

The Potential Protective Effects of Aqueous Extracts of Some Herbs on Renal Toxicity Induced by Formaldehyde in Experimental Rats

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ABSTRACT

The current study was performed to evaluate the protective effect of aqueous extracts of Corn silk and Asparagus *Officinalis* against renal toxicity induced by formaldehyde in rats. Thirty-six male albino rats with "Sprague Dawley" strain weighing (150±10 g) were used and split into 2 major groups, the first group (6 rats) fed on a basal diet and kept as a negative control, and the second group of 30 rats was injected with formaldehyde intraperitoneally (10 mg/kg BW /day on 14th to 28th day and divided into 5 subgroups. The first subgroup received a normal diet and acted as a positive control group. The second, third, fourth, and fifth categories consumed standard feed with oral dosages of 200 and 400 mg/Kg body weight per day of aqueous corn silk and asparagus aqueous extract respectively during a 28-day period. Body weight gain (BWG), Feed intake (FI), feed efficiency ratio (FER), and relative kidney weight were computed at the finish of the experiment. Assessment of some serum biochemical parameters, kidney tissues were analyzed for antioxidant/oxidant markers, and histopathology of kidneys were assessed. The results revealed that corn silk and Asparagus aqueous extracts improved the biological evaluation, kidney functions, liver functions, Serum electrolytes, antioxidant enzymes activity, and histopathology of kidneys compared to the positive group. In conclusion, the administration of corn silk and Asparagus aqueous extracts can lower the impacts of formaldehyde on kidneys.

Key words: Corn silk, Asparagus, Formaldehyde, kidney functions; antioxidant enzymes.

INTRODUCTION

A healthy life requires preservation of the kidney, one of the body's key organs that performs the detoxification process. Few herbs have been examined for nephroprotection, yet it is frequently linked to a decrease in proteinuria. Numerous natural products, medicinal plants, and dietary components have been investigated as possible nephron-protective agents (**Kasabe et al., 2012**). Nephrotoxicity is the malfunctioning of kidney-specific detoxification and excretion as a result of endogenous or exogenous toxins damaging or destroying renal function. The kidney is a significant control mechanism that maintains the body's homeostasis and is frequently being toxic by drug exposure. As such, the kidney is particularly vulnerable to xenobiotics. Alteration in glomerular hemodynamics, rhabdomyolysis, tubular cell toxicity, crystal nephropathy, inflammation, and thrombotic microangiopathy are a few of the mechanisms for drug-induced nephrotoxicity (**Kim and Moon, 2012**).

In its pure form, colorless formaldehyde (FA), which is extremely soluble in water, irritates. Owing to its widespread use in industries like the creation of building materials, textiles, the sterilization of goods, plastics, and cosmetics, FA is a contaminating substance that is frequently discovered in the environment (**Bakar et al., 2015**). Additionally, most morgues and anatomy departments employ FA to preserve cadavers. (**Raja, 2012**). FA is a crucial intermediate for the production of purines and other amino acids, and it is also created endogenously by the L-methionine metabolism, methanol, histamine, and methyl alanine (**Checkoway et al., 2015**). Reactive oxygen species can be produced and released as a result of the toxicity brought on by FA exposure through aerobic metabolism and inflammation (ROS) (**Gulec et al., 2006; Birben et al., 2012**). In albino rats that have been exposed to FA over a long period of time, the renal functions may gradually deteriorate. It's possible that urea, serum electrolytes, and creatinine are insufficient to detect early

kidney damage (**Olisah and Meludu, 2021**).

Corn silk (*Stigma maydis*) means that the stigmas come from the female flowers of corn, and fresh corn silk looks like soft silk threads about 10 – 20 cm long that are either light green or yellow-brown in color (**Sanusi et al., 2020**) and (**El-Seedy et al., 2022**) is generally discarded as waste due to the lack of utilization. It is important in attracting pollens, which are further transferred from the anther to the stigma during pollination (**Aukkanit et al., 2015**). It is also contained lipids, vitamins, minerals, proteins, carbohydrates, and volatile oils (**Chutima et al., 2020**). It also consists of various chemical components including polysaccharides, alkaloids, proteins, flavonoids, tannins, and steroids (**Dika et al., 2018**). It has bioactive constituents for example terpenoids, and flavonoids (**Vijitha and Saranya, 2017**). It has a variety of pharmacological actions that have been broadly described, including anti-inflammatory, anti-hyperlipidemic, anti-depressant, anti-diabetic, anti-fatigue, neuro-

protective, antioxidant, kaluretic impacts, and anticancer effects (**Jia et al., 2020**) and (**Chutima et al., 2020**).

Additionally, cystitis, kidney stones, edema, prostate disorders, urinary infections, bedwetting, obesity, and as a diuretic are all conditions that can be treated with corn silk. Moreover, it has been suggested that using corn silk can help treat pleurisy and inflammatory disorders brought on by oxidative stress (**Wang et al., 2012**).

Asparagus (*Asparagus Officinalis*) is the hard stem and woody bottom of the asparagus stalk, and it is an enriched source of bioactive compounds (**Huang et al., 2022**). It includes a variety of natural bioactive phytochemicals with diverse pharmacological actions (**Fathalipour et al., 2020**). Asparagus is utilized in salads, soups, and vegetable dishes, and it has various uses in European and Asian traditional herbal medicine (**Huang et al., 2008**). Pharmacological research has shown that *A. Officinalis* has anti-inflammatory, antifungal, antimutagenic, and diuretic effects.

Additionally, the plant's roots and sprouts are rich sources of oligosaccharides, flavonoids, steroidal saponins, and derivatives of amino acids (Tzatzarakis *et al.*, 2017). As per some research, *A. Officinalis* has biological properties that include reducing alcohol hangovers and defending liver cells from toxins (Kim *et al.*, 2009). *A. Officinalis* may be utilized to treat diabetic nephropathy (Somani *et al.*, 2012)

The goal of the current investigation was to assess the renal protective efficacy of aqueous extracts of *Asparagus Officinalis* and corn silk (*Stigma maydis*) on formaldehyde-induced kidney toxicity in experimental rats.

MATERIALS AND METHODS

Materials

Dried corn silk was obtained from Agriculture Seeds, Herbs and Medicinal plants Company, Cairo, Egypt. *Asparagus Officinalis* was obtained from Carrefour market, in Tanta, Egypt. Thirty-six male albino rats (*Sprague Dawley strain*) were obtained from the

Laboratory Animal Colony, Helwan, Cairo – Egypt, and weighting approximately between 150± 10g. Formaldehyde purity (37%) was supplied from Sigma chemical company. Corn oil and starch were purchased from the local market. Casein, cellulose, ingredients (vitamins, minerals), dextrin, L-cysteine, and choline chloride were obtained from the Cairo Company for Chemical Trading, Cairo, Egypt.

Methods

Preparation of corn silk extract

The plant extract was prepared according to the method used by Saheed *et al.*, (2015) exactly 100 g of corn silk was extracted by maceration in 1000 mL of distilled water. These heterogeneous mixtures were boiled with the aid of a heater for 4 hours at 100°C. After boiling the sample, the filtrate was separated from the corn silk residue with the aid of Whatman no. 1 filter paper, and a funnel was placed into a beaker for separation. The resulting filtrate was concentrated to dryness.

Preparation of *Asparagus Officinalis* extract (AOE)

For the preparation of AOE, plants were air-dried at room temperature (25 °C) the dried samples (100 g) were then extracted with 1 L of boiling distilled water for 2 h at 100 °C, and then the extracts were passed through a filter. The resulting filtrate was concentrated to dryness and stored at -20 °C until use, all the procedures were according to (Pal *et al.*, 2017).

