



## Prevalence of group B streptococcus among pregnant and non-pregnant women in Ismailia, Egypt

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### Abstract

*Streptococcus agalactiae* or group B streptococcus (GBS) is among the leading life-threatening infectious agents in neonates. It is also known to cause disease in pregnant women and immunocompromised patients. The aim of the current study was to determine GBS prevalence among pregnant and non-pregnant women in Ismailia, Egypt. Four-hundred sixty-two vaginal swabs were collected from pregnant and non-pregnant women. A total of 27.9% women were found as positive GBS carriers. In respect to pregnant participants, 26% of pregnant women were defined as asymptomatic GBS carriers versus 35.5% among non-pregnant participants. Age, parity, history of previous abortion or preterm labor were not significantly associated as potential risk factors for GBS carriage.

### 1. Introduction

*Streptococcus agalactiae* or group B streptococcus (GBS) is a commensal of the vagina and/or rectum of 10-30%

of pregnant women and up to 40% in non-pregnant adults (Schuchat, 1998; Verani et al., 2010). GBS is a principle etiological agent of sever neonatal infections such as sepsis, pneumonia, and meningitis (Schuchat, 1998; Verani et al., 2010).

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GBS infections manifest as early-onset disease during the first week of life and late-onset disease during the first three months after birth (Schuchat, 1998; Lindahl et al., 2005). GBS is also an important cause of disease in pregnant women, elderly, and immunocompromised patients with underlying conditions (Schuchat, 1998; Lindahl et al., 2005).

The most important risk factor of GBS transmission is maternal carriage (Verani et al., 2010). Vertical transmission during or after labor is the most predominant route of transmission (Verani et al., 2010). In 2010, the Centers for Disease Control and Prevention (CDC) (Verani et al., 2010) released revised guidelines which recommended culture-based GBS screening at 35 to 37 weeks of gestation followed by intrapartum antibiotic prophylaxis for positive carriers. Implementation of CDC guidelines resulted in significant decline in neonatal GBS disease in the US (Schrage and Verani, 2013).

Except for very few studies (Elbaradie et al., 2009; Shabayek et al., 2009; Shabayek et al., 2010; Shabayek and Abdalla, 2014; Shabayek et al., 2014), epidemiological data on GBS carriage and neonatal disease in Egypt still sparse. The aim of the current study was to determine GBS prevalence

among pregnant and non-pregnant women and evaluate potential risk factors associated with GBS carriage.

## **2. Materials and Methods**

### **2.1 Ethical approval**

All procedures involving human subjects were approved by the Research Ethics Committee at the Faculty of Pharmacy, Suez Canal University, Egypt (Reference number 201611RH1). An informed consent was obtained from all study participants.

### **2.2 Study participants**

This is a cross-sectional study where a total of 462 consecutive women (372 pregnant women, during their third trimester and 90 non-pregnant women) attending the Gynecological and Obstetric department at the University Hospital of Suez Canal University in the period from November 2016 to January 2017 were enrolled in the study.

### **2.3 Sample collection and transportation**

One vaginal swab was obtained from each participant as described by CDC (Verani et al., 2010). Afterwards, each swab was dipped into sterile tubes containing Amies Transport medium (Lab M) and stored at 4 °C until being processed.

#### 2.4 Demographic data collection

Demographic data including participants' age, parity, history of previous abortion and previous preterm labor were collected.

#### 2.5 GBS detection

GBS detection was done as per CDC recommendations (Verani et al., 2010) through a selective enrichment procedure where swabs were inoculated in Lim broth (Todd-Hewitt broth (Oxoid) supplemented with nalidixic acid 15 µg/ml and colistin 10 µg/ml) and incubated overnight at 37 °C in 5% CO<sub>2</sub>. Afterwards, samples were subcultured onto 5% sheep blood agar (Oxoid), supplemented with nalidixic acid 15 µg/ml and colistin 10 µg/ml, and CHROMagar™ StrepB agar (CHROMagar microbiology, France). Blood agar plates were incubated overnight at 37 °C in 5% CO<sub>2</sub> whereas CHROMagar™ StrepB plates were incubated overnight at 37 °C under anaerobic conditions. Colonies on blood agar plates were examined for hemolysis, CAMP test (named after Christie, Atkins, and Munch-Petersen), catalase, Hippurate hydrolysis, Bile esculin hydrolysis and Gram stain. Colonies on CHROMagar™ StrepB plates were examined for intense mauve coloration. Further confirmation was done using latex agglutination testing (Streptococcal grouping

kit, AVIPATH STREP, Omega Diagnostics Ltd.). This was done on both pure colonies and primary Lim broth overnight cultures (Park et al., 2001; Guerrero et al., 2004).

