



Synthesis and Antiproliferative Activity of Chalcone- Imide Derivatives Based on 3,4-Dichloro-1*H*-Pyrrole-2,5-dione



Mohamed. A. A. Radwan^{1,2*}, Fahad. M. Alminderej¹, Hala. E. M. Tolan³, Hanem. M. Awad⁴

¹Department of Chemistry, College of Science, Qassim University, Buraydah, Kingdom of Saudi Arabia.

²Applied Organic Chemistry Department, ³Department of Photochemistry, ⁴Tannins Chemistry Department, National Research Centre, El-Behouth St, Dokki, Cairo 12311, Egypt

Abstract

A series of chalcone imide derivatives, 4'-aminochalcones-based dichloromaleimides, was synthesized from the reaction of 1-(4-acetylphenyl)-3,4-dichloro-1*H*-pyrrole-2,5-dione with various substituted aldehydes, or by treating 4'-aminochalcone with 3,4-dichlorofuran-2,5-dione in an alternative path. The structures of chalcone imide derivatives were established using IR, ¹H NMR, ¹³C NMR, and mass spectroscopy. Antiproliferative effects of the newly synthesized compounds have been screened on two human cancer types via the MTT assay. Compounds with *p*-tolyl-1*H*-pyrrole-2,5-dione, and 4-bromophenyl-1*H*-pyrrole-2,5-dione derivatives, are highly active on the human liver cancer (HepG-2). On the other hand, all compounds were found to be more effective against breast cancer cells (MCF-7) than the positive control doxorubicin. The results of this work provide a basis for further research of selected chalcone-imide moiety as antiproliferative agents.

Keywords: chalcones, maleimides, pyrrole-2,5-dione, antiproliferative, cytotoxicity.

1. Introduction

Cancer is one of the main health problems in the world, and chemotherapy is one of the familiar medicines for the treatments of cancer [1]. Chemotherapy drugs have a cytotoxic attack on normal cells and therefore cause many strong side effects [2]. Hence, it is important to find anticancer medicines that possess cytotoxic and antiproliferative action in tumor cells lacking effect on normal cells. Despite the fact that several cytotoxic compounds have been established, a need to improve the use of other effective and safer chemotherapy drugs is still important [2]. Several compounds containing maleimide have been synthesized for this purpose, for example, chalcone-imide derivatives have been constructed and described for their cytotoxic activity on human CEM T lymphocytes, Molt 4/ C8, and murine L1210 cell lines [3]. As well, chalcones based on maleimide and dibromomaleimide have been synthesized and their antibacterial properties have been described [4-6]. Besides to the great

antimicrobial properties, maleimides had also been extensively investigated in medicine as antianxiety [7], anti-inflammatory [8], anticancer [9-11], and neuroprotective agents. Similarly, natural products with important units of maleimide have excellent biological activities, including antibacterial [12-19] and enzyme inhibitory activities [20-22]. In addition, the chalcone has shown a vital role in the biosynthesis of flavonoids and is ubiquitous in natural products. Moreover, chalcones have been investigated for their cytotoxic activity against several cancer types [23-28]. Furthermore, pyrrole is a public unit in several natural products, and many widespread drugs have a pyrrole moiety in their building. Pyrrole derivatives appeared as substructure units for some bioactive applicants [29]. Also, pyrrole derivatives showed antimicrobial [30], anti-viral [31], antitumor [32], antitubulin [33], antimalarial [34] and anticancer activities [35-37], Fig. 1.

*Corresponding author e-mail: m1radwan@yahoo.com; (Tel: +966530781050).

Receive Date: 15 June 2020, Revise Date: 08 July 2020, Accept Date: 12 July 2020

DOI: 10.21608/EJCHEM.2020.32862.2694

©2021 National Information and Documentation Center (NIDOC)

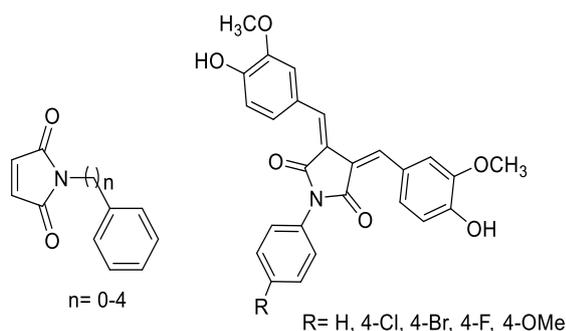


Figure 1. Bioactivemaleimide derivatives.

Extending our previous efforts that focused on the synthesis and development of biologically active compounds [38-50], in the current manuscript and in order to achieve constructive results, a combination between chalcones and maleimide was accomplished, sequentially a series of 4'-aminochalcone-based 3,4-dichloromaleimides was synthesized, and their cytotoxicity against breast cancer cells (MCF-7), and human hepatocellular carcinoma (HepG2) cells was investigated.

Experimental

Melting points were determined in open glass capillary tubes on an Electro Thermal Digital melting point apparatus (model: IA9100), and are uncorrected. Elemental analyses were carried out for elements C, H, N at Microanalytical Unit (at NRC). FT-IR spectral analyses (ν , cm^{-1}) were recorded on a Nexus 670 FTIR Nicolet spectrophotometer. ^1H and ^{13}C NMR spectra were measured on a Jeol 300 MHz in $\text{DMSO}-d_6$ using tetramethylsilane (TMS) as an internal standard (chemical shifts are expressed in ppm). Mass spectra were measured on a MAT Finnigan SSQ 7000 spectrometer, using the electron impact technique (EI). TLC was performed on silica gel aluminum sheets, 60 F₂₅₄ (E. Merck). Chalcones **4a-c** were synthesized according to reported procedure [51].

Synthesis of 1-(4-acetylphenyl)-3,4-dichloro-1H-pyrrole-2,5-dione (3).