Experimental design

Thirty-six adult male albino rats *Sprague Dawley* strain weighting (150± 10g) were housed in well-aerated cages under a hygienic condition and were fed on a basal diet according to Reeves *et al.*, (1993) for one week for adaptation. After this week, the rats were divided into two main groups: the first group (6 rats) fed on a basal diet and served as a negative control group. The second group (30 rats) was injected with formaldehyde purity (37%) intraperitoneally injection at a rate of 10 mg/kg BW/day for 14 days (Zararsiza *et al.*, 2006) to induce nephrotoxicity from the 14th to

28th day. The second group was divided into 5 subgroups as follows: subgroup (1) was received a normal feed only and served as a positive control group. Groups (2, 3): were intake standard meal with added corn silk (200 and 400 mg / Kg body weight) respectively orally for 28 days (Sepehri *et al.*, 2011). Groups (4, 5) were consumed major feed including *Asparagus Officinalis* (200 and 400 mg / Kg body weight) respectively orally for 28 days (Jashni *et al.*, 2016). Body weight and feed intake were checked once a week. At the end, animals were weighed, fasted overnight, and then sacrificed under very light anesthesia.

Biological evaluation

At the end of the experiment, feed intake, body weight gain, relative organs weight, and feed efficiency ratio were calculated according to Chapman *et al.*, (1959).

Biochemical analysis of serum

After the sacrifice of rats, blood samples were collected from the hepatic portal vein of each rat into dry clean centrifuge tubes.

Serum was carefully separated by centrifugation of blood samples at 3500 rounds per minute (rpm) for 15 minutes at room temperature, transferred into dry clean Eppendorf tubes, then kept frozen at -20°C for later determinations. Serum samples were used for the determination of blood glucose was determined according to **Trinder, (1969)**. Serum tumor necrosis factor- α (TNF- α) was assayed as an inflammatory biomarker (**Luo et al., 2005**). Uric acid, urea nitrogen, and creatinine were determined according to **Fossati et al., (1980)**, **Patton and Crouch (1977)**, **Bartels et al., (1972)** respectively. Aspartate aminotransferase (AST) was carried out according to the method of **Henry, (1974)** and **Yound, (1975)**. Alanine aminotransferase (ALT) was evaluated according to **Bergmeyer et al., (1986)**, serum (ALP) was estimated according to the colorimetric method of **Roy (1970)**, albumin according to **Drupt, (1974)**. A total protein limited according to **Sonnenwirth and Jaret, (1980)**. Globulin was calculated according to **Busher,**

(1990) using the following equation:

$$\text{Globulin} = \text{Total protein} - \text{Albumin.}$$

Creatinine clearance (CrCl) as a marker to glomerular filtration rate (GFR) was calculated from serum creatinine (SCr) and urine creatinine (UCr) levels and 24-hour urinary volumes according to **Inker and Perrone (2021)** using the following equation:

$$\text{GFR} = [\text{UCr} \times \text{V}] \div \text{SCr}$$

Sodium (Na⁺), potassium (K⁺), calcium (Ca⁺⁺) and Magnesium (Mg⁺⁺) were specified in the serum according to the method by **Berndt and Jackwerth, (1979)**.

Organ's sampling

Kidneys were carefully separated from all rats, washed with saline solution (0.9%), dried with filter paper, and individually weighted. A specimen from the kidneys was frozen at (-20°C) for preparing tissues homogenate to determine antioxidant activities. The homogenization was centrifuged at 1000 rpm for 10 min.

Assessment of antioxidant activities and lipid peroxidation in the kidney tissues:

Antioxidant indications were assessed such as Superoxide dismutase (SOD) according to **Nishikimi *et al.*, (1972)**. Catalase (CAT) by colorimetric assay as stated by **Sinha, (1972)**. Lipid peroxide (LPO) as malondialdehyde (MDA) was assessed-estimated by colorimetric assay as maintained by **Buege and Aust, (1978)**, Nitric oxide (NO) was determined according to **Cortas and Wakid, (1990)**.

Histopathological examination:

The kidney of each sacrificed rat was removed and fixed in a 10% neutral buffering formaldehyde solution with a pH of 7.5, then cleaned in xylol before being fixed in paraffin. For histological analysis, a 4-5 μm thick piece was cut and spotted with Hematoxylin and Eosin (H&E) (**Bancroft and Gamble, 2008**).

Statistical analysis:

One-way analysis of variance (ANOVA) was used, followed by the Duncan test, in SPSS software (18) to know the difference between means at $P < 0.05$. The data were presented as a

mean \pm standard deviation (SD) (**Snedecor and Cochran, 1989**).

RESULTS

Biological evaluation

Feed intake (FI), body weight gain % (BWG), and feed efficiency ratio (FER) were shown in Table 1 have significantly decreased in the positive Control group compared to the normal group. However, the other treated groups have revealed a significant increase in all of them compared with the positive control group, and the best results were found in group Corn Silk (400 mg /kg BW) in FI, BWG, and the superior results for FER was recorded to Corn Silk and Asparagus (400 mg /kg BW).

Relative kidney weight

As shown in (Table 2), relative kidney weight has been decreased in the positive control group compared with the negative control group. However, it was significantly increased in all treated groups compared with the positive control group. The best results were recorded in Corn Silk and Asparagus (400 mg/kg BW) as it recorded a significant increase in

relative kidney weight compared with other investigated groups.

Kidney functions

The data in Table (3) indicated that mean values of creatinine, urea, uric acid, and Creatinine clearance in the positive control group were significantly higher compared with the negative dominance group. All parameters in remedy categories significantly decreased ($P < 0.05$) compared to positive control set. The best findings in creatinine, urea and uric acid were found in the Corn Silk collection (400 mg/kg BW) while the preferable findings in Creatinine clearance were found in Corn Silk and Asparagus sets (400 mg/kg BW).

Serum electrolytes

Serum Na^+ and K^+ levels have increased in the renal dysfunction cohort versus the normal rats, as indicated in Table 4. In contrast to the ill rats, it was significantly lower in all different treatment groups. The most effective outcomes were noted with Corn Silk Set (400 mg/kg). While serum Ca^{++} and Mg^{++} levels in the G2 were lower than

those in the G1. Although it was significantly higher in all rats treated compared to the positive control, Corn Silk and Asparagus sets (400 mg/kg) gave good results.

Blood glucose

The data presented in Table 5 showed that the ill group's mean blood glucose values were significantly greater than those of the normal group. In comparison to the positive control rats, every parameter in the different remediations decreased significantly ($P < 0.05$). Corn Silk set (400 mg/kg BW) gave an improved result.

Liver functions

The data displayed in Table 6 showed that all remedy groups had considerably lower mean values of ALT, AST, and ALP compared to the ill group, while these values were significantly higher in the sick rats (G2) than in the healthy rats (G1). Good ALT, AST, and ALP results were seen in Corn Silk (400 mg/kg BW).

Serum Total protein, Albumin and Globulin

The results in Table 7 demonstrated that, in comparison to the G1, the mean value of total protein, albumin, and globulin significantly decreased in the G2. However, compared to the positive control group, every other investigated group experienced a considerable increase. When compared to the G2, the effects of asparagus at 200 mg/kg BW on total protein and albumin were non-significant. The groups treated with Corn Silk and Asparagus (400 mg/kg BW) had the highest levels of total protein, while the group treated with Corn Silk (400 mg/kg) had the best results for albumin and all of the treated groups had the great outcome for globulin due to their close similarity to the control group.