#### 2.6 GBS identification with a species-specific 23S rRNA Primer

DNA was extracted as described previously (Shabayek et al., 2010). Polymerase chain reaction (PCR) was done using the species-specific 23S rRNA primer sequences described by (Evans et al., 2008): *Forward* 5' AACAGCCTCGTATTTAAAATGATAG ATTAAC -3' and *Reverse* 5'- TCCTACCATGACACTAATGTGTC -3'. PCR was done according to (Kawata et al., 2004; Evans et al., 2008).

#### 2.7 Statistical analysis

Continuous variables were analyzed with two-tailed unpaired t-test while categorical data were analyzed with Chi-square and Fischer's exact test. A *p* value less than 0.05 was considered statistically significant.

### 3. Results

#### 3.1 GBS identification

All isolates were beta hemolytic, Lancefield Group B, catalase negative, Gram-positive cocci arranged in pairs or chains. All isolates showed deep intense mauve coloration on

CHROMagar™ StrepB plates and were positive for CAMP and Hippurate hydrolysis and negative for Bile esculin hydrolysis. The 23S rRNA fragment was amplified for all strains.

### 3.2 Prevalence

Among the 462 participants, 129 (27.9%) women were found as GBS- positive carriers. In respect to pregnant participants, 26% pregnant women (n= 97/372) were defined as asymptomatic GBS carriers versus 35.5% among non-pregnant participants (n= 32/90).

### 3.3 Demographics

The mean age of participants was  $29.5 \pm 6.9$ . The mean age of GBS-positive carriers was  $29.2 \pm 6.3$ . The mean age of GBS-negative carriers was  $29.7 \pm 7.1$ . GBS colonization rate was the highest in the age range from 21 – 30. However, GBS carriage was not significantly associated with older age ( $p = 0.74$ ). A similar scenario was observed for parity ( $p = 0.65$ ) as higher GBS colonization was detected for multigravida compared to primigravida or participants with no previous pregnancies. In addition, there was no statistically significant difference between GBS-positive and GBS-negative participants in respect to age ( $p = 0.45$ ) and parity ( $p = 0.48$ ). The same was true for history of

previous abortion ( $p = 0.38$ ) or preterm birth ( $p = 0.32$ ). These were not significantly associated with GBS carriage. Percentage of GBS carriage among study participants in respect to age and parity is shown in Figures 1 and 2. Demographic data is shown in Tables 1 and 2.

## 4. Discussion

The prevalence of GBS carriage in Ismailia, Egypt was 27.9% among all study participants including pregnant and non-pregnant women. This rate was comparable to previous work conducted in Ismailia, Egypt among pregnant and non-pregnant women where the observed colonization rates were found to be 25.3% and 27.3% (Shabayek et al., 2009; Shabayek et al., 2014). However, it was higher than that recorded in AlFayom, Egypt (Elbaradie et al., 2009) which was 17.89%.

