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# Treatment Effect of Oregano Essential Oil in Broiler Chickens Experimentally Infected with Avian Pathogenic *Escherichia coli* O27 with Emphasis on Hemogram, Serum Biochemistry, and Histopathology of Vital Organs



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#### Abstract

This study aimed to evaluate the efficacy of oregano oil 20% for the treatment of broiler chickens experimentally infected with avian pathogenic *Escherichia coli* O27 (APECO27) -local isolate. Hemogram, biochemical constituents, and histopathology of the internal organs were assessed. Two hundred chicks -19-days-old- were randomly assigned to four equal groups; G1 (normal control), G2 (Infected), G3 (oregano treated), and G4 (antibiotic treated). At the 19<sup>th</sup>, 20<sup>th</sup>, and 21<sup>st</sup> day of age, G2, G3, and G4 were intratracheally infected with 0.2ml of APECO27 (1×10<sup>7</sup> CFU/chick). On the 22<sup>nd</sup>, 23<sup>rd</sup>, and 24<sup>th</sup> day of age, G3 & G4 were treated with oregano 20% (2ml/5liters) and difloxacin (10mg/kg b.wt.) in drinking water, respectively. On the 3<sup>rd</sup>, 6<sup>th</sup>, 10<sup>th</sup>, and 21<sup>st</sup> day post-infection, blood and tissue samples were collected for laboratory investigations of the hemogram, serum biochemistry, and histopathology of the internal organs. Infected non-treated G2 chicks showed dullness, ruffled feathers, gasping and loss of appetite, while infected-treated G3 & G4 birds showed mild symptoms. The mortality rate was 30%, 8% and 4% in G2, G4 and G3, respectively. Chicks of G2 & G4 developed macrocytic hypochromic anemia, leukocytosis, heterophilia, and lymphopenia. Values of ALT, AST, creatinine, and uric acid were increased while protein profile was decreased. After oregano treatment, alterations were returned to normal levels. Histopathologically, chicks of G2 & G4 showed alterations in the internal organs. Finally, APECO27 caused alterations in hematology and serum biochemistry, and histopathology of the internal organs. Oregano could be used as antibiotic for the treatment of APECO27.

Keywords: Avian pathogenic E. coli O27; oregano oil 20%; hemogram; serum biochemistry; pathology, chickens.

# 1. Introduction

Avian colibacillosis caused by avian pathogenic Escherichia coli (APEC), is one of the most significant and widespread infectious diseases occurring in production birds. It is responsible for large financial losses for the poultry industry in Egypt and worldwide every year due to mortality, lost production, and condemnations [1]. Diagnosis of avian colibacillosis is primarily based on the clinical signs and characteristic lesions at necropsy, such as airsacculitis, perihepatitis, and pericarditis. If colibacillosis is suspected, isolation and identification of the pathogen is recommended [2]. The recorded E. coli serotypes are more than 1,000 but fewer have been implicated in poultry diseases, the most prevalent serotypes were O115, O142, O158, O55, O125, O114, O27, O20, and O15. Awad et al. [3] reported that by application of polymerase chain reaction (PCR), 83.3% (15/18) of the serotyped

isolates were confirmed to be E. coli, of which 93.3% (14/15), 46.6% (7/15), and 20% (3/15) of isolates harbored the iss, iutA, and fimH genes, respectively. These strains have specific virulence factors contributing to their ability to cause diseases in birds. Infections by these strains are mostly extra intestinal, with a majority being the cause of acute lung injury due to disruption of the lung endothelial and epithelial barriers.

Commercial antibiotics' nontherapeutic usage could cause tolerance or resistance in humans and animals. So, a worldwide ban has been forced on the usage of antibiotics in order to decrease antibioticresistant bacterial traits in poultry [4]. Oregano (*Origanum vulgare* L.), is a common species of genus origanum, Family Lamiaceae. Oregano essential oil is an aromatic herbal product that has been used as a growth promoter in the poultry farms [5]. Phenolic

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compounds such as carvacrol and thymol and their pre-cursors p-cymene and y-terpinene constitute about 78 to 82% of the total Oregano's volatile compounds. Such compounds have been demonstrated to have synergistic/additive effects, such as antiviral and antifungal properties [6,7], acaricidal activity [8], antimicrobial activities [9-13], and antioxidant [14,15] properties. Also, carvacrol-rich oregano oil has high antibiofilm and antivirulence activities against uropathogenic E. coli [16]. The antimicrobial activity of oregano essential oil has been explored in many in vitro assays which showed that carvacrol, thymol, and eugenol have high antimicrobial activity against pathogenic bacteria such as E. coli and Salmonella typhimurium which are potential risk factors of enteric infections [17]. Oregano could be a viable alternative to antibiotic growth promoters provided that there are no negative effects on the birds' health and the efficiency of controlling the pathogen load in the gastrointestinal tract. M'Barek et al. [18] on the other side mentioned that, carvacrol and other oregano compounds can be cytotoxic at high doses indicating the need for optimizing the concentration of oregano in feed to achieve maximum positive effects without cytotoxic effects.

It has been reported that chicken infected with *E. coli* revealed pancytopenia [19] and elevation of the activities of serum enzymes AST and ALT and values of serum uric acid, creatinine and globulin with decrease of serum protein and albumin [20]. Therefore, this work aimed to evaluate efficacy of oregano essential oil 20% for treatment of chicks experimentally infected with Avian Pathogenic *E. coli* O27 (APEC O27) strain- local isolate-. Hematological, serum biochemical and histopathological changes in the vital organs (lungs, trachea, liver, spleen and kidneys) were utilized for assessment.

#### 2. Material and Methods

#### 2.1. Ethics statement

This experiment was conducted according to guidelines for animal experimentation and the protocol approved by the Institutional Animal Care and Use Committee, National Research Centre, Dokki, Giza, Egypt, the protocol approval No.: 1276072021.

# 2.2. Avian Pathogenic Escherichia coli O27 (APEC 027)

Avian pathogenic *E. coli* serotype O27 –local isolate from lungs of broiler chickens- was used in this experimental infection. The APEC O27 was identified at Animal Health Research Institute, Dokki, Giza, Egypt. The result revealed that APEC O27 was highly pathogenic. The antibacterial sensitivity test for oregano oil has been determined at the concentration of 20% according to the antibacterial sensitivity test that has been done before using the method of Quinn et al. [21].

# 2.3. Oregano essential oil 20%

Oregano solution 20% (Ropadiar<sup>®</sup>) is derived from Oregano (*Origanum vulgare* L.), family Lamiaceae. It was purchased from ROPA Pharm International Co., Netherlands. It contains more than 60 active compounds. Carvacrol (62 - 68%), thymol (1 - 3%), and p-cymene (~ 8%)] are the major active components present in the *Origanum vulgare* essential oil. According to the manufacturer's instructions, oregano 20% was orally administered at a dose of 2 ml /5 liters in drinking water.

# 2.4. Antibacterial drug used

According to antibacterial sensitivity test, Difloxacin was the most sensitive antibacterial against APEC serotype O27. Diflobiotic® active principle is difloxacin. Each 1 ml contains Difloxacin hydrochloride 109.13 mg (equivalent to 100 mg Difloxacin base). Diflobiotic® was orally administered in drinking water at a dose of 10 mg/kg b.wt. of chicken according to the manufacturer's instructions. It was purchased from Atco Pharm Co., Egypt.

