that are activated when intracellular Ca<sup>2+</sup> levels rise. There are three subclasses of calcium-activated potassium channels, the large conductance channels, denoted BK<sub>Ca</sub> or maxi K, are probably the best characterized and most ubiquitous being expressed in all tissues with the exception of the heart. Two other subclasses are the intermediate, IK<sub>Ca</sub> and the small,  $SK_{Ca}$  conductance calcium-activated K<sup>+</sup> channels but these have not been described in any detail in the human myometrium<sup>(10)</sup>.

The  $BK_{Ca}$  or maxi K channel is the predominant  $K^+$  channel type encountered in non-pregnant and pregnant human myometrium<sup>(11,12)</sup>.

Channels belonging to this class are  $Ca^{2+}$  and voltage-dependent with channel open-state probability increasing with depolarization. BK<sub>Ca</sub> channels are associated with a variety of cellular functions including control of smooth muscle tone and neuronal firing<sup>(13)</sup>.

BK<sub>Ca</sub> channels exist in vivo as a complex of a-subunit (a 125 kDa protein) and a 31 kDa glycosylated regulatory protein, the  $\beta$ -subunit<sup>(14)</sup>. The mechanisms by which BK<sub>Ca</sub> channels sense Ca<sup>2+</sup> remain an enigma. Meera et al.<sup>(15)</sup> have reported that the myometrial  $\beta$ -subunit is functionally uncoupled from the  $\alpha$ subunit at resting intracellular Ca2+ levels and that as Ca<sup>2+</sup> rises inside the cell the  $\beta$ -subunit switches to acting as a  $Ca^{2+}$  sensor. In addition to the role of the  $\beta$ -subunit, the Ca<sup>2+</sup> sensing function of the BK<sub>Ca</sub> channel may also be attributable to the string of negatively charged aspartate residues that constitute the  $Ca^{2+}$  bowel located in the  $\alpha$ - subunit tail region<sup>(13)</sup>.

Modulation of BK<sub>Ca</sub> channels by hCG in Leydig cells from mature mouse testis was documented<sup>(16)</sup>, while still conflicting reports have appeared concerning the effects of hCG on these channels in the uterus. Moreover, their role in controlling myometrial contractility during pregnancy is still not clear.

In the present study, we addressed the question of whether the uterorelaxant effect of hCG on myometrial contractility would differ between different stages of pregnancy. Also the involvement of  $BK_{Ca}$  in such relaxant effect would be assessed and compared in the three trimesters of pregnancy.

## MATERIAL & METHODS

#### Animals:

Twenty four pregnant and 8 non pregnant rats of the local strain were used in the present study. They were brought from the Eye Research Institute and kept in the animal house of Kasr Al-aini faculty of Medicine, they had food and water ad libitum.

## Experimental procedure:

Pregnant and non-pregnant rats were killed by decapitation. The preparation of myometrial strips followed the method of Karsli et al. (2003)<sup>(17)</sup>. A full-length longitudinal abdominal incision was made to expose both uterine horns which were rapidly excised, carefully cleaned of surrounding connective tissue and opened longitudinally along the mesenteric border. Fetuses of the latestage pregnant rats were removed and non-uterine tissues were dissected and discarded. We obtained 4 longitudinal full-thickness myometrial muscle strips, measuring 4.5 mm in length and 1.5 mm in width, from each animal.

The strips were mounted vertically, one end of the strip was connected to the lower hook of the bath and the other end of the strip was connected to a force transducer. The strips were incubated in a 20 ml tissue Krebs-Henseleit bath containing physiological solution (composition in KCL 4.6; MgSO<sub>4</sub> mM: 1.16: NaH<sub>2</sub>PO<sub>4</sub> 1.16; CaCl<sub>2</sub> 2.5; NaCl 115.5; NaHCO<sub>3</sub> 21.9; and glucose 11.1.), which were aerated continuously with 95% oxygen and 5% carbon dioxide. The pH was kept at 7.4 and the temperature was