Serum tumor necrosis factor - α (α -TNF) and antioxidant enzymes (CAT, SOD, malondialdehyde (MDA) and nitric oxide (NO in kidney tissue.

Table 8 showed that compared to the healthy group (G1), the activities of catalase (CAT) and superoxide dismutase (SOD) considerably decreased in the unwell group (G2), but they

elevated in the therapy groups. Asparagus and Corn Silk (400 mg/kg BW) both had the best CAT results, while Corn Silk (400 mg/kg BW) had good SOD values. Further, the same table showed that, the mean values of TNF- α , MDA, and NO were significantly greater in the G2 than in G1; whereas their values were significantly lower in the other groups. Corn Silk (400 mg/kg) showed good results for TNF- α , MDA, and NO, whereas Corn Silk and Asparagus (400 mg/kg BW) showed the best improvements for MDA and NO.

Histological Results:

Histological sections of rat kidney showing the control group (A & B) with the normal histological structure of the renal cortex containing the glomeruli, proximal and distal convoluted tubules. The positive group (C & D) shows severe affection for the renal tubules with signs of tubular cell lining vacuolation (red arrow) and intratubular cast deposition (green arrow), tubular hydropic degeneration (yellow arrow) with congestion of the glomeruli (blue

arrow) and narrowing of the Bowman's space (black arrow). The Corn silk 200 (E & F) group shows congestion of the glomeruli (blue arrow) and intratubular hemorrhage (blue arrow). The Corn silk 400 group (G & H) shows fewer pathologic changes than the above group. The Asparagus 200 (I & J) shows tubular cell lining vacuolation (red arrow) with less marked glomerular congestion (blue arrow). The Asparagus 400 group (K & L) shows a more or less good picture than the above group.

DISCUSSION

The findings of the ongoing work are in line with **Wu et al., (2017)** who revealed that FA exposure induced a significant reduction in body weight in maternal mice, their offspring and body weights of newborns were significantly lower contrasted with the control group. These findings are in agreement with **Egwurugwu et al., (2018)** who observed that formalin-exposed rats reduced feeding habits causing weight loss contrasted with the normal control group. **Yogesh et al., (2013)** reported that the diets using corn

silk at different levels elevated the body weight gain of rats with kidney stones disease compared with a positive control group. In this respect, **Kim et al., (2017)** found that oral administration of Corn Silk produced significantly elevated body weight gain, feed intake and FER. **Ikpeazu et al., (2018)** mentioned that oral administration of a boiled aqueous solution of *Stigma maydis* produced significantly increased body weight of the extract-medicated animals when contrasted to the positive control group. The ongoing findings are in harmony with **Zhang et al., (2014)** who indicated that administration of aqueous extract *Asparagus Officinalis* significantly increased body weight compared with a positive control group.

The ongoing study's findings are in agreement with **Ha et al., (2018)** who found that rats administered with aqueous Corn silk extract showed increase of relative Kidney weights. Due to the fact that maize silk extract has a lot of phytochemicals that can produce diuresis and its diuretic action may lead to an elevation in

kidney weight. The present outcomes were supported by **Zhang et al., (2014)** who found that rats administered aqueous *Asparagus Officinalis L.* extract indicated a significant elevation in relative kidney weight contrasted with the untreated rats. This finding is emphasized by **Boj et al., (2003)** who demonstrated that statistically significant elevation in urea levels in the formaldehyde group contrasted with the normal rat group. The current results were supported by **Egwurugwu et al., (2018)** who indicated a statistically significant elevation in urea and creatinine's serum concentrations in the formalin-exposed rats when contrasted with the control group. Because several enzymes, notably NAD-dependent dehydrogenase formaldehyde, catalase, xanthine oxidase, and peroxidase, oxidize FA to formic acid. An increase in urea concentration has been linked to an elevation in the synthesis of these enzymes for the detoxification of FA. **Sepehri et al. (2011)** showed comparable outcomes, noting a significant reduction in serum creatinine in Corn silk + gentamicin groups

contrasted to gentamicin groups in rats. These findings are in line with **Tanideh et al., (2018)** who showed that rats whose administration of aqueous corn silk extract caused significantly increased of creatinine clearance when compared with the non-treated group. These findings are emphasized by **Poormoosavi et al., (2018)** Who reported that administration of *Asparagus Officinalis* induced toxicity in rats improved serum levels of creatinine and urea nitrogen when contrasted with the non-treated group.

The results are in line with those of **Egwurugwu et al. (2018)**, who found that formalin-exposed rats had statistically significant duration-dependent increases in serum potassium and sodium concentrations when contrasted to the healthy control group. The formalin's dehydration and thirst's impacts may also be to blame for the rise in serum electrolytes in exposed rats. Antidiuretic Hormone (ADH) release is stimulated by dehydration, which results in higher serum osmolarity and electrolytes.

The current findings are in line with **Hasanudin et al., (2012)** who mentioned that the rat Corn silk treated group significantly increased Na⁺ excretion and increased K⁺ excretion due to its diuretic impacts, whereby the urine flow was elevated as diuretic impacts can lead to water and solutes' loss in the blood. **Kamel et al., (2014)** investigated that Rutin one of the phenolic compounds in *Asparagus Officinalis* administration could offer preservation versus cisplatin-induced nephrotoxicity. Where a recorded significantly reduced the levels of serum sodium and potassium in comparison with the cisplatin-treated group. This finding is supported by **Tan et al., (2018)** mentioned that injection of FA-induced hyperglycemia due to Formaldehyde interacts with insulin to form Formaldehyde-insulin adducts, and these Formaldehyde -insulin adducts induced insulin deficiency.

The current findings, which **Hasanudin et al. (2012)** confirmed, showed that administering Corn Silk extract does not raise blood glucose levels.

These findings demonstrated that the mechanism of action of corn silk extract on glycemic metabolism is not through an increase in glycogen and inhibition of gluconeogenesis, but rather via an increase in insulin levels and recovery of the damaged cells. These results are in agreement with those found by **Ha et al., (2018)** and **El-seedy et al., (2022)** who revealed that significant decreases were recorded in glucose levels in rats fed on a diet containing corn silk. These results are in agreement with those found by **Dika et al., (2020)** who mentioned that giving corn silk led to a significant reduction in blood glucose concentrations in diabetic rats, because corn silk has many ingredients, including alkaloids, flavonoids, phenols, saponins, tannins, and sitosterol. The flavonoids in corn silk repair pancreatic β cells which can stimulate insulin secretion. These findings are consistent with those of **Zhang et al. (2014)** who found that *Asparagus Officinalis L.* effectively reduced hyperglycemia without increasing hepatic glycogen content or insulin

secretion. Other pathways include decreased serum glucagon, decreased levels of hepatic gluconeogenesis, decreased glucose absorption, and enhanced insulin sensitivity.

Results are supported by **Afrin *et al.*, (2016)** who cleared those activities of AST, ALP, and ALT enzymes were significantly increased in group treated intraperitoneally injected with formaldehyde in comparison to a normal control group. Because of its metabolic reactivity, formaldehyde is a necessary metabolic step in all cells and is used in the manufacture of some purines, amino acids, and thymidine.

The findings of the ongoing work are in line with **Ikpeazu *et al.*, (2018)** who found that dose-dependent increase was observed in AST and ALT significantly reduced in rats medicated with *Stigma maydis* (corn silk) when contrasted to control groups. These findings are in line with **Arba *et al.*, (2020)** who noted that treating rats on diet containing different levels of Corn silk led to a significant decrease in serum ALT,

ALP, and AST enzymes, as compared to the non-treated group. because corn silk contains phytochemicals that have beneficial effects, such as flavonoid compounds, which act as antioxidant agents and has a hepatoprotective effect.