Considering other Arabian-African countries in the neighborhood, a lower colonization rate was found in Tunisia (17%) (Ferjani et al., 2006) which is similar to that reported in AlFayom, Egypt. Considering other Arabian-Asian countries in the vicinity, similar rates were reported in Saudi Arabia (27.6%)

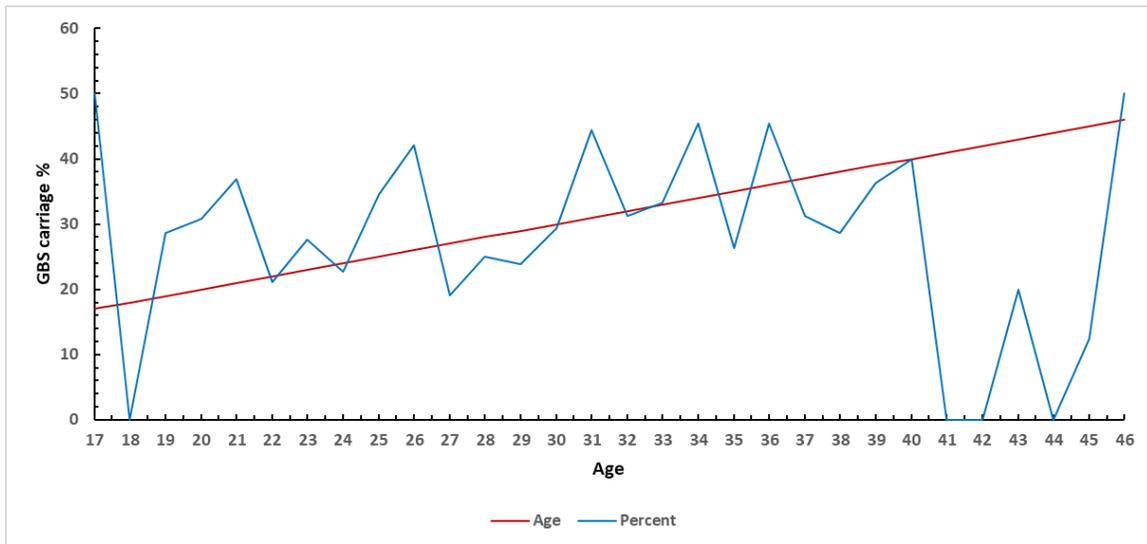


Figure 1: Percentage of GBS-positive participants in respect to age.

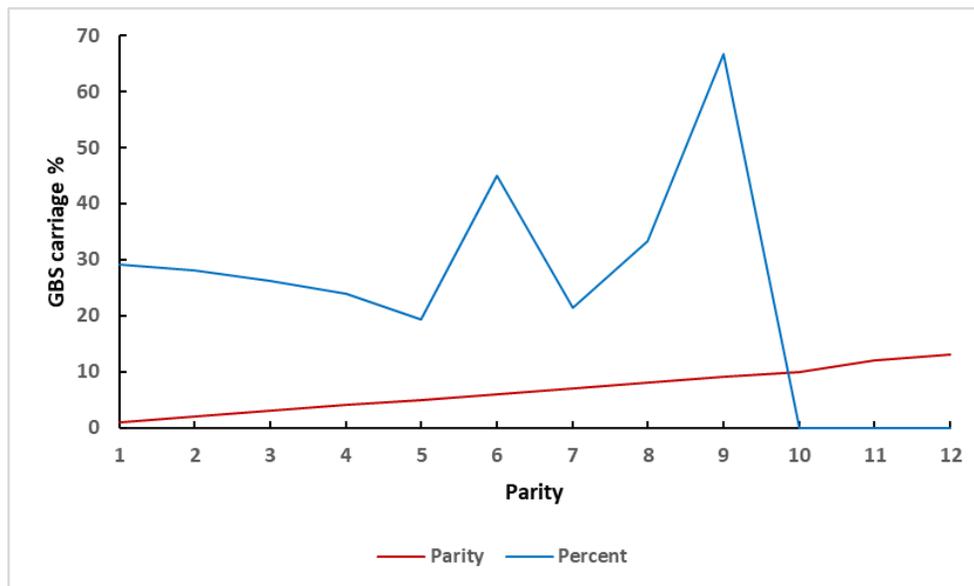


Figure 2: Percentage of GBS-positive participants in respect to parity.

