



Effect of Extremely Low Frequency Magnetic Field in Growth, CM Case, Electric Conductivity and DNA of *Aspergillus niger*



CrossMark

Sahar E. Abo-Neima^{1*} and Mohammad M. El-Metwally²

¹Department of physics, Faculty of Science, Damanhur University, Egypt.

²Department of Botany and Microbiology, Faculty of Science, Damanhur University, Damanhur, Egypt.

In the past years the extremely low frequency magnetic field (ELF-MF) of the millimeter range has been actively used in biology, medicine, biotechnology, in this study the impact of electromagnetic radiations in growth, carboxymethylcellulase activity (CM Case), cell conductivity and RAPD DNA pattern of *A. niger* were investigated. The *Aspergillus niger* fungus treated with magnetic field at constant intensity 50Hz, 10mT at different exposure times for five days (2 h/day) Results: the total biomass dry weight inhibited by 72.2% after the first 2 hr. ELF-MF showed also inhibition effect in CM Case activity of *A. niger*. The enzyme activity was decreased from 30 IU/ml to 22.5 IU/ml after only 2hr of exposure and the treated fungus lost about 50% of its CM Case activity after 10 hr. The treated fungus showed also a decrease in conductivity of fungal cells from 1.78×10^{-1} to 0.51×10^{-5} Siemens/meter. The study extended to detect the changes in RAPD patterns of *A. niger* DNA before and after exposure to the 50 Hz frequency. In conclusion, ELF-MF can be used as a safe method in control the activity of highly resistant fungi as *A. niger*.

Keywords : Electromagnetic radiation, CM Case, Electric conductivity, DNA, *A. niger*.

Introduction

Scientists have assumed for several years that the extremely low frequency magnetic field (ELF-MF) produced no major biological impact. Many scientific studies have confirmed in recent decades that extremely low frequency electrical and/or magnetic fields (< 300 Hz) may affect the biological systems [1, 2]. Such fields are generated from electric instruments, electric transmission lines, communication systems, and electronic appliances which affect all living forms in all occupational and residential environments.

In the past years ELF-EMF has been actively used in biology, medicine, biotechnology, etc. The actual mechanism by how this type of radiation effects biological systems is under intense study, But some explanations based on its effect in cell

membranes as the electromagnetic forces at the surface of a membrane could modify ligand-receptor interactions and, as result, would affect the state of the membrane molecules that control the cell transportation besides the conversion of the electromagnetic energy into acoustic-electric waves that affecting the metabolic processes inside the cell [3,4].

Aspergillus niger (*A. niger*) a fungus so widely distributed and ubiquitous in nature. This fungus can spread easily and colonize a wide range of substrates, causing different degree of deterioration and spoilage. Moreover, few studies have focused on the effect of magnetic field on growth and metabolisms of fungi have been published. So the objective of this research is exploring the knowledge of ELF-MF biological effects in *A. niger*.

*Corresponding author : Sahar_amr2002@yahoo.com; Tel : 01229631153

Received : 4/5/2020; accepted : 16/5/2020

DOI :10.21608/ejphysics.2020.28676.1042

©2021 National Information and Documentation Center (NIDOC)

Materials and Methods

Fungal isolate

The *A. niger* isolate used in this study obtained from fungal culture collection of Prof. Dr. Mohammad M. El-Metwally in Mycology Lab., Department of Botany and Microbiology, Faculty of Science, Damanhur University, Damanhur, Egypt.

The *A. niger* was cultivated in modified Czapek-Dox broth medium. The culture medium was prepared by dissolving of the following components into one liter of distilled water: di-potassium hydrogen phosphate KH_2PO_4 15g, $(\text{NH}_4)\text{SO}_4$ (5g), Calcium chloride 0.6 g, magnesium sulfate heptahydrate 0.6 g, iron (II) sulfate heptahydrate 0.005g, and 10 g of carboxy methyl cellulose (CMC). In 100 ml conical flasks 20ml of the medium sterilized by autoclavation and inoculated by 1ml of 10⁶ spore suspension.

Exposure facility system

The liquid medium were exposed to 50Hz, 10 mili Tesla (2 hr/day, for five days) homogenous electromagnetic field generated by four solenoids of 1500 turns each of electrically insulated 2.2mm copper wire, wound around a copper cylindrical chamber of 17cm external diameter as shown in Fig.1. Water was pumped in a copper jacket separating the wire winding and the chamber in order to keep the temperature of the chamber constant during the exposure period. The temperature of the flowing cooling water at the outlet of the jacket and the temperature inside the irradiation chamber were periodically

measured through the use of thermocouple thermometer, which can give readings for the temperature variations within $\pm 0.1^\circ\text{C}$. There was no measurable difference in temperature between the room and the irradiation chamber. The actual current passing in the solenoids was about 1Amper. The coils were connected to a variac fed from the mains (220Volt and 50Hz). The magnetic field exposure system was locally manufactured. The temperature during exposure was maintained by a thermostat in the range of 30°C . The temperature ($30 \pm 1^\circ\text{C}$) in the bottles was measured using a thin Type-K thermocouple thermometer having an accuracy of $\pm 0.2^\circ\text{C}$ and 0.8sec response time. Liquid cultures with *A. niger* was exposed in glass bottles (diameter of 60 mm with heights of 80 mm). Each glass bottle with samples was placed on a nonconductive stand with the height at the middle of the each coil.