#### 2.5. Chicks

A total number of 200 -one day old- chicks (Cobb 500) purchased from El-Wattania Co. were used in this experiment. Before the beginning of the experiment, five chicks were sacrificed randomly, and samples from liver, kidney, lungs, and trachea were examined bacteriologically for pathogenic *E. coli*. All the results were *E. coli* negative. These chicks were put under the required hygienic conditions, fed on a balanced commercial diet according to NRC [22], and supplied with clean water in sufficient quantities. The experimental chicks were subjected to traditional vaccine program for different viral diseases (Avian influenza, Newcastle, infectious bronchitis and infectious bursal disease) until the end of the experimental period.

#### 2.6. Experimental design

Experimental chicks were assigned to 4 equal groups, 50 chicks each (G1-G4). Chicks in G1 were used as normal control (non infected non treated). At the 19<sup>th</sup>, 20<sup>th</sup> and 21<sup>st</sup> day of age, chickens of G2, G3 and G4 were infected with local isolate APEC O27 via intra tracheal route at a dose of 0.2 ml of bacterial suspension in saline containing  $1 \times 10^7$  CFU/Chick according to the method of Rosenberger et al. [23]. At

the  $22^{nd}$ ,  $23^{rd}$  and  $24^{th}$  day of age, chickens of G3 (Oregano treated) and G4 (Antibiotic treated) were treated orally in drinking water with oregano 20% at a dose of 2 ml / 5 liter and Diflobiotic at a dose of 10 mg/kg b.wt., respectively. In all groups, clinical signs and mortality rate were recorded during the period of experiment. At the  $3^{rd}$ ,  $6^{th}$ ,  $10^{th}$ , and  $21^{st}$  day post-infection (dpi), reisolation of APEC O27 from the lung, trachea, and liver was performed, as well as blood and tissue samples were collected from 5 chickens in each group.

# 2.7. Reisolation of APEC O27 serotype

At the 3<sup>rd</sup>, 6<sup>th</sup>, 10<sup>th</sup>, and 21<sup>st</sup> dpi, samples from lung, trachea, and liver were taken under aseptic condition and incubated over night at 37°C in nutrient broth then loopfulls from inoculated broth were streaked on MacConkey's agar medium and Eosin Methylene Blue agar (Oxoid). The inoculated media were incubated at 37°C for 24–48h. *E. coli* colonies appeared pink colonies on MacConkey's agar and shiny metallic green colonies on Eosin Methylene Blue agar [24].

#### 2.8. Blood and tissue sampling

At the 3<sup>rd</sup>, 6<sup>th</sup>, 10<sup>th</sup>, and 21<sup>st</sup> dpi, blood and tissue samples were collected from all the experimental groups. Blood samples were collected from the wing vein of each bird and were divided into two portions. The first portion was collected into sterile tubes containing EDTA anticoagulant (Ethylene diamine tetra acetic acid) and used for evaluation of hematological investigations. The second portion was placed into a plain vacutainer tube, left to clot then centrifuged at 3000 rpm for 15 min. for separation of serum. Sera were stored at -20°C until used for biochemical analyses. Tissue specimens were collected from the lungs, trachea, liver, spleen, and kidneys of each bird for histopathological examination.

#### 2.9.1. Hematological examination

Complete blood picture consisted of erythrogram [red blood cell counts (RBCs), hemoglobin (Hb), packed cell volume (PCV), mean corpuscular volume (MCV), mean corpuscular hemoglobin (MCH) and mean corpuscular hemoglobin concentration (MCHC)], leukogram [total and differential White blood cell counts] and thrombocytes count. It was investigated according to the method of Weiss and Wardrop [25] using Natt and Herrick's solution as a diluent [26].

#### 2.9.2. Serum biochemical analysis

The activities of ALT and AST [27], and the levels of uric acid [28], creatinine [29], total proteins [30], and albumin [31] were determined spectrophotometrically (T80 UV/VIS PG instrument Ltd, UK) using commercial Test kits (Biodiagnostic Co., Egypt). Total globulins were calculated by subtracting the obtained value of albumin from the total proteins.

# 2.9.3. Histopathological examination

Tissue specimens were collected from the lungs, trachea, liver, spleen, and kidneys were immediately fixed in neutral buffered formalin 10%. Tissue specimens then were washed, dehydrated, cleared and embedded in paraffin. Paraffin blocks were sectioned at 4-5  $\mu$ m thickness and stained with hematoxylin and eosin (H&E) according to Suvarna et al. [32]. Slides were examined under a light microscope (Olympus B x50, Japan).

#### 2.10. Statistical analysis

All data were subjected to statistical analysis including the calculation of the mean  $\pm$ standard error. Differences between the normal control and the treated groups were tested for significance using one-way analysis of variance (ANOVA) followed by Duncan's multiple range test. The differences were considered significant at *P*<0.05 level [33] using Statistical Package for Social Sciences (SPSS) software version 20 computer program (*SPSS Inc, Chicago, IL, USA*).

# 3. Results

#### 3.1. Clinical signs and mortality rate

Clinical signs appeared on infected birds (G2) were dullness, ruffled feather, gasping, loss of appetite and depression, while after treatment (birds of G3 & G4) showed mild symptoms. Mortality rates among experimental infected chicks of different groups were recorded in Table (1).

# 3.2. Reisolation of APEC 027

At the  $3^{rd}$  dpi, APEC O27 was isolated from the lungs of G3 and G4, while it was isolated from G2 at the  $3^{rd}$ ,  $6^{th}$ , and  $10^{th}$  dpi.

	Chickens	Mortalities at different days post infection with					Mortality
	Number	<i>E. coli</i> serotype O27					rate (%)
Groups		1-3	4-6	7-10	11-21	Total	
Normal control (G1)	50	0	0	0	0	0	0
Infected (G2)	50	3	6	4	2	15	30%
Oregano treated (G3)	50	2	0	0	0	2	4%
Antibacterial treated G4)	50	2	1	1	0	4	8%

**Table 1.** Mortality rate of chickens experimentally infected with avian pathogenic *E. coli* O27 during the experimental periods.