**Table 1: Study participants.**

GBS status (number)	Age (mean $\pm$ SD)	<i>p</i> value	Parity	<i>p</i> value
GBS-positive (129)	29.17 $\pm$ 6.34	0.45	2.86 $\pm$ 1.89	0.48
GBS-negative (333)	29.70 $\pm$ 7.10		2.99 $\pm$ 1.84	

*Two-tailed unpaired t-test was calculated to evaluate the statistical differences between GBS -positive and GBS -negative participants for age and parity. P value < 0.05 is considered statistically significant.*

**Table 2: Detailed demographic data.**

Variable	GBS-positive	GBS-negative	<i>p</i> value
Age			
$\leq$ 20	9	24	0.74
21 - 30	71	188	
>30	49	121	
Parity			
No previous pregnancies	6	5	0.65
Primigravida	28	68	
Multigravida	95	260	
Previous abortion			
Yes	41	122	0.38
No	88	211	
Previous preterm labor			
Yes	11	41	0.32
No	118	292	

*Two-tailed Chi-square and Fischer`s exact tests were calculated to evaluate the statistical differences between GBS -positive and GBS -negative participants for age, parity, previous abortion, and history of preterm labor. P value < 0.05 is considered statistically significant.*

(El-Kersh et al., 2002). However, lower rates were demonstrated in Lebanon (17.7%) (Seoud et al., 2010), Kuwait (16.4%) (Al-Sweih et al., 2004), United Arab Emirates (10%) (Amin et al., 2002), and Iran (9.1%) (Namavar Jahromi et al., 2008). These later were also like those reported in AlFayom, Egypt (Elbaradie et al., 2009).

Nevertheless, all detected colonization rates were within the expected ranges. This was clear when considering the GBS carriage rates reported in the US and Europe showing a range between 6.5% and up to 36% (Campbell et al., 2000; Barcaite et al., 2008). According to (Stoll and Schuchat, 1998) comparable colonization rates can still be observed in developing and developed countries despite the geographical differences. According to (Benitz et al., 1999), differences in colonization rates are expectable across various geographical locations.

Moreover, (Benitz et al., 1999) demonstrated that variations in GBS colonization rates can also be attributed to ethnicity, parity, age, marital status, multiple sex partners, smoking and even to education level. In the present study, non-statistically significant associations were demonstrated between GBS carriage, age, and parity although

higher percentage of GBS carriage was observed with older age and multigravida.

Furthermore, (Philipson et al., 1995) reported that the true burden of GBS colonization might be underestimated by sampling technique. They demonstrated enhanced antenatal GBS detection when vaginal swabs were combined with rectal swabs. In the current study, only one vaginal swab was obtained from each participant.

It is noteworthy that the genera *Enterococcus*, *Staphylococcus* and *Candida* were the most predominant co-colonizers easily detected during GBS screening. Such observation was in concordance with (Kubota et al., 2002; Altoparlak et al., 2004). The possibility of GBS masking due to other predominant vaginal colonizers has been reported before (Ostroff and Steaffens, 1995; Bourbeau et al., 1997; Dunne and Holland-Staley, 1998). This highlights the importance of selective enrichment techniques for reliable detection of GBS in the vaginal swabs through inhibiting the overgrowth of other co-colonizers.

## 5. Conclusion

In the current study, GBS prevalence among pregnant and non-pregnant women in Ismailia, Egypt was 27.9% which is comparable to other regions in the world.

GBS is among the leading causes of neonatal mortality and morbidity worldwide. However, till now, no national guidelines or recommendations are available for GBS monitoring among Egyptian pregnant women. Further GBS investigations should be done to reveal the true burden and etiological agents of neonatal diseases and deaths in Egypt. This should also include deeper epidemiological characterization of the Egyptian GBS isolates in order to be considered in the ongoing vaccine clinical trials to combat neonatal GBS disease.