To a solution of 3,4-dichlorofuran-2,5-dione (2 mmol) **2** in acetic acid (10 ml), 4-acetyl aniline **1** (2 mmol) was added. The reaction mixture was refluxed for 4h. After completion of the reaction, the produced mixture was cooled, poured into ice/water and left for 3h. The resulted solid was filtered and crystallized from ethanol.

Yellow solid, (78% yield); m.p. 293-295 °C. FT-IR: 1740 (C=O), 1676 (2C=O). ^1H NMR (300 MHz, $\text{DMSO}-d_6$): 2.60 (3H, s, CH_3), 7.56 (2H, d, $J =$

8.3 Hz, Ar-H), 8.01 (2H, d, $J = 8.3$ Hz, Ar-H); ^{13}C NMR (125 MHz, $\text{DMSO}-d_6$): 26.83, 126.51, 129.03, 132.89, 134.96, 136.15, 161.87, 197.21. EI-MS (m/z): 283 [M]⁺. Anal. calcd. for $\text{C}_{12}\text{H}_7\text{Cl}_2\text{NO}_3$: C, 50.73; H, 2.48; N, 4.93%. Found: C, 50.78; H, 2.45; N, 4.97%.

General procedure for the synthesis of 3,4-dichloro-1-(4-(3-(4-aryl)acryloyl)phenyl)-1H-pyrrole-2,5-dione derivatives (5a-g).

Method A: A solution of 1-(4-acetylphenyl)-3,4-dichloro-1H-pyrrole-2,5-dione **3** (10 mmol), and appropriate aldehydes (10 mmol) in ethanol, was stirred under an ice bath for 5 min. The reaction was initiated by dropwise addition of 10 ml of 60 % sodium hydroxide solution in 30 min interval. To ensure the end of the reaction, the mixture was allowed for continuous stirring for another 2-3 h at room temperature. The reaction mixture was kept in a refrigerator for overnight and diluted with ice-cold distilled water (40 mL). The precipitated chalcone was filtered, washed well with cold water and air dried. Then, the chalcone was recrystallized from ethanol.

Method B. To a solution of 3,4-dichlorofuran-2,5-dione **2** (2 mmol) in acetic acid (10 ml), chalcones **4a-c** (2 mmol) was added. The reaction mixture was refluxed for 4h. The reaction left to cool, and poured into ice/water and stand up for 3h. The solid was filtered and crystallized from ethanol.

(E)-3,4-Dichloro-1-(4-(3-(4-nitrophenyl)acryloyl)phenyl)-1H-pyrrole-2,5-dione (5a). Yellow solid, Yield 89 %; m.p. 212-214°C. FT-IR: 1730 (C=O), 1670 (2C=O). ^1H NMR (300 MHz, $\text{DMSO}-d_6$): 7.58 (1H, d, $J = 15.6$ Hz, *E*-olefinic-H), 7.82 (1H, d, $J = 15.6$ Hz, *E*-olefinic-H), 8.05 (2H, d, $J = 8.3$ Hz, Ar-H), 8.10 (2H, d, $J = 8.6$ Hz, Ar-H), 8.13 (2H, d, $J = 8.3$ Hz, Ar-H), 8.29 (2H, d, $J = 8.6$ Hz, Ar-H); ^{13}C NMR ($\text{DMSO}-d_6$): 123.95, 125.94, 126.60, 129.58, 129.94, 132.94, 135.26, 136.51, 141.08, 141.47, 148.16, 161.87, 188.29. MS: m/z : 416 [M]⁺. Anal. calcd. for $\text{C}_{19}\text{H}_{10}\text{Cl}_2\text{N}_2\text{O}_5$: C, 54.70; H, 2.42; N, 6.71%. Found: C, 54.77; H, 2.35; N, 6.76%.

(E)-1-(4-(3-(4-Bromophenyl)acryloyl)phenyl)-3,4-dichloro-1H-pyrrole-2,5-dione (5b). Yellow solid, Yield 83 %; m.p. 206-208°C. FT-IR: 1720 (C=O), 1665 (2C=O). ^1H NMR (300 MHz, $\text{DMSO}-d_6$): 7.56 (2H, d, $J = 8.1$ Hz, Ar-H), 7.68 (2H, d, $J = 8.3$ Hz, Ar-H), 7.72 (1H, d, $J = 15.4$ Hz, *E*-olefinic-H), 7.84 (2H, d, $J = 8.3$ Hz, Ar-H), 7.97 (1H, d, $J = 15.4$ Hz, *E*-olefinic-H), 8.27 (2H, d, $J = 8.1$ Hz, Ar-H); ^{13}C NMR (125 MHz, $\text{DMSO}-d_6$): 122.71, 126.59, 129.43, 130.87, 131.92, 132.92, 135.05, 136.58, 138.10,

138.43, 143.02, 161.88, 188.24. EI-MS (m/z): 449[M⁺]. Anal. calcd. for C₁₉H₁₀BrCl₂NO₃: C, 50.59; H, 2.23; N, 3.11%. Found: C, 50.64; H, 2.20; N, 3.14%.

(*E*)-3,4-Dichloro-1-(4-(3-(4-chlorophenyl)acryloyl)phenyl)-1*H*-pyrrole-2,5-dione (5c). Yellow solid, Yield 83 %; m.p. 195-197°C. FT-IR: 1725 (C=O), 1668 (2C=O). ¹H NMR (300 MHz, DMSO-*d*₆): 7.50 (2H, d, *J* = 8.2 Hz, Ar-H), 7.56 (2H, d, *J* = 8.4 Hz, Ar-H), 7.73 (1H, d, *J* = 15.8 Hz, *E*-olefinic-H), 7.91 (2H, d, *J* = 8.4 Hz, Ar-H), 7.95 (1H, d, *J* = 15.8 Hz, *E*-olefinic-H), 8.26 (2H, d, *J* = 8.2 Hz, Ar-H); ¹³C NMR (125 MHz, DMSO-*d*₆): 122.67, 126.59, 128.99, 129.44, 130.68, 132.91, 133.60, 135.04, 135.25, 136.83, 142.92, 161.89, 188.32. EI-MS (m/z): 405 [M⁺]. Anal. calcd. for C₁₉H₁₀Cl₃NO₃: C, 56.12; H, 2.48; N, 3.44%. Found: C, 56.17; H, 2.42; N, 3.39%.

