



IMPACT OF GINSENG (*PANAX GINSENG*) ON GROWTH PERFORMANCE, BLOOD BIOCHEMICAL PARAMETERS AND ANTIOXIDATIVE STATUS OF JAPANESE QUAIL

Asmaa Sh. ELnaggar¹, M.E.Ghonime² and Enass Abd El-khalek³

¹Dep. of Anim. and Poul. Prod., Fac. of Agric., Damanhour Univ., Damanhour, Egypt

²Dept. of Anim. and Poul. Prod., Fac. of Agric. and Nat. Reso., Aswan Uni.

³Poult. Pro. Dep., Fac. of Agric. (El-Shatby), Alexandria Univ., Alexandria, Egypt

Corresponding author: Asmaa Sh. Elnaggar; Email: asmaa.elnaggar@agr.dmu.edu.eg

Received: 14/03/2022

Accepted: 24 /04/2022

ABSTRACT: The effect of dietary inclusion of varying levels of ginseng powder as a natural antioxidant on the productive performance, hematological and blood biochemical parameters of blood, and antioxidative status of Japanese quails. A total of 300 unsexed Japanese quails, aged one week were randomly divided into five experimental groups. The experiment lasted five weeks. The first group fed the basal diet (control); while the 2nd, 3rd, 4th, and 5th groups were fed the basal diet supplemented with 0.5, 1.0, 1.5, and 2.0 g ginseng/ Kg of diet, respectively. The results showed that quails fed the basal diet supplemented with different levels of ginseng (treated groups) had significantly better live body weight, body weight gain, feed conversion ratio, economic efficiency, and production index compared with the control group. In comparison to the control group, supplementation of various ginseng lowered serum levels of cholesterol and low-density lipoprotein (LDL), while RBCs count, hemoglobin, PCV, glucose, globulin, and antioxidant enzymes activity were significantly increased. Additionally, ginseng supplementation increased *lactobacillus* sp. while total bacterial count (*E. coli*, and *proteus*) decreased compared with the control group. In conclusion, supplementing Japanese quails with ginseng improved their growth performance, physiological characteristics, and antioxidative status.

Key words: Ginseng, Quail, Productive performance, Bacterial count, Antioxidative status

INTRODUCTION

Recently, throughout the world, including in Egypt, the Japanese quail (*Coturnix japonica*) as an alternative part to chicken as a source of eggs and meat is considered of great economic importance (El-Daly *et al.*, 2014). Quail is one of the smallest types of birds farmed for meat and eggs, and it is also of scientific importance as a laboratory animal (Vali *et al.*, 2005). The production of quail in Egypt is increasing commercially very quickly, due to the interest in selecting new breeds that have a high body weight and rapid growth and require a small space for growth and maintenance with very low funding compared to other species of poultry (Abd El-Azeem *et al.*, 2019). Quail meat is considered the best for patients with high blood pressure, as it is characterized by low fat content, very delicious, and healthy (Elnaggar, *et al.*, 2021). The major challenge in the long-term sustainability of quail production remains the cost of dietary protein and the supply of essential amino acids (Wickramasuriya *et al.*, 2015; Rezaeipour *et al.*, 2016).

Due to their plant-derived (phyto-additive) characteristics and growth-promoting benefits, several therapeutic herbs have become increasingly popular in recent decades as an alternative to antibiotics (Abd El-Hack *et al.*, 2020; Alagawany *et al.*, 2020; Ebrahim *et al.*, 2020). Many herbs and plant extracts possess antimicrobial activities and antioxidant properties that make them useful as natural animal feed additives (Kim *et al.*, 2010; Huang *et al.*, 2012).

Ginseng is a highly valued and popular medicinal plant that belongs to the Araliaceae family and belongs to the genus *Panax* (Helms, 2004). Microorganism fermentation of medicinal

herbs such as red ginseng improved their pharmacological efficacy and neuroprotective activity by modifying the components of medicinal herbs (Zhao *et al.*, 2016). For a long time, ginseng has been utilized as one of the most well-known herbal treatments (Kiefer and Pantuso, 2003). Ginseng saponins (GS), also known as ginsenosides, are thought to be one of the main active ingredients in the root (Zhang *et al.*, 2008), with a wide range of pharmacological activities such as anti-inflammatory (Wang *et al.*, 2013), antioxidant (Lee *et al.*, 2012), and immunomodulatory properties. Ginseng plays role in regulating blood lipid and decreasing blood sugar levels, cancer prevention, liver, kidney, heart protection, and immunostimulatory effects (Sohn *et al.*, 2008). To date, animal experiments study has shown that ginseng reduced blood pressure and improve cardiovascular dysfunction (Kang *et al.*, 1995) and had a relaxing effect on vascular smooth muscle and anti-inflammatory properties as well as anti-stress effect (Peng *et al.*, 1995). Therefore, the objectives of the present study were to study the feeding effect of ginseng supplementation in Japanese quail diets on productive performance, physiological characteristics, and antioxidative status.

MATERIAL AND METHODS

The current experiment was conducted at the Animal and Poultry Research Centre (El-Bostan Farm), which is part of the Animal and Poultry Production Department, Faculty of Agriculture, Damanhour University. All treatments and birds care procedures were approved by the Institutional Animal Care and Use Committee in Damanhour University, Egypt. Authors declare that the procedures imposed on the birds were

Ginseng, Quail, Productive performance, Bacterial count, Antioxidative status

carried out to meet the Directive 2010/63/EU of the European Parliament and of the Council of 22 September 2010 on the protection of animals and birds used for scientific purposes.

Experimental design

A total number of 300 one week unsexed Japanese quail chicks, aged one week and averaged 41.3 g body weight (BW) were randomly divided into five experimental groups (60 chicks each). Each group was subdivided into 6 replicates (10 chicks each). The first treatment was fed the basal diet without any supplementation (control), while the 2nd, 3rd, 4th and 5th treatments were supplied with different levels of ginseng 0.5, 1.0, 1.5, and 2.0 g/Kg diet, respectively. The ginseng material used in the experiment was obtained in powder form a commercial source (local market) in Damanshour City, Beheira Governorate, and analyzed in the laboratory, the most important major compounds are shown in Table (1). The experimental diets were formulated according to NRC (1994). During the starter (7-21 d) and grower stages (22-42d), chicks were fed a basal diet comprising (22.9 and 21.4%) of crude protein and (3042 and 3103 kcal/kg) of metabolizable energy, respectively.

Housing and management

The birds were kept under the same environmental and managerial conditions. All birds were wing banded and housed in wire cages (40×50×40 cm). The birds were reared under variable temperatures, the initial temperature in the farm was maintained at 35°C for the first 2 days and then reduced linearly by 0.5°C per day from the third to the sixth day of age. From the 7th day until 42 days of age, the temperature was slowly reduced until reaching the comfort temperature (24°C). The light regime used was 20 L: 4D/ day

(artificial light). Chicks were fed the experimental diets *ad libitum* and clean fresh water was available. The experiment lasted five weeks after 1st week at 42 days of age.