maintained at 37°C. The solution was prepared daily for each experiment. Myometrial strips of rats were allowed to equilibrate for 20 min. The characteristics of the contraction including frequency and amplitude were recorded by an isometric force transducer of the Intracel computer using analyzing software Phys. 4 (Intracel company; England). When the contractions of the strips became

regular, the contraction frequency and amplitude were recorded as baseline activity, then the following drugs, hCG (1 iu/ ml bath solution), TEA (10 mM) as well as hCG in the presence of TEA were added according to the experimental protocol. Each strip was exposed to only one agent. Drugcontaining solutions were prepared immediately before the experiment.

#### **Experimental protocol:**

A total of 128 myometrial strips were obtained from pregnant and non pregnant rats, and were randomly assigned to 1 of 4 main groups (n=32 strips, each group): Group I: non-pregnant group

**Group II:** pregnant 1<sup>st</sup> trimester group (at day 3 - 4 of pregnancy). **Group III:** pregnant 2<sup>nd</sup> trimester group (at day 11-12 of pregnancy). **Group IV:** pregnant 3<sup>rd</sup> trimester group (at day 20- 21 of pregnancy)<sup>(18)</sup>.

After spontaneous myometrial contractions were induced in the Krebs-Henseleit physiological solution, each group was further subclassified into 4 subgroups (n=8 strips, each subgroup):

Subgroup 1, control group: frequency and amplitude of contraction were recorded 20 minutes after equilibration without addition of any drugs to the bathing solution.

Subgroup 2, hCG group: frequency and amplitude of contraction were recorded 20 minutes after addition of 1 iu/ ml hCG to the bathing solution<sup>(19)</sup>.

Subgroup 3. TEA group: frequency and amplitude of contraction were recorded 20 minutes after addition of 10 mM tetraethyl ammonium [TEA] to the bathing solution<sup>(20)</sup>.

Subgroup 4, hCG and TEA group: TEA was added to the bathing solution 20 minutes before adding hCG and after another 20 minutes, recordings of frequency and amplitude of contraction from uterine strips were carried out.

Drugs: hCG was purchased from Organon Co- Switzerland. TEA was purchased from Sigma Chemical Co., St Louis, Mo, USA hCG and TEA were dissolved in distilled water.

#### **Statistical Analysis:**

The results were expressed as mean values ± SD. Comparisons between subgroups in each main group or between subgroups in different main

groups were carried out by one-way analysis of variance (ANOVA) for multiple comparisons. A P value < 0.05 was regarded as statistically significant.

## RESULTS

Results are shown in tables (1- 3) and figures (1- 10).

Spontaneous myometrial contractions were less frequent and having less amplitude in pregnant uterine non treated strips from any of the three trimesters when compared with non-pregnant, non treated strips. P<0.001 (pregnant 1<sup>st</sup>, 2<sup>nd</sup>, and 3<sup>rd</sup> trimesters versus non-pregnant) as regard the frequency of contraction, P<0.005 (pregnant 1<sup>st</sup> and  $2^{nd}$ trimesters versus non-pregnant) and P<0.001 (pregnant 3<sup>rd</sup> trimester versus non-pregnant) as regard the amplitude of contraction.

Effect of hCG on the contractility of isolated uterine muscle strips from pregnant and

non-pregnant female rats:

Comparing the frequency of contraction between subgroups in each main group revealed that addition of hCG to the bathing solution significantly decreased frequency of contraction in uterine strips from all groups when compared with non treated control strips in each corresponding group (P<0.05 in non-pregnant strips). This decrease was more significant in pregnant than non pregnant strips.

As regards the effect of hCG on amplitude of contraction, it reduced amplitude of contraction in nonpregnant strips but this reduction did not reach statistical significance. hCG significantly decreased amplitude of contraction in 1<sup>st</sup>, 2<sup>nd</sup> and 3<sup>rd</sup> trimester pregnant strips, P< 0.005, P<0.05 and P< 0.001 respectively when compared to non treated strips in each group.