(*E*)-3,4-Dichloro-1-(4-(3-(2-hydroxyphenyl)acryloyl)phenyl)-1*H*-pyrrole-2,5-dione (5d). Yellow solid, Yield 83 %; m.p. 221-223°C. FT-IR: 1345 (OH), 1725 (C=O), 1668 (2C=O). ¹H NMR (300 MHz, DMSO-*d*₆): 6.9-7.15 (3H, m, Ar-H), 7.51 (1H, d, *J* = 8.1 Hz, Ar-H), 7.56 (2H, d, *J* = 8.4 Hz, Ar-H), 7.74 (1H, d, *J* = 15.6 Hz, *E*-olefinic-H), 7.98 (2H, d, *J* = 8.4 Hz, Ar-H), 8.10 (1H, d, *J* = 15.8 Hz, *E*-olefinic-H), 10.67 (1H, OH, D₂O exchange); ¹³C NMR (125 MHz, DMSO-*d*₆): 123.14, 126.48, 127.87, 129.49, 132.39, 133.21, 135.13, 135.27, 136.44, 141.13, 154.46, 161.78, 188.61. EI-MS (m/z): 387 [M⁺]. Anal. calcd. for C₁₉H₁₁Cl₂NO₄: C, 58.79; H, 2.86; N, 3.61%. Found: C, 58.85; H, 2.82; N, 3.60%.

(*E*)-3,4-Dichloro-1-(4-(3-(4-hydroxyphenyl)acryloyl)phenyl)-1*H*-pyrrole-2,5-dione (5e). Yellow solid, Yield 83 %; m.p. 199-201°C. FT-IR: 1335 (OH), 1720 (C=O), 1660 (2C=O). ¹H NMR (300 MHz, DMSO-*d*₆): 7.38 (2H, d, *J* = 8.2 Hz, Ar-H), 7.51 (2H, d, *J* = 8.4 Hz, Ar-H), 7.71 (1H, d, *J* = 15.8 Hz, *E*-olefinic-H), 7.93 (2H, d, *J* = 8.4 Hz, Ar-H), 8.03 (1H, d, *J* = 15.8 Hz, *E*-olefinic-H), 8.23 (2H, d, *J* = 8.2 Hz, Ar-H), 10.34 (1H, OH, D₂O exchange); ¹³C NMR (125 MHz, DMSO-*d*₆): 123.32, 127.11, 127.95, 129.66, 131.32, 134.40, 135.31, 135.58, 136.40, 142.08, 155.12, 161.46, 188.22. EI-MS (m/z): 387 [M⁺]. Anal. calcd. for C₁₉H₁₁Cl₂NO₄: C, 58.79; H, 2.86; N, 3.61%. Found: C, 58.83; H, 2.80; N, 3.63%.

(*E*)-3,4-Dichloro-1-(4-(3-(*p*-tolyl)acryloyl)phenyl)-1*H*-pyrrole-2,5-dione (5f). Yellow solid, Yield 83 %; m.p. 185-187°C. FT-IR: 1715 (C=O), 1658 (2C=O). ¹H NMR (300 MHz, DMSO-*d*₆): 2.43 (3H, s, CH₃), 7.33 (2H, d, *J* = 8.2 Hz, Ar-H), 7.46 (2H, d, *J* = 8.4 Hz, Ar-H), 7.74 (1H, d, *J* = 15.2 Hz, *E*-olefinic-H), 7.92 (2H,

d, *J* = 8.4 Hz, Ar-H), 7.98 (2H, d, *J* = 8.2 Hz, Ar-H), 8.01 (1H, d, *J* = 15.2 Hz, *E*-olefinic-H); ¹³C NMR (125 MHz, DMSO-*d*₆): 22.44, 123.16, 126.18, 128.35, 129.11, 131.81, 132.69, 133.78, 135.52, 136.12, 137.38, 141.93, 161.76, 188.41. EI-MS (m/z): 385 [M⁺]. Anal. calcd. for C₂₀H₁₃Cl₂NO₃: C, 62.20; H, 3.39; N, 3.63%. Found: C, 62.25; H, 3.32; N, 3.66%.

(*E*)-3,4-Dichloro-1-(4-(3-(thiophen-2-yl)acryloyl)phenyl)-1*H*-pyrrole-2,5-dione (5g).

Yellow solid, Yield 83 %; m.p. 167-169°C. FT-IR: 1712 (C=O), 1655 (2C=O). ¹H NMR (300 MHz, DMSO-*d*₆): 7.27 (1H, m, thiophene H-4), 7.54 (2H, d, *J* = 8.1 Hz, Ar-H), 7.57 (1H, d, thiophene H-3), 7.71 (1H, d, *J* = 15.2 Hz, *E*-olefinic-H), 7.79 (1H, d, thiophene H-4), 7.98 (2H, d, *J* = 8.1 Hz, Ar-H), 8.06 (1H, d, *J* = 15.2 Hz, *E*-olefinic-H); ¹³C NMR (125 MHz, DMSO-*d*₆): 120.19, 126.50, 128.77, 129.22, 130.78, 132.90, 133.20, 134.87, 136.89, 137.18, 139.64, 161.88, 187.91. EI-MS (m/z): 377 [M⁺]. Anal. calcd. for C₁₇H₉Cl₂NO₃S: C, 53.99; H, 2.40; N, 3.70; S, 8.48%. Found: C, 54.07; H, 2.37; N, 3.72; S, 8.42%.