Mycelium growth inhibition

The percentage of mycelium growth inhibition was computed from the equation (1):

$$\text{Inhibition \%} = \frac{\text{Average growth of control fungal colony} - \text{Average growth of treated fungal colony}}{\text{Average growth of control fungal colony}} \times 100 \quad (1)$$

Effect of magnetic radiation on *A. niger*

Effect on growth

The fungal mycelium was harvested after every 24 hr of growth, separated from the culture liquid by filtration through a Whatman No. 1 filter paper. The mycelial pellet was repeatedly washed with distilled water and dried at 80°C overnight. The dry weight of the fungus was calculated by using the following equation (2):

$$\text{Dry weight} = (\text{weight of filter paper} + \text{mycelium}) - (\text{weight of filter paper}) \quad (2)$$

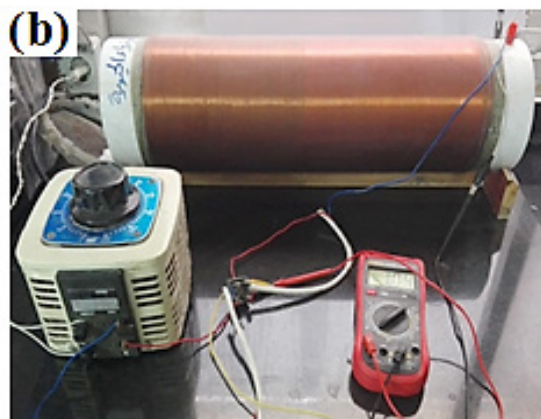
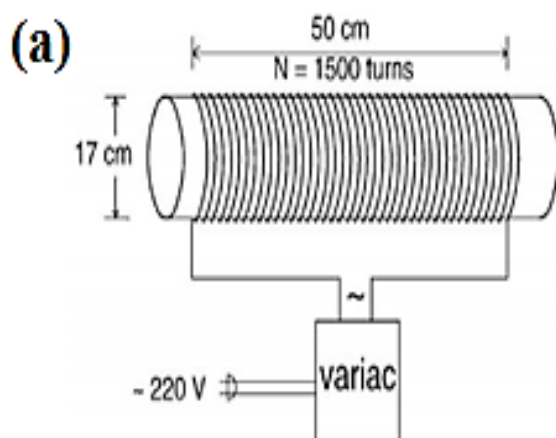


Fig.1. Schematic diagram (a) and photo (b) for exposure facility system.

Assay of carboxymethyl cellulase (CMCase) activity

CMCase activity was determined using the method recommended by Acharya et al. (2008) [5]. The reaction mixture contained 0.5 ml of 0.5% of CMC substrate prepared in 0.5M sodium acetate buffer pH 5.5 and 0.5 ml of enzyme extract. The control sample contained the same amount of substrate and 0.5 ml of the enzyme solution heated at 100°C for 15 min. Both the experimental and control samples were incubated at 50°C for 30 min. At the end of the incubation period, tubes were removed from the water bath, and the reaction was terminated by addition of 3ml of 3, 5-dinitrosalicylic acid (DNSA) reagent per tube [6]. The tubes were incubated for 5 min in a boiling water bath for color development and then were cooled rapidly. The activity of reaction mixture was measured against a blank sample at wavelength of 540 nm. The concentration of glucose released by enzymes was determined by comparing against a standard curve constructed similarly with known concentration of glucose. Unit enzyme activity was defined as the amount of enzyme required for liberating 1 μ M of glucose per milliliter per minute.

*Conductivity measurements of *A. niger* cells*

For conductivity measurement, a fungal sample (1ml) was placed into sterile micro centrifuge tube and centrifuged at 14,000 rpm at 4°C for 15 min. The pellet was then harvested and resuspended in a 1 ml volume of sterile deionized

water. The tube was then centrifuged and the pellet was washed with deionized water twice more, before finally being resuspended in sterile deionized water. The dielectric measurements were carried out for the samples at frequency range 50Hz using a loss Factor Meter type HIOKI 3532 LCR Hi TESTER; version 1.02, Japan, and cell types (PW950/60) manufactured by Philips as shown in Fig. 2. The cell has two parallel square platinum black electrodes of 0.8 cm side each, and area 0.64cm², with an inter-electrode distance of 1cm. During the measurements both the cell and the sample were kept at 25°C in an incubator (Kottermann type 2771, Germany). Each run was repeated three times.