# 3.4. Hematological findings

## 3.4.1. Erythrogram

Erythrocytes' count and Hb concentration revealed significant (P < 0.05) decrease in both infected (G2) and antibiotic treated (G4) groups at the 6th, 10th, and 21st dpi compared to normal control (G1) and oregano (G3). Values of PCV in infected chickens (G2) exhibited gradual decrease from the 3rd dpi till the 21st dpi which was significant (P < 0.05) at the 6<sup>th</sup> and 10<sup>th</sup> dpi compared to the other treated groups. PCV values in the antibiotic treated (G4) birds revealed a marked (P < 0.05) decrease at the 10<sup>th</sup> dpi only compared to both groups of normal control (G1) and oregano treated (G3). At the 6<sup>th</sup>, 10<sup>th</sup> and 21<sup>st</sup> dpi, MCV values indicated significant (P < 0.05) increase while MCHC revealed marked (P < 0.05) decrease in both groups of infected (G2) and antibiotic treated (G4) compared to the other groups (G1 & G3). MCH showed no significant changes in all groups during the experiment except at the  $21^{st}$  dpi only there was significant (*P*<0.05) increase in both infected and antibiotic groups. Evaluation of red cell indices revealed presence of macrocytic hypochromic type anemia in infected birds of G2 and antibiotic treated birds of G4. (Table 2)

# 3.4.2. Leukogram

Total WBCs and heterophil counts in both infected (G2) and antibiotic treated (G4) chicks were significantly (P<0.05) increased allover experimental period in comparison with the normal control (G1). Oregano treated chicks of (G3) revealed significant (P<0.05) increase in total WBCs and heterophils counts only at the 3rd, and 6th dpi. Chicks of G2 & G4 showed significant lymphopenia from the 3<sup>rd</sup> dpi till the end of the experiment in comparison with the other groups. Monocytes revealed significant (P < 0.05) increase in infected (G2) and treated groups (G3 & G4) on the 6<sup>th</sup> and 21<sup>st</sup> dpi. Eosinophil count was significantly (P < 0.05) increased in the infected group at the 6<sup>th</sup> and 21<sup>st</sup> dpi compared to the other groups. Basophils showed significant (P < 0.05) increase in infected group (from the 3<sup>rd</sup> till the 21<sup>st</sup> dpi), and treated groups (at the 6<sup>th</sup> and 10<sup>th</sup> dpi). (Table 3) 3.4.3. Thrombocytes

Compared to control group (G1), chickens of the infected group (G2) revealed a significant decrease in thrombocyte count from the  $6^{th}$  dpi till the end of the experiment, while treated groups (G3&G4) thrombocytes count slightly decreased at the  $10^{st}$  dpi (Table 3).

**Table 2.** Erythrogram of chickens treated with oregano 20% after infection with avian pathogenic *E. coli* O27 during the different experimental periods (Mean  $\pm$  SE, N=5)

Parameters	Periods	Groups				
	(dpi)	Normal Control	Infected	Oregano (20%)	Antibacterial treated	
		(G1)	(G2)	treated (G3)	(G4)	
Red blood cell count	0	2.01±0.03 <sup>a</sup>	1.97±0.04 <sup>a</sup>	2.08±0.12 <sup>a</sup>	1.99±0.08 <sup>a</sup>	
(×10 <sup>6</sup> /µl)	3	2.03±0.09 <sup>a</sup>	1.93±0.05 <sup>a</sup>	2.12±0.07 <sup>a</sup>	2.01±0.05 <sup>a</sup>	
	6	2.11±0.05 <sup>a</sup>	1.88±0.01 <sup>b</sup>	$2.04 \pm 0.06^{a}$	1.91±0.04 <sup>b</sup>	
	10	2.20±0.08 <sup>a</sup>	1.84±0.03 <sup>b</sup>	2.27±0.07 <sup>a</sup>	1.97±0.04 <sup>b</sup>	
	21	2.34±0.05 <sup>b</sup>	2.03±0.03 <sup>d</sup>	2.51±0.06 <sup>a</sup>	2.19±0.05°	
Packed Cell volume	0	27.80±0.26 <sup>a</sup>	28.22±0.18 <sup>a</sup>	27.84±0.24 <sup>a</sup>	28.16±0.34 <sup>a</sup>	
(%)	3	28.08±0.36 <sup>a</sup>	27.60±0.39 <sup>a</sup>	28.14±0.32 <sup>a</sup>	28.10±0.37 <sup>a</sup>	
	6	28.44±0.09 <sup>a</sup>	27.28±0.12 <sup>b</sup>	28.14±0.17 <sup>a</sup>	27.94±0.27 <sup>a</sup>	
	10	28.64±0.20 <sup>a</sup>	26.64±0.16°	28.46±0.27 <sup>a</sup>	27.54±0.21 <sup>b</sup>	
	21	28.12±0.33 <sup>a</sup>	27.62±0.16 <sup>a</sup>	28.52±0.29 <sup>a</sup>	27.96±0.11 <sup>a</sup>	
Hemoglobin (g/dl)	0	9.00±0.07 <sup>a</sup>	9.16±0.13 <sup>a</sup>	9.18±0.09 <sup>a</sup>	9.16±0.11 <sup>a</sup>	
	3	9.20±0.17 <sup>a</sup>	8.72±0.11 <sup>a</sup>	8.98±0.20 <sup>a</sup>	8.62±0.16 <sup>a</sup>	
	6	9.10±0.14 <sup>a</sup>	7.94±0.12 <sup>b</sup>	9.28±0.16 <sup>a</sup>	7.86±0.10 <sup>b</sup>	
	10	9.08±0.14 <sup>a</sup>	7.08±0.05 <sup>c</sup>	9.14±0.05 <sup>a</sup>	7.76±0.17 <sup>b</sup>	

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	21	9.04±0.13 <sup>a</sup>	8.28±0.12 <sup>b</sup>	9.40±0.10 <sup>a</sup>	8.56±0.13 <sup>b</sup>
Mean corpuscle	0	138.43±2.12 <sup>a</sup>	143.19±3.01ª	135.68±8.37 <sup>a</sup>	142.42±5.80 <sup>a</sup>
volume (fl)	3	139.24±6.95 <sup>a</sup>	143.54±3.96 <sup>a</sup>	133.31±2.61 <sup>a</sup>	140.43±4.08 <sup>a</sup>
	6	135.16±2.77 <sup>b</sup>	145.13±0.66 <sup>a</sup>	138.38±3.56 <sup>ab</sup>	146.80±2.81ª
	10	130.61±3.87 <sup>b</sup>	145.09±2.27 <sup>a</sup>	125.67±3.14 <sup>b</sup>	140.11±2.13 <sup>a</sup>
	21	120.42±3.22°	136.16±1.53 <sup>a</sup>	113.95±1.52°	127.76±2.34 <sup>b</sup>
Mean corpuscle hemoglobin (pg)	0	44.81±0.56 <sup>a</sup>	46.48±1.21 <sup>a</sup>	44.74±2.77 <sup>a</sup>	46.38±2.16 <sup>a</sup>
	3	45.63±2.39 <sup>a</sup>	44.39±1.53 <sup>a</sup>	42.51±0.88 <sup>a</sup>	43.10±1.54 <sup>a</sup>
	6	43.26±1.22 <sup>a</sup>	42.25±0.77 <sup>a</sup>	45.74±2.06 <sup>a</sup>	41.33±1.18 <sup>a</sup>
	10	41.45±1.61 <sup>a</sup>	38.59±0.92ª	40.40±1.35 <sup>a</sup>	39.52±1.30 <sup>a</sup>
	21	38.44±0.82°	40.81±0.52 <sup>a</sup>	37.58±0.87°	39.08±0.42 <sup>ab</sup>
Mean corpuscle hemoglobin concentration (g/dl)	0	32.38±0.22 <sup>a</sup>	32.45±0.31ª	32.97±0.21ª	32.53±0.27 <sup>a</sup>
	3	32.75±0.20 <sup>a</sup>	31.60±0.28 <sup>bc</sup>	31.90±0.47 <sup>ab</sup>	30.67±0.25°
	6	32.00±0.50ª	29.11±0.42 <sup>b</sup>	32.99±0.71ª	28.14±0.42 <sup>b</sup>
	10	31.71±0.49 <sup>a</sup>	26.58±0.31°	32.13±0.46 <sup>a</sup>	28.20±0.75 <sup>b</sup>
	21	31.96±0.63ª	29.97±0.34 <sup>b</sup>	32.97±0.41ª	30.61±0.36 <sup>b</sup>