Effect of TEA on the contractility of isolated uterine muscle strips from pregnant and

non-pregnant female rats:

The large conductance  $Ca^{2+}$  sensitive K<sup>+</sup> channels blocker, tetraethyl ammonium, TEA, was used to study the role of these channels in regulating basal contractility in myometrial smooth muscle. We found that TEA significantly increased frequency of contraction in non pregnant, 1<sup>st</sup>, 2<sup>nd</sup> and 3<sup>rd</sup> trimester pregnant strip P< 0.001, P<0.001, P<0.001 and P<0.005 respectively.

The same finding was obtained on observing the effect of TEA on amplitude of contraction, it significantly increased amplitude of contraction in all uterine strips P<0.01, P<0.005, P<0.5 and P<0.05 respectively when compared with control non-treated strips in each group.

Effect of TEA on hCG induced relaxations in isolated uterine muscle strips from pregnant and non-pregnant female rats:

In an attempt to examine whether the blockade of the large conductance Ca<sup>2+</sup> sensitive K<sup>+</sup> channels interferes with hCG-induced relaxation of myometrial smooth muscles, incubation of uterine strips with TEA before addition of hCG to the bathing solution was performed. We obtained different responses on frequency and amplitude of contraction in different trimester strips. Both drugs when added together did not alter any of the two parameters of contraction in nonpregnant and 1<sup>st</sup> trimester pregnant uterine strips.

On the other hand in  $2^{nd}$  and  $3^{rd}$  trimester pregnant uterine strips, adding hCG and TEA together decreased frequency of contraction which was significant in  $3^{rd}$  trimester uterine strips P<0.05, when compared

to control non treated uterine  $3^{rd}$  trimester strips. Both drugs together reduced the amplitude of contraction in  $2^{nd}$  and  $3^{rd}$  trimester pregnant strips and this reduction was significant in both trimesters, P< 0.05 and P< 0.001.

Table	1:	frequency	(cycles/	min.)	and	amplitude	(screen	units)	of	contraction	of
uterine	e str	rips from co	ontrol nor	1-pregr	nant,	pregnant 1s	<sup>st</sup> , 2 <sup>nd</sup> and	d 3 <sup>rd</sup> tr	imes	sters.	

	Control non- pregnant	Pregnant 1 <sup>st</sup> trimester	Pregnant 2 <sup>nd</sup> trimester	Pregnant 3 <sup>rd</sup> trimester
Frequency	$1.19 \pm 0.12$	$0.89 \pm 0.17$ P < 0.001 *	$0.71 \pm 0.12$ P < 0.001 *	$0.86 \pm 0.17$ P < 0.001 *
Amplitude	19.25 ±2.12	15.63 ±1.92 P < 0.005 *	15.5 ±2.14 P < 0.005 *	$15 \pm 1.85$ P < 0.001 *

\* Significant when compared with non-pregnant control group.

**Table 2:** frequency (cycles/ min.) and amplitude (screen units) of contraction of uterine strips from non- pregnant and pregnant 1<sup>st</sup> trimester

	Non-pregnant				Pregnant 1 <sup>st</sup> trimester				
	Control	hCG	TEA	hCG +	Control	hCG	TEA	hCG +	
Frequency	1.19 ± 0.12	$1.03 \pm 0.13$ P < 0.05*	$1.64 \pm 0.14$ P < 0.001*	1.21 ± 0.12	0.89 ± 0.17	$0.58 \pm 0.18$ P < 0.005 *	1.29 ± 0.18 P < 0.001*	0.94 ± 0.17	
Amplitude	19.25 ± 2.12	16.75± 1.91	22.88 ±3.27 P < 0.01*	19.38 ±2.39	15.63 ± 1.92	12.5 ± 1.77 P < 0.005 *	19.13 ± 2.23 P < 0.005 *	14.25 ± 1.67	

\* Significant when compared with the corresponding control.

Control: no drugs added to the bathing solution.

hCG: hCG 20 iu were added to the bathing solution.