DNA analysis

Isolation of DNA

DNA was extracted from 50 mg of fresh cultures of fungi (either control or exposed cultures) according to this method which is an improved method of the standard phenol/chloroform method [7]. To extract the DNA from fungi, 1ml cell suspension was centrifuged at 8000 rpm for 2min, to pellet cells. After removing the supernatant, the cells were washed with 400 μ l STE Buffer (100mM NaCl, 10mM Tris/HCl, 1mM EDTA, pH 8.0) twice. Then the cells were centrifuged at 8000 rpm for 2min. The pellets were resuspended in 200 μ l TE buffer (10mM Tris/HCl, 1mM EDTA, and pH 8.0). Then 100 μ l Tris-saturated phenols (pH 8.0) were

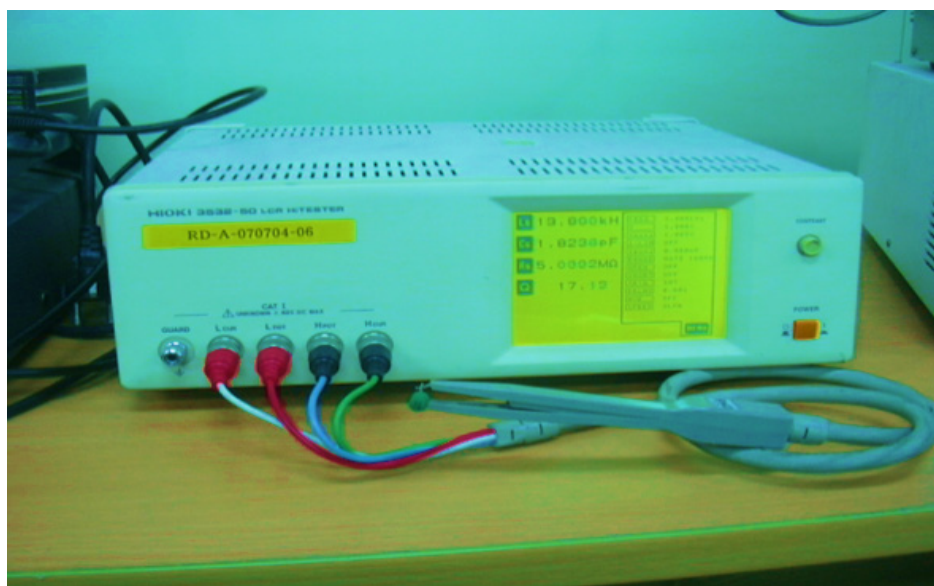


Fig. 2. Conductivity measurements (Hioki, 3532-50 LCR Hitester).

added to these tubes, followed by a vortex-mixing step of 60 second to lyse cells. The samples were subsequently centrifuged at 13000 rpm for 5 min at 4°C to separate the aqueous phase from the organic phase. 160 µl upper aqueous phases were transferred to a clean 1.5 ml tube. 40µl TE buffer were added to make 200µl and mixed with 100µl chloroform and centrifuged for 5min at 13000 rpm at 4°C Lysate was purified by chloroform extraction until a white interface was no longer present; this procedure might have to be repeated three times. 160µl upper aqueous phases were transferred to a clean 1.5ml tube. 40 µl TE and 25µl Proteinase K (modified instead of 5µl RNAs) were added and incubated at 37°C for 10min to digest RNA. Then 100µl Chloroform were added to the tube, mixed well and centrifuged for 5min at 13000 rpm at 4°C .150µl upper aqueous phase were transferred to a clean 1.5ml tube. The aqueous phase contained purified DNA and was directly used for the subsequent experiments or stored at 20°C. The purity and yield of the DNA were assessed spectrophotometrically by calculating the A260/A280 ratios and the A260/A280 values to determine protein impurities and DNA concentrations [8].

Quantization of DNA sample

Two types of methods were used to measure the amount of nucleic acid in a preparation:

I) Spectrophotometer quantization of the extracted DNA

The ratio between the reading of optical density for the DNA samples at 260 nm and 280 nm (OD260/OD280) provides an estimate of the purity of the nucleic acid was done as follow [9]. 1ml TE (Tris–EDTA) buffer was taken in a cuvette and calibrate the spectrophotometer at 260nm as well as 280nm. 10 µl of each DNA sample was added to 900 µl TE buffer and mixed well. TE buffer was used as a blank in the other cuvette of the spectrophotometer. The OD260 and OD280 ratio was calculated. The following comments were considered that the ratio between “1.8-2.0” denotes that the absorption in the UV range is due to nucleic acids and a ratio lower than 1.8 indicates the presence of proteins and/or other UV absorbers. Also a ratio higher than 2.0 indicates that the samples may be contaminated with chloroform or phenol. In either case (<1.8

or > 2.0) it is advisable to reprecipitate the DNA. The amount of DNA was quantified using the formula [9] in equation (3):

$$DNA\ concentration\ \left(\frac{\mu g}{ml}\right) = \frac{[OD_{260} \times dilution\ factor \times 50 \frac{\mu g}{ml}]}{100} \quad (3)$$

II) Quantization of the DNA quality by means of agarose gel

This method is based on comparing the DNA extracted sample with DNA of known concentrations and was done as follows: Prepare a 0.8% agarose gel. Add 1l of 6X gel loading dye to 2-3l of each DNA sample before loading the wells of the gel. Addition of dye allows us to note the extent to which the samples might have migrated during electrophoresis, so that it can be halted at an appropriate stage. Load at least 2 wells with uncut, good quality DNA or any previously quantified DNA samples (50ng and 100ng) as molecular weight standards. Run the submarine electrophoretic gel at 70Volt till the dye has migrated one-third of the distance in the gel. DNA can be visualized using a UV trans-illuminator and quantified in comparison with the fluorescent yield of the standards.