**Table 3.** Leukogram of chickens treated with oregano 20% after infection with avian pathogenic *E. coli* O27 during the different experimental periods (Mean  $\pm$  SE, N=5)

$ \begin{array}{ c c c c c c c c c c c c c c c c c c c$	Parameters	Periods	Groups				
$ \begin{array}{ c c c c c c c c c c c c c c c c c c c$		(dpi)	Normal Control	Infected	Oregano (20%)	Antibacterial treated	
$ \begin{array}{c c c c c c c c c c c c c c c c c c c $			(G1)	(G2)	treated (G3)	(G4)	
$ \begin{array}{c cell \ count \ (\times 10^3/\mu l) \\ \hline & 3 & 19.68 \pm 0.21^b & 24.10 \pm 0.50^a & 24.56 \pm 0.26^a & 24.35 \pm 0.44^a \\ \hline & 6 & 20.11 \pm 0.25^d & 25.11 \pm 0.52^a & 21.55 \pm 0.26^c & 20.46 \pm 0.36^b \\ \hline & 10 & 20.63 \pm 0.25^c & 24.62 \pm 0.52^a & 21.08 \pm 0.26^{bc} & 23.49 \pm 0.36^b \\ \hline & 10 & 20.30 \pm 0.23^b & 22.09 \pm 0.24^a & 20.46 \pm 0.27^b & 19.16 \pm 0.22^c \\ \hline & 19.16 \pm 0.22^c & 7.44 \pm 0.26^a & 7.41 \pm 0.19^a & 7.30 \pm 0.19 \\ \hline & (\times 10^3/\mu l) & 3 & 7.63 \pm 0.14^b & 12.89 \pm 0.12^a & 12.92 \pm 0.17^a & 12.64 \pm 0.29^a \\ \hline & 6 & 7.53 \pm 0.31^d & 13.27 \pm 0.27^a & 10.01 \pm 0.25^c & 11.48 \pm 0.37^b \\ \hline & 10 & 8.30 \pm 0.20^c & 13.15 \pm 0.42^a & 8.77 \pm 0.18^c & 9.85 \pm 0.27^b \\ \hline & 10 & 8.30 \pm 0.20^c & 13.15 \pm 0.42^a & 10.00 \pm 0.21^a & 10.14 \pm 0.23^a \\ \hline & (\times 10^3/\mu l) & 3 & 10.11 \pm 0.11^a & 8.95 \pm 0.47^b & 9.32 \pm 0.18^b & 9.29 \pm 0.15^b \\ \hline & 6 & 10.61 \pm 0.19^a & 0.244 \pm 0.20^c & 0.144 \pm 0.20^b & 10.064 \pm 0.99^c \\ \hline \end{array} $	Total White blood	0	19.31±0.17 <sup>a</sup>	19.44±0.23 <sup>a</sup>	19.23±0.25 <sup>a</sup>	19.18±0.15 <sup>a</sup>	
$ \begin{array}{c ccccccccccccccccccccccccccccccccccc$	cell count (×10 <sup>3</sup> /µl)	3	19.68±0.21 <sup>b</sup>	24.10±0.50 <sup>a</sup>	24.56±0.26 <sup>a</sup>	24.35±0.44ª	
$ \begin{array}{c ccccccccccccccccccccccccccccccccccc$		6	20.11±0.25 <sup>d</sup>	25.11±0.52 <sup>a</sup>	21.55±0.26°	20.46±0.36 <sup>b</sup>	
$ \begin{array}{c ccccccccccccccccccccccccccccccccccc$		10	20.63±0.25°	24.62±0.52ª	21.08±0.26 <sup>bc</sup>	23.49±0.36 <sup>b</sup>	
$ \begin{array}{c c c c c c c c c c c c c c c c c c c $		21	20.30±0.23 <sup>b</sup>	22.09±0.24 <sup>a</sup>	20.46±0.27 <sup>b</sup>	19.16±0.22°	
$ \begin{array}{c ccccccccccccccccccccccccccccccccccc$	Heterophils	0	7.45±0.17 <sup>a</sup>	7.44±0.26 <sup>a</sup>	7.41±0.19 <sup>a</sup>	7.30±0.19	
$ \begin{array}{c ccccccccccccccccccccccccccccccccccc$	$(\times 10^{3}/\mu l)$	3	7.63±0.14 <sup>b</sup>	12.89±0.12 <sup>a</sup>	12.92±0.17 <sup>a</sup>	12.64±0.29 <sup>a</sup>	
$ \begin{array}{c ccccccccccccccccccccccccccccccccccc$		6	7.53±0.31 <sup>d</sup>	13.27±0.27 <sup>a</sup>	10.01±0.25°	11.48±0.37 <sup>b</sup>	
$ \begin{array}{c ccccccccccccccccccccccccccccccccccc$		10	8.30±0.20 <sup>c</sup>	13.15±0.42 <sup>a</sup>	8.77±0.18°	9.85±0.27 <sup>b</sup>	
$ \begin{array}{c c} Lymphocytes \\ (\times 10^{3}/\mu l) \end{array} \begin{array}{c c} 0 & 9.77\pm 0.08^{a} & 9.64\pm 0.14^{a} & 10.00\pm 0.21^{a} & 10.14\pm 0.23^{a} \\ \hline 3 & 10.11\pm 0.11^{a} & 8.95\pm 0.47^{b} & 9.32\pm 0.18^{b} & 9.29\pm 0.15^{b} \\ \hline 6 & 10.00\pm 0.18^{a} & 0.24\pm 0.206 & 0.14\pm 0.22b \\ \hline \end{array}$		21	7.88±0.13 <sup>b</sup>	12.11±0.16 <sup>a</sup>	7.94±0.17 <sup>b</sup>	8.36±0.18 <sup>b</sup>	
$(\times 10^{3} / \mu l) \qquad 3 \qquad 10.11 \pm 0.11^{a} \qquad 8.95 \pm 0.47^{b} \qquad 9.32 \pm 0.18^{b} \qquad 9.29 \pm 0.15^{b} \\ (\sim 10^{-6} / 0.18^{3} - 0.20^{c} - 0.14 + 0.20^{c} - 0.$	Lymphocytes	0	9.77±0.08 <sup>a</sup>	9.64±0.14 <sup>a</sup>	10.00±0.21ª	10.14±0.23 <sup>a</sup>	
	(×10 <sup>3</sup> /µl)	3	10.11±0.11 <sup>a</sup>	8.95±0.47 <sup>b</sup>	9.32±0.18 <sup>b</sup>	9.29±0.15 <sup>b</sup>	
$6  10.69\pm0.18^{\circ\circ}  8.24\pm0.39^{\circ\circ}  9.14\pm0.22^{\circ\circ}  10.06\pm0.08^{\circ\circ}$		6	10.69±0.18 <sup>a</sup>	8.24±0.39°	9.14±0.22 <sup>b</sup>	10.06±0.08 <sup>bc</sup>	