These results are in line with those reported by **Al-Snafi (2015)**, who claimed that an aqueous asparagus extract's administration reduced serum levels of ALT, AST, and ALP. According to **Poormoosavi *et al.* (2018)**, *Asparagus Officinalis* aqueous extract significantly reduced the serum levels of (AST) and (ALT). *Asparagus Officinalis*'s tissue-protective properties are likely achieved by reducing G-SH depletion, limiting lipid peroxidation, and boosting antioxidative ability. As a result, *Asparagus Officinalis* is a viable option for defending tissues (such as the liver and kidneys) against numerous toxins due to its potency and effectiveness.

These outcomes are in line with **Chinedu *et al.*, (2017)** who showed that short-term exposure to formalin induced a significant

decrease in the mean serum levels of albumin, and total protein when compared with before formalin exposure. In the subjects, these findings are in line with **Ha et al., (2018)** who cleared those rats medicated with Corn Silk was increased total protein and albumin when compared with rat untreated rat group.

These results agree with that **Persoz et al., (2010)** The researchers reported that FA has a stimulating impact on cytokines such as TNF- α , which affects inducible nitric oxide synthase (iNOS) metabolism. **Egwurugwu et al., (2018)** who indicated that extended exposure to formalin can trigger several pathophysiological circumstances, such as inflammatory disorders by interfering in the natural killer (NK) cells and TNF's levels.

These outcomes are in line with **Sener et al., (2015)** who showed that rats in the formaldehyde group had higher NO than controls due to formaldehyde being is thought to increase NO synthesis through inducible nitric oxide synthase (iNOS) activity. Increased

synthesis of NO and superoxide anion radicals lead to increased concentrations of peroxy-nitrite and causes further damage. **Sener et al., (2015)** found that administration of formaldehyde into the rats resulted in an impaired antioxidant defense in the renal tissues, as shown by a significant reduction in the activity of antioxidant enzymes such as SOD and CAT in the formaldehyde group as compared to normal controls.

These outcomes are in consistent with those noted by **Sepehri et al., (2011)** who discovered that mice fed with maize silk extract had greater levels of SOD activity in their kidneys and attributed these results to the antioxidant and free radical-scavenging properties of corn silk, which includes phenolic compounds. Presented results are in the line with results **Al-Snafi, (2015)** who showed a protective effect of asparagus on nephrotoxicity in rats compared with the positive control, the content of serum malondialdehyde (MDA) were lower and the content of superoxide dismutase (SOD)

was higher in asparagus treated group. This might be accomplished by reducing the production of free radicals and increasing the activity of free radical scavengers.

Shi et al., (2012) provided evidence that formaldehyde caused structural alterations in glomeruli and proximal convoluted tubules, which were more severely impacted than the distal ones. The findings are corroborated by their findings. This was explained by the fact that after the poisonous chemical has been filtered by the glomeruli, it initially contacts the proximal convoluted tubules. The present study's outcomes are in line with those of **Treesh et al. (2014)**, who discovered that the kidney tissues from the FA-injected group exhibited dilation in Bowman's capsule, congestion in the glomeruli and between the renal tubules, as well as vacuolation in the lining epithelium's cytoplasm of the proximal convoluted tubules with loss of their brush border. According to **Sepehri et al., (2011)**, rats treated with corn silk shown indications of tube regeneration and protection from

gentamicin-induced interstitial nephritis. Additionally, according to **Poormoosavi et al., (2018)**, the majority of the identified abnormalities in renal morphology were ameliorated in rats treated with bisphenol A after receiving asparagus officinalis extract. This was shown by little glomerular congestion, cell edema with largely intact renal tubules, and acute tubular necrosis levels that did not significantly differ from the control group owing to the decreased lipid peroxidation and enhanced antioxidant capacity. The flavonoids and polyphenols' exitance in *Asparagus Officinalis* extract may be responsible for its antioxidant qualities.

CONCLUSION

Biochemical and histopathological findings demonstrate that formaldehyde has harmful adverse effects on experimental animals. The findings showed that the tissues and functions of the kidneys were improved by employing high doses of aqueous extracts of corn silk and asparagus.

REFERENCES

- Afrin M; Amin T; Karim R and Islam MR (2016):**
Effects of formaldehyde intoxication on liver of Swiss albino mice. *Journal of Agriculture and Veterinary Science*, 9(9): 76-81.
- Al-Snafi AE (2015):**
The Pharmacological Importance of *Asparagus Officinalis* - A Review. *Journal of Pharmaceutical Biology*, 5(2): 93-98.
- Arba PR; Hady AT; Tika LS; Jasn O and Mabrurotul M (2020):**
Hepatoprotective effect of corn silk infusion in male wistar rats. *Eksakta journal of sciences and data analysis*, 1(1);51- 55
- Aukkanit N; Kemngoen T and Ponharn N (2015):**
Utilization of corn silk in low fat meatballs and its characteristics. *Proc. Soc. Behav. Sci*, 197, 1403–1410.
- Bancroft JD and Gamble M (2008):**
Theory and Practice of Histological Techniques. 6th Edition, Churchill Livingstone, Elsevier, China.
- Bakar E; Ulucam E; Cerkezkayabekir A (2015):**
Protective effects of proanthocyanidin and vitamin E against toxic effects of formaldehyde in kidney tissue. *Biotech. Histochem*, 90(1):69–78.
- Bartels H; Bohnzer M and Heierli C (1972):**
Serum creatinine determination without protein precipitation. *Clinical chemistry Acta*, 37: 193.
- Bergmeyer HU; Horder M and Rey J (1986):**
Approved recommendation on IFCC methods for the measurement of catalytic enzymes. Part 3: IFCC method for alanine aminotransferase. *J. Clin.*

Chem. Clin. Biochem., 24:
481–495.

European Journal of Paediatric Dentistry, 4(1):16-20.

Berndt H and Jackwerth E (1979):

Determination of Li, Na, K, Mg and Ca with a mechanized flame photometric micro-method. Mechanized micro-method ("injection method") of flame photometry (atomic absorption--atomic emission) for the determination of serum electrolytes and trace elements (Fe, Cu, Zn). *J Clin Chem Clin Biochem* 17(2):71-6.

Buege JA and Aust SD (1978):

Microsomal lipid peroxidation, *Methods in Enzymology*, 52: 302–310.

Busher JT (1990):

Serum albumin and globulin. Clinical methods: the history, physical and laboratory examinations. 3rd edition. Boston. Chapter: 101.

Birben E; Sahiner UM; Sackesen C; Erzurum S and Kalayci O (2012):

Oxidative stress and antioxidant defense. *World Allergy Organ J.* 5(1):9–19.

Chapman DG; Gastilla R and Campbell TA (1959):

Evaluation of protein in food. I. A. Method for the determination of protein efficiency ratio. *Can. j. Biochem. Physio.* 1(37): 679- 686.

Boj JR; Marco I; Cortés O and Canalda C (2003):

The acute nephrotoxicity of systemically administered formaldehyde in rats.

Checkoway H; Dell LD; Boffetta P; Gallagher AE; Crawford L; Lees PS and Mundt KA (2015):

Formaldehyde exposure and mortality risks from acute myeloid leukemia and other lymphohematopoietic malignancies in the US National Cancer Institute cohort

study of workers in formaldehyde industries. *J. Occup. Environ. Med*, 57(7):785–94.