Random amplified polymorphic DNA technique (RAPD-PCR)

Five primers were tested Codes, nucleotide sequences and G+C percentages of tested primers used in the RAPD reaction are shown in Table 1. The reaction was carried out in a DNA Thermocycler (MJ Research Inc. USA). Reactions without DNA were used as negative controls. A stock buffered solution containing 250µl of 10×PCR buffer, 12.5µl at a concentration of 125 milimole (mM) for each dATP, dTTP, dGTP, Dctp and 100µl of 25Mm MgCl2 was prepared in 1.5ml Eppendorf tube. The primers were used at a concentration of 20 icogram, and double distilled water was added to bring the volume of the stock buffer solution to 1.5 ml. A volume of 5µl of the target DNA was added to 44µl of the stock solution in PCR tubes and mixed by vortexing. A volume of 1µl of Taq DNA polymerase (Perkin Elmer, Amersham, USA) was used at a concentration of 5.0 units. To avoid evaporation of the PCR mixture during the high temperature of the thermal cycling profiles. A drop of mineral oil was added to cover the reaction mixture [10].

TABLE 1. Code, nucleotide sequence and G+C (%) of arbitrary primers used in the random amplified polymorphic DNA (RAPD) reaction.

Primer	Sequence 5'-3'	(%) G+C	Annealing (°C)
P1	ATGCCC CTG T	60	32
P2	GAG CCA GTG TCT GCT TTG	56	30
P3	CGC TGT CGC C	80	28
P4	GAG CCA GTG TCT GCT TTG	56	30
P5	TGAAGGGGGGAACCCCTGTG	65	62

All PCR amplification reactions were carried out in a final volume of 50µl. The thermal cycling profiles were as follow a 2-min incubation at 95°C, followed by 40 cycles of 94°C for 1min, 55°C for 30 sec and 72°C for 45sec, and a final incubation at 72°C for 10min. Thermal profiles were performed on a Techne PHC-2 thermal cycler (Techne, Princeton, N.J.U.S.A) following amplifications, 20µL from each PCR reaction containing amplified product were loaded onto gels of 1.5% Seakemagarose (FMC Bio product, Rockland ME., U.S.A) and electrophoresed. The gels were stained with ethidium bromide and the PCR products were identified following visualization under UV light [9].

DNA Electrophoresis

For all samples, the amplified DNA (15µl) was electrophoresed using electrophoresis unit (wide mini-sub-cell GT Bio-Rad) on 2% agarose containing ethidium bromide (0.5µl/ml), at a constant 75volt and 60mA, and visualized with UV trans-illuminator.

Gel analysis

DNA gel was scanned for band, using gel documentation system (AAB Advanced American Biotechnology 1166 E. Valencia Dr. Unit 6 C, Fullerton, CA 92631). The different molecular weights of bands were determined against a DNA standard (100bp DNA Ladder, Stragene, and Canda) with molecular weights. 100, 200, 300, 400, 500, 600, 700, 800, 900, and 1000bp. The similarity level was determined by un-weighted pair group method based on arithmetic mean (UPGMA) [11].

Results

Effect of magnetic radiation in growth of *A. niger*

When *A. niger* treated with ELF-MF at constant intensity 50 Hz, 10mT at different

exposure times for five days (2h/day) the data in Fig. 3 revealed that as exposure time increased the total biomass dry weight decreased. After only 2 hr of exposure the fungal growth inhibited by 72.2% and the inhibition degree increased with exposure time to complete inhibition after 10 hr of exposure.

Effect of magnetic field in CMCase activity of *A. niger*

The data in Fig. 4 showed the inhibition effect of magnetic radiation in CMCase activity of *A. niger*. The enzyme activity was decreased from 30 U/ml to 22.5 U/ml after only 2hr of exposure and the treated fungus lost about 50% of its CMCase activity after 10 hr.

Effect of magnetic radiation in cell conductivity of *A. niger*

The results in Fig. 5 show a decrease in conductivity of fungal cells with increase in time of exposure from 1.78×10^{-1} S/m in control to 0.98×10^{-3} S/m after 2 hr of exposure 0.52×10^{-5} after 8 hr of exposure in the 4th day.

Effect of magnetic radiation in DNA of *A. niger*

Figure 6 show electrophoresis RAPD patterns for the nuclear DNA extracted from *A. niger* before and after exposure to magnetic field (B=10mT, 50Hz) after 6 days (2hr /day). These patterns were detected using different random primers, P1) 5/-ATGCCC CTGT-3/ P2) 5/- CGC TGT CGC C-3/ P3) 5/- GAG CCA GTG TCT GCT TTG -3 /P4) 5/- GAG CCA GTG TCT GCT TTG-3/ P5) 5/-TGAAGGGGGGAACCCCTGTG-3/ respectively. The RAPD pattern for exposed and unexposed *A. niger* in Fig.6. Reveal the appearance of new bands in the amplified DNA for five primers the results shows for control *A. niger* five bands, as a result of exposure to magnetic field for 120 min this number of bands have been decreased to three bands. For P1 the

disappearance of two bands between 900pb and 1000pb and for P2 the appearance of one band between 800pb and 900pb respectively, for P3 the appearance of three bands one of them between 300pb and 400pb and the other appears between 700pb and 800pb and one band at 1000pb. For P4 the appearance of two bands one of them at

300pb and the other between 500 pb and 600pb. For P5 show the appearance of two stuck thick fragments with each other between 700pb & 800pb and 900pb & 1000pb, all five primers exhibit the appearance of new bands as a result of exposure which may be considered as a marker for genetic alterations in the DNA of the exposed microorganism.