TEA: TEA 10 mM were added to the bathing solution.

hCG + TEA: both drugs were added to the bathing solution.

	Pregnant 2 <sup>nd</sup> trimester				Pregnant 3 <sup>rd</sup> trimester			
	Control	hCG	TEA	hCG + TEA	Control	hCG	TEA	hCG + TEA
Frequency	0.71 ± 0.12	$0.46 \pm 0.13$ P < 0.005*	$1.11 \pm 0.2$ P < 0.001*	0.6 ± 0.13	0.86 ± 0.17	$0.59 \pm 0.16$ P < 0.005*	$1.2 \pm 0.2$ P < 0.005*	0.66 ± 0.16 P<0.05*
Amplitude	15.5 ± 2.14	$12.63 \pm 1.92$ P < 0.05*	18.13 ± 2.79 P < 0.05*	$12.63 \pm 2.26$ P < 0.05*	15 ± 1.85	11.13± 1.25 P < 0.001*	16.88 ± 2.1 P < 0.05*	11.38 ± 1.41 P<0.001*

**Table 3:** frequency (cycles/ min.) and amplitude (screen units) of contraction of uterine strips from pregnant  $2^{nd}$  and  $3^{rd}$  trimesters

\* Significant when compared with the corresponding control.

Control: no drugs added to the bathing solution.

hCG: hCG 20 iu were added to the bathing solution.

TEA: TEA 10 mM were added to the bathing solution.

hCG + TEA: both drugs were added to the bathing solution.



Figure 1: frequency (cycles/ min.) and amplitude (screen units) of contraction of uterine strips from control non-pregnant, pregnant 1<sup>st</sup>, 2<sup>nd</sup> and 3<sup>rd</sup> trimesters \* Significant when compared with non-pregnant control group.



Figure 2: amplitude of contraction (screen units) of uterine strips from nonpregnant, pregnant  $1^{st}$ ,  $2^{nd}$  and  $3^{rd}$  trimesters

Control: no drugs added to the bathing solution.

hCG: hCG 20 iu were added to the bathing solution.

TEA: TEA 10 mM were added to the bathing solution.

*hCG* + *TEA*: both drugs were added to the bathing solution.

\* Significant when compared with the corresponding control.



Figure 3: frequency of contraction (cycles/ min.) of uterine strips from nonpregnant, pregnant  $1^{st}$ ,  $2^{nd}$  and  $3^{rd}$  trimesters

Control: no drugs added to the bathing solution.

hCG: hCG 20 iu were added to the bathing solution.

TEA: TEA 10 mM were added to the bathing solution.

*hCG* + *TEA*: both drugs were added to the bathing solution. \* Significant when compared with the corresponding control.

Significani when compared with the corresponding control.



trimester pregnant uterus



Figure 9: Effect of hCG + TEA on  $2^{nd}$  trimester pregnant uterus



3<sup>rd</sup> trimester pregnant uterus

## DISCUSSION

The uterorelaxant effect of human chorionic gonadotropin is regarded as an important mediator in maintenance of uterine quiescence during pregnancy, while the high conductance, calcium activated potassium channels were shown to play a pivotal role in modulating myometrial excitability.

We aimed to compare the involvement of  $BK_{Ca}$  channel function in the response of uterine contractility to hCG between different trimesters of pregnancy.

The results of the present study revealed that frequency and amplitude of contraction decreased significantly in pregnant strips when compared to non-pregnant ones.