Chinedu IA; Chukwuemeka OE; Nwabunwanne OV; Ndumnworo OD; Chidiebere OE; Igube NA; Kalu AU and Emmanuel AB (2017):

Effect Of Short-Term Exposure to Formalin on Kidney Function. *European Journal of Biomedical AND Pharmaceutical sciences*, 4 (12): 51-55

Chutima L; Chutipa N; Kamol D; Tharit M; Pana Ch and Sontaya L (2020):

Phytochemical analysis of baby corn silk extracts. *Journal of ayurveda and integrative medicine* ,11(3):344-351.

Cortas NK and Wakid NW (1990):

Determination of inorganic nitrate in serum and urine by a kinetic cadmium-reduction method. *Clin Chem* .36: 1440-3.

Dika L; Esa RV; Novita SP and Grispenjas SP (2018):

Zea mays l to the decreasing blood glucose levels in animal trial (rat) with diabetes mellitus: systematic review. The 9 *International nursing conference*,428-431.

Dika L; Ketut S and Abu B (2020):

The Effect of Corn Silk Ethanol Extract (Zea Mays. L) on Decreasing the Blood Glucose Levels. *International Journal of Nursing and Health Services*, 3(1);96-100.

Drupt F (1974):

Depression of human serum albumin." *Farm. Boil.*, 9: 222-229.

Egwurugwu JN; Ohamaeme MC; Ekweogu C N; Ngwu E E; Ugwuezumba PC; Ogunnaya FU; Azudialu BC; Izunwanne DI; Nwamkpa P; Elendu MU and Eberendu IG (2018):

Effects of Formalin Inhalation on Physical Characteristics and Renal Profile of Albino Wistar.

Asian Journal of Medicine and Health, 12(4):1-11.

El-Seedy GM; sahloul TM and Ismail FA (2022):

The Therapeutic Effect of Corn Silk on Rats with Kidney Stones. *Journal of research in the fields of specific education*.8(38);863-885.

Fathalipour M; Delnavazi M; Safa O; Zarifinia N and Rafiee B (2020):

Antioxidant and antinociceptive effects of hydroalcoholic root extract of *Asparagus officinalis* L. *Physiology and Pharmacology*, 24(4):322-330.

Fossati P; Prencipe L and Berti G (1980):

Use of 3,5-dichloro-2-hydroxybenzene-sulfonic acid/4-aminophenazone chromogenic system in direct enzymatic assay of uric acid in serum and urine. *Clinical chemistry*, 26: 227-231.

Gulec M Gurel A and Armutcu F (2006):

Vitamin E protects against oxidative damage caused by formaldehyde in the liver and plasma of rats. *Mol. Cell. Biochem*, 290(1-2):61-7.

Ha AW; Kang HJ; Kim SL; Kim MH and Kim WK (2018):

Acute and Subacute Toxicity Evaluation of Corn Silk Extract. *Prevention Nutrition and Food Science*, 23(1):70-76.

Hasanudin K; Hashim P and Mustafa S (2012):

Corn Silk (*Stigma Maydis*) in Healthcare: A Phytochemical and Pharmacological Review. *17(8): 9697-9715*.

Henry RJ (1974):

Clinical Chemist principles and Technics, 2nd edition, Hagers town (MD), Harcer, Row, p.8802.

Huang XF; Lin YY and Kong LY (2008):

Steroids from the roots of *Asparagus officinalis* and

their cytotoxic activity, *J. Integr. Plant Biol.* 50 (6) ; 717–722.

[e.com/contents/calculation-of-the-creatinine-clearance](https://www.e.com/contents/calculation-of-the-creatinine-clearance)

Huang Y; Sridhar K and Tsai p (2022):

Enzymatically hydrolysed asparagus (*Asparagus officinalis* L.) hard-stem exhibits the ability to inhibit angiotensin-converting enzyme (ACE). *International Journal of Food Science and Technology*, 57, 3196–3203

Jia YC; Xin Y S; and Jian M O (2020):

Modulation of calcium oxalate crystal growth and protection from oxidatively damaged renal epithelial cells of corn silk polysaccharides with different molecular weights. *Hindawi Oxidative Medicine and Cellular Longevity*, (19),1-19.

Ikpeazu O; Ugbogu EA; Emmanuel O; Ikonke CU and Okoro B (2018):

Evaluation of the Safety of Oral Intake of Aqueous Extract of *Stigma Maydis* (Corn Silk) In Rats. *Acta Sci. Pol. Technol. Aliment*, 17(4): 387–397

Jashni HK; Jahromi HK; Ranjbary AG; Jahromi ZK and Kherameh ZK (2016):

Effects of aqueous extract from *Asparagus officinalis* L. roots on hypothalamic-pituitary-gonadal axis hormone levels and the number of ovarian follicles in adult rats. *J Reprod BioMed*,14(2):75-80.

Inker LA and Perrone RD (2021):

Calculation of the creatinine clearance. In: Sterns RH, ed. UpToDate. Updated June 11, 2021. Accessed June 24, 2021. <https://www.uptodat>

Kamel KM; Abd El-Raouf OM; Metwally SA; Abd El-Latif HA and El-Sayed ME (2014):

Hesperidin and Rutin, Antioxidant Citrus Flavonoids, Attenuate Cisplatin-Induced Nephrotoxicity in Rats. *J. of biochemical and molecular toxicology*,28(7): 312-319.

Kasabe P; Patil P; Kamble D and Dandge P (2012):

Nutritional, Elemental Analysis and Antioxidant Activity of Garden Cress (*Lepidium sativum* L.) Seeds. *Int J Pharm Pharm Sci*, 4 (3): 392-395.

Kim S and Moon A (2012):

Drug-Induced Nephrotoxicity and Its Biomarkers. *Biomolecules and Therapeutics. Biomol Ther*, 20(3):268-272.

Kim BY; Cui ZG; Lee S R; Kim SJ; Kang HK and Lee YK (2009):

Effects of *Asparagus officinalis* extracts on liver cell toxicity and ethanol metabolism. *J. Food Sci*, 74 (7).

Kim SR; Ha AW; Choi HJ; Kim SL; Kang HJ; Kim MH and Kim WK (2017):

Corn silk extract improves benign prostatic hyperplasia in experimental rat model. *Nutrition Research and Practice*. 11(5):373-380.

Luo L; Zhang Z and Lifeng M (2005):

Determination of recombinant human tumor necrosis factor- α in serum by chemiluminescence imaging. *Analytica Chimica Acta*. 539, (1–2); 277-282.

Nishikimi M; Dunmore ODF; Chidiebere OE; Igube NA; Kalu AU; Rae N A and Yagi K (1972):

The occurrence of superoxide anion in the action of reduced phenazine methosulphate and molecular oxygen. *Biochemical and Biophysical Research Communications*, 46: 849-853.

Olisah MC and Meludu S (2021):

A Comparative Study of Serum Cystatin C, Serum

Electrolytes, Urea and Creatinine in Early Detection of Kidney Injuries in Albino Rats Exposed to Formaldehyde. *IPS Journal of Toxicology*,1(1): 1-4.

Pal S; Sarkar K; Nath PP; Mondal M; Khatun A and Paul G (2017):

Bisphenol S impairs blood functions and induces cardiovascular risks in rats, *Toxicology Reports*, 4 ,560–565.

Patton CJ and Crouch SR (1977):

Spectrophotometric and kinetics investigation of the Berthelot reaction for the determination of ammonia: *Analytical Chemistry*, 49(3):464-469.