Collected performance data

Productive performance including individual live body weight (LBW, g), body weight gain (BWG, g), and feed consumption (FC, g) were recorded throughout the trial period (1-6 wk of age). For each replicate within treatment groups, feed conversion ratio (feed/gain ratio, FCR) was calculated according to the formula:

$$\text{FCR} = \text{FC (g)} / \text{BWG (g)}.$$

Also, the survival rate was calculated. The economic efficiency of experimental diets was estimated by (Zeweil, 1996) as the ratio between income and total feed cost during the experimental growth period. The price of the diets and ginseng supplements was calculated according to the local market price at the same time as the experiment in 2020 by the Egyptian pound (L.E.).

$$\text{Economic efficiency} = (\text{Net revenue} / \text{Total feed cost}) * 100$$

$$\text{Net revenue} = \text{Total revenue} - \text{Total feed cost}.$$

Total revenue: the selling price of the obtained live weight. The production index value was calculated throughout the experimental period (7-42 d) of age according to Hubbard broiler management guide 1999

$$\text{EPEI} = \frac{\text{BW (kg)} \times \text{SR}}{\text{PP} \times \text{FCR}} \times 100 \text{ Where:}$$

EPEI = European Production Efficiency Index.

BW = Body weight (kg)

SR = Survival rate (100% - mortality) PP = Production Period (days)

FCR = Feed conversion ratio (kg feed / kg gain).

Blood collection and hemato-biochemical parameters

At the age of 6 weeks, blood samples (approximately 2 ml) were fasted and collected between 8.00 and 9.00 a.m. from the brachial vein of a random sample of 6 quails /group into heparinized and un-heparinized under vacuum tubes. Coagulated blood samples were centrifuged at 4,000 rpm for 15 min, and the clear serum was separated (isolated) and stored (stocked) frozen at -20°C until biochemical analysis. Non-coagulated blood, a part of each sample was used to assess the hematological parameters including red blood cells count (RBCs), hemoglobin (Hb), the percentage of packed cell volume (PCV) were also measured according to (Feldman *et al.* 2000). Plasma samples were separated from blood by centrifugation at 4,000 rpm for 15 min and stored at -20°C .

Plasma total proteins, albumin, were measured by using special kits delivered from sentinel CH. Milano, Italy by means of spectrophotometer according to guidelines and recommendation of (Armstrong and Carr 1965; Doumas *et al.* 1971). Plasma globulin level was calculated by subtracting the total protein and albumin since the fibrinogen usually comprises a negligible fraction (Sturkie, 1986). Albumin to globulin ratio was also calculated according to (Fossati and Prencipe 1982), plasma glucose concentration was measured by the method of (Trinder 1969). The activity of serum alanine aminotransferase (ALT), were estimated according to Reitman and (Frankle 1957). The creatinine level was measured according to the method explained by (Husdan and Rapoport 1968). Serum triglycerides concentrations were tested using specific kits from CAL-TECH Diagnostics, INC, (CAL) Chino,

California, U.S.A. According to the recommendations of (Bogin and Keller 1987), serum total cholesterol was estimated using the specific kits, high-density lipoprotein (HDL) was measured according to the method outlined by (Lopes-Virella *et al.* 1977), and low-density lipoprotein (LDL) was calculated by the formula of (Raya *et al.* 2014). The activity of malondialdehyde (MDA) in the blood was measured using the method reported by (Placer *et al.* 1966). The calorimetric method of (Koracevic *et al.* 2001) was used to assess blood total antioxidant capacity (TAC). Serum glutathione peroxidase activity (GPX) was determined according to the colorimetric method of (Bauer 1982). Serum glutathione activity (GSH) was determined according to (Ellman 1959). (Misra and Fridovich 1972) method for measuring serum superoxide dismutase (SOD) was used.

Slaughter traits

At the end of the experimental period (42 d of age), six quails from each group were randomly taken, fasted for 12 hours, individually weighed, and slaughtered to complete bleeding. Abdominal fat was removed from the gizzard and abdominal region and individually weighed for each carcass. Edible giblets (liver, heart, and gizzard) were individually separated and weighed. Pancreas, spleen and gland bursa of fabricius were individually separated and weighed, and relative weight to live (pre-slaughtered) and calculated for each organ relative to pre-slaughtered weight. The length of the small intestine was also measured. Dressing percentages were calculated as follows:

Dressing % = (dressing weight/ pre-slaughter live body weight) * 100

Ginseng, Quail, Productive performance, Bacterial count, Antioxidative status

Where: Dressing weight = weight of empty carcass without head + edible giblets part.

Bacterial count

During slaughtering procedures, six birds from each treatment were taken randomly for the collection of cecal content samples. Then, samples were subjected to analyzed for total bacterial count (TBC, cfu x 10⁶), *Lactobacillus Spp.* (cfu x 10³), *Escherichia coli* as colony-forming unit (cfu x 10³), and proteus (cfu x 10³) using modified methods described by (Baurhoo *et al.* 2007), which differed only in used agars

Statistical analysis

The analysis of variance for completely randomized design experiments using general linear models procedure of SAS (2006) examined the effects of dietary treatments. The model used was:

$$Y_{ij} = \mu + T_i + e_{ij}$$

Where Y_{ij} = is the dependent variable; μ = the general mean; T_i = the fixed effect of treatment and e_{ij} = random error. The significance of differences among means were determined using Duncan's new multiple range test (Duncan, 1955) at $P \leq 0.05$.

RESULTS AND DISCUSSION

Productive performance traits

Effects of dietary inclusion with different levels of ginseng on growth performance of Japanese quail were shown in Table (2). Varying levels of ginseng supplementation resulted in a significant ($P=0.001$) increase in live body weight (LBW) at 3 and 6 weeks of age. Overall, ginseng treatments raised BW at 3 weeks of age by 6.9, 14.6, 19.3, and 14.5 % of the control value with the 0.5, 1.0, 1.5, and 2.0 % treatments, respectively. Also, ginseng , treatments raised BW at 6

weeks of age by 15.7, 23.0, 23.5, and 20.5 %, respectively, with the best BW being at the levels of 1.0 and 1.5 %. BWG was increased in a level-dependent way ($P=0.001$) from 1st to 3rd week and a similar trend was seen with LBW, which reached 109.8, 121.9, 130.3, and 121.3 % of the control group, respectively. During the second period from 3th to 6th week BWG was increased in a level-dependent way ($P=0.003$), and a similar trend was seen with BW, which reached 115.5, 120.4, 115.2, and 115.1 % of the control group, respectively. During the total period from 1st to 6th week, BWG was increased in a level-dependent way ($P=0.001$), and a similar trend was seen with BW, which reached 112.7, 121.2, 122.1, and 117.9 % of the control group, respectively.

At the time periods indicated above, it is clear from the data analysis of variance that different levels of ginseng, overall quail chick feed consumption was lowered not significantly. The improvement in weight gain associated with the reduction in feed consumption resulted in a significantly ($P=0.007$, 0.003, and 0.001) improved FCR (Table 2), during the first period the 0.5, 1.0, 1.5, and 2.0 % treatments enhanced FCR by 17.1, 19.9, 23.5, and 18.8%, respectively. During the second period improved FCR by 18.1, 21.7, 17.4, and 14.8%, respectively. During the third period improved FCR by 18.1, 21.7, 17.4, and 14.8%, respectively, compared to the control group, with the best FCR observed at the 1.0 and 1.5 g/kg levels. Data from Table (3) shows the survival rate (%), where it is noted that when compared to the control group, SR (%) were improved significantly ($P=0.004$) with ginseng treatments, reaching 103.2, 103.1, 101.3, and 103.2 %, respectively.