Our results agree with many previous observations which stated that during pregnancy uterine contractility is normally held in abeyance till parturition, when it gets gradually activated giving rise to powerful rhythmic contractions



culminating in delivery of the fetus. The progesterone withdrawal theory was the earliest theory that explained shift in myometrial activity during parturition, it ascribed the relative quiescence of the pregnant uterus to the presence of progesterone and that the decline in its level at term releases from inhibition<sup>(21)</sup>. mvometrium Researches have proved that hCG<sup>(7)</sup> and  $Bk_{Ca}$  channels<sup>(1)</sup> may be involved in the mechanisms underlying the inhibition of uterine activity during pregnancy. The conversion of the silent pregnant uterus to highly excitable at term had been explained by many observations like the significant decrease in hCG receptors at labor myometrium tissue<sup>(22)</sup>, also the  $BK_{Ca}$  channels were found to be either absent or considerably altered and in their physiological pharmacological properties at labor, they have their identical conductance and K<sup>+</sup> selectivity but exhibit no Ca<sup>++</sup> voltage sensitivity, allowing or cytoplasmic Ca<sup>++</sup> levels to rise without activating a counteracting K<sup>+</sup> outward current<sup>(23)</sup>.

Changes in electrical activity of the uterus may be considered also in the dramatic change in myometrial contractility at term, Sims et al. (1982)<sup>(24)</sup> observed a decrease in myometrial cell resting potential in rats at term. Conversion of complex action potentials to simple spikes at labor was also documented<sup>(25)</sup>. Many compounds have been implicated also such as prostaglandins, oxytocin and heteromeric G protein family<sup>(26)</sup>.

The present study showed that hCG decreased frequency and amplitude of contraction in non pregnant and pregnant uterine strips and that inhibitory effect was more significant in the  $3^{rd}$  trimester pregnant strips. This suggests that the quiescent effect of hCG on the myometrium gets more prominent in pregnant uteri in the  $3^{rd}$  trimester.

In accordance with our suggestion is that of Slattery et al.  $(2001)^{(18)}$  who highlighted a potential role of hCG in maintaining uterine quiescence in the 3<sup>rd</sup> trimester.

In addition, in the study of Rzucidlo et al.  $(1998)^{(27)}$ , the authors measured the unoccupied binding sites of hCG receptors in the porcine myometrial tissue at different stages of pregnancy, it was significantly higher in the 1<sup>st</sup> trimester than 2<sup>nd</sup> and 3<sup>rd</sup> trimester pregnant uteri, this means that more receptors were occupied in the 2<sup>nd</sup> and 3<sup>rd</sup> trimesters, which supports our observation in the present study that hCG has a more prominent effect in the 2<sup>nd</sup> and 3<sup>rd</sup> trimester pregnant uterine strips.

Blocking the BK<sub>Ca</sub> channels in our experiment enhanced contractility of non-pregnant and pregnant uterine strips, as it allows intracellular accumulation of Ca<sup>2+</sup> without K<sup>+</sup> outflow, i.e. depolarizing the cells and hence increasing contractile activity. This finding concurs with the data from the report of Sanborn, (2000)<sup>(28)</sup>, who stated that BK<sub>Ca</sub> channels blocking increased contractile activity in rat and human myometrium. Also the study of Kafali et al. (2002)<sup>(29)</sup> who observed that TEA increased spontaneous myometrial contraction in pregnant rats.

The other important finding in the present work, is that addition of hCG with the Ca<sup>2+</sup> sensitive  $K^+$  channel blocker, TEA, did not alter frequency

and amplitude of contraction in nonpregnant and  $1^{st}$  term pregnant uteri, but significantly decreased the same parameters in  $2^{nd}$  and  $3^{rd}$  trimester pregnant myometrial strips when compared to control non treated strips in each group.

According to the report of Doheny et al.  $(2003)^{(30)}$ , who observed that hCG increased open state probability of the BK<sub>Ca</sub> channels and blocking of these channels significantly attenuated the relaxant effect exerted by hCG in human myometrium of pregnant women undergoing cesarean section. It was expected in our study that incubation of uterine strips with TEA would block the relaxant effect of hCG, this was observed in non-pregnant and 1<sup>st</sup> trimester pregnant strips, while in the 2<sup>nd</sup> and 3<sup>rd</sup> trimester pregnant strips addition of TEA did not block the effect of hCG which still significantly inhibited uterine contractility, this suggests that hCG exerts stronger effect in 2<sup>nd</sup> and 3<sup>rd</sup> trimesters which could be mediated by other mechanisms than  $BK_{Ca}$  channels.