Persoz C; Achard S; Leleu C; Momas I; Seta N (2010):

An *in vitro* model to evaluate the inflammatory response after gaseous formaldehyde exposure of lung epithelial cells.

Toxicol Lett ,195(2-3): 99–105.

Poormoosavi SM ; Najafzadehvarzi H; Behmanesh MA and Amirgholami R (2018):

Protective effects of *Asparagus officinalis* extract against Bisphenol A- induced toxicity in Wistar rats. *Journal Toxicology Reports*, (5):427-433.

Raja DS (2012).

Potential health hazards for students exposed to formaldehyde in the gross anatomy laboratory. *Journal Environ Health*, 74(6): 36-40.

Reeves P; Rossow K and Lindlauf J (1993):

Development and testing of the AIN-93 purified diets for rodents: results on growth, kidney calcification and bone mineralization in rats and mice. *Journal of Nutrition*, 123(11): 1923-1931.

Roy SE (1970):

Colorimetric determination of serum alkaline

phosphatase". *J. Clin. Chem.*, 16:431-432.

gentamicin-induced nephrotoxicity in rat. *Comp Clin Pathol* 20:89-94.

Saheed S; Oladipipo AE; Abdulazeez AA; Olarewaju SA; Ismaila NO; Emmanuel IA and Aisha AY (2015):

Toxicological evaluations of *Stigma maydis* (corn silk) aqueous extract on hematological and lipid parameters in Wistar rats. *Toxicol. Rep.* 2, 638-644.

Shi YQ; Chen X; Dai J; Jiang ZF; Li N; Zhang BY and Zhang ZB (2012):

Toxicology and Industrial Health. Published online before print November 16.

Sanusi BM; Lawal S and Glori DCH (2020):

Corn silk from waste material to potential therapeutic: a mini review. *FUW Trends in Science & Technology Journal* ,5(3), 816 - 820.

Sinha KA (1972):

Colorimetric Assay of Catalase. *Analytical Biochemistry*, 47: 389-394.

Snedecor GW and Cochran WG (1989):

Statistical Methods. 8th ed., Iowa State University Press, Ames, Iowa 50014, USA.

Sener U; Uygur R; Kurt O; Caglar V; Gelincik I and Gurel A (2015):

Protective Effects of Curcumin against Formaldehyde-induced Renal Toxicity in Rats, *West Indian Med J*, 69 (5): 338.

Somani R; Singhai AK; Shivgunde P and Jain D (2012):

Asparagus Racemosus Willd (Liliaceae) Ameliorates Early Diabetic Nephropathy in STZ Induced Diabetic Rats. *Journal Experimental Biology*,50(7):469-75

Sepehri G; Derakhshanfar A and Zade FY (2011):

Protective effects of corn silk extract administration on

Sonnenwirth A and Jaret L (1980):

Grad Wholes Clinical Laboratory Methods and Diagnosis, 258-259.

Tan T; Zhang Y; Luo W; Lv J; Han C; Hamlin JNR; Luo H; Li H; WanY; Yang X; Song W; Tong Z (2018):

Formaldehyde induces diabetes-associated cognitive impairments. *Journal Official Publication of the Federation of American Societies for Experimental Biology*,32(7): 3669-3679

Tanideh N; Zarifi F; Rafiee S; Khastkhodaei M; Hosseinabadi OK; Tarkesh F; Kherad Z; Taghi MM; Kamali M; Shekarkhar G; Jahromi M and Zarif F (2018):

Effect of Methanolic Extract of Corn Silk on Cisplatin-Induced Nephrotoxicity in Rats. *Galen Medical Journal*, (7).

Treesh SA; Eljaafari HA; Darmun EI; Abu-Aisha AM; alwaer FS; Eltubuly RA; Elghedamsi M And Aburawi S M (2014):

Histological Study on The Effect of Formaldehyde on Mice Liver and Kidney and Possible Protective Role of Selenium. *Journal of Cell and Tissue Research* ,14(2): 4201-4209.

Trinder P (1969):

Determination of glucose in blood using glucose oxidase with an alternative oxygen acceptor. *Ann. Clin.Biochem*, (6): 24-25.

Tzatzarakis MN; Karzi V; Vakonaki E; Goumenou M; Kavvalakis M and Stivaktakis P (2017):

Bisphenol A in soft drinks and canned foods and data evaluation, *Food Addit. Contam*,10 (2): 85–90.

Vijitha TP and Saranya D (2017):

Corn silk -amedicinal boon. *International Journal of Chemtech Research*, 10(10):129-137.

Wang GQ; Lu S; Liu J and Wang C (2012):

Anti-inflammation effects of corn silk in a rat model of carrageenan-induced

pleurisy. *Inflammation*
.35(3): 822–827.

Wu D; Zhirong Jiang MM; Bing Gong MD; Yue Dou MM; Mingxuan Song MM; Xiaoxia Song MM and Yu Tian MM (2017):

Vitamin E Reversed Apoptosis of Cardiomyocytes Induced by Exposure to High Dose Formaldehyde During Mice Pregnancy. *the International Heart Journal Association*,58(5): 769-777.

Yogesh PT; Kanchan AG; Sheetal DK; Kishori GA and Pradeep BP (2013):

Antirolithic activity of corn silk extract in mice. *International Journal of Universal Pharmacy and Bio Sciences*,2(4): 65-77.

Yound, D.S. (1975):

Determination of Got. *J. Chem*, 21-1

Zararsiza I; Sonmez MF; Yilmaz RH; Tasa U; Kusa I; Kavaklia A and Sarsilmaza M (2006):

Effects of v-3 essential fatty acids against formaldehyde-induced. *Journal Toxicology and Industrial Health*, (22): 223 229.

Zhang W; Wu W; Wang Q; Chen Y and Yue G (2014):

The juice of asparagus by-product exerts hypoglycemic activity in streptozotocin-induced diabetic rats. *Journal of Food Biochemistry*, 38(5): 509-517.

Table (1): Protective effect of aqueous extracts of Corn silk and Asparagus on feed intake (FI), body weight gain (BWG %) and feed efficiency ratio (FER) in rats with renal toxicity (mean \pm SD)

Parameters Groups	FI (g) for 28 days	BWG gm (%) preceeding period	FER
-ve Control (G1)	604.33 \pm 2.16 ^a	44.55 \pm 2.84 ^a	0.080 \pm .006 ^a
+ve Control (G2)	412.50 \pm 1.87 ^e	20.72 \pm 2.92 ^e	0.063 \pm .005 ^d
Corn silk 200 mg/ kg BW(G3)	531.70 \pm 2.53 ^d	32.38 \pm 2.36 ^{cd}	0.070 \pm .006 ^c
Corn Silk 400 mg /kg BW (G4)	571.36 \pm 1.97 ^b	41.17 \pm 2.04 ^b	0.075 \pm .008 ^b
Asparagus 200 mg /kg BW (G5)	532.0 \pm 3.03 ^d	29.78 \pm 1.97 ^d	0.068 \pm .007 ^c
Asparagus 400 mg /kg BW (G6)	550.33 \pm 7.11 ^c	35.09 \pm 3.39 ^c	0.072 \pm .008 ^{bc}

Means in the same column with completely different letters are significantly different at $p < 0.05$.

Table (2): Protective effect of aqueous extracts of Corn silk and Asparagus on relative kidney weight in rats with renal toxicity (mean \pm SD)

Parameters Groups	Relative kidney weight gm %
-ve Control (G1)	1.88 \pm 0.023 ^a
+ve Control (G2)	1.00 \pm 0.015 ^d
Corn silk 200 mg /kg BW (G3)	1.77 \pm 0.015 ^c
Corn Silk 400 mg(G4)	1.82 \pm 0.015 ^b
Asparagus 200 mg/ kg BW (G5)	1.75 \pm 0.029 ^c
Asparagus 400 mg /kg BW (G6)	1.80 \pm 0.015 ^b

Means in the same column with completely different letters are significantly different at $p < 0.05$.