## 6. References

- Al-Sweih, N., Maiyegun, S., Diejomaoh, M., Rotimi, V., Khodakhast, F., Hassan, N., et al. (2004). Streptococcus agalactiae (Group B Streptococci) Carriage in Late Pregnancy in Kuwait. *Med. Princ. Pract.* 13, 10-14.
- Altöparlak, U., Kadanali, A., and Kadanali, S. (2004). Genital flora in pregnancy and its association with group B streptococcal colonization. *Int J Gynaecol Obstet* 87(3), 245-246. doi: 10.1016/j.ijgo.2004.08.006.
- Amin, A., Abdulrazzaq, Y.M., and Uduman, S. (2002). Group B Streptococcal Serotype Distribution of Isolates from Colonized Pregnant Women at the Time of Delivery in United Arab Emirates. *J. Infect.* 45, 42-46.
- Barcaite, E., Bartusevicius, A., Tameliene, R., Kliucinskas, M., Maleckiene, L., and Nadisauskiene, R. (2008). Prevalence of maternal group B streptococcal colonisation in European countries. *Acta Obstet Gynecol Scand* 87(3), 260-271. doi: 10.1080/00016340801908759.
- Benitz, W.E., Gould, J.B., and Druzin, M.L. (1999). Risk factors for early-onset group B streptococcal sepsis: estimation of odds ratios by critical literature review. *Pediatrics* 103(6), e77. doi: 10.1542/peds.103.6.e77.
- Bourbeau, P.P., Heiter, B.J., and Figdore, M. (1997). Use of Gen-Probe AccuProbe Group B Streptococcus Test to Detect Group B Streptococci in Broth Cultures of Vaginal-Anorectal Specimens from Pregnant Women: Comparison with Traditional Culture Method. *J. Clin. Microbiol* 35(1), 144-147.
- Campbell, J.R., Hillier, S.L., Krohn, M.A., Ferrieri, P., Zaleznik, D.F., and Baker, C.J. (2000). Group B streptococcal colonization and serotype-specific immunity in pregnant women at delivery. *Obstet*

- Gynecol* 96(4), 498-503. doi: 10.1016/s0029-7844(00)00977-7.
- Dunne, W.M., and Holland-Staley, C.A. (1998). Comparison of NNA Agar Culture and Selective Broth Culture for Detection of Group B Streptococcal Colonization in Women. *J. Clin. Microbiol* 36(8), 2298-2300.
- El-Kersh, T.A., Al-Nuaim, L.A., Kharfy, T.A., Al-Shammary, F.J., Al-Saleh, S.S., and Al-Zamel, F.A. (2002). Detection of Genital Colonization of Group B Streptococci During Late Pregnancy. *Saudi. Med. J.* 23(1), 56.
- Elbaradie, S.M., Mahmoud, M., and Farid, M. (2009). Maternal and neonatal screening for Group B streptococci by SCP B gene based PCR: a preliminary study. *Indian J Med Microbiol* 27(1), 17-21.
- Evans, J.J., Bohnsack, J.F., Klesius, P.H., Whiting, A.A., Garcia, J.C., Shoemaker, C.A., et al. (2008). Phylogenetic relationships among *Streptococcus agalactiae* isolated from piscine, dolphin, bovine and human sources: a dolphin and piscine lineage associated with a fish epidemic in Kuwait is also associated with human neonatal infections in Japan. *J Med Microbiol* 57(Pt 11), 1369-1376. doi: 10.1099/jmm.0.47815-0.
- Ferjani, A., Ben Abdallah, H., Ben Saida, N., Gozzi, C., and Boukadida, J. (2006). Vaginal Colonization of the *Streptococcus agalactiae* in Pregnant Women in Tunisia: Risk Factors and Susceptibility of Isolates to Antibiotics [Abstract]. *Bull. Soc. Pathol. Exot.* 99(2), 99-102.
- Guerrero, C., Martínez, J., Menasalvas, A., Blázquez, R., Rodríguez, T., and Segovia, M. (2004). Use of direct latex agglutination testing of selective broth in the detection of group B streptococcal carriage in pregnant women. *Eur J Clin Microbiol Infect Dis* 23(1), 61-62. doi: 10.1007/s10096-003-1052-x.
- Kawata, K., Anzai, T., Senna, K., Kikuchi, N., Ezawa, A., and Takahashi, T. (2004). Simple and rapid PCR method for identification of streptococcal species relevant to animal infections based on 23S rDNA sequence. *FEMS Microbiol Lett* 237(1), 57-64. doi: 10.1016/j.femsle.2004.06.015.
- Kubota, T., Nojima, M., and Itoh, S. (2002). Vaginal bacterial flora of pregnant