The discrepancy between the observation of Doheny et al, that  $BK_{Ca}$  channel blocking could block the inhibitory effect of hCG on myometrial contraction at those females undergoing cesarean section, and our observation that TEA did not exert the same action in the 2<sup>nd</sup> and 3<sup>rd</sup> trimesters may be attributed to the fact that the myometrial strips in their study were taken from uteri at term which are expected to have fewer hCG receptors with unremarkable effect for hCG<sup>(22)</sup>.

The suggested mechanisms for the uterorelaxant effect of hCG

included alterations in myometrial gap junctions and hence the electrical coupling between muscle cells, hCG was found to decrease connexin 43, an integral protein in myometrial gap junction, in addition it decreased the assembly of gap junctions and also permeability in their cultured myometrial cells<sup>(31)</sup>. hCG was found to inhibit uterotropic eicosanoids<sup>(22)</sup> and reduce intracellular calcium availability<sup>(32)</sup>. Data also showed that hCG may mediate its action through increasing myometrial biosynthesis of progesterone in rats<sup>(33)</sup>. By these suggested mechanisms hCG can exert its uterorelaxant effect despite the block of BK<sub>Ca</sub> channels.

In some groups in our study, the effects of the drugs used, on either frequency or amplitude of contraction, were not exactly the same. It is known that whether excitatory or inhibitory compounds can alter the frequency or amplitude of contraction or even the baseline of the recording. Quite often low concentrations produce changes frequency, while intermediate in concentrations can affect amplitude of contraction and higher concentrations may change the baseline<sup>(34)</sup>. As we implemented one concentration for each drug in our study, this could explain that frequency and amplitude of contraction did not always show the same significance in change by each drug used.

**Conclusion:** the present study pointed to the highly significant uterorelaxant effect of hCG in the  $3^{rd}$  trimester. It also confirmed that activation of BK<sub>Ca</sub> channels could be one of the mechanisms underlying this uterorelaxant effect.

### REFERENCES

- 1- Khan RN, Ball MB, Arulkumaran S and Ashford MLJ. Potassium channels in human myometrium. Experimental Physiology 2001, 86 (2): 255-264.
- 2- Webley GE, Luck MR and Hearn JP. Stimulation of progesterone secretion by cultured human granulose cells with melatonin and catecholamines. J Reprod Fertil 1988, 84: 669-677.
- **3- Pierce JG and Parsons TF.** Glycoprotein hormones: structure and function. Annu Rev Biochem 1981, 50: 466-495.
- 4- Gam L and Latiff A. SDS-PAGE Electrophoretic Property of Human Chorionic Gonadotropin (hCG) and its βsubunit. Int J Biol Sci 2005, 1(3): 103–109.
- 5- Reshef E, Lei ZM, Rao ChV, Pridham DD, Chegini N and Luborsky JL. The presence of gonadotropin receptors in nonpregnant human uterus, human placenta, fetal membranes and deciduas. J Clin Endocrinol Metab 1990, 70: 421-430.
- 6- Kornyei J, Lei ZM and Rao Ch V. Human myometrial smooth muscle cells are novel targets of direct regulation by human chorionic gonadotropin. Biol Reprod 1993, 49:1149-1157.
- 7- Eta E, Ambrus G and Rao Ch V. Regulation of human myometrial contractions by human chorionic gonadotropin. J Clin Endocrinol Metab 1994, 79 (6): 1582-1586.