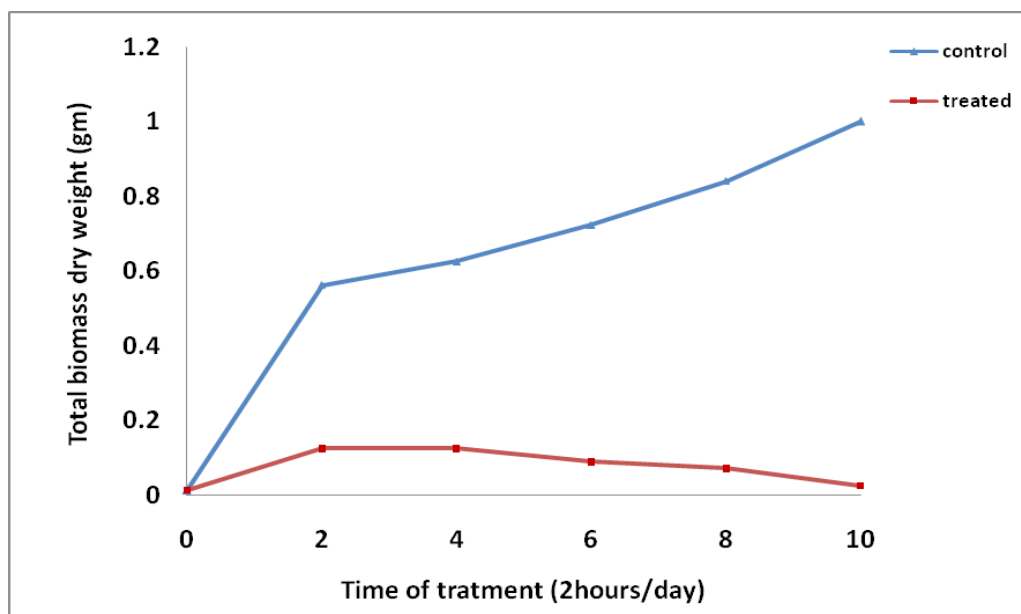


Fig. 3. Total biomass dry weight of fungus *A. niger* in treated fungi as compared to negative control.

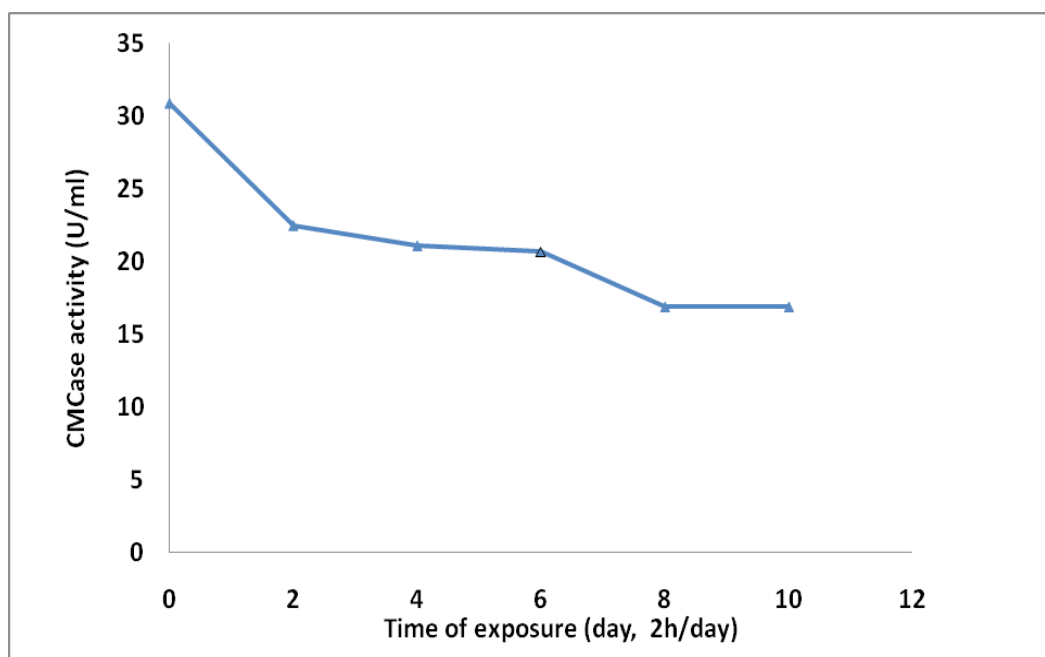


Fig. 4. Effect of magnetic radiation in CMCase activity of *A. niger* (B = 10 mT).