10 $10.36\pm0.18^{a}$ $8.90\pm0.17^{c}$ $10.11\pm0.09^{ab}$ $9.70\pm0.08^{b}$		10	10.36±0.18 <sup>a</sup>	8.90±0.17°	10.11±0.09 <sup>ab</sup>	9.70±0.08 <sup>b</sup>	
21 10.56±0.16 <sup>a</sup> 7.02±0.17 <sup>c</sup> 10.54±0.17 <sup>a</sup> 8.97±0.15 <sup>b</sup>		21	10.56±0.16 <sup>a</sup>	7.02±0.17°	10.54±0.17 <sup>a</sup>	8.97±0.15 <sup>b</sup>	
Monocytes 0 0.90±0.05 <sup>a</sup> 0.88±0.05 <sup>a</sup> 0.94±0.09 <sup>a</sup> 0.92±0.08 <sup>a</sup>	Monocytes	0	0.90±0.05ª	$0.88 \pm 0.05^{a}$	0.94±0.09 <sup>a</sup>	0.92±0.08 <sup>a</sup>	
$(\times 10^{3}/\mu l)$ 3 $0.99\pm 0.08^{a}$ $1.00\pm 0.09^{a}$ $0.90\pm 0.07^{a}$ $1.20\pm 0.07^{a}$	(×10 <sup>3</sup> /µl)	3	0.99±0.08 <sup>a</sup>	1.00±0.09 <sup>a</sup>	0.90±0.07ª	1.20±0.07 <sup>a</sup>	
$6  1.00 \pm 0.09^{b}  1.89 \pm 0.15^{a}  1.25 \pm 0.13^{a}  1.74 \pm 0.14^{a}$		6	1.00±0.09 <sup>b</sup>	1.89±0.15 <sup>a</sup>	1.25±0.13 <sup>a</sup>	1.74±0.14 <sup>a</sup>	
10 $1.02\pm0.18^{a}$ $1.19\pm0.11^{a}$ $1.31\pm0.12^{a}$ $1.26\pm0.11^{a}$		10	1.02±0.18 <sup>a</sup>	1.19±0.11 <sup>a</sup>	1.31±0.12 <sup>a</sup>	1.26±0.11 <sup>a</sup>	
21 $1.15\pm0.11^{b}$ $1.66\pm0.12^{a}$ $1.05\pm0.08^{b}$ $0.99\pm0.07^{b}$		21	1.15±0.11 <sup>b</sup>	1.66±0.12 <sup>a</sup>	1.05±0.08 <sup>b</sup>	0.99±0.07 <sup>b</sup>	
Eosinophils 0 $0.42\pm0.04^{b}$ $0.43\pm0.07^{ab}$ $0.39\pm0.06^{a}$ $0.46\pm0.05^{a}$	Eosinophils	0	0.42±0.04 <sup>b</sup>	0.43±0.07 <sup>ab</sup>	0.39±0.06 <sup>a</sup>	0.46±0.05 <sup>a</sup>	
$(\times 10^{3}/\mu l)$ 3 $0.44\pm 0.04^{a}$ $0.48\pm 0.01^{a}$ $0.54\pm 0.06^{a}$ $0.52\pm 0.05^{a}$	(×10 <sup>3</sup> /µl)	3	0.44±0.04 <sup>a</sup>	0.48±0.01 <sup>a</sup>	0.54±0.06 <sup>a</sup>	0.52±0.05 <sup>a</sup>	
		6	0.44±0.04 <sup>b</sup>	0.74±0.10 <sup>a</sup>	0.49±0.05 <sup>b</sup>	0.57±0.06 <sup>b</sup>	
10 $0.45\pm0.05$ $0.55\pm0.05^{a}$ $0.46\pm0.01^{a}$ $0.45\pm0.01^{a}$		10	0.45±0.05	0.55±0.05ª	0.46±0.01ª	0.45±0.01ª	
21 $0.45\pm0.01^{b}$ $0.53\pm0.05^{a}$ $0.50\pm0.04^{b}$ $0.45\pm0.03^{b}$		21	0.45±0.01 <sup>b</sup>	0.53±0.05ª	0.50±0.04 <sup>b</sup>	0.45±0.03 <sup>b</sup>	
Basophils 0 0.66±0.05 <sup>a</sup> 0.62±0.03 <sup>a</sup> 0.60±0.06 <sup>a</sup> 0.67±0.06 <sup>a</sup>	Basophils	0	0.66±0.05 <sup>a</sup>	0.62±0.03ª	0.60±0.06 <sup>a</sup>	0.67±0.06 <sup>a</sup>	
$(\times 10^{3}/\mu)$ 3 $0.49\pm0.07^{a}$ $0.76\pm0.10^{ab}$ $0.69\pm0.10^{b}$ $0.68\pm0.01^{b}$	$(\times 10^{3}/\mu l)$	3	0.49±0.07 <sup>a</sup>	0.76±0.10 <sup>ab</sup>	0.69±0.10 <sup>b</sup>	0.68±0.01 <sup>b</sup>	
$6   0.44 \pm 0.07^{b}   0.95 \pm 0.09^{a}   0.69 \pm 0.08^{b}   1.08 \pm 0.07^{b}$		6	0.44±0.07 <sup>b</sup>	0.95±0.09 <sup>a</sup>	0.69±0.08 <sup>b</sup>	1.08±0.07 <sup>b</sup>	
10 $0.54\pm0.05^{b}$ $0.84\pm0.07^{a}$ $0.42\pm0.07^{b}$ $0.48\pm0.08^{b}$		10	0.54±0.05 <sup>b</sup>	$0.84{\pm}0.07^{a}$	0.42±0.07 <sup>b</sup>	0.48±0.08 <sup>b</sup>	
21 0.28±0.05 <sup>b</sup> 0.75±0.11 <sup>a</sup> 0.40±0.09 <sup>b</sup> 0.42±0.07 <sup>b</sup>		21	0.28±0.05 <sup>b</sup>	0.75±0.11 <sup>a</sup>	0.40±0.09 <sup>b</sup>	0.42±0.07 <sup>b</sup>	
Platelets     0     5.28±0.07 <sup>a</sup> 5.26±0.08 <sup>a</sup> 5.49±0.12 <sup>a</sup> 5.41±0.08 <sup>a</sup>	Platelets	0	5.28±0.07 <sup>a</sup>	5.26±0.08 <sup>a</sup>	5.49±0.12 <sup>a</sup>	5.41±0.08 <sup>a</sup>	
$(\times 10^{3}/\mu l)$ 3 5.61±0.15 <sup>a</sup> 5.35±0.10 <sup>a</sup> 5.68±0.11 <sup>a</sup> 4.41±0.10 <sup>a</sup>	(×10 <sup>3</sup> /µl)	3	5.61±0.15 <sup>a</sup>	5.35±0.10 <sup>a</sup>	5.68±0.11 <sup>a</sup>	4.41±0.10 <sup>a</sup>	
6 $6.85\pm0.09^{a}$ $6.11\pm0.27^{b}$ $6.61\pm0.17^{ab}$ $6.49\pm0.10^{a}$		6	6.85±0.09 <sup>a</sup>	6.11±0.27 <sup>b</sup>	6.61±0.17 <sup>ab</sup>	6.49±0.10 <sup>a</sup>	
10 $7.05\pm0.09^{a}$ $5.95\pm0.10^{c}$ $6.61\pm0.13^{b}$ $6.59\pm0.08^{b}$		10	7.05±0.09 <sup>a</sup>	5.95±0.10°	6.61±0.13 <sup>b</sup>	6.59±0.08 <sup>b</sup>	
21 $6.21\pm0.06^{a}$ $5.39\pm0.18^{c}$ $5.91\pm0.09^{ab}$ $5.62\pm0.19^{a}$		21	6.21±0.06 <sup>a</sup>	5.39±0.18°	5.91±0.09 <sup>ab</sup>	5.62±0.19 <sup>a</sup>	

Means with different superscripts (a, b, c) in the same row are significantly different at P<0.05.