- women colonized with group B streptococcus. *J Infect Chemother* 8(4), 326-330. doi: 10.1007/s10156-002-0190-x.
- Lindahl, G., Stalhammar-Carlemalm, M., and Areschoug, T. (2005). Surface Proteins of *Streptococcus agalactiae* and Related Proteins in Other Bacterial Pathogens. *Clin. Microbiol. Rev.* 18(1), 102 - 127.
- Namavar Jahromi, B., Poorarian, S., and Poorbarfehee, S. (2008). The prevalence and adverse effects of group B streptococcal colonization during pregnancy. *Arch Iran Med* 11(6), 654-657.
- Ostroff, R.M., and Steaffens, J.W. (1995). Effect of Specimen Storage, Antibiotics, Feminine Hygiene Product on the Detection of Group B Streptococcus by Culture and the STREP B OIA Test [Abstract]. *Diagn. Microbiol. Infect. Dis.* 22(3), 253-259.
- Park, C.J., Vandell, N.M., Ruprai, D.K., Martin, E.A., Gates, K.M., and Coker, D. (2001). Detection of group B streptococcal colonization in pregnant women using direct latex agglutination testing of selective broth. *J Clin Microbiol* 39(1), 408-409. doi: 10.1128/jcm.39.1.408-409.2001.
- Philipson, E.H., Palermino, D.A., and Robinson, A. (1995). Enhanced antenatal detection of group B streptococcus colonization. *Obstet Gynecol* 85(3), 437-439. doi: 10.1016/0029-7844(94)00412-7.
- Schrag, S.J., and Verani, J.R. (2013). Intrapartum antibiotic prophylaxis for the prevention of perinatal group B streptococcal disease: experience in the United States and implications for a potential group B streptococcal vaccine. *Vaccine* 31 Suppl 4, D20-26. doi: 10.1016/j.vaccine.2012.11.056.
- Schuchat, A. (1998). Epidemiology of Group B Streptococcal Disease in The United States: Shifting Paradigms. *Clin. Microbiol. Rev.* 11(3), 497 - 513.
- Seoud, M., Nassar, A.H., Zalloua, P., Boghossian, N., Ezeddine, J., Fakhoury, H., et al. (2010). Prenatal and neonatal Group B Streptococcus screening and serotyping in Lebanon: incidence and implications. *Acta Obstet Gynecol Scand* 89(3), 399-403. doi: 10.3109/00016340903560008.

- Shabayek, S., and Abdalla, S. (2014). Macrolide- and tetracycline-resistance determinants of colonizing group B streptococcus in women in Egypt. *J Med Microbiol* 63(Pt 10), 1324-1327. doi: 10.1099/jmm.0.077057-0.
- Shabayek, S., Abdalla, S., and Abouzeid, A.M. (2010). Comparison of scpB gene and cfb gene polymerase chain reaction assays with culture on Islam medium to detect Group B Streptococcus in pregnancy. *Indian J Med Microbiol* 28(4), 320-325. doi: 10.4103/0255-0857.71821.
- Shabayek, S., Abdalla, S., and Abouzeid, A.M. (2014). Serotype and surface protein gene distribution of colonizing group B streptococcus in women in Egypt. *Epidemiol Infect* 142(1), 208-210. doi: 10.1017/s0950268813000848.
- Shabayek, S.A., Abdalla, S.M., and Abouzeid, A.M. (2009). Vaginal carriage and antibiotic susceptibility profile of group B Streptococcus during late pregnancy in Ismailia, Egypt. *J Infect Public Health* 2(2), 86-90. doi: 10.1016/j.jiph.2009.03.004.
- Stoll, B.J., and Schuchat, A. (1998). Maternal carriage of group B streptococci in developing countries. *Pediatr Infect Dis J* 17(6), 499-503. doi: 10.1097/00006454-199806000-00013.
- Verani, J.R., McGee, L., and Schrag, S.J. (2010). Prevention of perinatal group B streptococcal disease--revised guidelines from CDC, 2010. *MMWR Recomm Rep* 59(Rr-10), 1-36.