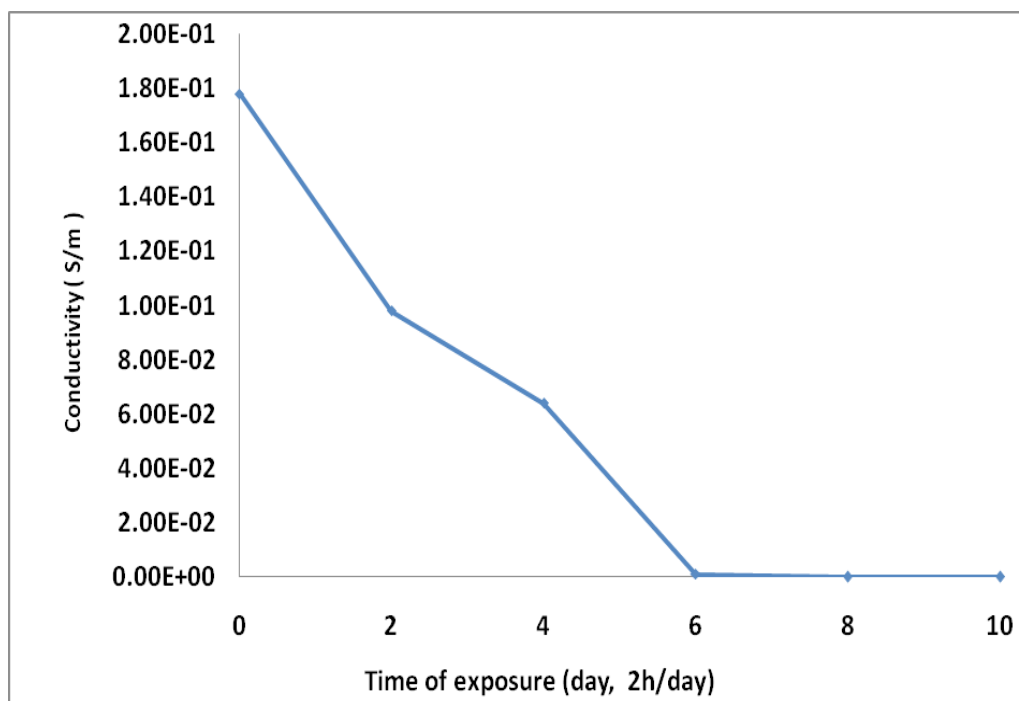


Fig. 5. conductivity of *A.niger* fungi exposed to magnetic fields.

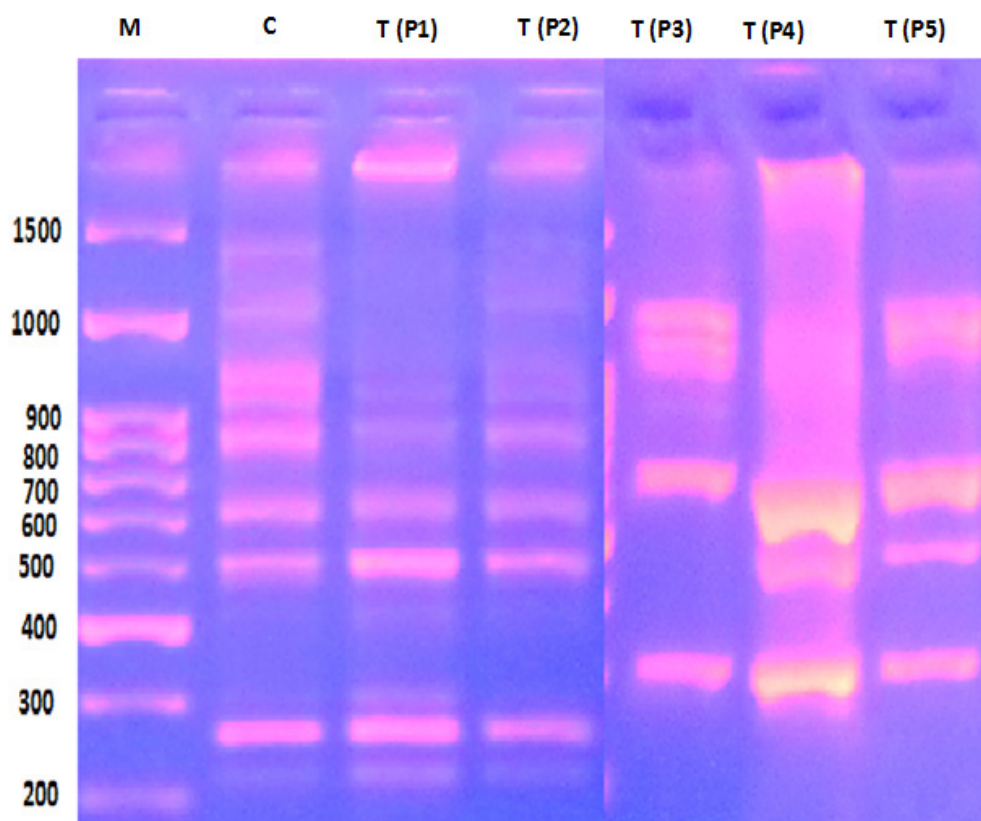


Fig .6. Electrophoresis RAPD Profiles of *non-irradiated* and irradiated *A. niger* with magnetic field M=DNA ladder (DNA marker), C=DNA of control sample,T= treated sample. (p1: primer 1,p2: primer 2, p3: primer 3, p4: primer 4 and p5: primer 5).