Cell culture conditions

The cells of human liver carcinoma (HepG-2), and human breast adenocarcinoma (MCF-7) were purchased from the American Type Culture Collection (Rockville, MD). All cells were maintained in a DMEM medium, which was supplemented with 10% of heat-inactivated fetal bovine serum (FBS), 100U/ml of each of penicillin and streptomycin. The cells were grown at 37°C in a humidified atmosphere of 5% CO₂.

MTT antiproliferative assay

The antiproliferative activities on the HepG-2, and MCF-7 human cancer cell lines were estimated, using the 3-[4,5-dimethyl-2-thiazolyl]-2,5-diphenyl-2*H*-tetrazolium bromide (MTT) assay, which was grounded on the reduction of the tetrazolium salt by the mitochondrial dehydrogenases in viable cells [52, 53]. The cells were dispensed in a 96 well sterile microplate (3 x 10⁴ cells/well), followed by their incubation at 37°C with a series of different concentrations of 10 μl of each compound or Doxorubicin® (positive control, in DMSO) for 48 h in a serum free medium prior to the MTT assay. Subsequently, the media were carefully removed, 40 μL of MTT (2.5 mg/mL) were added to each well, and then incubated for an additional 4 h. The purple formazan dye crystals were solubilized by the addition of 200 μL of DMSO. The absorbance was measured at 570 nm applying a SpectraMax® Paradigm® Multi-Mode microplate reader. The relative cell viability was

expressed as the mean percentage of viable cells relative to the untreated control cells. All experiments were conducted in triplicate and were repeated on three different days. All the values were represented as mean \pm SD. The IC_{50} s were determined by the SPSS probit analysis software program (SPSS Inc., Chicago, IL).

Naturally, your paper should start with a concise and informative title. Do not use abbreviations in it. Next, list all authors with their first names or initials and surnames (in that order). Indicate the author for correspondence using the third menu option. Present addresses can be inserted using a normal footnote (on the 'Text' menu). After having listed all authors' names, you should list their respective affiliations. Link authors and affiliations using superscript lower case letters.

Results and Discussion

Chemistry

Continuing our previous efforts to develop successful anticancer agents, a series of 4'-aminochalcones-based on dichloro-*1H*-pyrrole-2,5-dione **5a-g** has been designed and synthesized. Consequently, 4-acetyl aniline **1** reacted with 3,4-dichlorofuran-2,5-dione **2** in acetic acid at refluxed temperature for 4h to give 1-(4-acetylphenyl)-3,4-dichloro-*1H*-pyrrole-2,5-dione **3** in a good yield. The structure of **3** was elucidated according to spectral data IR, NMR, and mass analysis. Spectral data are in a good agreement with the proposed structure (Scheme 1). The IR (KBr, cm^{-1}) spectra of this compound released absorption bands at 1676-1740 cm^{-1} for three carbonyl groups. Similarly, 1H -NMR data of compound **3** shows the CH_3 protons signal at 2.60 ppm and the aromatic signal at 7.56–8.01 ppm and the ^{13}C -NMR indicated that there were three carbonyl carbons at 161 and 197 ppm.

Subsequently, chalcone-imides **5a-g** were synthesized by Claisen–Schmidt condensation of compound **3** with appropriate aldehydes. The structures of chalcone derivatives **5a-g** were elucidated according to spectral data IR, NMR, and mass analysis. Spectral data are in a good agreement with the suggested structure. For example, the IR spectra of compound **5a** released absorption bands at 1670-1730 cm^{-1} for three carbonyl groups. Also, 1H -NMR data of compound **5a** exhibited two doublet signals at 7.58-7.82 ppm of α,β -unsaturated protons in addition to aromatic signals at 8.05-8.29 ppm. The ^{13}C -NMR revealed three carbonyl carbons at 161.86 and

188.29 ppm. In addition, the DEPT experiment (Fig. 2) revealed six odd protons, (CH), (two α,β -unsaturated protons plus eight aromatic protons with two para-substituted benzene rings). The J -coupling ($J = 15.6$ Hz) of compound **5a-g** was established their *E*-isomers. Furthermore, an alternative high coast route for the synthesis of compound **5a-g** was carried out, but with low yield, (Scheme 2).

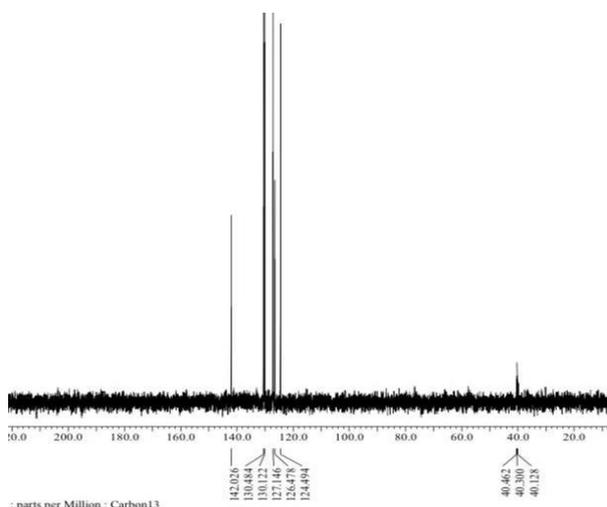
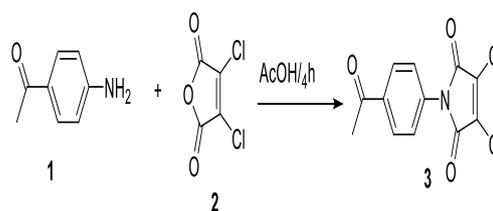
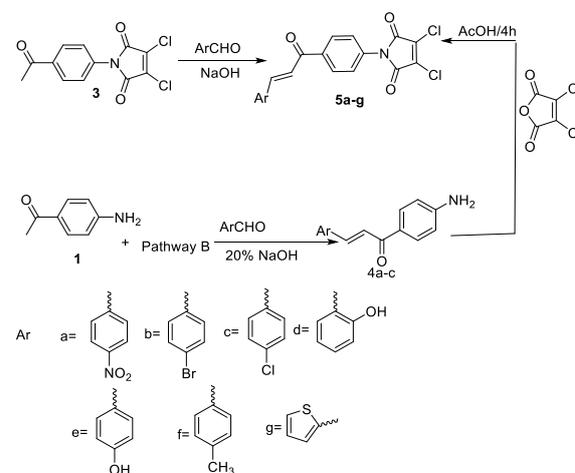


Fig 2. DEPT-NMR of compound **5a**



Scheme 1. Synthetic routes to maleimide **3**



Scheme 2. Synthetic routes to chalcone-imide derivatives **5a-g**.