- 8- Ticconi C, Piccione E, Belmonte A, Rao Ch V. HCG-A new kid on the block in prematurity prevention. J Matern Fetal Neonatal Med 2006, 19(11):687-92
- **9- Jan LY and Jan YN.** Structural elements involved in specific K+ channel functions. Annual Review of Physiology 1992, 54: 537-555.
- **10- Vergara C, Latorre R, Marrion NV and Adelman JP**. Calcium activated potassium channels. Current Opinion in Neurobiology 1998, 8: 321-329.
- 11- Perez GJ, Toro L, Erulkar SD, and Stefani E. Characterisation of large-conductance, calciumactivated potassium channels from human myometrium. Am J Obstet Gynecol 1993, 168: 652-660.
- 12- Khan RN, Smith SK, Morrison JJ and Ashford MLJ. Properties of large conductance potassium channels in human myometrium during pregnancy and labor. Proceedings of the Royal Society B 1993, 251: 9-15.
- **13-** Schreiber M and Salkoff L. A novel calcium-sensing domain in the BK channel. Biophysical Journal 1997, 73: 1355-1363.
- 14- Knaus HG, Garcia-Calvo M, Kaczorowski G and Garcia M. Subunit composition of the high conductance calcium activated potassium channel from smooth muscle; a representative of the mslo and slowpoke family of potassium channels. J Biol Chem 1994b, 269: 3921-3924.
- 15- Meera P, Wallner M, Jiang Z and Toro L. A calcium switch

1994.

for the functional coupling between a (hslo) and  $\beta$ -subunit of (Kv, <sub>Ca</sub> $\beta$ ) of maxi K channels. FEBS Letters 1996, 283: 84-88.

- 16- Desaphy J, Rogier C and Joffre M. Modulation of K+ conductances by Ca2+ and human chorionic gonadotrophin in Leydig cells from mature rat testis J Physiol. 1996,15;495(Pt 1): 23–35.
- 17- Karlsi B, Kayacan N, Kucukyavuz Z and Mimaroglu C. Effects of local anaesthetics on pregnant uterine muscle. Pol J Pharmacol 2003, 55 (1):51-56
- 18- Longo M, Jain V, Vedernikov YP et al. Effect of nitric oxide and carbon monoxide on uterine contractility during human and rat pregnancy. 1999, 181 (4): 981-988.
- 19- Slattery MM, Brennan C, O'Leary MJ and Morrison JJ. Human chorionic gonadotropin inhibition of pregnant human myometrial contractility. Br J Obstet Gynaecol 2001, 108 (7): 704-708.
- **20- Hughes SJ and Hollingsworth M.** The lack of a role for potassium channel opening in the action of relaxin in the rat isolated uterus; a comparison with levcromakalim and salbutamol. Br J Pharmacol 1996, 117(7): 1435-42.
- **21- Csapo AI.** Progesterone block. Am J Anat 1956, 98: 273-291.
- 22- Zuo J, Lei ZM and Rao Ch V. Human myometrial chorionic gonadotropin/ luteinizing hormone receptors in preterm and term deliveries. J Clin Endocrinol

metab 79 (3): 907-911.

- 23- Matharoo-Ball, Ashford MLJ, Arulkumaran S and Khan RN. Down regulation of the  $\alpha$  and  $\beta$ subunits of calcium activated potassium channel in human myometrium with parturition. Biology of reproduction 2003, 68: 2135-2141.
- 24- Sims SM, Daniel EE and Garfield RE. Improved electrical coupling in uterine smooth muscle is associated with increased numbers of gap junctions at parturition. J Gen Physiol 1982, 80: 353-375.
- 25- Bengtsson B, Chow EMH and Marshall JM. Activity of circular muscle of uterus at different times in pregnancy. A J Physiol 1984, 246: C216-C223.
- 26- Tahara M, Morishige K, Sawada K et al. RhoA/Rho kinase cascade is involved in oxytocin induced rat uterine contraction. Endocrinology 2002, 143(3): 920-929.
- 27- Rzucidlo SJ, Weigl RM and Tilton JE. Myometrial LH/hCG receptors during the estrous cycle and pregnancy in pigs. Animal Reproduction Science 1998, 51: 249-257.
- 28- Sanborn BM. Relationship of ion channel activity to control of myometrial calcium. J Soc Gynecol Investig 2000, 7: 4-11.
- 29- Kafali H, Kaya T, Gursoy S, Bagcivan I, Karadas B and Sarioglu Y. The role of K<sup>+</sup> channels on the inhibitor effect of sevoflurane in pregnant rat myometrium. Anesth Analg 2002, 94(1): 174-178.