The results obtained in this study corresponded with the results of (Muwalla and Abuirmeileh 1990) showed that supplementation of 0.25% of powder Panax ginseng increased body weight gain. Also, Yan *et al.* 2010 noticed that body weight gain was improved with the dietary treatment of powder ginseng adventitious root meal at 0.1% in broiler chickens. Saha *et al.* 2011 studied that the positive effect of ginseng on the growth performance of treated groups could be attributed to natural plant extracts containing a variety of bioactive ingredients that have intrinsic abilities to improve digestion and stimulate enzyme activity. Chung and Choi 2016 also found that mortality and weight gain were most enhanced in the groups fed diets supplemented with 1% fermented red ginseng powder combined with red koji. Additionally, the presence of ginsenoside in the Panax ginseng complex contributed to improving the parameters evaluated by its antimicrobial and antioxidant potential, as well as, assist the absorption of nutrients by increasing the permeability of the small intestinal mucosa as confirmed by Zhang *et al.* (2008) & Kim *et al.* (2010). Therefore, using ginseng powder is considered a key strategy to support gut health and to optimize digestive functions, thereby improving growth performance. While, Özcan (2016) found that supplementing Japanese quails with 5 and 10 mg/kg ginseng root extract had no effect on body weight. The use of 5% Panax ginseng reduced feed consumption but had no effect on feed conversion ratio (Sohn *et al.*, 2008). Kim *et al.* (2014) reported that supplementing with 3% red ginseng marc markedly decreased mortality of broiler.

Hematological characteristics

Hematological parameters of ginseng-treated quails were shown in Table (3). All hematological indices were within the range of reference values. Red blood cells (RBCs) counts increased (P=0.001) gradually as ginseng levels increased. The increases of RBCs were 32.4, 33.2, 32.4, and 31.8 % compared with the control. It is noted that when compared to the control group, Hb concentration (g/dl) was significantly increased (P=0.002) with ginseng treatments, reaching 129.2, 126.0, 135.7, and 135.9 %, respectively. Also, the PCV % value was increased significantly (P=0.003) with ginseng treatments, reaching 107.3, 123.3, 112.7, and 124.4 %, respectively. The improvement in the number of RBCs may be due to the role of ginseng in stimulating the bone marrow to synthesis erythropoietin (Jensen *et al.*, 2000; Karadeniz and Altintas, 2008). Panax ginseng administration significantly enhanced haemoglobin concentrations, RBCs, WBCs count, and lymphocyte numbers in rats, according to Simsek *et al.* (2007). Xi *et al.* (2017) showed that adding different types of ginseng to poultry diets may enhance the hematological parameters in poultry. While Yan *et al.* (2010) indicated that ginseng supplementation did not affect RBCs and WBCs counts and increased lymphocyte levels. Also, in the study with laying hens (Kim *et al.*, 2015), WBCs, hemoglobin levels and hematocrit were not affected and RBCs count was improved with ginseng supplementation. Yener *et al.* (2021) found that ginseng plant supplementation did not affect erythrocytes, hematocrit, lymphocytes, and increased hemoglobin, leukocytes.

Ginseng, Quail, Productive performance, Bacterial count, Antioxidative status

Protein profile

Protein profiles (total protein, and globulin) as shown in Table (3) were significantly increased ($P=0.001$) by the ginseng treatments of quails. The percentages of those increases in total protein compared with control were 12.3, 13.5, 8.2, and 12.2 %, respectively. Likewise, the percentages of those increases in globulin compared with control were 23.3, 33.3, 22.2, and 30.4 %, respectively. Data from the same table showed the differences in the serum albumin and albumin/globulin ratio concentration were not statistically significant with ginseng treatments compared to the control group.

Some blood biochemical parameters

Data from Table (3) shows the quail's blood analysis, where it is noted that when compared to the control group, plasma glucose levels were increased significantly ($P=0.001$) with ginseng treatments, reaching 35.5, 30.4, 26.8, and 28.8 %, respectively. With ginseng treatment, the alanine aminotransferase (ALT) level decreased not significantly with the four ginseng levels. The normal values recorded here for ALT (which reflect insignificant differences compared with control values) were indicative of the normal functioning of the livers of the experimental quails. Different levels of ginseng inclusion in the diets appeared to considerably significantly reduced serum concentrations of creatinine, reaching 78.9, 79.7, 74.8, and 71.5 % of control quails, respectively.

Panax ginseng powder extract supplementation raised total protein, ALT, and AST levels in Japanese quails (Osfor, 1995), but had no effect on urea or creatinine levels in serum. Saponin enhanced the active component serum production, according to Ruda Kewich

and Benishin (2001). Ginseng plant supplementation enhanced AST, ALT, and GGT levels in rats (Karakus *et al.*, 2011), but had no effect on AST and ALT levels in laying hens (Kang *et al.*, 2016). According to Sohn *et al.* (2008), supplementing with 0.5 and 1 percent Sibirya ginseng leaf had no effect on AST, ALT, albumin, total protein, and increased glucose in blood serum. Song *et al.* (2004) found that supplementing with ginseng, a polysaccharide derived from the root of Panax ginseng C.A. Meyer, had no effect on serum AST and ALT activity or albumin levels, similar to the current study.

Lipid profile

In respect of serum triglycerides concentration, the present results showed a lower non-significant of quails supplemented by different ginseng levels compared with the control group. However, serum total cholesterol and low-density lipoprotein (LDL) showed significantly decreased ($P=0.001$ and 0.004) differences in ginseng treatments compared with the control group. They were reduced by 87.4, 88.2, 88.8, and 89.0 % and by 91.8, 92.8, 95.6, and 92.9 %, respectively. On the other hand, ginseng therapies had a substantial ($P=0.002$) effect on blood HDL levels. When compared to the control group with the four levels of ginseng, blood high-density lipoprotein was raised by 14.9, 14.9, 11.3, and 17.2 %, respectively.

In our study, dietary ginseng significantly decreased total cholesterol and it had no effect on serum triglyceride levels in quails, according to several studies (Hassan *et al.*, 2012; Yildirim *et al.*, 2013), ginseng root powder had no effect on serum triglyceride and cholesterol levels. However, several researchers (Muwalla and Abuirmeileh, 1990) found

that dietary ginseng supplementation lowered serum total cholesterol and LDL cholesterol levels in avian species via impairing hepatic cholesterogenesis. When compared to a diet without ginseng, Qureshi *et al.* (1983) found that ginseng supplementation reduced hydroxyl methylglutaryl-CoA (HMG-CoA) reductase activity and cholesterol 7-hydroxylase activity, and that ginsenoside (saponins) are the bioactive agents for the suppression of cholesterogenesis and lipogenesis. The ginseng plant also reduced serum triglyceride and cholesterol levels, according to several researchers (Jang *et al.*, 2007; Yan *et al.*, 2010; Kim *et al.*, 2015; Özcan, 2016). According to Kim *et al.* (1985), ginseng saponin can partially restore LDL biosynthesis in fowl fed a high cholesterol diet. The saponin's solubilizing activity may aid in the elimination of fats from the circulation.

Antioxidant status

Table (3) shows the effect of ginseng supplements on antioxidant levels in the blood serum of treated quails. Ginseng treatments enhanced ($P=0.001$) blood total antioxidant capacity (TAC) by 2.9, 3.5, 4.2, and 3.4 % with the four levels, respectively compared with the control group. Glutathione activity (GSH), Glutathione peroxidase activity (GPx), and superoxide dismutase (SOD) showed significantly increased ($P=0.001$, $P=0.002$, and 0.003 , respectively) by 11.7, 17.1, 3.0, and 5.1 %, and by 0.87, 0.52, 0.95, and 1.43 %, and by 8.4, 7.3, 8.0, and 4.2 % compared with the control group with the four levels of ginseng, respectively. On the other hand, ginseng therapies had a substantial effect ($P=0.001$) on blood malondialdehyde (MDA) levels. When compared to the control group with the four levels of

ginseng, blood MDA was reduced by 26.7, 32.7, 30.9, and 32.6 %, respectively. The results in the current study are similar to the results reported previously for ginseng extract and demonstrate the antioxidant evidenced by its free radical scavenging. Zhang *et al.* (2008) found that ginseng saponins (GS), i.e., ginsenosides, are believed to be one of the major active ingredients in the root. The antioxidant capacity of ginseng extract may have the potential to be used as an effective dietary antioxidant to prevent oxidative stress-related diseases suggested by Oh *et al.* (2010). The improvement of growth performance may be to the total phenolic content and antioxidant activity of ginseng (Chen *et al.*, 2010; Deng *et al.*, 2013). Ginseng possesses a variety of pharmacological activities, including anti-inflammatory, antioxidant, and immune-modulatory activities (Wang *et al.*, 2013; Lee *et al.*, 2012).