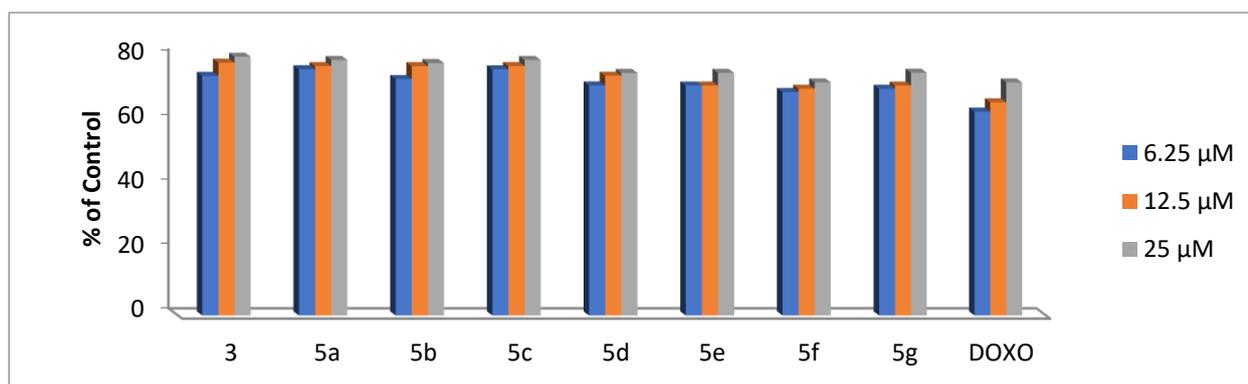


Fig. 3: Dose-dependent antiproliferative data of the compounds on the HepG-2 human cancer type, according to the MTT assay after 48 h of exposure.

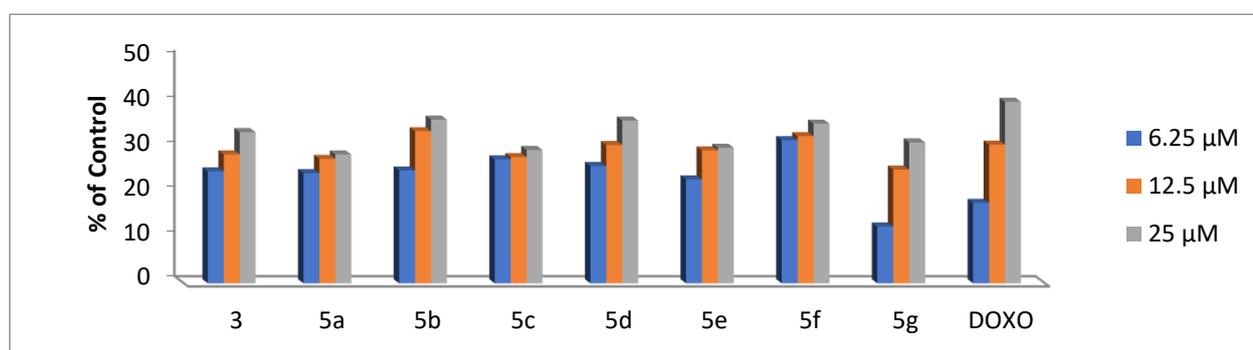


Fig. 4: Dose dependent antiproliferative data of the compounds on the MCF-7 human cancer type according to the MTT assay after 48 h of exposure

Antiproliferative activity

Selected compounds were tested *in vitro* for their antiproliferative activities on the HepG-2 and MCF-7 human cancer cell lines through the employment of the MTT assay. The percentages of the viable cells and their IC_{50} values were measured and were, subsequently, assessed with those of the control, Doxorubicin (table, Figs. 3, and 4). The results obtained revealed that all compounds showed dose-dependent antiproliferative activity against both cells (Figs. 3, and 4). Constructed deduction from these outcomes is that, in assessment with the positive control Doxorubicin, for human liver cancer (HepG-2), compounds 5f, 5b, and 5d, respectively, have comparable activities; the rest of the compounds had slightly less activities than the positive control (table, Fig. 3). In breast cancer cells (MCF-7); all compounds are more potent relative to the positive control (table 1, Fig.4).

Table 1: The antiproliferative IC_{50} values of the compounds according to the MTT assay on the two human cell types.

| Compounds | IC_{50} (μ M) \pm SD | |
|-------------|-------------------------------|----------------|
| | HepG-2 | MCF-7 |
| 3 | 30.8 \pm 3.1 | 7.5 \pm 0.5 |
| 5a | 32.1 \pm 3.1 | 7.7 \pm 0.9 |
| 5b | 29.1 \pm 2.9 | 7.9 \pm 0.6 |
| 5c | 31.1 \pm 2.6 | 7.6 \pm 0.5 |
| 5d | 29.6 \pm 2.8 | 9.0 \pm 1.1 |
| 5e | 31.7 \pm 3.1 | 9.2 \pm 0.9 |
| 5f | 28.1 \pm 2.5 | 10.0 \pm 1.1 |
| 5g | 35.1 \pm 3.3 | 9.4 \pm 0.9 |
| Doxorubicin | 28.5 \pm 1.9 | 10.3 \pm 0.8 |

Conclusion

In the current manuscript, a series of promising chalcone-imides, 5a–g was synthesized, characterized and their anticancer activity against HepG-2, and MCF-7 human cancer cell lines was investigated.