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# 3.5. Biochemical findings

Infected group (G2) exhibited a significant (P<0.05) increase in the activities of AST and ALT compared to normal control group starting from the 3<sup>rd</sup> till the end of the experimental periods. In both groups of treated chicks (G3 and G4), values of ALT activity significantly (P<0.05) elevated at the 3<sup>rd</sup> and 6<sup>th</sup> dpi, while values of AST activity were significantly

(P < 0.05) increased at the 3<sup>rd</sup>, 6<sup>th</sup> and 10<sup>th</sup> dpi then returned to the normal control values thereafter (Fig. 1)

Serum creatinine and uric acid levels in chicks of infected (G2) and antibiotic treated (G4) were increased from the 3<sup>rd</sup> till the 10<sup>th</sup> dpi then returned to normal levels at the 21<sup>st</sup> dpi compared to chicks of control (G1) and oregano treated (G3). (Fig. 1)



**Fig. 1.** Alanine aminotransferase (ALT) (A), aspartate aminotransferase (AST) (B), creatinine (C), and uric acid (D) levels in the serum of chickens treated with oregano after infection with avian pathogenic *E. coli* O27 during the different experimental periods (Mean $\pm$ SE, N=5). Means with different letters (a, b, c, d) at the same period are significant at *P*<0.05.

From the  $3^{rd}$  dpi till the end of the experiment, values of total serum proteins, albumin, total globulins and A/G ratio significantly (*P*<0.01) decreased in infected (G2) and antibiotic treated chickens (G4) compared to normal control ones. Oregano treated

chickens recorded marked decrease in the serum protein profile at the  $3^{rd}$  and  $6^{th}$  dpi. These parameters restored normal levels from the  $10^{th}$  day to the end of the experimental period. (Fig. 2)



**Fig. 2.** Serum Protein profile; total proteins (A), albumin (B), total globulins (C), and Albumin/globulins (D) levels in the serum of chickens treated with oregano after infection with avian pathogenic *E. coli* O27 during the different experimental periods (Mean  $\pm$  SE, N=5). Means with different letters (a, b, c) at the same period are significant at *P*<0.05.

# 3.6. Pathological findings

# 3.6.1. Post-mortem findings

No gross lesions were observed in chickens of the normal control group and oregano treated group all over the experimental period. Infected and antibiotic treated groups showed slight congestion at the 3rd and the 6th dpi in some lungs, livers, and kidneys. At the 10th and 21st dpi, severe congestion of lungs, liver, spleen and kidney was seen. *3.6.2. Microscopic findings* 

No microscopic alterations were observed in the lung, trachea, liver, spleen, and kidneys of the normal control group (G1) during different experimental periods (**Figs. 3A-7A**).

**Lungs:** Microscopic examination of lung sections in infected group (G2), at the 3<sup>rd</sup> and 6<sup>th</sup> dpi, showed congestion of the peri air blood capillaries (**Fig. 3B**). At the 10<sup>th</sup> and 21<sup>st</sup> dpi, lung showed congestion of the peri air blood capillaries and interstitial blood vessels (**Fig. 3C**). Lungs of oregano group (G3) showed slight congestion of air capillaries in all periods (**Fig. 3D**). Lungs of antibiotic treated group (G4), showed congestion of the peri air blood capillaries at the 3<sup>rd</sup> and 6<sup>th</sup> dpi (**Fig. 3E**). By the 10<sup>th</sup> dpi, lung

demonstrated congestion of the peri air blood capillaries and interstitial blood vessels (**Fig. 3F**). By the end of experiment, lungs showed normal histological picture in all groups except infected group.

Trachea: In the infected group, (Fig. 4B-C) hyperplasia in the mucosal tracheal epithelium and edema of the submucosa was the constant finding in the trachea at the 3<sup>rd</sup> and 6<sup>th</sup> dpi (Fig. 4B). At the 6<sup>th</sup> & 21st dpi, proprial mononuclear cells infiltration with congestion of the blood vessels was observed in trachea (Fig. 4C). Also, at the 10<sup>th</sup> and 21<sup>st</sup> dpi, hyperplasia in the mucosal epithelium was detected in trachea of oregano group (Fig. 4D). In antibiotic treated group; at the 3<sup>rd</sup> dpi, trachea showed hyperplasia in the lining mucosa with increase in the number of goblet cells (Fig. 4E). At the 6<sup>th</sup> &10<sup>th</sup> dpi, trachea displayed necrosis in the mucosal lining epithelium with mononuclear cells infiltration in the lamina propria (Fig. 4F). By the end of the experiment, trachea showed normal histological picture in all groups except infected group.

Liver: Microscopic examination of liver sections in infected group (Fig. 5B-D), at the 3<sup>rd</sup> day post infection (dpi), showed congestion of hepatic sinusoids, central veins and portal BVs (Fig. 5B). At the  $6^{th}$  dpi, some livers revealed congested central vein (Fig. 5C), by the 10<sup>th</sup> dpi, the liver showed aggregation of mononuclear cells infiltrations forming minute foci with multiple areas of diffuse hemorrhages (Fig. 5D). At the 6<sup>th</sup> dpi, after treated with oregano, livers showed normal hepatic parenchyma with normal hepatic architecture (Fig. 5E). However, antibiotic treated group; at the 3<sup>rd</sup> till 10<sup>th</sup> dpi, liver showed congested central vein with pericentral mononuclear cells infiltrations (Fig. 5F). By the end of experiment, histological picture of livers appeared normal in all groups except the infected group.

**Spleen:** In infected group (**Fig. 6B-C**), congestion of splenic red pulp was the constant finding in the spleen at the 3<sup>rd</sup> and 6<sup>th</sup> dpi (**Fig. 6B**). Also, hyperplasia of ellipsoidal reticular cells in the subcapsular sinuses and around capillaries was observed at the 10<sup>th</sup> dpi. Moreover, some spleens showed thickened muscular layer of blood vessels (**Fig. 6C**). In oregano group, spleen displayed focal areas of hemorrhage at the 6<sup>th</sup> dpi (**Fig. 6D**), and at the 10<sup>th</sup> dpi, spleen showed dilated blood vessels with thickened wall (**Fig. 6E**). At the 3<sup>rd</sup> and 10<sup>th</sup> dpi, spleen of antibiotic treated group demonstrated thickened muscular layer of the blood vessels (**Fig. 6F**). By the end of experiment histological picture of spleens appeared normal in all groups except the infected group.