Carcass characteristics

Table (4) shows the quail's carcass characteristics measurements. Dressing (%) were increased significantly ($P=0.001$) with ginseng treatments, reaching 106.4, 104.2, 103.5, and 104.6 %, respectively. On the other hand, ginseng therapies had a substantial ($P=0.001$) effect on abdominal fat (%), that reduced by 30.0, 26.7, 33.3, and 23.3 %, compared with control, respectively. On the other hand, liver, heart, gizzard, pancreas, spleen, and bursa of Fabricius weight percentages were not significantly different among experimental groups. Table (4) summarizes the effects of ginseng treatments on intestine length (cm) in quails. Ginseng treatments enhanced the intestine length; hence, this leads to increase its inner surface and thus increasing the absorption of digested

Ginseng, Quail, Productive performance, Bacterial count, Antioxidative status

nutrients that are transported to all cells of the body through the blood. This improvement in the relative weight of the bursa of Fabricius may be due to the increase in the number of lymphocytes due to the ginseng containing effective compounds such as saponins and polysaccharide acidic sugars, which have a positive effect on immune functions and strengthen the immune system in the body of the live bird (Ilsley *et al.*, 2005; Dong *et al.*, 2007). Kim *et al.* (2002) indicated that 5% *Panax ginseng* leaves had no effect on carcass yield. Sohn *et al.* (2008) reported that 0.5 and 1% ginseng increased bursa Fabricius weight but did not affect the liver and spleen weight. Ginseng supplementation had important role in the meat palatability because it affects the first impressions of the meat by consumers. In the study of Yener *et al.* (2021) found that carcass and the relative weight percentages of heart, liver, bursa Fabricius, and abdominal fat were not affected by supplementation of ginseng plant. However, the relative weight percentages of the gizzard and spleen were increased in the doses of powder red ginseng root.

Bacterial count

Data for the cecal bacterial count are shown in Table (5). All the supplemented ginseng treatments had significantly increased ($P=0.002$) *Lactobacillus sp.* compared with the control group; either in terms of TBC or the pathogenic bacteria species (*E. coli* and proteus). Our results showed a significant reduction in the number of pathogenic bacteria, indicating the role of ginseng as a good source to relieve bacterial disorders in the digestive tract of quails. *Lactobacillus spp.* is an indicator microorganism for flora that mediates excellent digestive system functioning in chicken. These

bacteria make short-chain fatty acids, which create an acidic environment that prevents the growth of bacteria that produce odor. *Lactobacillus* species are also hypothesized to prevent *E. coli* from producing poisonous amines by secreting antienterotoxins. The antibacterial properties of ginseng have been attributed to a number of ginseng components (Tan and Vanitha, 2004). The beneficial effects of ginseng on the growth performance of rabbits were mainly attributed to the alteration of the intestinal microbial, increased enzyme secretion, improved immune response, reducing biological stress, increasing antioxidant activity, and consequently improving health status (Brugalli, 2003, Fascina *et al.*, 2012). Park *et al.* (2009) and Yan *et al.* (2011) found that a significant reduction in the number of pathogenic bacteria. It is well known that the beneficial bacterial community in the digestive tract could enhance and maintain the structural and functional integrity of the epithelial lining mucosa, and affect the immune system. Furthermore, many studies have shown that ginseng's immunomodulatory properties may be related to the actions of its ginseng polysaccharides (Xi *et al.*, 2017). Many plants have a favorable effect on bacteria by stimulating their selective development and metabolic activity in the caecum, as well as decrease the number of potentially harmful species like *E. coli*. For example, maternal seaweed derived polysaccharide supplementation reduced the number of *E. coli* and salmonella in weaned pigs' colon and caecum (Leonard *et al.*, 2011).

Economic efficiency and production index

Results of economic efficiency in Table (6) shows the total revenue, net revenue (NR), economic efficiency, relative

Asmaa Sh. ELnaggar¹ et al.

economic efficiency (%), and European production efficiency index (EPEI) that were used to assess the economic nutritional programs altered net revenue, which is represented by the equation $NR = \text{total revenue} - \text{total feed cost}$, indicating that net revenue increased as dietary nutrient levels increased by supplementing with ginseng. The best economic efficiency and relative economic efficiency (%) were obtained present study, the broiler was evaluated in terms of EPEF, which combines body weight and livability, according to Yener *et al.* (2021). In terms of livability and EPEF, dietary red ginseng root powder supplementation had no significant impacts. In laying hens, a similar result was achieved (Yildirim *et al.*, 2013).

CONCLUSION

Based on the results of the present study, it could be concluded that all ginseng

viability of various levels of ginseng. Supplementing with ginseng in the

with 2.0 g /kg diets than the other treatments. Therefore, it is advised to include these ingredients in the poultry diet to improve performance and health status, which will benefit the owners of commercial farms to raise the economic value. Contrary to the results of the

treatments enhanced productive performance, physiological traits, antioxidant status, bacterial count, carcass characteristics as well as economic efficiency and production index of Japanese quails. In spite of the favorable results of all dietary ginseng supplementation at the middle levels (1.0, and 1.5 g/ kg diet), seems to be better than the low and high levels.

Table (1): The most important major compounds of powder ginseng

Proximate analysis of ginseng	%
Moisture	9.6
Ash	5.1
Crude protein	11.6
Lipids	5.5
Crude fiber	7.8
Carbohydrate	55.0
Oils	5.4
Fatty acid content of ginseng	
Meristic	11.7
Palmitic	26.1
Stearic	3.8
Oleic	30.3
Linoleic	23.9
Linolenic	3.7

Ginseng, Quail, Productive performance, Bacterial count, Antioxidative status

Table (2): Effect of dietary inclusion with different levels of ginseng on growth performance of Japanese quail

Dietary supplementations	Ginseng levels (g/kg diet)						
	Control	0.5	1.0	1.5	2.0	SEM	<i>p</i> value
Items	Body weight (g)						
1 wk	41.3	40.9	41.6	40.9	42.1	1.89	0.085
3 wk	117.9 ^c	126.0 ^b	135.1 ^a	140.7 ^a	135.0 ^a	2.90	0.001
6 wk	199.1 ^c	230.3 ^a	244.9 ^a	245.8 ^a	240.0 ^a	8.00	0.001
	Body weight gain (g)						
1-3 wk	76.6 ^c	84.1 ^b	93.4 ^a	99.8 ^a	92.9 ^a	6.77	0.001
3-6 wk	91.2 ^b	105.3 ^a	109.8 ^a	105.1 ^a	105.0 ^a	9.11	0.003
1-6 wk	167.8 ^c	189.1 ^b	203.3 ^a	204.9 ^a	197.9 ^a	10.00	0.001
	Feed consumption (g feed / bird/ period)						
1-3 wk	212.0	200.0	210.0	212.0	209.0	3.00	0.098
3-6 wk	530.0	500.0	500.0	505.0	520.0	2.90	0.978
1-6 wk	742.0	700.0	710.0	717.0	729.0	11.09	0.190
	Feed conversion ratio (g feed/ g body weight gain)						
1-3 wk	2.77 ^a	2.38 ^b	2.22 ^b	2.12 ^b	2.25 ^b	0.980	0.007
3-6 wk	5.81 ^a	4.76 ^b	4.55 ^b	4.80 ^b	4.95 ^b	0.876	0.003
1-6 wk	4.42 ^a	3.70 ^b	3.49 ^c	3.50 ^c	3.68 ^b	0.065	0.001
Survival rate (%)	95.9 ^b	99.0 ^a	98.9 ^a	97.1 ^a	99.0 ^a	2.77	0.004

^{a,b,c} Means in the same row followed by different superscripts are significantly different at ($p \leq 0.05$). SEM, Standard error of means; BW, body weight; BWG, weight gain; FC, Feed consumption; FCR, Feed conversion ratio.