Compounds **5f**, **5b**, and **5d**, were highly active on the human liver cancer (HepG-2). On the other hand, all compounds are more effective against breast cancer cells (MCF-7) than the positive control doxorubicin. The results of this manuscript offer a basis for further research on selected chalcone-imide derivatives as antiproliferative agents (especially for breast cancer cells (MCF-7)).

In the view of SAR correlations compounds **5f**, **5b**, and **5d**, with 4-methyl, 4-bromo, and 2-hydroxyphenyl linked to chalcone-imides were more active on the human liver cancer (HepG-2). While all compounds are more effective against breast cancer cells (MCF-7) than the positive control doxorubicin specially compound **5a** with 4-nitophenyl substituted.

Funding

This research received no external funding”.

Consent for Publication

Not applicable.

Conflict of Interests

The authors declare no conflict of interests, financial or otherwise.

Acknowledgements

The authors are grateful to the National Research Centre and Qassim university for the facilities provided.

References

- Gallorini M., Cataldi A., and Di Giacomo V., Cyclin-dependent kinase modulators and cancer therapy. *biopharmaceuticals and gene therapy*, 26, 377-391 (2012). DOI:10.1007/bf03261895
- Joseph A., Shah C.S., Kumar S.S., Alex A.T., Maliyakkal N., Moorkoth S., and Mathew J.E., Synthesis, in vitro anticancer and antioxidant activity of thiazazole substituted thiazolidin-4-ones. *Acta pharmaceutica*, 63, 397-408 (2013). doi:10.2478/acph-2013-0028
- Jha A., Chandrani M., Alfred J.R., Erik D.C., Jan B., and James P.S., Cytostatic activity of novel 4'-aminochalcone-based imides. *Bioorg med Chem Lett*, 17, 4545-4550 (2007). doi.org/10.1016/j.bmcl.2007.05.094
- Patel J.R., and Dholakiya B.Z., Synthesis, characterization and antimicrobial activity of 4'-aminochalcone-based dibromomaleimides. *Der pharma chemica*, 3, 458-466 (2011).
- Patel J.R., and Dholakiya B.Z., Synthesis of 1-(4-((E)-3-arylacryloyl) phenyl)-3,4-dibromo-1H-pyrrole-2,5-diones and screening for anti-Candida and antituberculosis activity. *Medicinal chemistry research*, 21, 1977-1983 (2012). DOI 10.1007/s00044-011-9718-x
- Patel J.R., Malani M.H., and Dholakiya B.Z., Silica sulfuric acid-catalyzed Claisen-Schmidt condensation of 1,3,4 trisubstituted pyrrole 2,5 dione to chalcones. *Research on chemical intermediates*, 38, 2371-2381 (2012). DOI 10.1007/s11164-012-0553-6
- Chen X.L., Zhang L.J., Li F.G., Fan Y.X., Wang W.P., Li B.J., and Shen Y.C., Synthesis and antifungal evaluation of a series of maleimides. *Pest Manag. Sci.*, 71, 433-440 (2015). doi.org/10.1007/s10340-012-0466-6
- Li W., Fan Y.X., Shen Z.Z., Chen X.L., and Shen Y.C., Antifungal activity of simple compounds with maleic anhydride or dimethylmaleimide structure against Botrytis cinerea. *J. Pestic. Sci.*, 37, 247-251 (2012). doi.org/10.1584/jpestics.D11-054
- Shen Z.Z., Fan Y.X., Li F.G., Chen X.L., and Shen Y.C., Synthesis of a series of N-substituted dimethylmaleimides and their antifungal activities against Sclerotinia sclerotiorum. *J. Pest Sci.*, 86, 353-360 (2013). doi.org/10.1007/s10340-012-0466-6
- Jens R.A., Irma K.B., Beata A.C., Anthony C.W.A., Edward H.D., Stephen S.B., Edward T.C., and Vito F.D., The synthesis and biological evaluation of two analogues of the C-Riboside showdomycin. *Aust. J. Chem.*, 58, 86-93 (2005). doi.org/10.1071/CH04273
- Yongxian F., Yuele Lu., Xiaolong C., Babu T., Xing-Cong L., and Yinchu S., Anti-Leishmanial and Cytotoxic Activities of a Series of Maleimides: Synthesis, Biological Evaluation and Structure-Activity Relationship. *Molecules*, 23, 2878 (2018). doi: 10.3390/molecules23112878
- Thomas B., and Stephan A.S., Showdomycin as a versatile chemical tool for the detection of pathogenesis-associated enzymes in bacteria. *J. Am. Chem. Soc.*, 132, 6964-6972 (2010). doi.org/10.1021/ja909150y
- Wu M.D., and Cheng M.J., Maleimide and maleic anhydride derivatives from the mycelia of *Antrodia cinnamomea* and their nitric oxide inhibitory activities in macrophages. *J. Nat. Prod.*, 71, 1258-1261 (2008). doi: 10.1021/np070634k
- Wael A.Z., Clarisse B.F., and Fondja Y., Aqabamycins, A-G. Novel nitro maleimides