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Table (3): Protective effect of aqueous extracts of Corn silk and Asparagus on kidney functions in rats with renal toxicity (mean \pm SD)

Parameters Groups	Creatinine (mg/dl)	Urea (mg/dl)	Uric acid (mg/dl)	Creatinine Clearance (ml/min)
-ve Control(G1)	0.54 \pm 0.05 ^f	22.53 \pm 3.56 ^f	1.75 \pm 0.14 ^e	1.31 \pm 0.097 ^a
+ve Control(G2)	1.31 \pm 0.13 ^a	97.30 \pm 1.46 ^a	3.35 \pm 0.17 ^a	0.350 \pm 0.035 ^e
Corn silk 200 mg/kg BW(G3)	0.84 \pm 0.05 ^c	51.51 \pm 3.22 ^c	2.55 \pm 0.14 ^c	0.79 \pm 0.098 ^c
Corn Silk 400 mg/kg BW(G4)	0.65 \pm 0.05 ^e	29.41 \pm 4.07 ^e	2.00 \pm 0.12 ^d	1.10 \pm 0.108 ^b
Asparagus 200 mg/kg BW(G5)	0.94 \pm 0.04 ^b	61.13 \pm 3.88 ^b	2.77 \pm 0.12 ^b	0.67 \pm 0.092 ^d
Asparagus 400 mg/kg BW(G6)	0.74 \pm 0.04 ^d	40.42 \pm 3.97 ^d	2.03 \pm 0.19 ^d	1.00 \pm 0.097 ^b

Means in the same column with completely different letters are significantly different at $p < 0.05$.

Table (4): Protective effect of aqueous extracts of Corn silk and Asparagus on serum electrolytes in rats with renal toxicity (mean \pm SD)

Parameters Groups	Sodium (Na ⁺) mEq/L	potassium (K ⁺) mEq/L	Calcium (Ca ⁺⁺) mg/dl	Magnesium (Mg ⁺⁺) mg/dl
-ve Control (G1)	142.85 \pm 1.30 ^e	4.61 \pm 0.15 ^e	9.83 \pm 0.17 ^a	3.03 \pm 0.10 ^a
+ve Control (G2)	160.64 \pm 3.69 ^a	5.97 \pm 0.14 ^a	7.78 \pm 0.24 ^e	2.51 \pm 0.07 ^d
Corn silk 200 mg/kg BW (G3)	154.09 \pm 1.17 ^b	5.32 \pm 0.09 ^c	8.63 \pm 0.24 ^c	2.68 \pm 0.09 ^c
Corn Silk 400 mg/kg BW (G4)	147.26 \pm 1.84 ^d	5.01 \pm 0.10 ^d	9.50 \pm 0.10 ^b	2.86 \pm 0.04 ^b
Asparagus 200 mg/kg BW (G5)	153.99 \pm 1.20 ^b	5.55 \pm 0.16 ^b	8.29 \pm 0.16 ^d	2.65 \pm 0.05 ^c
Asparagus 400 mg/kg BW (G6)	150.62 \pm 1.41 ^c	5.42 \pm 0.16 ^{bc}	9.33 \pm 0.16 ^b	2.77 \pm 0.08 ^b

Means in the same column with completely different letters are significantly different at $p < 0.05$.

Table (5): Protective effect of aqueous extracts of Corn silk and Asparagus on blood glucose in rats with renal toxicity (mean ± SD)

Parameters Groups	Blood Glucose (mg/dl)
-ve Control (G1)	77.75±3.68 ^e
+ve Control (G2)	137.00±2.28 ^a
Corn silk 200 mg/ kg BW(G3)	90.00±3.68 ^c
Corn Silk 400 mg/ kg BW (G4)	83.41±2.90 ^d
Asparagus 200 mg/ kg BW (G5)	100.00±3.68 ^b
Asparagus 400 mg/ kg BW (G6)	93.66±2.50 ^c

Means in the same column with completely different letters are significantly different at p<0.05.

Table (6): Protective effect of aqueous extracts of Corn silk and Asparagus on Liver functions in rats with renal toxicity (mean ± SD)

Parameters Groups	ALT(I/U)	AST(I/U)	ALP(I/U)
-ve Control (G1)	25.08±3.55 ^e	97.37±2.44 ^f	268.33±3.20 ^e
+ve Control (G2)	55.08±3.26 ^a	207.95±2.88 ^a	314.91±3.72 ^a
Corn silk 200 mg/ kg BW(G3)	46.16±2.69 ^b	150.04±3.68 ^c	292.91±3.20 ^b
Corn Silk 400 mg/ kg BW (G4)	29.16±3.35 ^d	106.66±3.98 ^e	272.66±2.80 ^d
Asparagus 200 mg/kg BW (G5)	49.08±1.42 ^b	171.16±1.43 ^b	294.00±2.82 ^b
Asparagus 400 mg/kg BW (G6)	36.08±2.69 ^c	130.20±3.70 ^d	281.91±3.04 ^c

Means in the same column with completely different letters are significantly different at p<0.05.

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Table (7): Protective effect of aqueous extracts of Corn silk and Asparagus on total protein, albumin and globulin in rats with renal toxicity (mean ± SD)

Parameters Groups	T. Protein (g/dl)	Albumin (g/dl)	Globulin(g/dl)
-ve Control (G1)	8.30±1.48 ^a	5.12±1.49 ^a	3.18±1.00 ^a
+ve Control (G2)	4.25±0.82 ^c	2.90±0.041 ^d	1.35±0.050 ^b
Corn silk 200 mg/kg BW (G3)	6.25±2.27 ^b	3.76±0.045 ^{b^c}	2.49±1.00 ^a
Corn Silk 400 mg/kg BW (G4)	7.40±1.34 ^{ab}	4.50±0.041 ^{ab}	2.90±0.000 ^a
Asparagus 200 mg/kg BW (G5)	5.80±0.83 ^{bc}	3.02±0.008 ^{cd}	2.78±0.020 ^a
Asparagus 400 mg/kg BW (G6)	6.75±0.82 ^{ab}	3.89±0.059 ^b	2.86±0.030 ^a

Means in the same column with completely different letters are significantly different at $p < 0.05$.