- 30- Doheny HC, Houlihan DD, Ravikumar N, Smith TJ and Morrison JJ. Human chorionic gonadotropin relaxation of human pregnant myometrium and activation of the BK<sub>Ca</sub> channel. J Clin Endocrinol Metab 2003, 88 (9): 4310-4315.
- **31- Ambrusi G, Rao Ch V.** Novel regulation of pregnant human myometrial smooth muscle gap junctions by human chorionic gonadotropin. Endocrinol 1994. 135: 2772-2779
- 32- Singh M, Zuo J, Li X et al. Decreased expression of functional human chorionic

gonadotropin/luteinizing hormone receptor gene in human uterine leiomyomas. Biol Reprod 1995, 53: 591-597.

- **33-** Bonnamy P-J, Benhaim A and Leymarie P. Human chorionic gonadotropin affects tissue levels of progesterone and cyclic adenosine 3',5'-monophosphate in the metestrus rat in vitro. Biol Reprod 1989, 40: 511-516.
- **34-** Crankshaw DJ and Gaspar V. Effects of prostanoids on the rat's myometrium in vitro during pregnancy. Biology of reproduction 1992, 46: 392-400.

## تأثير العلاقة بين هرمون الحمل و قنوات البوتاسيوم التي تتأثر بالكالسيوم على انقباض الرحم في المراحل المختلفة للحمل

ان هرمون الحمل (hCG) يلعب دورا هاما فى انبساط عضلة الرحم اثناء الحمل، و قد صممت هذه الدراسة لبحث ما اذا كان تأثير الhCG على عضلة الرحم يختلف اثناء مراحل الحمل، كذلك اذا كانت هناك علاقة بين هذا التأثير و بين قنوات البوتاسيوم التى تتأثر بالكالسيوم و ما اذا كانت هذه العلاقة تختلف اثناء المراحل المختلفة للحمل اجريت هذه الدراسة على ٣٢ فأرة تم تقسيمها الى ٤ مجموعات، المجموعة الاولى ٨ غير حوامل، المجموعة الثانية ٨ حوامل فى اليوم ٢٤ مجموعة الثالثة ٨ حوامل فى اليوم ٢١-٢١ و المجموعة الرابعة ٨ حوامل فى اليوم ٢٠-٢. فى كل مجموعة تم قياس معدل و ارتفاع الانقباضات فى عضلة الرحم، اما الانقباضات التلقائية، او بعد اضافة ٠٠ وحدة دولية من الAGD ، او بعد اضافة ١٠ ميللى مول من الAED الذى يلغى عمل قنوات البوتاسيوم او بعد اضافة كلا من الAGD و المحموعة الرابع.

و قد أظهرت النتائج ان معدل و ارتفاع الانقباضات التلقائية في رحم المجموعات الحوامل كان اقل بدلالة احصائية من قيمتهما في رحم المجموعة غير الحوامل. كذلك اظهرت النتائج ان الhCG ادى الى انخفاض هذين المؤشرين في كل المجموعات و لكن الانخفاض كان أكثر دلالة احصائية في مجموعة اخر الحمل، بينما ادى ال TEA الى ارتفاع هذين المؤشرين ارتفاعا ذا دلالة احصائية في كل المجموعات. اظهرت النتائج ايضا ان اضافة كلا من الhCG و الAET ادى الى عدم تغير اى من معدل او ارتفاع انقباض عضلة الرحم في المجموعتين الاولى والثانية بينما أدى الى انخفاض ذى دلالة احصائية في هذين المؤشرين في مجموعتي الفئران الحوامل في وسط و اخر الحمل.

تشير نتائج هذه الدراسة الى ان الhCG لـه تأثير مثبط على عضلة الرحم و خاصة في اخر الحمل كذلك تشير الى ان قنوات البوتاسيوم التي تتأثر بالكالسيوم قد تلعب دورا في طريقة عمل ال hCG.