- from a marine *Vibrio* species: I. taxonomy, fermentation, isolation and biological activities. *J. Antibiot.*,63, 297-301 (2010). doi: 10.1038/ja.2010.34
15. Frederic Z., and Alain V., Synthesis and antimicrobial activities of N-substituted imides. *IL Farmaco*,57,421-426 (2002).
16. David C., and Emmanuelle S.S., Monohalogenated maleimides as potential agents for the inhibition of *Pseudomonas aeruginosa* biofilm. *Biofouling*,26,379-385 (2010).doi.org/10.1080/08927011003653441
17. López S.N., Castelli M.V., de Campos F., Corrêa R., Cechinel F.V., Yunes R.A., Zamora M.A., Enriz R.D., Ribas J.C., Furlán R.L., and Zacchino S.A. In vitro Antifungal Properties, Structure-activity Relationships and Studies on the Mode of Action of *N*-Phenyl, *N*-Aryl, *N*-Phenylalkyl Maleimides and Related Compounds. *Arzneimittelforschung*,55,123-132 (2005). DOI:10.1055/s-0031-1296833
18. Slavica A., Dib I., and Nidetzky B., Selective modification of surface-exposed thiol groups in *Trigonopsis variabilis* D-amino acid oxidase using poly (ethylene glycol) maleimide and its effect on activity and stability of the enzyme. *Biotechnol. Bioeng*,96,9-17 (2007).DOI:10.1002/bit.21181
19. Manas K.S., Debjani D., and Dulal P., Pyrene excimer fluorescence of yeast alcohol dehydrogenase: A sensitive probe to investigate ligand binding and unfolding pathway of the enzyme. *Photochem. Photobiol.*,82,480-486 (2006).doi.org/10.1562/2005-09-26-RA-698
20. Nara L.M., Gislaïne F., and Carla S., N-antipyrene-3,4-dichloromaleimide, an effective cyclic imide for the treatment of chronic pain: the role of the glutamatergic system. *Anesth. Analg.*,110,942-950 (2010). doi: 10.1213/ANE.0b013e3181cbd7f6
21. Khan M.I., Baloch, M.K., and Ashfaq, M., Biological aspects of new organotin (IV) compounds of 3-maleimidopropionic acid. *J. Organomet. Chem.*,689,3370-3378 (2004). doi:10.1016/j.jorganchem.2004.07.049
22. Sosabowski J.K., Matzow T., Foster J.M., Finucane C., Ellison D., Watson S.A., and Mather S.J., Targeting of CCK-2 receptor-expressing tumors using a radiolabelled divalent gastrin peptide. *J. Nucl. Med.*,50,2082-2089 (2009). doi: 10.2967/jnumed.109.064808
23. Zhuang C., Zhang W., Sheng C., Zhang W., Xing C., and Miao Z., Chalcone: A Privileged structure in medicinal chemistry. *Chem. Rev.*,117,7762-7810 (2017). doi: 10.1021/acs.chemrev.7b00020
24. Jandial D.D., Blair C.A., Zhang S., Krill L.S., Zhang Y.B., and Zi X., Molecular targeted approaches to cancer therapy and prevention using chalcones. *Curr. Cancer Drug Targets*,14,181-200 (2014).doi:10.2174/1568009614666140122160515
25. Sharma R., Kumar R., Kodwani R., Kapoor S., Khare A., Bansal R., Khurana S., Singh S., Thomas J., Roy B., and et al., A review on mechanisms of anti tumor activity of chalcones. *Anti-Cancer Agents Med. Chem.*,16,200-211 (2016). doi:10.2174/1871520615666150518093144
26. Sahu N.K., Balbhadra S.S., Choudhary J., and Kohli D.V., Exploring pharmacological significance of chalcone scaffold: A review. *Curr. Med. Chem.*,19,209-225 (2012). doi:10.2174/092986712803414132
27. Mahapatra D.K., Bharti S.K., and Asati V., Anti-cancer chalcones: Structural and molecular target perspectives. *Eur. J. Med. Chem.*, 98,69-114 (2015). doi: 10.1016/j.ejmech.2015.05.004
28. Dhivare R.S., Rajput S.S., Chemical Science Review and Letters Synthesis and antimicrobial evaluation of some novel bis-heterocyclic chalcones from cyclic imides under microwave irradiation. *Chem Sci Rev Lett.*, 4, 937-944 (2015).
29. Sortino M., Cechinel-Filho V., Corrêa R., Zacchino S., *N*-Phenyl and *N*-phenylalkyl-maleimides acting against *Candida* spp.: Time-to-kill, stability, interaction with maleamic acids. *Bioorg Med Chem.*,16, 560-568 (2008).
30. Basha S.S., Ramachandra-Reddy P., Padmaja A., Padmavathi V., Mouli K.C., and Vijaya T., Synthesis and antimicrobial activity of 3-aryloxy-4-heteroaryl pyrroles and pyrazoles. *Med. Chem. Res.*, 24, 954-964 (2015). https://doi.org/10.1007/s00044-014-1169-8.
31. He X.Y., Lu L., Qiu J., Zou P., Yu F., Jiang X.K., Li L., Jiang S., Liu S., and Xie L., *Bioorg. Med. Chem.*, 21, 7539-7584 (2013). https://doi.org/10.1016/j.bmc.2013.04.046.
32. Wang M., Ye C., Liu M., Wu Z., Li L., Wang C., Liu X., and Guo H., Synthesis and antitumor activity of 5-(5-halogenated-2-oxo-1H-pyrrolo[2,3-b]pyridin-(3Z)-ylidenemethyl)-2,4-dimethyl-1H-pyrrole-3-carboxamides. *Bioorg. Med. Chem. Lett.*, 25, 2782-2787