**Kidneys:** Histopathological examination of kidney sections of infected group (**Figs. 7B-7D**), revealed congestion of interstitial blood vessels at the 3<sup>rd</sup> dpi in

infected group (**Fig. 7B**). Also, degenerative changes in the renal epithelium were seen. At the 6<sup>th</sup> dpi, slight degeneration of some renal tubules was noticed, as well as hemorrhage in the interstitial tissue of kidney was detected (**Fig. 7C**). By the 10<sup>th</sup> dpi, kidney showed hemorrhages in the interstitial tissue (**Fig. 7D**). At the 3<sup>rd</sup> dpi, kidneys of oregano group demonstrated necrosed renal tubules and focal areas of hemorrhage (**Fig. 7E**). At the 3<sup>rd</sup> dpi, kidneys of the antibiotic treated group displayed congestion of the interstitial blood vessels (**Fig. 7F**). By the end of experiment, kidneys histological picture appeared normal in all groups except the infected group.

#### 4. Discussion

This study was conducted to evaluate the efficacy of oregano oil 20% for treatment of chicks -19-day-old experimentally infected with APEC O27 -local isolate-. Hematological, serum biochemical, and pathological alterations of the vital organs were investigated during the experimental periods.

Avian colibacillosis is one of the most common diseases causing significant economic losses to the poultry industry in the world [34]. Clinical signs demonstrated in the infected group began at the 3rd dpi in the form of dullness, ruffled feather, gasping, huddling together, loss of appetite and depression. Mortality rates among infected group, oregano treated and antibiotic treated were 30%, 4% and 8%, respectively. Similar results were previously observed by De Carli et al. [35].

Data of erythrogram revealed significant decrease in RBCs and Hb concentration beginning at the 3<sup>rd</sup> dpi and macrocytic hypochromic anemia was recorded. Similar results were previously mentioned by Huff et al. [36]. The decrease in hematological parameters may be due to break down of erythrocytes by hemolytic enzymes found in endotoxins that produced by E. coli [37] led to decrease in the values of RBCs count, and PCV and hemoglobin concentration [25]. Observation of the erythrocytic indices revealed an increase in the values of MCV and decrease in MCHC which indicated that chicks were suffering from macrocytic hypomochromic anaemia. Saini [38] reported normocytic normochromic anemia in E. coli infection in broiler chicken. But, Petrov et al. [39] reported a decrease in MCV in the offspring of E. coli infected rabbits as well as in the experimentally infected rabbits. Hag et al. [40] reported decrease in MCV, MCH and MCHC in E. coli infection in pigeons. The erythrogram had a tendency to be affected till the end of experiment which indicated that APEC O27 affected strongly the erythrogram of chickens.

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**Fig. 3.** Lung of broiler chickens: (A) Normal control group; showing, normal parenchyma of parabronchus and air capillaries, (B-D) Infected group; (B) showing congestion of the peri air blood capillaries (arrows) at the 3<sup>rd</sup> day post infection (dpi), (C) showing congestion of the peri air blood capillaries (arrow) and interstitial blood vessel (arrow head) at the 10<sup>th</sup> dpi, and (D) showing congestion of the interstitial blood vessel (arrow head) at the 21<sup>st</sup> dpi, (E) Oregano treated group; showing congestion of the air capillaries (arrows) at the 6<sup>th</sup> dpi, and (F) Antibiotic treated group; showing congestion of the peri air blood capillaries (arrow) and interstitial blood vessel (arrow head) at the 10<sup>th</sup> dpi, and (F) Antibiotic treated group; showing congestion of the peri air blood capillaries (arrow) and interstitial blood vessel (arrow head) at the 10<sup>th</sup> dpi, (H&E, x200)



**Fig. 4.** Trachea of broiler chickens: (A) Normal control group; showing normal mucosa (H&E, x200), (B-C) Infected group; (B) showing hyperplasia in the mucosal epithelium (arrow) and edema of the submucosa (arrow head) at the  $3^{rd}$  day post infection (dpi) (H&E, x200), (C) showing proprial mononuclear cells infiltration (arrow head) with congested blood vessel (arrow) at the  $6^{th}$  dpi (H&E, x200), (D) Oregano group; showing hyperplasia in the mucosal epithelium at the  $21^{st}$  dpi (H&E, x400), (E-F) Antibiotic group; (E) showing hyperplasia in the lining mucosa (arrow) with goblet cell formation (arrow head) at the  $3^{th}$  dpi (H&E, x200), and (F) showing necrosis in the mucosal lining epithelium (arrow) with mononuclear cells infiltrations in the lamina propria (arrow head) at the  $10^{th}$  dpi (H&E, x400).



Fig. 5. Liver of broiler chickens: (A) Normal control group; showing healthy parenchyma of hepatocytes, central vein, and blood sinusoids (H&E x400), (B-D) Infected group; (B) showing congestion of the hepatoportal blood vessel (arrow) at the 3rd day post infection (dpi) (H&E x200), (C) showing congested central vein (arrow head) at the 6th dpi (H&E x400), and (D) showing large focal area of mononuclear cells infiltration (arrow head) at the 10th dpi (H&E x400), (E) Oregeno group; showing normal hepatic parenchyma at the 6<sup>th</sup> dpi (H&E x400), and (F) Antibiotic group; showing congested central vein (arrow head) with pericentral mononuclear cells infiltration (arrow) at the 3<sup>rd</sup> dpi (H&E x200).

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**Fig. 6.** Spleen of broiler chickens: (A) Normal control group; showing normal follicles, white pulps, and red pulps, (H&E, x200), (B-C) Infected group; (B) showing congestion of the blood vessel, (arrow) at the  $3^{rd}$  day post infection (dpi), (H&E, x200), and (C) showing thickened muscular layer of the blood vessel (arrows) at the  $10^{th}$  dpi (H&E, x400), Oregano group (D-E); (D) showing focal areas of hemorrhage (arrow) at the  $6^{th}$  dpi, and (E) showing dilated blood vessel with thickened wall (arrow) at the  $10^{th}$  dpi (H&E, x400), and (F) Antibiotic group, showing thickened muscular layer of the blood vessel (arrow) at the  $3^{rd}$  dpi (H&E, x400).



**Fig. 7.** Kidney of broiler chickens: (A) Normal control group; showing normal parenchyma of renal glomeruli and renal tubules (H&E, x400), (B-D) Infected group; (B) showing congestion of the interstitial blood vessel (arrow) at the 3<sup>rd</sup> day post infection (dpi) (H&E x400), and (C) showing hemorrhages in the interstitial tissue (arrow head) at the 6th dpi (H&E x400), and (D) showing hemorrhages in the interstitial tissue (arrow head) at the 10<sup>th</sup> dpi, (H&E, x200), (E) Oregano group; showing necrosed renal tubules and focal area of hemorrhage (arrow) at the 6<sup>th</sup> dpi (H&E, x400), and (F) Antibiotic group; showing congestion of the interstitial blood vessel (arrow) at the 3<sup>rd</sup> dpi (H&E, x400).