Discussion

The number of studies of the effect of electromagnetic waves on microorganisms has increased significantly in the last decades. But little studies have been done in fungi [12, 13]. In the present study, the data confirm that ELF-MF have the potential to reduce the growth of *A. niger* when exposed to magnetic field at 50 Hz, 10mT. The results indicate a sharp decreased in the total biomass dry weight with variation of exposure time as compared with control culture which can explained by changes in metabolic activity or electrostatic properties of the cell surface or both which appears in the decreased in fungal cell conductivity. But knowledge about possible effect low frequency or static magnetic fields on fungi is very confusing, Nagy and Fischl (2004) [14] for example, it has been reported that magnetic fields produce morphological changes on the conidia of *Aspergillus puniceus* and *Alternaria alternata*, as well as changes in colony pigmentation of *Aspergillus niger* [15]. In contrast, Ruiz-Gómez et al. (2004) [16] demonstrated that magnetic fields have no effect on fungal growth, and others found that magnetic fields depressed it [17]. In the same connection, Growth and sporulation of phytopathogen microscopic fungi were studied under a static magnetic field. The applied flux densities were 0.1, 0.5 and 1mT. The magnetic field decreased the growth of colonies by 10% using this flux density region. At the same time, the number of the developed conidia of *Alternaria alternata* and *Curvularia inaequalis* increased by 68-133 percent, but the number of *Fusarium oxysporum* conidia decreased by 79-83 percent [18]. Al-Mayah and Ali concluded that, when magnetic field are applied at constant frequency, with high energy and for a sufficiently long period of time, their thermal effect is most likely dominant and kills fungal cells[19]. Generally, the negative or positive effect of ELF-MF on microorganisms depends on the strength and frequency of the electromagnetic field applied, and microbial strain used [20, 21].

Fungal cells contain charged ions, polarized molecules and electric fields which obey the laws of electromagnetism and thermodynamics. For example, every cell possesses a resting transmembrane potential, and the absence of such a potential is clear evidence of a non-functional dead cell. The impact of the magnetic field energy lies in the stimulus to the events

of significant changes in the characteristics of metabolic organisms; these are changes in the exchange of ions through the cell membrane [22]. The main theories based on the possible effects on the permeability of the ionic channels in the membrane; this can affect ion transport into the cells and result in biological changes [23, 24, 25] as well as ATPase activity [26]. The magnetic fields can interact with atom and molecules of the cell to produce free radicals, which are able to diffuse enough to interact with the cell and cause damage for the fungal cell which prevent cell to cell communications that leads to cell died. So, when we try to explain the decrease and increase in biomass, it will regarded to the effect of magnetic field on the calcium signal transduction [27, 28].

The decrease in cell conductivity of exposed fungus culture is an indicator for the inhibition of fungal cells, which explain the mechanism of interaction of ELF-MF with fungi i.e. as a result of exposure time increased the heating effect of ELF-MF could be the result of Faraday induction this means that the electric fields and currents leads to charge movements, which is related to the electrical parameters of the biological cell so living cell changes under the influence of ELF-MF exposure; that affects in cell structure and composition which leads to decreased the cell conductivity.

Cellini et al. (2008) [29] studied the genetic effect of electromagnetic fields on bacterial DNA when exposed to 50 Hz, DNA fingerprinting revealed no obvious differences among the DNA patterns at each conditions of study. Potenza et al., 2004 [13] when recording DNA changes during RAPD-PCR studies found that exposure to magnetic fields radiation can induce DNA alterations normalized in organisms with cellular protective responses.

Conclusion

Application of magnetic field induces changes in the metabolism characteristic of *A. niger*. These changes indicate the role of ELF-MF in inhibition the growth, CMCase productivity of the fungus and made variations in DNA molecular structure. This result open a wide door on alternative and save physical method in fungal growth control.

Declaration of interest

The authors report no conflicts of interest.