Table (3): Effect of dietary inclusion with different levels of ginseng on hemato-biochemical parameters of Japanese quail

Dietary supplementations	Ginseng levels (g/kg diet)						SEM	p value
	Control	0.5	1.0	1.5	2.0			
Items	Hematological traits							
RBCs (10 ⁶ /cmm ³)	3.40 ^b	4.50 ^a	4.53 ^a	4.50 ^a	4.48 ^a	0.811	0.001	
Hb (g/dl)	10.30 ^b	13.31 ^a	12.98 ^a	13.98 ^a	14.00 ^a	0.909	0.002	
PCV (%)	31.32 ^c	33.60 ^b	38.62 ^a	35.31 ^b	38.95 ^a	1.990	0.003	
Blood biochemical constituents								
T. protein (g/dl)	5.7 ^b	6.4 ^a	6.5 ^a	6.2 ^a	6.4 ^a	2.88	0.001	
Albumin(g/dl)	3.0	3.1	2.9	2.9	2.9	1.09	0.065	
Globulin(g/dl)	2.7 ^b	3.3 ^a	3.6 ^a	3.3 ^a	3.5 ^a	0.998	0.001	
A/G ratio	1.11	0.92	0.80	0.87	0.82	0.087	0.060	
Glucose (mg/dl)	140.3 ^b	190.1 ^a	183.0 ^a	177.9 ^a	180.7 ^a	8.98	0.001	
ALT (IU/L)	64.0	59.7	59.7	60.0	61.2	9.98	0.083	
Creatinine (mg/dl)	1.23 ^a	0.97 ^b	0.98 ^b	0.92 ^b	0.88 ^b	0.098	0.002	
Triglycerides (mg/dl)	138.3	101.0	102.0	107.7	104.9	18.91	0.082	
Cholesterol (mg/dl)	213.6 ^a	186.7 ^b	188.3 ^b	189.7 ^b	190.1 ^b	23.91	0.001	
HDL (mg/dl)	47.0 ^c	54.0 ^a	54.0 ^a	50.3 ^b	55.1 ^a	8.99	0.002	
LDL (mg/dl)	138.9 ^a	127.5 ^b	128.9 ^b	132.8 ^b	129.0 ^b	11.9	0.004	
Indicators of antioxidative status								
MDA (µg/ml)	118.7 ^a	87.0 ^b	79.9 ^b	82.0 ^b	80.1 ^b	0.065	0.001	
TAC (nmol/L)	406.6 ^c	418.3 ^b	419.0 ^b	423.7 ^a	420.5 ^a	29.9	0.001	
GPX (mmol/L)	877.7 ^b	885.3 ^a	882.3 ^a	886.0 ^a	890.3 ^a	12.90	0.002	
GSH (U/MI)	59.1 ^b	66.0 ^a	69.2 ^a	68.0 ^a	62.1 ^a	0.098	0.001	
SOD (U/ml)	245.7 ^b	266.3 ^a	263.7 ^a	265.3 ^a	255.9 ^a	8.99	0.003	

^{a,b,c} Means in the same row followed by different superscripts are significantly different at (p≤ 0.05).

SEM, standard error of means; RBCs, red blood cell; PCV, packed cell volume; HDL, high-density lipoprotein; LDL, low-density lipoprotein; MDA, malondialdehyde; TAC, total antioxidant capacity; GPx, glutathione peroxidase; GSH, glutathione; SOD, superoxide dismutase.

Table (4): Effect of dietary inclusion with different levels of ginseng on carcass characteristics of Japanese quail

Dietary supplementations	Ginseng levels (g/kg diet)						
	Control	0.5	1.0	1.5	2.0	SEM	<i>p value</i>
Items	Carcass characteristics						
Dressing (%)	69.21 ^b	73.62 ^a	72.55 ^a	71.62 ^a	72.42 ^a	9.87	0.001
Liver (%)	1.63	1.72	1.70	1.77	1.67	0.89	0.098
Heart (%)	0.92	0.89	0.87	0.84	0.87	0.07	0.341
Gizzard (%)	1.64	1.67	1.69	1.71	1.69	0.34	0.187
Abdominal fat (%)	0.30 ^a	0.21 ^b	0.22 ^b	0.20 ^b	0.23 ^b	0.05	0.001
Pancreas (%)	0.53	0.45	0.41	0.48	0.44	0.09	0.082
Spleen (%)	0.041	0.042	0.040	0.041	0.039	0.01	0.198
Bursa of Fabricius(%)	0.07	0.07	0.06	0.08	0.08	0.01	0.098
Intestine length (cm)	66.6	71.6	76.3	80.0	66.6	1.91	0.230

^{a,b}. Means in the same row followed by different superscripts are significantly different at ($p \leq 0.05$) SEM, standard error of means.

Table (5): Effect of dietary inclusion with different levels of ginseng on bacterial counts in cecal contents of Japanese quail

Dietary supplementations	Ginseng levels (g/kg diet)						
	Control	0.5	1.0	1.5	2.0	SEM	<i>p value</i>
Items	Bacterial counts						
TBC (cfu x 10 ⁶)	4.91 ^a	3.73 ^b	3.77 ^b	3.83 ^b	3.61 ^b	0.891	0.001
<i>Lactobacillus</i> (cfu x 10 ³)	1.11 ^b	1.55 ^a	1.77 ^a	1.70 ^a	1.92 ^a	0.993	0.002
<i>E. Coli</i> (cfu x 10 ³)	1.40 ^a	0.87 ^b	0.73 ^b	0.77 ^b	0.80 ^b	0.911	0.001
<i>Proteus</i> (cfu x 10 ³)	0.88 ^a	0.47 ^b	0.53 ^b	0.43 ^b	0.51 ^b	0.068	0.003

^{a,b}. Means in the same row followed by different superscripts are significantly different at ($p \leq 0.05$) SEM, standard error of means; TBC, total bacterial count.