- (2015).<https://doi.org/10.1016/j.bmcl.2015.05.017>
33. Da C., Telang N., Barelli P., Jia X., Gupton J.T., Mooberry S.L., and Kellogg G.E., Pyrrole-Based Antitubulin Agents: Two Distinct Binding Modalities Are Predicted for C-2 Analogues in the Colchicine Site. *ACS Med. Chem. Lett.*, 3, 53-57 (2012). <https://doi.org/10.1021/ml200217u>.
34. Mahajan D.T., Masand V.H., Patil K.N., Hadda T.B., and Rastija V., Integrating GUSAR and QSAR analyses for antimalarial activity of synthetic prodiginines against multi drug resistant strain. *Med. Chem. Res.*, 22, 2284-2292 (2013). <https://doi.org/10.1007/s00044-012-0223-7>.
35. Williams I.S., Joshi P., Gatchie L., Sharma M., Satti N.K., Vishwakarma R.A., Chaudhuri B., and Bharate S.B., Synthesis and biological evaluation of pyrrole-based chalcones as CYP1 enzyme inhibitors, for possible prevention of cancer and overcoming cisplatin resistance. *Bioorg. Med. Chem. Lett.*, 27, 3683-3687 (2017). <https://doi.org/10.1016/j.bmcl.2017.07.010>.
36. Moreno J.S., Agas D., Sabbieti M.G., Magno M.D., Migliorini A., and Loreto M.A., Synthesis of novel pyrrolyl-indomethacin derivatives. *Eur. J. Med. Chem.*, 57, 391-397 (2012). <https://doi.org/10.1016/j.ejmech.2012.09.008>.
37. Dyson L., Wright A.D., Young K.A., Sakoff J.A., and McCluskey A., Synthesis and anticancer activity of focused compound libraries from the natural product lead, oroidin. *Bioorg. Med. Chem.*, 22, 1690-1699 (2014). <https://doi.org/10.1016/j.bmc.2014.01.021>.
38. Radwan M. A., El-Sherbiny M. Synthesis and antitumor activity of indolylpyrimidines: marine natural product meridianin D analogues. *Bioorg Med Chem.*, 15, 1206-1211(2007)
39. Radwan M. A., Ragab E. A., Sabry N. M., El-Shenawy S. M. Synthesis and biological evaluation of new 3-substituted indole derivatives as potential anti-inflammatory and analgesic agents. *Bioorg Med Chem.*, 15, 3832-41 (2007).
40. Radwan M.A.A., Ragab E.A., Shaaban M.R., El-Nezhawy A. Application of (2Z)-3-dimethylamino-2-(1H-indole-3-carbonyl) acrylonitrile in the synthesis of novel 3-heteroarylindoles: condensed meridianine analogs, *ARKIVOC*, (vii), 281-293 (2009).
41. El-Nezhawy A. O., Eweas A. F., Radwan M. A. El-Naggar T. B. Synthesis and Molecular Docking Studies of Novel 2-Phenyl-4-Substituted Oxazole Derivatives as Potential Anti-cancer Agents. *Journal of Heterocyclic Chemistry*, 53, 271-279 (2016).
42. Fakhr I. M., Radwan M. A., el-Batran S., Abd el-Salam O. M., el-Shenawy, S. M. Synthesis and pharmacological evaluation of 2-substituted benzo[b]thiophenes as anti-inflammatory and analgesic agents. *Eur J Med Chem*, 44, 1718-25 (2009).
43. Radwan M. A., Shehab M. A., El-Shenawy S. M. Synthesis and biological evaluation of 5-substituted benzo [b] thiophene derivatives as anti-inflammatory agents. *Monatshefte für Chemie*, 140, 445-50 (2009).
44. Radwan M. A. A., Alminderej F. M., Awad H. M. One-Pot Multicomponent Synthesis and Cytotoxic Evaluation of Novel 7-Substituted-5-(1H-Indol-3-yl)Tetrazolo[1,5-a] Pyrimidine-6-Carbonitrile. *Molecules*, 25, 255 (2020).
45. Muhammad Z., Radwan M. A. A., Farghaly T., Gaber H., Elaasser M. Synthesis and antitumor activity of novel [1,2,4,5]-tetrazepino[6,7-b] indole derivatives: Marine natural product hyrtioreticuline C and D analogues. *Mini-Reviews in Med. Chem.* 1979-86 (2019).
46. El Nezhawy A., Gaballah, S., Radwan M. A. A. Studying the reactivity of (phthalazin-1(2H)-on-2-yl)methyl trichloroacetimidate towards different C- and O-nucleophiles. *Tetrahedron Letters*, 50, 6646-6650(2009).
47. El Nezhawy A., Radwan M. A. A., Gaballah S., Synthesis of chiral N-(2-(1-oxophthalazin-2(1H)-yl) ethanoyl)- α -amino acid derivatives as antitumor agents. *Arkivoc*, 12 119-130 (2009).
48. Radwan M. A. A., Abbas E., Synthesis of some pyridine, thiopyrimidine, and isoxazoline derivatives based on the pyrrole moiety. *Monatshefte für Chemie*, 140, 229-233 (2009).
49. Ghorab M., Radwan M. A. A., Taha N., Amin N., Shehab M., Faker I., Dapson in heterocyclic chemistry, part II: Antimicrobial and antitumor activities of some novel sulfone biscompounds containing biologically active thioureido, carbamothioate, quinazoline, imidazolidine, and thiazole moieties. *Phosphorus, Sulfur and Silicon and the Related Elements*, 183, 2906-2917 (2008).

-
50. Fakhr I., Hamdy N., Radwan M. A. A., Ahmed Y., Synthesis of new bioactive benzothiophene derivatives. *Egyptian J. of Chem.*, 47, 201-215 (2004).
51. Said A.H., El-Fekya H., Abd El-Fattah N.A., Mohd I., and Mohamed N.Z., Design, synthesis and in vitro antitumor activity of novel phthalazin-1,4-dione/chalcone hybrids and phthalazin-1,4-dione/pyrazoline hybrids. *Journal of Chemical and Pharmaceutical Research*, 7, 1154-1166 (2015).
52. Asmaa F. Kassem I.F., Nassar M.T., Hanem M.A., and Wael A., Synthesis and Anticancer Activity of New ((Furan-2-yl)-1,3,4-thiadiazolyl)-1,3,4-oxadiazole Acyclic Sugar Derivatives. *Chem Pharm Bull., (Tokyo)*, 67, 888-895 (2019). doi: 10.1248/cpb.c19-00280.
53. Mokedda E.H., Ebtehal S., Nesreen S., Hazem A., Hanem M.A., Efficient and easy synthesis of new Benzo[h]chromene and Benzo[h]quinoline derivatives as a new class of cytotoxic agents. *Journal of Molecular Structure*, 1195, 702-711 (2019). doi.org/10.1016/j.molstruc.2019.05.081