References

- [1] Volpe P. Interaction of zero-frequency and oscillating magnetic fields with biostructures and biosystems. *Photochemical and Photobiological Sciences*, 2003; 2:637-48
- [2] Simko M. Induction of cell activation processes by low frequency electromagnetic fields. *Scientific World Journal*, 2004; 4 (2):4–22.
- [3] Dobson ST, Pierre TJ, Weiser HG, Fuller MR. Changes in paroxysmal brainwave patterns of epileptics by weak field magnetic stimulation. *Bioelectromagnetics J.* 2000; 21: 423-30.
- [4] Pirogova E, Vojisavljevic V, Cosic I. Biological Effects of Electromagnetic Radiation, in Biomedical Engineering, *Vukovar In-Tech.* 2009; 87–06.
- [5] Acharya, S. N.; Thomas, J. E.; Basu, S. K., 2008. Fenugreek, an alternative crop for semiarid regions of North America. *Crop Sci.* 2008; 48 (3): 841-53.
- [6] Shazia KM, Hamid M, Ammad AF, Ikram-U. Optimization of Process Parameters for the Biosynthesis of Cellulases by *Trichoderma Viride*. *Pak. J. Bot.* 2010; 42(6): 4243-251.
- [7] Neumann G, van Noord G. Self-monitoring with reversible grammars. In: *Proceedings of the 14th International Conference on Computational Linguistics (COLING)*. 1992; p.700-06.
- [8] Hai-Rong C, Ning J. Extremely Rapid extraction of DNA from bacteria and yeasts. *Biotechnol. Lett.* 2006; 28: 55-59.
- [9] Kaushlesh K, Yadav MS, Sarad KM, Nasruddin Alam MD, Khurshid AM molecular characterization of cellulose degrading bacteria on the basis of 16s rRNA. 2012. *Journal of Recent Advances in Applied Sciences (JRAAS)*. 2012; 27:80-92.
- [10] National research centre for spices (ICAR), Kerala, p 63 Fevzi B (2001): Random Amplified Gujarat Agricultural university, Sardar krishinagar, Dantiwada Karkachi NE, Gharbi S, Kihal M, Hemmi JE (2010) *Biological* 44 H.P.
- [11] Eberswalde FH (2009): Orduung fur das praktische Studiensemester des student-ganges Landschaftsnutzung und Naturschutz (Bachelor of Science). Anlage 2 zur studien-und prufungsordnung. URL: 6F60AFB-D670-4160-882D-BEB783396722/outline/WWW.hnce.de/app/so.asp?o=obj/E6F60AFB-D670-4160-882D-BEB783396722/outline/ LN-BSc-SPO-WS-2009-Anlage-2- Praktikumsordnung.pdf.
- [12] Nagy P. The effect of low inductivity static magnetic field on some plant pathogen fungi, *Journal of Central European Agriculture*, 2005; 6:167-72.
- [13] Potenza L, Saltarelli R, Polidori E, Ceccaroli P, Amicucci A, Zeppa S, Zambonelli A, Stocchi V. Effect of 300 mT static and 50 Hz 0.1 mT extremely low frequency magnetic fields on *Tuber borchii* mycelium. *Can. J. Microbiol.* 2012; 58(10):1174–182.
- [14] Nagy P, Fischl G. Effect of static magnetic field on growth and sporulation of some plant pathogenic fungi. *Bioelectromagnetics*, 2004; 25(4): 316–18. doi:10.1002/bem.20015. PMID: 15114642.
- [15] Sadauskas KK, Lugauskas AY, M AI. Effects of constant and pulsating low-frequency magnetic field on microscopic fungi. *Mikol.* 1987; 21:160-62.
- [16] Ruiz-Gómez MJ, Prieto-Barcia MI, Ristori-Bogajo E, Martínez-Morillo M. Static and 50 Hz magnetic fields of 0.35 and 2.45 mT have no effect on the growth of *Saccharomyces cerevisiae*. *Bioelectrochemistry*. 2004; 64(2):151-5.
- [17] Novák J, Strašák L, Fojt L, Slaninová I, Vetterl V. Effects of low-frequency magnetic fields on the viability of yeast *Saccharomyces cerevisiae*. *Bioelectrochemistry*, 2007; 70 (1): 115–21. doi:10.1016/j.bioelechem.2006.03.029. PMID: 16713383.
- [18] Pal M. Random forest classifier for remote sensing classification. *Int. J. Remote Sens.* 2005; 26(1): 217-22.
- [19] Al-Mayah A, Eman TA. Mobile microwave effect on bacterial antibiotic sensitivity. *Bas, J. Vet. Res.* 2010; 10 (2):89-03.
- [20] Fojt L, Strašák L, Vetterl V, Šmarda J. Comparison of the low-frequency magnetic field effects on bacteria *Escherichia coli*, *Leclercia adecarboxylata* and *Staphylococcus aureus*, *Bioelectrochemistry*, 2004, 63: 337-41.
- [21] Justo OR, Perez VH, Alvarez DC, Alegre RM. Growth of *Escherichia coli* under extremely low-frequency electromagnetic fields. *Applied Biochemistry and Biotechnology*, 2006; 134:155-63.

-
- [22] Li J, Menzel WP, Sun F, Schmit TJ, Gurka JJ. AIRS subpixel cloud characterization using MODIS cloud products. *J. Appl. Meteorol.* 2004; 43: 1083–94.
 - [23] Matthews EK. British Medical Bulletin, Calcium and membrane permeability. *Microbio. Gene J.* 1986; 42: 391-97.
 - [24] Galvanoskis J, Sandblom J. Theoretical studies of the effects of low – frequency field on the magnitude of oscillation. *Bioenergy J.* 1998; 46: 161-74.
 - [25] Shckorbatov YG, Shakhbazov VG, Gorobets NN, Kiyko VI, Zhuravleva LA, Navrotskaya VV. The Influence of Electromagnetic Radiation of Millimeter and Centimeter Range on Human Epithelial Cells. *Proceedings of 10th International Crimean Conference “Microwave and Telecommunication Technology” Crimea;* 2000, 557–558.
 - [26] Richard G, Stevens, The effect of low inductivity static magnetic field on some plant pathogens fungi. *J. Cent. Eur. Agric.* 2005; 6: 167-71.
 - [27] Blank M, Goodman R. Stimulation of stress response by low frequency electromagnetic fields: possibility of direct interaction with DNA. *Bioelectromagnetics J.* 2000; 28: 168-72.
 - [28] May AE, Snoussi SR, Miloud NB, Maatouk LB, Aissa RB. Foodborne Effect of static magnetic field on cell growth, viability, and differential gene expression in salmonella. pathog, *Dis. J.* 2009; 6: 547-52.
 - [29] Cellini L, Grande R, Di Campli E. Bacterial response to the exposure of 50 Hz electromagnetic fields. *Bioelectromagnetics*, 2008; 29: 302–11.