Table (6): Effect of dietary inclusion with different levels of ginseng on economic efficiency and production index of Japanese quail

Dietary supplementations	Ginseng levels (g/kg diet)						
	Control	0.5	1.0	1.5	2.0	SEM	<i>p</i> value
Items	Carcass characteristics						
Birds price (7 d) (LE)	1.45	1.45	1.45	1.45	1.45	--	--
FC (kg/chick)	0.742	0.700	0.71	0.717	0.729	11.09	0.190
Price/ Bird (L.E)	7.5	7.5	7.5	7.5	7.5	--	--
Feed cost L.E/Kg+ feed additive	5.57	5.25	5.33	5.38	5.47	--	--
Body weight gain (g/ bird)	167.8 ^c	189.1 ^b	203.3 ^a	204.9 ^a	197.9 ^a	10.00	0.001
Total cost	8.67 ^a	8.35 ^b	8.00 ^c	7.94 ^c	7.84 ^c	0.987	0.001
Total Revenue (LE)	12.0	12.0	12.0	12.0	12.0	--	--
Net Revenue (LE)	3.33 ^c	3.65 ^b	4.00 ^a	4.06 ^a	4.16 ^a	0.890	0.001
Economic efficiency	0.38 ^c	0.44 ^b	0.50 ^a	0.51 ^a	0.53 ^a	0.065	0.001
Relative economic efficiency (%)	100.0	113.8	130.2	133.1	138.2	--	--
Production index	8.43 ^c	11.80 ^b	15.26 ^a	15.02 ^a	15.97 ^a	2.90	0.001

^{a,b,c} Means in the same row followed by different superscripts are significantly different at ($p \leq 0.05$); SEM, standard error of means.

REFERENCES

- Abd El-Azeem, N. A. A., Madkour, M., Aboelazab, O. M., & El-Wardany, I. 2019.** Physiological responses of Japanese quail breeders to age at mating and silver nanoparticles administration. *International Journal of Veterinary Science*, 8(2), 67-72.
- Abd El-Hack, M. E., Elnesr, S. S., Alagawany, M., Gado, A., Noreldin, A. E., & Gabr, A. A. 2020.** Impact of green tea (*Camellia sinensis*) and epigallocatechin gallate on poultry. *World's Poultry Science Journal*, 76(1), 49-63.
- Alagawany, M., Farag, M. R., Sahfi, M. E., Elnesr, S. S., Alqaisi, O., El-Kassas, S., ... & Abd E-Hack, M. E. 2020.** Phytochemical characteristics of Paulownia trees wastes and its use as unconventional feedstuff in animal feed. *Animal Biotechnology*, 1-8.
- Armstrong, W. D., & Carr, C. W. 1965.** Physiological chemistry: laboratory directions. Burgess.
- Bauer, J. D. 1982.** Clinical laboratory methods. Mosby, USA.
- Baurhoo, B., Phillip, L., & Ruiz-Feria, C. A. 2007.** Effects of purified lignin and mannan oligosaccharides on intestinal integrity and microbial populations in the ceca and litter of broiler chickens. *Poultry science*, 86(6), 1070-1078.
- Bogin, E., & Keller, P. 1987.** Application of clinical biochemistry to

- medically relevant animal models and standardization and quality control in animal biochemistry. *J. Clin. Chem. Clin. Biochem*, 25, 873-878.
- Brugalli, I. 2003.** Alimentação alternativa: a utilização de fitoterápicos ou nutracêuticos como moduladores da imunidade e desempenho animal. Simpósio sobre manejo e nutrição de aves e suínos, 1, 167-182.
- Chen, C. Y. O., Ribaya-Mercado, J. D., McKay, D. L., Croom, E., and Blumberg, J. B. 2010.** Differential antioxidant and quinone reductase inducing activity of American, Asian, and Siberian ginseng. *Food Chemistry*, 119(2), 445-451.
- Chung, T. H., & Choi, I. H. 2016.** Growth performance and fatty acid profiles of broilers given diets supplemented with fermented red ginseng marc powder combined with red koji. *Brazilian Journal of Poultry Science*, 18, 733-738.
- Deng, G. F., Lin, X., Xu, X. R., Gao, L. L., Xie, J. F., & Li, H. B. 2013.** Antioxidant capacities and total phenolic contents of 56 vegetables. *Journal of functional foods*, 5(1), 260-266.
- Dong, X. F., Gao, W. W., Tong, J. M., Jia, H. Q., Sa, R. N., & Zhang, Q. 2007.** Effect of polysavone (alfalfa extract) on abdominal fat deposition and immunity in broiler chickens. *Poultry science*, 86(9), 1955-1959.
- Doumas, B. T., Watson, W. A., & Biggs, H. G. 1971.** Albumin standards and the measurement of serum albumin with bromocresol green. *Clinica chimica acta*, 31(1), 87-96.
- Duncan, D. B. 1955.** Multiple range and multiple "F" test. *Bio-metrics*. 11, 1-42.
- Ebrahim, A. A., Elnesr, S. S., Abdel-Mageed, M. A. A., & Aly, M. M. M. 2020.** Nutritional significance of aloe vera (*Aloe barbadensis* Miller) and its beneficial impact on poultry. *World's Poultry Science Journal*, 76(4), 803-814.
- El-Daly, E. F., Abd El-Azeem, N. A., El-Wardany, I., Abd El-Gawad, A. H., & Hemid, A. E. A. 2014.** Effect of different anti-stress feed additives on some blood metabolites and lymphoid organs histology in laying Japanese quail. *World Applied Sciences Journal*, 29(12), 1574-1585.
- Ellman, G. L. 1959.** Tissue sulfhydryl groups. *Archives of biochemistry and biophysics*, 82(1), 70-77.
- Elnaggar, A. Sh.; R., A. M. Ali and E. A. El-Said. 2021.** Complementary effect of black pepper and turmeric on productive performance and physiological response of Japanese quail. *Egypt. Poult. Sci. Vol. (41) (I): (77-91)*.
- Fascina, V. B., Sartori, J. R., Gonzales, E., Carvalho, F. B. D., Souza, I. M. G. P. D., Polycarpo, G. D. V., ... & Pelícia, V. C. 2012.** Phytogetic additives and organic acids in broiler chicken diets. *Revista Brasileira de Zootecnia*, 41(10), 2189-2197.
- Feldman, B., Zinkl, J., & Jain, N. 2000.** *Schalm's veterinary hematology*. Philadelphia, PA: Lippincott Williams and Wilkins.
- Fossati, P., & Prencipe, L. 1982.** Serum triglycerides determined colorimetrically with an enzyme that produces hydrogen peroxide. *Clinical chemistry*, 28(10), 2077-2080.