There was a significant increase in the total WBCs and absolute heterophils, eosinophils and basophiles in the infected groups as compared to control group. Leukocytes play an important role in infection and Leukocytosis immune response. is mainly encountered in localized or generalized infections, tissue necrosis, acute hemorrhages and acute hemolysis resulted from APEC O27 infection and its endotoxin [25]. Heterophils are the first line of defense which attack and engulf microorganisms. Heterophilia is a normal response to bacterial E. coli infection [25]. The observed heterophilia in the present experiment may be due to APEC infection that increased the inflammatory response as myeloperoxidase activity and production of cytokines, such as IL1β, TNF-α, IL-6 of lung tissues [41]. This was confirmed by necrosis and hemorrhages in visceral organs on the basis of histopathological findings. Lymphopenia was recorded in our study. Lymphopenia may be due to stress of infection with APEC O27 and its endotoxin which stimulate the adrenal gland to secrete corticosteroid hormones, causing destruction of lymphocytes. The endogenous glucocorticoids cause heterophilia associated with lymphopenia. [42, 43]

There was a decrease in thrombocytes count in infected groups. It is may be due to endotoxins produced by APEC O27 [37] which was confirmed histopathological finding of congestion and hemorrhages in the internal organs.

Biochemical studies revealed significant increase in the serum AST and ALT activity of the infected group. Similar results have been reported by Kumari et al. [44,45] who worked on E. coli infection in broiler chickens. These increases could be attributed to hepatocellular damage. In the present study, the elevation of the hepatic enzymes activities was supported by the histopathological findings of the liver. These quantitative differences in the results of serum AST and ALT between the infected group and oregano treated group suggested that hepatoprotective and cardio protective effects of oregano essential oil supplementation that contain bioactive components (such as carvacrol, thymol and p-cymene) have antioxidant and antibacterial effects [12,13,15] which restored the normal enzymes activity compared to infected group.

The level of creatinine and uric acid is known to reflect the state of glomerular filtration rate and kidney functions. The present work showed a significant increase in serum creatinine and uric acid levels that may be attributed to kidney damage caused by the effect of APEC and its toxin on the kidney tissue. This result was confirmed by degenerative changes in the renal epithelium, slight degeneration of some renal tubules and hemorrhage in the interstitial tissue of infected group. Our results agree with Kumari et al. [44,45]. After treatment with oregano, the creatinine showed within the normal level. It may be due to its bioactive components which have antioxidant and antibacterial effects [12,13,15]. So, this result revealed that, the oregano has nephroprotective effect. In antibiotic group, creatinine level retained to the normal after the long time (at the 21<sup>st</sup> dpi). It is may be due to the adverse effect of both endotoxins produced by APEC O27 [37,44,45] and residue antibiotic (Difloxacin) on the kidney functions [46].

Regarding serum protein profile, results revealed significant reduction in the concentrations of total proteins, albumin, total globulins and A/G ratio in the serum of both infected and antibiotic treated chickens which started after infection with APEC O27 till the end of the experiment. However, serum protein profile of oregano treated chickens exhibited decreases from the 3<sup>rd</sup> till the 6<sup>th</sup> dpi then returned to the normal levels indicating the hepatoprotective effect of oregano essential oil. Similar results have been previously reported in chickens infected by APEC [45,47]. Hypoproteinemia may be due to liver damage which causes failure in plasma protein synthesis, and renal affection which leads to protein loss [48]. The liver is a site for all protein fractions synthesis except gamma globulin. In the present study, damage to liver and kidney as evidenced by gross and histopathological studies were observed, leading to decrease in total serum proteins and albumin concentration. In oregano treated chicken, protein profile restored normal levels from the 10<sup>th</sup> (dpi) to the end of the experimental period indicating the hepatoprotective, antioxidant and anti-inflammatory effects of oregano essential oil which contains bioactive phenolic compounds; such as carvacrol, thymol and p-cymene that have antibacterial and antioxidant activities [12,13,15] Our results disagree with Saini [38] who recorded hyperglobulinemia associated with liver cirrhosis, hepatitis, and Kupffer cells proliferation in broiler chicks infected with APEC.

Regarding the microscopical examination, lungs of chickens experimentally infected with APEC O27 noticed severe congestion of pulmonary blood vessels and capillaries with hemorrhage which were prevalent in most cases. These microscopical findings were previously mentioned by Reese et al. [49]. Hyperplasia in the mucosal tracheal epithelium and edema of the submucosa were the constant findings at the 3rd and 6<sup>th</sup> dpi for infected group. Moreover, proprial mononuclear cells infiltration with congested blood vessels were observed at the 6<sup>th</sup> till 21<sup>st</sup> dpi. Microscopical examination of the examined liver of chicks experimentally infected with APEC O27, revealed congestion of some portal blood vessels with inflammatory mononuclear cells infiltration. These findings were in complete agreement with Koutsianos et

al. [50]. At the 3rd and 6th dpi, congestion of splenic red pulp was the constant finding in infected and antibiotic treated groups. At the 10th dpi, hyperplasia of ellipsoidal reticular cells in the subcapsular sinuses and around capillaries in spleen was observed. In spleen of oregano group, focal areas of hemorrhage were observed at the 6<sup>th</sup> dpi and dilated blood vessels with thickened wall were observed at the 10<sup>th</sup> dpi. The obtained microscopic pictures were mostly compatible with the result of Antão et al. [51]. Histopathological examination of kidney sections of infected group revealed congestion of interstitial blood vessels at the 3rd dpi. Also, degenerative changes in the renal epithelium were seen. At the 6<sup>th</sup> and 10<sup>th</sup> dpi, slight degeneration of some renal tubules and hemorrhage in the interstitial tissue were noticed. These observations were previously mentioned by Rodriguez-Siek et al. [52], and Ewers et al. [53]. At the 3<sup>rd</sup> dpi, (before treatment), kidneys showed that necrosed renal tubules and focal areas of hemorrhage in oregano group, and congestion of the interstitial blood vessels in antibiotic treated group. It is may be due to the effect of APEC infection and its toxin on the kidneys. Similar results were recorded by Kumari et al. [44,45]. By the end of the experiment, kidneys showed normal histological picture in all groups of treatment except the infected group.

After treatment of infected chickens with oregano essential oil 20%, blood picture, liver and kidney functions were returned to the normal level. In addition, lung, trachea, liver, spleen, and kidneys were within the normal histopathological structures. It may be due to oregano essential oil content of bioactive phenolic compounds such as carvacrol, thymol and p-cymene that have antibacterial, anti-inflammatory and antioxidant activities [12,13,15,54]. Such compounds decrease the activity of reactive oxygen species levels that are produced by macrophage cells stimulated by APEC lipopolysaccharide [55]. The antibacterial activity has been recorded due to the synergetic effect of carvacrol and thymol constituents of oregano essential oil [56].

#### 5. Conclusion

Experimental infection of chickens with APEC O27 caused macrocytic hypochromic anemia, leukocytosis, heterophilia, and lymphopenia, and alterations in the liver and kidney functions, as well as histopathological changes in the internal organs. These changes ranged from moderate to slight after treatment with antibiotic and returned to the normal levels after treatment with oregano essential oil 20%. Supplementation of oregano essential oil 20% could ameliorate the severity of the pathogenicity caused by APEC O27. So, it could be used as antibiotic, and hepato- and nephroprotective agent. It could be recommended using oregano essential oil 20% for treatment in poultry farms rather than antibiotics.

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### 8. Conflict of Interest

There are no conflicts to declare.

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