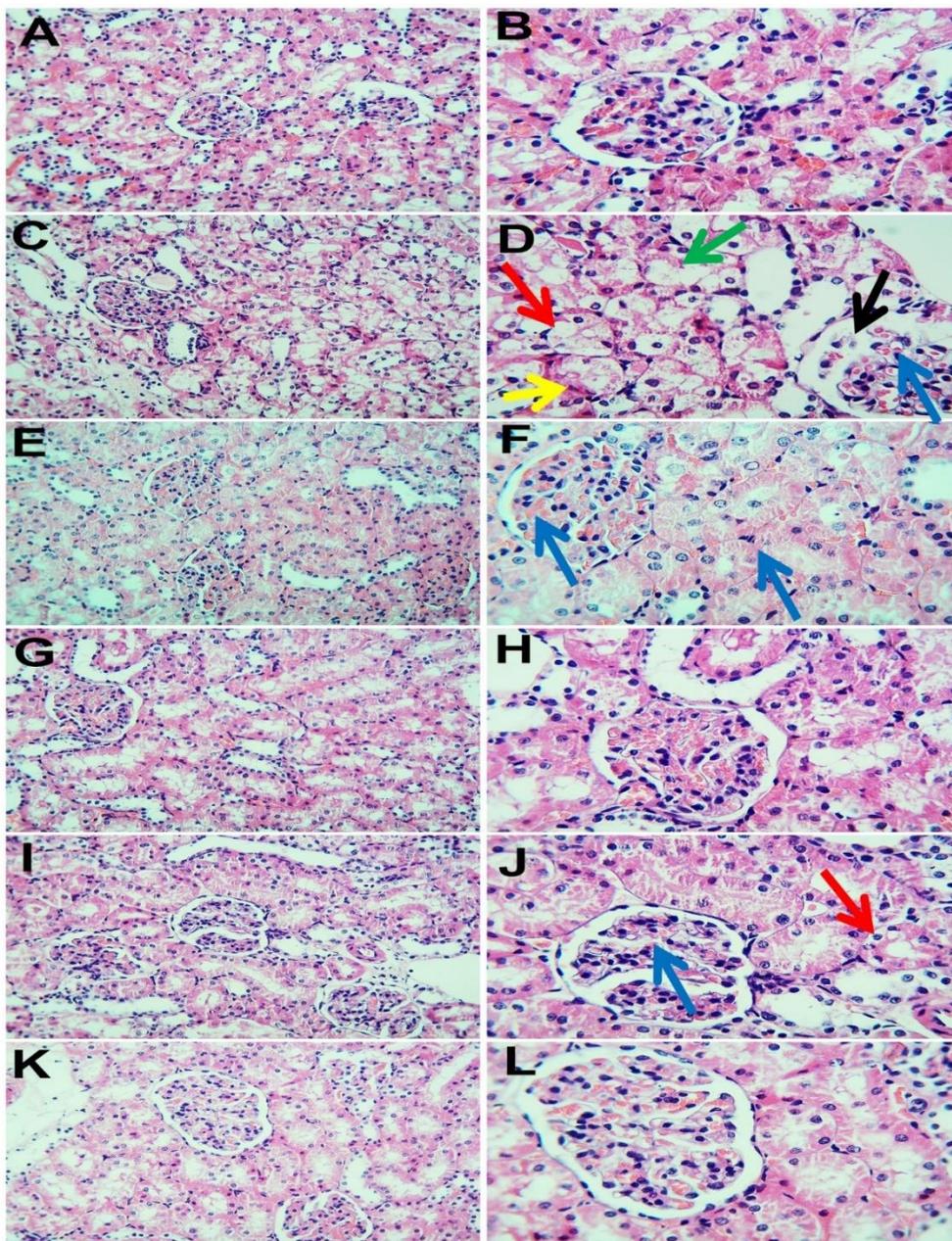
Table (8): Protective effect of aqueous extracts of Corn silk and Asparagus on serum tumor necrosis factor α and CAT, SOD, MDA and NO in kidney tissues of rats with renal toxicity (mean ± SD)

Parameters Groups	Serum TNF- α (pg/ml)	CAT (U/g.t)	SOD (U/g.t)	MDA (nmol/g.t)	NO (μ mol/g.t)
-ve Control (G1)	21.60±1.80 ^f	2.94±0.09 ^a	148.40±2.0 ^a	9.99±1.62 ^e	2.26±0.14 ^d
+ve Control (G2)	90.15±4.05 ^a	1.15±0.21 ^e	61.21±2.0 ^f	23.35±0.74 ^a	6.07±0.31 ^a
Corn silk 200 mg/ kg BW(G3)	51.15±2.15 ^c	2.09±0.19 ^c	112.25±3.0 ^d	16.82±0.62 ^c	4.11±0.20 ^b
Corn Silk 400 mg/kg BW (G4)	28.65±2.95 ^e	2.48±0.17 ^b	133.09±3.0 ^b	12.35±1.39 ^d	3.06±0.20 ^c
Asparagus 200 mg /kg BW (G5)	59.45±3.05 ^b	1.68±0.04 ^d	85.76±3.0 ^e	19.60±0.21 ^b	4.38±0.26 ^b
Asparagus 400 mg /kg BW (G6)	36.30±3.76 ^d	2.32±0.20 ^{bc}	123.80±3.0 ^c	13.35±1.17 ^d	3.22±0.21 ^c

Means in the same column with completely different letters are significantly different at $p < 0.05$.

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Microscopic images of hematoxylin and eosin (H&E x 400) stained kidney sections showing (A) Normal (-ve) Control group. (B) formaldehyde (+ve) Control group. (C) formaldehyde + Corn silk (200 mg /kg) group. (D) formaldehyde +Corn silk (400 mg/ kg BW) group. (E) formaldehyde+ Asparagus (200 mg/kg) group (F) formaldehyde+ Asparagus (400 mg/kg BW)

التأثيرات الوقائية المحتملة للمستخلصات المائية لبعض الاعشاب على التسمم الكلوي المحدث بالفورمالدهيد في جردان التجارب

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أجريت الدراسة الحالية لمعرفة التأثير الوقائي للمستخلصات المائية لكلا من حريرة الذرة والهلبيون ضد السمية الكلوية التي يسببها الفورمالدهيد في جردان التجارب . تم استخدام ستة وثلاثون من ذكور الجرذان تتراوح أوزانهم بين (150 ± 10 جم) قسمت إلي مجموعتين رئيسيتين. المجموعة الأولى (6 جردان) تغذت علي الغذاء القياسي كمجموعة ضابطه ساليه المجموعة الثانية (30 جرد) تم حقنها بالفورمالدهيد في الغشاء البروتوني بمقدار 10 مجم / كجم من وزن الجسم / يوم من اليوم الرابع عشر إلي اليوم الثامن والعشرون وتم تقسيم هذه المجموعة إلي خمس مجموعات فرعية . مجموعه (1) تتغذى علي الغذاء القياسي كمجموعة ضابطة موجبة . مجموعة (2,3) تغذت علي الغذاء القياسي بالإضافة إلي جرعات من المستخلصات المائية لحريرة الذرة (200 و 400 ملجم / كجم من وزن الجسم) عن طريق الفم لمدة 28 يوما .مجموعة (4,5) تغذت علي الغذاء القياسي بالإضافة إلي جرعات من المستخلصات المائية للهلبيون (200 و 400 ملجم / كجم من وزن الجسم) عن طريق الفم لمدة 28 يوما . في نهاية التجربة تم حساب المأخوذ الغذائي، الزيادة المكتسبة في الوزن، معدل كفاءة الغذاء و الوزن النسبي للكلية. كما تم تقدير بعض النماذج البيوكيميائية في السيرم و مضادات الأكسدة والعوامل المؤكسدة في أنسجة الكلية كما أجري الفحص الهيستوباثولوجي لأنسجة الكلية . أظهرت النتائج أن المستخلصات المائية لحريرة الذرة والهلبيون أدت إلى تحسين في التقييم البيولوجي ووظائف الكلية ووظائف الكبد و إلكتروليت المصل والإنزيمات المضادة للأكسدة و الفحص الهستوباثولوجي لأنسجة الكلية مقارنة بالمجموعة الضابطة الموجبة لذلك يمكن أن نستنتج أن تناول المستخلصات المائية لكلا من حريرة الذرة والهلبيون يمكن أن تقلل من تأثير الفورمالدهيد الضارة على الكلية.

الكلمات المفتاحية: حريرة الذرة ، الهلبيون ، الفورمالدهيد ، وظائف الكلية ، الأنزيمات المضادة للأكسدة.