- Hassan, M. R., Choe, H. S., & Ryu, K. S. 2012.** A comparison of feeding multi-probiotics and fermented ginseng byproducts on performance, intestinal microflora and immunity of broiler chicks. *Korean Journal of Poultry Science*, 39(4), 253-260.
- Helms, S. 2004.** Cancer prevention and therapeutics: Panax ginseng. *Alternative Medicine Review*, 9(3).
- Huang, C. W., Lee, T. T., Shih, Y. C., & Yu, B. 2012.** Effects of dietary supplementation of Chinese medicinal herbs on polymorphonuclear neutrophil immune activity and small intestinal morphology in weanling pigs. *Journal of Animal Physiology and Animal Nutrition*, 96(2), 285-294.
- Hubbard, S. A. S. 1999.** Broiler management guide. *Hubbard SAS, Quintin, France*.
- Husdan, H., and Rapoport, A. 1968.** Estimation of creatinine by the Jaffe reaction: a comparison of three methods. *Clinical chemistry*, 14(3), 222-238.
- Ilsley, S. E., Miller, H. M., & Kamel, C. 2005.** Effects of dietary quillaja saponin and curcumin on the performance and immune status of weaned piglets. *Journal of animal science*, 83(1), 82-88.
- Jang, H. D., Kim, H. J., Cho, J. H., Chen, Y. J., Yoo, J. S., Min, B. J., and Kim, I. H. 2007.** Effects of dietary supplementation of fermented wild-ginseng culture by-products on egg productivity, egg quality, blood characteristics and ginsenoside concentration of yolk in laying hens. *Korean Journal of Poultry Science*, 34(4), 271-278.
- Jensen, G. S., Ginsberg, D. I., Huerta, P., Citton, M., & Drapeau, C. 2000.** Consumption of *Aphanizomenon flos-aquae* has rapid effects on the circulation and function of immune cells in humans. *Journal of the American Nutraceutical Association*, 2, 50-58.
- Kang, H. K., Park, S. B., & Kim, C. H. 2016.** Effect of dietary supplementation of red ginseng by-product on laying performance, blood biochemistry, serum immunoglobulin and microbial population in laying hens. *Asian-Australasian journal of animal sciences*, 29(10), 1464.
- Kang, S. Y., Schini-Kerth, V. B., & Kim, N. D. 1995.** Ginsenosides of the protopanaxatriol group cause endothelium-dependent relaxation in the rat aorta. *Life Sciences*, 56(19), 1577-1586.
- Karadeniz, A., and Altintas, L. 2008.** Effects of panax ginseng on fluoride-induced haematological pattern changes in mice. *Fluoride*, 41(1), 67.
- Karakus, E., Karadeniz, A., Simsek, N., Can, I., Kara, A., Yildirim, S., ... and Kisa, F. 2011.** Protective effect of Panax ginseng against serum biochemical changes and apoptosis in liver of rats treated with carbon tetrachloride (CCl₄). *Journal of hazardous materials*, 195, 208-213.
- Kiefer, D. S., & Pantuso, T. 2003.** Panax ginseng. *American family physician*, 68(8), 1539-1542.
- Kim, B. K., Hwang, I. E., Kang, S. S., Shin, S. H., Woo, S. C., Kim, Y. J., & Hwang, Y. H. 2002.** Effects of dietary Panax ginseng, *Dioscorea Japonica* and oriental medicine refuse on productivity of Korean native chicken. *Journal of Animal Science and Technology*, 44(3), 297-304.
- Kim, I. C., Yang, J. H., & Hur, S. S. 2010.** Characterization of a loess

Ginseng, Quail, Productive performance, Bacterial count, Antioxidative status

- module for manufacturing loess red ginseng. *Journal of Ginseng Research*, 34(4), 282-287.
- Kim, J. W., Lee, H. B., & Joo, C. N. 1985.** Biochemical studies on ginseng saponin (XXV) ginseng saponin-protein interaction. *Korean Journal of Biochemistry*, 18, 453-458.
- Kim, S. H., Kwon, C. H., & Nakano, I. 2014.** Detoxification of oxidative stress in glioma stem cells: mechanism, clinical relevance, and therapeutic development. *Journal of Neuroscience Research*, 92(11), 1419-1424.
- Kim, S., Jiang, Y., and Song, J. 2015.** The effects of interest and utility value on mathematics engagement and achievement. In K. A. Renninger, M. Nieswandt, & S. Hidi (Eds.), *Interest in mathematics and science learning* (pp. 63-78). Washington, DC: American Educational Research Association.
- Koracevic, D., Koracevic, G., Djordjevic, V., Andrejevic, S., & Cosic, V. 2001.** Method for the measurement of antioxidant activity in human fluids. *Journal of clinical pathology*, 54(5), 356-361.
- Lee, H., Kim, J., Lee, S. Y., Park, J. H., & Hwang, G. S. 2012.** Processed Panax ginseng, sun ginseng, decreases oxidative damage induced by tert-butyl hydroperoxide via regulation of antioxidant enzyme and anti-apoptotic molecules in HepG2 cells. *Journal of ginseng research*, 36(3), 248.
- Leonard, S. G., Sweeney, T., Bahar, B., Lynch, B. P., & O'Doherty, J. V. 2011.** Effect of dietary seaweed extracts and fish oil supplementation in sows on performance, intestinal microflora, intestinal morphology, volatile fatty acid concentrations and immune status of weaned pigs. *British Journal of Nutrition*, 105(4), 549-560.
- Lopes-Virella, M. F., Stone, P., Ellis, S., & Colwell, J. A. 1977.** Cholesterol determination in high-density lipoproteins separated by three different methods. *Clinical chemistry*, 23(5), 882-884.
- Misra, H. P., & Fridovich, I. 1972.** The role of superoxide anion in the autoxidation of epinephrine and a simple assay for superoxide dismutase. *Journal of Biological chemistry*, 247(10), 3170-3175.
- Muwalla, M. M., & Abuirmeileh, N. M. 1990.** Suppression of avian hepatic cholesterogenesis by dietary ginseng. *The Journal of nutritional biochemistry*, 1(10), 518-521.
- NRC, 1994.** National Research Council, *Nutrient requirements of poultry*. 9.ed. Washington: National Academy. 176p.
- Oh, C. H., Kim, G. N., Lee, S. H., Lee, J. S., & Jang, H. D. 2010.** Effects of heat processing time on total phenolic content and antioxidant capacity of ginseng Jung Kwa. *Journal of Ginseng Research*, 34(3), 198-204.
- Osfor M. M. H. 1995.** Some biochemical and nutritional studies on the effect of Panax ginseng powder on adult Japanese quails. *Pol J Food Nutr Sci*, 4/45, 73-79.
- Özcan, M. A. 2016.** Japon bildircinlarının rasyonlarına Panax Ginseng kök ekstrakt ilavesinin yumurta verimi ve bazı kan parametreleri üzerine etkileri. *Ordu Üniversitesi Bilim ve Teknoloji Dergisi*, 6(2), 68-74.
- Park, B. J., Lim, Y. S., Lee, H. J., Eum, W. S., Park, J. S., Han, K. H., and Lee, K. S. 2009.** Anti-oxidative effects of *Phellinus linteus* and red ginseng extracts on oxidative stress-induced

- DNA damage. *BMB reports*, 42(8), 500-505.
- Peng, C. F., Li, Y. J., Li, Y. J., & Deng, H. W. 1995.** Effects of ginsenosides on vasodilator nerve actions in the rat perfused mesentery are mediated by nitric oxide. *Journal of pharmacy and pharmacology*, 47(7), 614-617.
- Placer, Z. A., Cushman, L. L., & Johnson, B. C. 1966.** Estimation of product of lipid peroxidation (malonyl dialdehyde) in biochemical systems. *Analytical biochemistry*, 16(2), 359-364.
- Qureshi, A. A., Abuirmeileh, N., Din, Z. Z., Ahmad, Y., Burger, W. C., & Elson, C. E. 1983** Suppression of cholesterogenesis and reduction of LDL cholesterol by dietary ginseng and its fractions in chicken liver. *Atherosclerosis*, 48(1), 81-94.
- Raya, A. H., Sherif, E., Rabie, M. H., & Bedair, H. F. 2014.** Effect of dietary supplementation with dried garlic and thyme on growth performance of Japanese quail. *Journal of Animal and Poultry Production*, 5(2), 73-85.
- Reitman, S., and Frankel, S. O. 1957.** A colorimetric method for the determination of serum glutamic oxalacetic and glutamic pyruvic transaminases. *American journal of clinical pathology*, 28(1), 56-63.
- Rezaeipour, V., Barsalani, A., & Abdollahpour, R. 2016.** Effects of phytase supplementation on growth performance, jejunum morphology, liver health, and serum metabolites of Japanese quails fed sesame (*Sesamum indicum*) meal-based diets containing graded levels of protein. *Tropical animal health and production*, 48(6), 1141-1146.
- Rudakewich, M., Ba, F., & Benishin, C. G. 2001.** Neurotrophic and neuroprotective actions of ginsenosides Rb1 and Rg1. *Planta Medica*, 67(06), 533-537.
- Saha, M., Chowdhury, S. D., Hossain, M. E., Islam, M. K., & Roy, B. 2011.** Organic water additive on growth performances, hematological parameters and cost effectiveness in broiler production. *Journal of Animal Science and Technology*, 53(6), 517-523.
- SAS, 2006.** SAS/STAT User's guide statistics. SAS institute INC., Cary. NC, USA.
- Simsek, N., Karadeniz, A., & Karaca, T. 2007.** Effects of the *Spirulina platensis* and *Panax ginseng* oral supplementation on peripheral. *Revue Méd. Vét*, 158(10), 483-488.
- Sohn, S. H., Jang, I. S., Moon, Y. S., Kim, Y. J., Lee, S. H., Ko, Y. H., ... & Kang, H. K. 2008.** Effect of dietary siberian ginseng and eucommia on broiler performance, serum biochemical profiles and telomere length. *Korean Journal of Poultry Science*, 35(3), 283-290.
- Song, J. Y., Akhalaia, M., Platonov, A., Kim, H. D., Jung, I. S., Han, Y. S., & Yun, Y. S. 2004.** Effects of polysaccharide ginsan from *Panax ginseng* on liver function. *Archives of pharmacal research*, 27(5), 531-538.
- Sturkie, P. D. 1986.** *Avian Physiology*, Springer-Verlag, New York, NY., pp. 103-121.
- Tan, B. K., & Vanitha, J. 2004.** Immunomodulatory and antimicrobial effects of some traditional Chinese medicinal herbs: a review. *Current medicinal chemistry*, 11(11), 1423-1430.
- Trinder, P. 1969.** Determination of glucose in blood using glucose oxidase with an alternative oxygen

- acceptor. *Annals of clinical Biochemistry*, 6(1), 24-27.
- Vali, N., Edriss, M. A., & Rahmani, H. R. 2005.** Genetic parameters of body and some carcass traits in two quail strains. *International Journal of Poultry Science*, 4(5), 296-300.
- Wang, L., Yang, X., Yu, X., Yao, Y., & Ren, G. 2013.** Evaluation of antibacterial and anti-inflammatory activities of less polar ginsenosides produced from polar ginsenosides by heat-transformation. *Journal of agricultural and food chemistry*, 61(50), 12274-12282
- Wickramasuriya, S. S., Yi, Y. J., Yoo, J., Kang, N. K., & Heo, J. M. 2015.** A review of canola meal as an alternative feed ingredient for ducks. *Journal of animal science and technology*, 57(1), 1-9.
- Xi, Q. Y., Jiang, Y., Zhao, S., Zeng, B., Wang, F., Wang, L. N., and Zhang, Y. L. 2017.** Effect of ginseng polysaccharides on the immunity and growth of piglets by dietary supplementation during late pregnancy and lactating sows. *Animal Science Journal*, 88(6), 863-872.
- Yan, L., Meng, Q. W., & Kim, I. H. 2011.** The effect of an herb extract mixture on growth performance, nutrient digestibility, blood characteristics and fecal noxious gas content in growing pigs. *Livestock Science*, 141(2-3), 143-147.
- Yan, L., Meng, Q. W., Lee, J. H., Wang, J. P., & Kim, I. H. 2010.** Effects of dietary wild-ginseng adventitious root meal on growth performance, blood profiles, relative organ weight and meat quality in broiler chickens. *Asian-Australasian Journal of Animal Sciences*, 24(2), 258-263.
- Yener, Y., Yalcin, S., & Colpan, I. 2021.** Effects of dietary supplementation of red ginseng root powder on performance, immune system, caecal microbial population and some blood parameters in broilers. *Ankara Üniversitesi Veteriner Fakültesi Dergisi*, 68(2), 137-145.
- Yildirim, A., Sen, M. I., Sekeroglu, A., & Eleroglu, H. 2013.** Effects of Korean ginseng (*Panax ginseng* CA Meyer) root extract on egg production performance and egg quality of laying hens. *South African Journal of Animal Science*, 43(2), 194-207.
- Zeweil, H. S. 1996.** Enzyme supplements to diets of growing Japanese quails. *Egypt. Poult. Sci. J*, 16, 535-557.
- Zhang, Q. H., Wu, C. F., Duan, L., and Yang, J. Y. 2008.** Protective effects of total saponins from stem and leaf of *Panax ginseng* against cyclophosphamide-induced genotoxicity and apoptosis in mouse bone marrow cells and peripheral lymphocyte cells. *Food and chemical toxicology*, 46(1), 293-302.
- Zhao, P., Li, H., Lei, Y., Li, T., Kim, S., & Kim, I. 2016.** Effect of fermented medicinal plants on growth performance, nutrient digestibility, fecal noxious gas emissions, and diarrhea score in weanling pigs. *Journal of the Science of Food and Agriculture*, 96(4), 1269-1274.

المخلص العربي
تأثير إستخدام نبات الجينسنج علي الاداء الانتاجي ،خصائص الدم البيوكيميائية وخصائص
الاكسدة علي السمان الياباني

أسماء شوقي النجار^١ ؛ محمد السيد غنيم^٢ ؛ إيناس عبد الخالق محمود^٣
^١قسم الإنتاج الحيواني والداخلي - كلية الزراعة - جامعة دمنهور
^٢قسم الإنتاج الحيواني والداخلي ، كلية الزراعة والموارد الطبيعية ، جامعة أسوان
^٣ قسم إنتاج الدواجن- كلية الزراعة (الشاطبي)- جامعة الأسكندرية

أجريت هذه الدراسة في وحدة بحوث الدواجن بمزرعه البستان، قسم الإنتاج الحيواني والداخلي، كلية الزراعة - جامعة دمنهور وكان الهدف منها دراسة التأثيرات الناتجة عن إضافة مستويات مختلفة من نبات الجينسنج علي الصفات الانتاجية والكفاءة الاقتصادية ومعامل الانتاج الاوربي والصفات الفسيولوجية وانزيمات الاكسدة لكناكيت السمان الياباني النامي تم استخدام ٣٠٠ كتكوت من كتاكيت السمان الياباني عشوائيا بداية من عمر أسبوع إلى خمسة معاملات تجريبيه بكل معاملة ٦٠ طائر في ستة مكررات بكل منها ١٠ كتاكيت علي النحو التالي: المجموعة الاولى هي المقارنه (الكنترول) وكانت تغذت علي العليقة بدون إضافات ؛ والمعاملات الاربعه الاخرى تغذت علي العليقه الأساسيه مع اضافته الجينسنج بمستويات ٥،٠ - ١,٠٠ - ١,٥٠ - ٢,٠٠ جم /كجم علف علي التوالي. أظهرت النتائج حدوث زيادة معنوية في وزن الجسم الحي ومعدل الزيادة في وزن الجسم مع تحسن في الكفاءة الغذائية والكفاءة الاقتصادية في المجموعات التي غذيت علي المستويات المختلفة من الجينسنج بالمقارنة بمجموعة الكنترول. لوحظ أيضا وجود زيادة في مستوى تركيز انزيمات الاكسدة في سيرم الدم في المجموعات المغذاه علي الجينسنج بالمستويات المختلفة مقارنة بالكنترول. وقد خلصت نتائج الدراسة إلي أن إضافة المستويات المختلفة من الجينسنج كان لها تأثير ايجابي ومعنوي علي الأداء الإنتاجي وقياسات الدم ,وانزيمات الاكسدة في كتاكيت السمان وكانت افضل معاملته هي ١,٠٠ - ١,٥٠ جم / كجم علف افضل نسبة اضافة من الناحية الاقتصادية.